Topological data analysis quantifies biological nano-structure from single molecule localization microscopy

Pike, Jeremy; Khan, Abdullah; Pallini, Chiara; Thomas, Steven; Mund, Markus; Ries, Jonas; Poulter, Natalie; Styles, Iain

DOI:
10.1093/bioinformatics/btz788

License:
Creative Commons: Attribution (CC BY)

Citation for published version (Harvard):

Link to publication on Research at Birmingham portal

General rights
Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

• Users may freely distribute the URL that is used to identify this publication.
• Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
• Users may use extracts from the document in line with the concept of ‘fair dealing’ under the Copyright, Designs and Patents Act 1988 (?)
• Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy
While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Download date: 23. Jun. 2020
Bioimage informatics
Topological data analysis quantifies biological nano-structure from single molecule localization microscopy

Jeremy A. Pike 1,2,*, Abdullah O. Khan 1,2, Chiara Pallini 1,2, Steven G. Thomas 1,2, Markus Mund 3,4, Jonas Ries 3, Natalie S. Poulter 1,2 and Iain B. Styles 1,5,*

1 Centre of Membrane Proteins and Receptors (COMPARE), Universities of Birmingham and Nottingham, Midlands, UK, 2 Institute of Cardiovascular Sciences, College of Medical and Dental Sciences, University of Birmingham, Birmingham B15 2TT, UK, 3 Cell Biology and Biophysics Unit, European Molecular Biology Laboratory (EMBL), Heidelberg 69117, Germany, 4 Department of Biochemistry, University of Geneva, 1211 Geneva 4, Switzerland and 5 School of Computer Science, University of Birmingham, Birmingham B15 2TT, UK

*To whom correspondence should be addressed.

Associate Editor: Robert Murphy

Received on December 12, 2018; revised on September 3, 2019; editorial decision on October 11, 2019; accepted on October 17, 2019

Abstract

Motivation: Localization microscopy data is represented by a set of spatial coordinates, each corresponding to a single detection, that form a point cloud. This can be analyzed either by rendering an image from these coordinates, or by analyzing the point cloud directly. Analysis of this type has focused on clustering detections into distinct groups which produces measurements such as cluster area, but has limited capacity to quantify complex molecular organization and nano-structure.

Results: We present a segmentation protocol which, through the application of persistence-based clustering, is capable of probing densely packed structures which vary in scale. An increase in segmentation performance over state-of-the-art methods is demonstrated. Moreover we employ persistent homology to move beyond clustering, and quantify the topological structure within data. This provides new information about the preserved shapes formed by molecular architecture. Our methods are flexible and we demonstrate this by applying them to receptor clustering in platelets, nuclear pore components, endocytic proteins and microtubule networks. Both 2D and 3D implementations are provided within RSMLM, an R package for pointillist-based analysis and batch processing of localization microscopy data.

Availability and implementation: RSMLM has been released under the GNU General Public License v3.0 and is available at https://github.com/JeremyPike/RSMLM. Tutorials for this library implemented as Binder ready Jupyter notebooks are available at https://github.com/JeremyPike/RSMLM-tutorials.

Contact: j.a.pike@bham.ac.uk or i.b.styles@bham.ac.uk

Supplementary information: Supplementary data are available at Bioinformatics online.

1 Introduction

Single molecule localization microscopy (SMLM) is a super-resolution fluorescence imaging technique capable of localizing individual molecules to approximately 20 nm. Since its introduction (Betzig et al., 2006; Hess et al., 2006; Rust et al., 2006), SMLM has matured as a technology and is now routinely used to probe biological nano-structure and processes for a range of biological applications (Leterrier et al., 2015; Mund et al., 2018). After performing localization, the data from a SMLM experiment is represented by a set of spatial coordinates, each corresponding to a single detection, that form a point cloud. This can be analyzed either by rendering an image from these coordinates and using image-based analysis methods, or by analyzing the point cloud directly. Strategies for the latter have focused on the concept of clustering, either by analyzing the spatial statistics of the point cloud to confirm the presence of clustered molecules (Owen et al., 2010; Sengupta et al., 2011; Veatch et al., 2012), or by grouping individual detections into distinct clusters (Andronov et al., 2016; Levet et al., 2015; Owen et al., 2010). This latter approach allows per-cluster statistics such as area and detection density to be calculated.

