

## Ultrasound capsule endoscopy with a mechanically scanning micro-ultrasound

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1                   **Ultrasound capsule endoscopy with a mechanically scanning micro-**  
2   **ultrasound: a porcine study**

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1     **Abstract**

2     Wireless capsule endoscopy has been used for the clinical examination of the gastrointestinal (GI) tract  
3     for two decades. However, most commercially available devices only utilise optical imaging to examine  
4     the GI wall surface. Using this sensing modality, pathology within the GI wall cannot be detected.  
5     Microultrasound ( $\mu$ US) using high-frequency ( $>20$ MHz) ultrasound can provide a means of transmural or  
6     cross-sectional image of the GI tract. Depth of imaging is approximately 10 mm with a resolution of  
7     between 40-120  $\mu$ m that is sufficient to differentiate between subsurface histological layers of the various  
8     regions of the GI tract. Ultrasound capsule endoscopy (USCE) uses a capsule equipped with  $\mu$ US  
9     transducers that are capable of imaging below the GI wall surface, offering thereby a complementary  
10    sensing technique to optical imaging capsule endoscopy. In this work, a USCE device integrated with a  
11    ~30 MHz ultrasonic transducer was developed to capture a full 360° image of the lumen. The  
12    performance of the device was initially evaluated using a wire phantom, indicating an axial resolution of  
13    69.0  $\mu$ m and lateral resolution of 262.5  $\mu$ m. Later, *in vivo* imaging performance was characterised in the  
14    oesophagus and small intestine of anaesthetized pigs. The reconstructed images demonstrate clear layer  
15    differentiation of the lumen wall. The tissue thicknesses measured from the B-scan images show good  
16    agreement with ex vivo images from the literature. The high-resolution ultrasound images in the *in vivo*  
17    porcine model achieved with this device is an encouraging preliminary step in the translation of these  
18    devices towards future clinical use.

19  
20    **Keywords:** Capsule endoscopy, high-frequency ultrasound, micro-ultrasound capsule endoscopy, *in*  
21    *vivo* porcine animal study.

## INTRODUCTION

Conventional endoscopy assesses the gastrointestinal (GI) tract using a flexible probe equipped with either an optical imaging sensor (and a light source) or an ultrasound-emitting device (Dimagno et al. 1980; Valdastrì et al. 2012). Acquired images of the internal GI structure assist in the diagnosis of numerous conditions. However, conventional endoscopy requires the use of highly trained staff, is uncomfortable for patients, sometimes necessitating the use of sedatives or even anaesthetics. Furthermore, it cannot readily investigate all parts of the GI tract, e.g., the small intestine (Cummins et al. 2019). Capsule endoscopy (CE) also integrates optical sensors within an easily-swallowed capsule form factor, typically 11 mm in diameter and 24 - 33 mm long; with a mass of 3 grams, that allows minimally invasive inspection of the entire GI tract for up to six hours, constrained by the battery life (Iddan et al. 2000; Moglia et al. 2009; Wang et al. 2013). These devices are capable of acquiring a considerable amount of image data of the GI tract, which assists in the diagnosis of a wide range of gastrointestinal disease. However, similar to conventional endoscopy, the assessment remains reliant on camera imaging. This limits diagnostic utility to superficially visible manifestations of diseases which are often late stage and difficult to interpret due to visual overlap (Hamilton 2012).

An alternative to optical endoscopy is ultrasound endoscopy (EUS), which is a routinely used clinical imaging modality for the examination of the oesophageal and stomach linings as well as the walls of the upper and lower GI tract. Conventional EUS typically employs ultrasound frequencies in the range 5–18 MHz, corresponding to axial resolutions of 0.2–0.8 mm and depths of 2–8 cm, respectively (McNally 2010). Lower frequency (5-20 MHz) imaging enables observation of organs located beyond the wall of the gastrointestinal tract whereas higher frequencies (>20 MHz) may provide more detailed images of the gut wall at the expense of lower depth penetration (Correia 2009; Wang et al. 2017a). High-frequency imaging is usually achieved through the insertion of high-resolution ultrasound, also known as micro-ultrasound, mini-probes into conventional endoscopes via the biopsy channel (Schembre et al. 2005).