Clustering strategies commonly used for SMLM datasets estimate local detection density and construct clusters from the detections with density above a specified threshold. DBSCAN and
Ripley’s K-based clustering estimate density using the number of neighbouring detections within a specified distance (Ester et al., 1996; Owen et al., 2010), whereas Voronoi diagram-based clustering uses the area of the tiles in the associated tessellation (Andronov et al., 2016; Levet et al., 2015). The free parameters, a density threshold and sometimes a distance scale, can be set manually or automatically using mean cluster density (Levet et al., 2015) or Monte-Carlo simulations (Andronov et al., 2016). If assumptions can be made about the distribution and shape of the clusters, a Bayesian engine can be used to set parameters (Rubin-Delanchy et al., 2015). However, biological data is complex, often containing structures of significantly varying density. For such data a single density threshold is not sufficient and a multi-scale approach is required. Clustering algorithms can be repeated, using different parameter values, to segment structures at different densities, for example cells, organelles and protein clusters (Levet et al., 2015). An alternative approach to density thresholding is to identify clusters based on persistence, or topographic prominence (Chazal et al., 2013). This strategy has shown promise for SMLM datasets in the context of Ripley’s K-based clustering (Griffie et al., 2015, 2017).

A further limitation of current clustering approaches is that topological information and higher order structure is not considered. Topological data analysis (TDA) provides a robust mathematical framework for probing the topology, or shape, of a point cloud. In this work we employ, and extend, methods from TDA, specifically persistence-based clustering (Chazal et al., 2013) and persistent homology (Edelsbrunner et al., 2002; Ghrist, 2007; Zomorodian and Carlsson, 2005), to quantify clustering and topological structure within SMLM datasets at a range of scales and densities. We demonstrate their ability to outperform existing methods and reveal new insight into biological nano-structure. Our clustering workflow is used to show a decrease in the area of platelet integrin α2β1 clusters when the tyrosine kinase Syk is inhibited. Additionally our persistent homology methodology is used to quantify topological structure for endocytic proteins and nuclear pore complex components. The tools are made available to the community as an R package.

2 System and results

2.1 Persistence-based clustering outperforms existing approaches

In common with many existing approaches, the first step in persistence-based clustering is the calculation of an underlying density estimate, \( f \) (Chazal et al., 2013). Detections are assigned to local maxima within the density estimate by following the gradient of the density along a specified graph, a collection of points (detections) and lines linking pairs of points. This approach is known as mode density along a specified graph, a collection of points (detections) maxima within the density estimate by following the gradient of the density; \( P \). For each candidate the persistence is defined as the difference between the birth density and the death density, \( P = f_b - f_d \). Finally a persistence threshold, \( \tau \), is specified and candidate clusters with \( P < \tau \) are merged to a neighbouring candidate with \( P \geq \tau \) which cannot be linked to a candidate with \( P \geq \tau \) are considered to be noise. This clustering scheme is named the Topological Mode Analysis Tool (ToMATo) and is analogous to local thresholding of the density estimate (Supplementary Figs S1 and S2). Further algorithmic details can be found in the Supplementary Methods and the original publication (Chazal et al., 2013). A key advantage of ToMATo is the ability to segment clusters which are close together even if they vary in density.

To evaluate ToMATo on SMLM data we generated realistic simulated dSTORM datasets of Gaussian clusters. Low, high and mixed (a mixture of high and low) density clusters were generated, either in close proximity or well separated (Supplementary Fig. S3). Local detection density was estimated by counting the number of detections within a fixed radius, and candidate clusters were constructed from the graph linking all detections within the same search radius (Fig. 1a and b). With our implementation of ToMATo there are two free parameters; the search radius and the persistence threshold, \( \tau \). To enable the selection of a suitable persistence threshold a scatter plot of the death and birth densities for each candidate cluster can be plotted, this is known as a ToMATo (persistence) diagram (Fig. 1c). Further guidance on parameter selection can be found in the Supplementary Methods.

We compared persistence-based clustering to existing routinely used approaches, specifically DBSCAN (Ester et al., 1996), Ripley’s K-based clustering (Owen et al., 2010; Rubin-Delanchy et al., 2015) and Voronoi tessellation (Andronov et al., 2016; Levet et al., 2015). A range of free parameters were used for each simulation scenario and algorithm. Performance was quantified as the percentage of correctly assigned detections and averaged over repeated simulations (Supplementary Fig. S4). ToMATo significantly outperforms these existing approaches in challenging scenarios when clusters are close together (Fig. 1e). For the easier simulations where clusters are well separated it performs equally well. Moreover ToMATo is less sensitive to small changes in the choice of free parameters (Fig. 1f).

In biological applications it is common to have structures which are close together. These results demonstrate that persistence-based clustering is the highest performing and most stable of the tested algorithms for SMLM cluster analysis under these conditions. To better understand the sources of error for the ToMATo approach we classified detection assignment errors as either false positives, false negatives or incorrect cluster assignments (Supplementary Table S1). For scenarios where clusters were well separated the prevalent source of error was noise detections being assigned to clusters (false positives). This is because the overall error rate is low and randomly positioned noise detections which lie within the vicinity of clusters are not easily identified. As expected, when clusters are close together the prevalent source of error was incorrect cluster assignment.

To better understand the benefits and limitations of ToMATo clustering for SMLM data further simulations were performed where either the cluster separation, density, or signal to noise ratio (SNR) was varied (Supplementary Figs S5–S7). In the latter two cases clusters were well separated. For each algorithm a range of parameters were tested but fixed for the variations in cluster separation, density or SNR. When cluster separation was varied ToMATo outperformed the other tested algorithms with a large maximal performance increase of 36% at 60 nm cluster separation.

When cluster density was varied ToMATo performed at least as well as the other tested algorithms. When the SNR was varied Ripley’s K-based clustering was the highest performing algorithm with a small maximal performance increase of 3.2% over ToMATo. This highlights a limitation of the ToMATo approach. If the SNR varies significantly within your data we recommend optimizing experimental conditions to reduce variation and pre-filtering detections based on density (Supplementary Fig. S8).

Persistence-based clustering is not limited to a particular structure, or shape, and is capable of segmenting non-circular objects such as microtubules, or even whole cells (Supplementary Fig. S8). However when working with fibrous structures specialized techniques such as those developed by Peters et al. (2017) should be considered.

2.2 Syk inhibition reduces the area of integrin α2β1 clusters in platelets

To demonstrate the use of persistence-based clustering on real SMLM datasets we segment nano-structures of integrin α2β1 in platelets seeded on collagen fibres (Fig. 2a and b). Integrin α2β1, a platelet collagen receptor, accumulates at collagen fibres in spread platelets as shown in Supplementary Figure S9 (Poulter et al., 2017). Within a single platelet there are areas with sparse or tightly packed α2β1 clusters due to differences in the underlying collagen
distribution. This therefore represents a difficult multi-density segmentation problem. Stable platelet adhesion to collagen via z2/1 under flow conditions is dependent on the presence of the cytoskeletal adapter protein talin, which links integrins to the actin cytoskeleton (Beckerle et al., 1989; Petrich et al., 2007). We disrupted the cortical actin organization using the tyrosine kinase Syk inhibitor PRT060318 (Jau mouille et al., 2014; Reilly et al., 2011), which we hypothesized would interfere with z2/1 clustering on collagen fibres. Syk inhibition results in a significant reduction in mean cluster area, however, no significant difference in cluster density was observed (Fig. 2c). A visual comparison with other segmentation methods is shown in Supplementary Figure S10 and computational time is compared in Supplementary Table S2.

2.3 Persistent homology quantifies topological nano-structure

SMLM data is information rich and contains more structural insight than is available through cluster analysis alone. Here we use persistent homology to extract complementary topological information from the data (Edelsbrunner et al., 2002; Ghrist, 2007; Zomorodian and Carlsson, 2005). The concept of a graph can be extended to a higher dimensional structure, known as a simplicial complex. A simplicial complex is constructed from points, lines, triangles, tetrahedrons and equivalent higher order structures, collectively known as simplices. For SMLM data we build the simplicial complex on top of the point cloud formed by the detection list. There are a variety of methods for building complexes but this work focuses on the Rips complex, an abstract simplicial complex chosen for its efficient computation and storage. If each point within a candidate simplex (two points for a line and three for a triangle) is within a search distance, a point cloud can be computed as a function of search distance, or scale. We will refer to this collection of features as the topological configuration. First order features correspond to the number of connected components in the complex, second order to the number of holes or loops. When working with 3D datasets, the third order features are enclosed voids.

Computing the topological structure within the data at a single scale is not very informative; any given feature could be unstable due to small variations in scale and it is not possible to capture multi-scale structure. To overcome this the Rips complex is computed for a range of scales, a process known as a filtration (Supplementary Video S1 and Fig. 3a and b). The birth scale of a feature is defined as the scale at which the feature forms in the filtration, and the greater the distance from the diagonal of the diagram, the greater the distance from the diagonal of the diagram. Fragile features, typically noise, will have low persistence and be located close to the diagonal. Therefore thresholding features by persistence selects only the most robust, a procedure known as persistent homology (Edelsbrunner et al., 2002; Ghrist, 2007; Zomorodian and Carlsson, 2005).