1 These probes have improved axial and lateral image resolution relative to conventional EUS frequencies,  
2 and are capable of producing highly-detailed submucosal images that include information on the structure  
3 and tissue composition.

4 Following the example of clinical utility set by EUS, the inclusion of ultrasound imaging into capsule  
5 endoscopy is a desirable means of improving the diagnostic capabilities of CE (Cummins et al. 2019).  
6 Ultrasound capsule endoscopy (USCE) has been proposed by several groups (Correia 2009; Lay et al.  
7 2019; Lee et al. 2014; Memon et al. 2015; Memon et al. 2016; Wang et al. 2017a). These efforts include  
8 the integration of ring arrays composed of multiple capacitive micromachined ultrasonic transducers  
9 (CMUTs) that operate with frequencies comparable to conventional endoscopy, providing a 360° image  
10 of the organs surrounding the GI tract (Memon et al. 2015; Memon et al. 2016; Wang et al. 2017a). An  
11 example of such a device is shown in Fig. 1. The development of micro-ultrasound capsule endoscopy  
12 ( $\mu$ USCE) capable of producing 360° cross-sectional ultrasound B-scan images is of particular interest due  
13 to high-resolution images that can be achieved of the constituent layers of the GI tract (Panes et al. 2013).  
14 Cross-sectional (i.e. transmural) imaging allows assessment of disease extent across the GI wall.  $\mu$ USCEs  
15 reported to date include a rotating mirror as shown in Fig. 1(a) (Iddan 2010), unfocused 10 MHz  
16 transducer (Lee et al. 2014), focused 30 MHz transducers (Lay et al. 2019), and proposed array-based  
17 methods as shown in Fig. 1(b) (Arneson et al. 2014; Lay et al. 2018). The rotational mirror solution arises  
18 complexity and difficulty in manufacture and assembly of the acoustic mirror. The array-based solution  
19 inherits the technical challenges in the manufacture of high frequency, miniaturized, circumferential  
20 transducer arrays. The rate of development of  $\mu$ USCE technology has been slow due to the challenges  
21 associated with the integration and miniaturization required to realize these capsules.

22 However, mechanical scanning of the transducer can provide the clinician with a high-resolution,  
23 transmural 360° view of the tissue layers of the GI tract and is therefore of immediate interest as it can  
24 potentially provide highly detailed cross sectional images of the GI wall. Compared to the other two  
25 solutions, mechanical rotating transducer, Fig. 1(c), simplifies device fabrication and electronics and is

1 therefore particularly useful for USCE in its early development, paralleling to the development of the  
2 similar solution in intravascular ultrasound (Yu et al. 2017). In a previous study, we demonstrated a  
3 mechanically scanned ultrasound capsule, shown in Fig. 1(c), that showed potential for clinical  
4 applications based on experimental data from *ex vivo* tests (Wang et al. 2017b). The new capsule was  
5 characterized on the bench using wire phantoms. Successful *in vivo*, high-resolution ultrasound results are  
6 also reported for the cross-sectional imaging the small intestine and oesophagus of porcine animal models.

## 7 MATERIALS AND METHODS

### 8 Capsule Device

9 The structure of the proof of concept  $\mu$ USCE device is shown in Fig. 1(d) and consists of a micromotor,  
10 a transducer holder and an ultrasound transducer encased in a biocompatible shell with an acoustically  
11 transparent window. The USCE shell of diameter 10 mm and length 30 mm was made from  
12 biocompatible poly (methyl methacrylate) PMMA, ( $Z=3.2$  MRayl). This size agrees to the majority of  
13 video capsule endoscopes on the market, 11 mm in diameter and 24–33 mm in length (Stewart et al.  
14 2017). The USCE is connected to external instrumentation that drives the motor and controls ultrasound  
15 transduction via a Teflon tube of outer diameter 1.7 mm and inner diameter 1.1 mm (Shenzhen Woer  
16 Heat-Shrinkable Material Co., Ltd. China). A tethered design was used at this stage to ease retrieval of the  
17 device during *in vivo* trials and provide control for the operation of the ultrasonic transducer and the  
18 motor due to the difficulty in sourcing a suitably sized commercially available integrated circuit (IC) that  
19 could be integrated inside the capsule. The rotation of the motor (TT Motor Shenzhen Industrial Co, Ltd,  
20 China) was controlled by varying the amplitude and direction of the input DC current. The motor rotated  
21 the transducer  $360^\circ$  in an oscillatory manner, alternately clockwise and counterclockwise in every 0.2  
22 seconds with a  $\pm 1.2$  V input. This leads to a frame rate of 5 fps for the system.