Before performing persistent homology it is advantageous to perform a cluster analysis to segment the data into nano-structures (Supplementary Fig. S11). This helps with interpretation of the results, and also reduces the computational cost (Supplementary Table S3). For each cluster a filtration is performed and a persistence diagram calculated. The number of features above the persistence threshold can then be counted for each cluster. SMLM data is inherently noisy and complex; containing labelling artifacts, significant localization uncertainty and substantial false positive rates. Persistent homology is robust to small perturbations in detection localization (Cohen-Steiner et al., 2007) but not necessarily to the addition of noisy detections, or removal of true detections. To help overcome this we have developed an extension to the standard persistent homology workflow specific to SMLM. Instead of computing the persistence diagram for all detections within a cluster we sub-sample the cluster detections (with replacement) and repeatedly calculate the topological configuration for a specified persistence
threshold. The sample size is set to the number of detections within the cluster. The sampling probability, $p$, can be weighted by detection measurements inherent to SMLM such as localization uncertainty; $p = e^{-w}$, $w$ is the normalized detection localization uncertainty, scaled between 0 and 1 for all detections within the cluster. $\mu$ is a constant which throughout this study is set to $\mu = -\ln 0.1 = 2.3$. This was set such that the detection with the lowest localization uncertainty is ten times more likely to be sampled than the detection with the highest localization uncertainty. The most common configuration, the consensus and an agreement value, $\alpha$, are returned. $\alpha$ corresponds to the percentage of sub-sampled clusters which return the consensus result and provides a confidence level for the specified topological configuration.

To test our persistent homology-based workflow for SMLM data, realistic synthetic datasets were generated. Molecules were distributed according to one of three scenarios; (i) complete spatial randomness (CSR), (ii) Gaussian clusters or (iii) circular rings with molecules evenly distributed on the circumference. We used ToMato to segment the clusters and subsequently performed our persistent homology workflow to produce persistence diagrams (Fig. 3d and e and Supplementary Video S2). A single persistence threshold was applied to all scenarios and the number of holes for each cluster was calculated using both the full diagrams, and the sub-sampled consensus approach (Fig. 3f). Guidance for selecting appropriate persistence thresholds can be found in the Supplementary Methods and Supplementary Figure S12. The sub-sampled consensus approach produced significantly fewer incorrect classifications. Furthermore, by selecting only clusters with $\alpha > 90\%$ the percentage of incorrect classifications was reduced further. As expected no significant difference in cluster area was found between Gaussian and ring clusters. Therefore for these simulations standard cluster statistics cannot distinguish between Gaussian and ring clusters, whereas persistent homology clearly reveals the key topological difference. Several clusters were found in the CSR scenario due to artifacts such as multiple blinking events. However no CSR clusters with holes and $\alpha > 90\%$ were found demonstrating the robustness of our approach to noise.
The topology of clusters, as defined by our workflow, can be used to filter clusters for downstream analysis. For example we can select all clusters with a single hole (\(x > 90\%\)) and find the average radial distribution as shown for the ring simulations in Figure 3f. Our workflow can also be applied to 3D SMLM datasets as demonstrated for equivalent simulations in Supplementary Figure S13 and Supplementary Video S3.

2.4 Topological analysis of nuclear pore and endocytic proteins

To evaluate our novel methods on real data we focused on structures for which the topology has already been well characterized through image-based particle averaging. This is appropriate as it facilitates a robust validation of the proposed workflow. Specifically we choose Nup107, a component of the nuclear and cytoplasmic rings of nuclear pore complexes ([Li et al., 2018; Otsuka et al., 2016; Szymborska et al., 2013]), and three different proteins of the yeast endocytic machinery; Las17, Edel and Sla1 ([Mund et al., 2018]).

Persistence-based clustering was used as a pre-processing step to segment either the nuclear pore complex, or endocytic sites (Supplementary Figs S14–S16). For Nup107 both 2D and 3D dSTORM datasets were analyzed. For the 2D Nup107 dataset the resulting persistence diagram shows a large number of holes above the threshold (Fig. 4a and b). As expected a large proportion of the clusters have a single hole topology (Fig. 4c). However even for the sub-sampled approach, with \(x > 90\%\) filtering, a significant proportion of clusters (39%) are found to have no holes. These clusters could be a result of many scenarios including clustered molecules not in a pore complex, imaging artifacts, unspecific labelling and variation in pore alignment. Filtering of clusters with a single hole results in an average radial profile with a peak at 50 nm, reproducing the results of imaged-based particle averaging (Fig. 4d) ([Szymborska et al., 2013]). No radial structure is apparent when clusters with no holes are averaged. This implies these clusters are correctly assigned by the persistent homology workflow.

When imaged in 3D both the cytoplasmic and nuclear rings of the complex must be considered. Both rings should have the topology of a cylinder and contain a single hole and no enclosed voids. When our method is applied to 3D data we observe large numbers of structures with no topological features (81%). However a significant number of clusters have single holes (17%) and a small percentage have two holes (1%) (Supplementary Fig. S16). No structures had enclosed voids. The quoted percentages are for the sub-sampled approach with \(x > 90\%\) filtering.

In a recent study the actin nucleation promoting factor Las17, was shown to have a clear ring profile when many endocytic sites are averaged, whereas the coat protein Sla1 does not ([Mund et al., 2018]). It was also shown that Edel is recruited to sites in the early stages of endocytosis, and clusters are not uniform in size and shape, hence a ring like topology is not clear from image-based particle averaging without filtering of clusters. We take data from this study,
TDA quantifies biological nano-structure

3 Implementation

3.1 RSMLM: R package for analysis of SMLM datasets

To complement this study we have released RSMLM, an R package for the pointillist-based analysis of SMLM data. This package includes the methods described in this paper for persistence-based clustering (ToMATo), alongside DBSCAN, Voronoi tessellation and Ripley’s K-based clustering. Utility methods for persistent homology including the sub-sampling consensus approach are provided. There is also the capacity to simulate dSTORM data. This library will provide an adaptable framework for analysing and batch processing both 2D, and 3D, SMLM datasets. Binder ready Jupyter notebook tutorials are provided to facilitate easy use of the package. The functionality of the library can also be included within KNIME workflows using simple R-snippets (Berthold et al., 2009). This enables users without any scripting knowledge to access the core functionality.

3.2 Persistence-based clustering

Persistence-based clustering, or ToMATo, was originally developed by (Chazal et al., 2013) and implemented for this work as follows. Detection density estimates were calculated by counting the number of other detections within a specified radius. This was implemented using the R package dbscan which uses the C++ library ANN to employ a k-d tree framework for efficient computation. Density modes were calculated using a mode seeking approach and a Rips graph (Koontz et al. 1976). Modes were then merged or designated as noise based on a specified persistence threshold. This was implemented by adapting C++ scripts (GPlv3) described in Chazal et al. (2013). C++ functions were incorporated into RSMLM using the R
package Rcpp. For algorithm comparison the search radius ranged from 1 to 50 nm, and the persistence threshold from 0 to 50 detections. Further algorithmic details and the implementations of other clustering algorithms are described in the Supplementary Methods.

3.3 Persistent homology
Rips filtrations and persistence diagrams were computed using the R package TDA which employs the C++ library GDUDH (Fasy et al., 2014; Maria et al., 2014). When the density of points in a persistence diagram was too high for a simple plot, features were grouped into a joint histogram with 1 nm² bins and displayed using a heat-map. Visualizations of Rips complexes were created using plex-viewer (https://github.com/atausz/plex-viewer) and the Java library javaPlex (Adams et al., 2014). Filtration movies were created using POV-Ray.

3.4 dSTORM simulations
Fluorophore blinking characteristics were modelled using a geometric distribution with probability of transition to the dark state set to 0.5 (Lee et al., 2012). Simulated molecules were bound to an average of 5 fluorophores, randomly distributed between molecules. The localization uncertainty for each blinking event was determined using a normal distribution centred on the molecule position. Standard deviation for localization uncertainty was set using a log-normal distribution with mean 2.8 and standard deviation 0.28 (experimentally measured parameters for AlexaFluor 647) (Mollazadeh et al., 2017). Detection rate for blinking events was set to 70%. 10% false detections (noise) were added and distributed randomly across the field of view. Further simulation parameters are described in the Supplementary Methods.