23 The  $\mu$ US transducer was fabricated from a piezocrystal  $\text{LiNbO}_3$  to operate at a resonant frequency  $\sim 30$   
24 MHz, with a press-focused focal distance of 8 mm and the diameter of 3 mm. Two matching layers were

1 designed for the transducer to compensate for acoustic impedance mismatch between tissue and  
2 piezoelectric material. The first matching layer ( $Z = 7.3 \text{ MRayl}$ ) with  $14 \mu\text{m}$  thickness was made of  
3 Insulcast 501 and Insulcure 9 (American Safety Technologies, Roseland, NJ), and  $2\text{-}3 \mu\text{m}$  silver particles  
4 (Sigma-Aldrich Inc., St. Louis, MO). The second layer was made by depositing an  $18 \mu\text{m}$  thick Parylene  
5 C ( $Z = 2.5 \text{ MRayl}$ ) layer. E-solder 3022 (Von Roll Isola, New Haven, CT) with an acoustic impedance of  
6  $5.9 \text{ MRayl}$  was used as the backing layer.

7 The transducer is placed in a PMMA holder that aligns it with both the central longitudinal axis of the  
8 capsule and the acoustic window. The distance from the front face of the transducer to the capsule outer  
9 surface was about  $6 \text{ mm}$ . During rotation, the transducer emits and receives ultrasound through an  
10 acoustic window in the capsule shell. The acoustic window is made from polydimethylsiloxane (PDMS)  
11 to minimize the reflection and attenuation during the ultrasound transmission. To further optimize the  
12 acoustic transmission, a layer of  $18 \mu\text{m}$  thick Parylene C was coated on both surfaces of the PDMS  
13 material.

14 All components and bonding were achieved with medical-grade epoxy (Epoxy Technologies, Billerica,  
15 MA, USA). The final  $\mu\text{USCE}$  device is shown in Fig. 2(a). The quality of the seal and the mechanical  
16 robustness of empty capsules were quantified prior to insertion into porcine animal models to verify  
17 safety.

## 18 **Imaging Platform**

19 An imaging platform was developed specifically for the evaluation of the proposed  $\mu\text{USCE}$  device as  
20 shown in Fig. 2. A pair of metal-oxide-semiconductor field effect transistors (MOSFETs) (TC6320,  
21 Supertex Inc., Sunnyvale, CA) was employed for the pulse generation. The MOSFETs were excited by  
22 two drivers (EL7158, Intersil Corporation, Milpitas, CA), and a field programmable FPGA device  
23 (Cyclone-V 5CGXFC7D7F31C8N, Altera Corporation, San Jose, CA). The excitation pulses sent from  
24 the imaging platform are single cycle,  $30\text{MHz}$  center frequency, and  $\pm 60 \text{ V}$  amplitude. In the data  
25 acquisition circuitry, a low-noise amplifier (AD8331, Analog Devices, Canton, MA) was used to amplify

1 the echo signals. The gain was set to 45 dB during the data acquisition process. An analog filter was  
2 designed for anti-aliasing filtering to remove high-frequency noise. A 12-bit ADC (AD9230, Analog  
3 Devices) with a maximal sampling rate of 250 mega-samples per second (MSPS), was employed to  
4 digitize the echoic signals. The ultrasound data was then transferred to a computer via a USB 3.0 interface  
5 (CYUSB3014, Cypress, San Jose, CA). The data acquisition process was described in detail previously  
6 by our group (Wang et al. 2017b), the process diagram is shown in Fig. 2(c).

7 The control of the motor back and forth rotation was realized through two analog switch channels to  
8 control the supply of positive and negative power inputs. The circuit diagram and control time sequences  
9 of the positive and negative power paths and relay path are shown in Fig. 3.

## 10 **Animal Study**

11 To demonstrate the effectiveness of the device, *in vivo* studies were performed in the oesophagus and  
12 small intestine of two anaesthetised female Landrace pigs. This model was chosen because of the  
13 anatomical and physiological similarities between the GI tract of pigs and humans (Kararli 1995; Swindle  
14 et al. 2012; Ziegler et al. 2016). The experiments were conducted after approval by the Animal Welfare  
15 and Ethical Review Board of the Roslin Institute (Roslin, Midlothian, EH25 9RG) and under UK Home  
16 License (Project Licence: P3F417A41) in accordance with the Animal (Scientific Procedures) Act 1986.

17 Two four-month old female Landrace pigs weighing 48 and 51 kg were used in the studies. Food was  
18 withheld for 12 hours before anaesthesia to obtain a relatively empty proximal intestine and access to  
19 water was maintained until the pre-anaesthetic medication was injected intramuscularly. This medication  
20 was midazolam ( $0.5 \text{ mg kg}^{-1}$ ), ketamine ( $5 \text{ mg kg}^{-1}$ ) and medetomidine ( $2.5 \mu\text{g kg}^{-1}$ ). As with previous  
21 work (Lay et al. 2019), anaesthesia was induced with isoflurane (IsoFlo, Zoetis, Surrey, UK) vaporized in  
22 oxygen and nitrous oxide administered via a Bain breathing system and facemask. A cannula was placed  
23 in the auricular vein and after intravenous propofol, the trachea was intubated. Anaesthesia was  
24 maintained with isoflurane delivered in an oxygen/air mixture via a circle breathing system. Ringer's  
25 lactate solution (Aquapharm No 11, Animalcare, York, UK) was administered at  $10 \text{ ml kg}^{-1} \text{ hr}^{-1}$  throughout

1 each study to maintain fluid and electrolyte levels. During anaesthesia, the lungs of the pigs were  
2 mechanically ventilated to maintain normocapnia and vital signs were monitored throughout the  
3 experiment by veterinary anaesthetists.

4 Two capsule prototypes were created and each was tested in both the oesophagus and small intestine of  
5 the anaesthetised pigs where ultrasonic echoes were collected during experiments. Because of the  
6 constrained shared space with anaesthetic intubation, a shortened, wide-diameter endotracheal tube was  
7 inserted transorally into the oesophagus. This allowed rapid, direct access to the proximal oesophagus.  
8 During the study, the capsule was inserted at least 36 cm beyond the mouth as measured by the length of  
9 tether inserted down the endotracheal tube. Image data from the capsule was collected at three different  
10 positions, separated from each other by 2 cm as measured from the length of tether manually pulled from  
11 the oesophagus. An artificial stoma was surgically created on the pig's lateral side to directly access the  
12 small intestine. The capsule was inserted directly into the small intestine through the artificial stoma with  
13 the pig lying on the other side under general anaesthetic. Three different points were imaged in the small  
14 intestine and were measured to be 60 cm, 40 cm and 20 cm from the entrance of the stoma as measured  
15 by the length of tether manually pulled from the stoma. The capsule was fixed at these points during  
16 imaging through the use of surgical tape that attached the tether to the surrounding skin. The distances  
17 were marked with tape before the experiments. A saline drip (1-2 drips/s) applied to the entrance of the  
18 stoma and oesophagus ensured lubricity to facilitate capsule insertion and acoustic coupling between the  
19 capsule and the surrounding tissue.