3.5 Real datasets
Methodological details for the dSTORM imaging of integrin α2β1 in platelets is described in the Supplementary Methods. The 2D and 3D SMLM Nup107 data was recently published and described in (Li et al., 2018) where Nup107–SNAP Alexa Fluor 647 was imaged using dSTORM in U-2 OS cells. 3D localization was achieved using an experimental PSF model. SMLM datasets for Las17, Ede1 and Sla1 were recently described in (Mund et al., 2018). In short endocytic proteins were endogenously tagged with the photoconvertible protein mMaple (McEvoy et al., 2012) in budding yeast strains and imaged in high-throughput using a homebuilt SMLM imaging system (Deschamps et al., 2016). Image acquisition for α2β1 dSTORM was performed for formalin fixed A549 cells immunofluorescence labelled for α-tubulin (Alexa-647 secondary) as previously described in Khan et al. (2017). Detections in the integrin α2β1, α-tubulin and Nup107 datasets were filtered by intensity with a minimum value of 1000 photons. For all datasets detections that were found in consecutive frames within a distance of 73 nm were grouped into a single detection.

4 Discussion
As single molecule methods for high throughput data acquisition (Beghin et al., 2017) and sample labelling (Khan et al., 2017) improve, there is a parallel need for automated, robust analytical approaches. The mathematical field of topological data analysis provides a powerful framework for structural analysis of SMLM data. We have introduced tools using both persistence-based clustering (Chazal et al., 2013) and persistent homology (Ghrist, 2007) to quantify biological nano-structure. Empirical evidence suggests ToMATo provides superior clustering performance over commonly used algorithms for SMLM data when clusters are close together, even if clusters vary in density (Andronov et al., 2016; Ester et al., 1996; Levet et al., 2015; Owen et al., 2010). We argue that biological nano-structure is rarely simple and for many applications these conditions will be commonplace, as we show for clustering of collagen receptors in platelets. Moreover selection of free parameters is less sensitive for persistence-based clustering, and can be guided by ToMATo diagrams. Whilst Bayesian approaches (Rubin-Delanchy et al., 2015) to parameter setting are very powerful when the underlying nano-structure can be modeled in advance, it is not suitable for exploratory applications where the nano-structure is not known a priori.

A disadvantage of the ToMATo approach when compared to Voronoi tessellation-based clustering is that there are two free parameters to set, whereas tessellation only has one which can be normalized. The core concepts of this work could also be transferred to other clustering approaches which could be extended to threshold on persistence (Griffith et al., 2015, 2017). For example Voronoi tessellation could be used to calculate the density estimate. The dual graph of the tessellation could also be used to link detections resulting in an algorithm with only one free parameter, providing an interesting avenue for further research.

Persistent homology is designed to reveal topological structure within pointlist datasets and has natural applications for the analysis of SMLM data, as also demonstrated in parallel work (Hofmann et al., 2018). Our framework moves beyond per cluster statistics such as area and density to reveal new information about the underlying topological features within segmented nano-structures. Importantly, this is a multi-scale approach, revealing unbiased structural information at a range of distance scales simultaneously. Furthermore we have developed a novel extension of the standard persistent homology workflow which is based on sub-sampling of detections. This extension ensures the method is robust to SMLM artifacts and provides a confidence value for any specific result. Validation of these approaches was performed using nuclear pore and endocytic site proteins. Similar structural information can be gained from SMLM data by image-based particle averaging (Mund et al., 2018; Szymborska et al., 2013). However persistent homology can be used in cases where particle averaging methods may fail, for example, where the underlying structures vary in shape or size but maintains a constant topology. Persistent homology can also quantify the structure of individual clusters, not just average ensembles, and is suitable for studying smaller datasets. Finally image-based analysis workflows are typically complex with many pre-processing steps specific to the application. Our streamlined workflow requires minimal adaptation for different datasets as demonstrated throughout this study.

Our methods for persistence-based clustering and persistent homology have been implemented and validated in both 2D and 3D. This framework is applicable for a wide range of problems, revealing new topological information and unprecedented insight into biological nano-structure not attainable from existing tools.

Acknowledgements
The authors gratefully acknowledge the equipment and support provided by Deirdre Kavanagh and the Centre of Membrane Proteins and Receptors (COMPARE), Universities of Birmingham and Nottingham, Midlands, UK. Steve P Watson provided many useful comments and advice.

Funding
This work was funded by the Centre of Membrane Proteins and Receptors (COMPARE), Universities of Birmingham and Nottingham, Midlands, UK; and from the British Heart Foundation through the Chair award [CH03/ 003 to Steve P. Watson].

Conflict of Interest: none declared.

References