20

21

## RESULTS

22

### Characterisation of $\mu$ USCE

23

24

25

The pulse-echo response of the transducer was measured to characterise its performance. It was  
mounted on a holder and immersed in a tank filled with deionized water. A glass reflector was placed a  
distance of 8 mm from the transducer, corresponding with its focal length. The centre frequency of the

1 ultrasound transducer was measured from the pulse-echo waveform to be 28.15 MHz and the -6 dB  
2 bandwidth is about 50.1%.

3 The spatial resolution of the USCE was characterized using a wire phantom consisting of five 25  $\mu\text{m}$   
4 diameter tungsten wires (Advent Research Materials Ltd, Oxford, UK) spaced 1.4 mm apart. These wires  
5 were placed close to the focal point of the transducer in the capsule and the radial distances of all 5 wires  
6 to the capsule shell were in a range of 1.5-2.5 mm. Fig. 4(a) shows the wire phantom image acquired  
7 using the proposed  $\mu\text{USCE}$  device with a dynamic range of 50 dB. The -6 dB (FWHM, full-width half-  
8 maximum) axial and lateral resolutions of the  $\mu\text{USCE}$  device were 69.0  $\mu\text{m}$  and 262.5  $\mu\text{m}$ , respectively.

### 9 **Small Intestine Imaging**

10 B-scan data of the small intestine was successfully recorded during the *in vivo* experiments. An  
11 example of a post-processed B-scan image of the small intestine, which is reconstructed from 797 sets of  
12 A-scan data, is shown in Fig. 5, demonstrating clear layer differentiation of the lumen wall structure.

13 The sublayers of the small intestine, i.e. mucosa, submucosa, muscularis propria and serosa, can be  
14 clearly observed from the B-scan image (Fig. 5). The tissue sublayer thicknesses represented in the B-  
15 scan image correlates with hematoxylin and eosin (H&E) slides and *ex vivo* ultrasound images of the  
16 porcine small intestine from the literature (Cox et al. 2017). During the experiment, the small intestine  
17 tissue tended to envelop and compress itself against the capsule shell. Hence, the ring down artifact  
18 caused by the capsule shell reduced the visibility of the mucosa layer and varied the sublayer thickness.

### 19 **Oesophagus Imaging**

20 B-scan data of the oesophagus was successfully recorded during the *in vivo* experiments. An example  
21 of a post-processed image, which is reconstructed from 876 sets of A-scan data, is shown in Fig. 6.  
22 Physiologically, the natural corrugations of the tissue of an empty oesophagus form longitudinal folds;  
23 some of these folds flatten out and compress the capsule during the passage. This can be directly observed  
24 from Fig. 6: clear layer differentiation can be seen at the bottom-left of the image where the tissue folded

1 away from the capsule, while ring down artifacts shows at the top-right of the image led to poor layer  
2 differentiation except for the deep layer (i.e. Adventitia). Moreover, pure ring down artifact can be  
3 noticed at the bottom-right of the image. This is thought to be caused by air and poor acoustic coupling  
4 between the capsule and the tissue. More interestingly, an oval-shaped feature can be noticed at the right  
5 of the image, which is thought to be a vein or lymph node, as the size and imaging characteristics  
6 corresponding to relevant anatomical literature (Saar 1962).

## 7 **DISCUSSION AND CONCLUSIONS**

8 Capsule endoscopy has revolutionized the endoscopic imaging through miniaturization, enabling  
9 examination of the entire length of the GI tract with minimal discomfort to patients. However, most  
10 commercially available capsule devices still use white light imaging to image the GI tract, limiting  
11 diagnosis to just surface examination. Some advanced capsules have been proposed for non-optical  
12 diagnostic biomarkers (Cummins et al. 2019) and therapeutic applications (Stewart et al. 2017).

13 Therefore, this study investigated the optimisation of the scan method and further evaluated the device  
14 with *in vivo* animal tests. This paper expands on this earlier work (Wang et al. 2017b) on two fronts: (1)  
15 by describing the next generation of this technology and (2) by reporting on the tests of our capsule in  
16 anaesthetised pigs and achievement of *in vivo* high-resolution ultrasound images. On the technological  
17 side, a simpler mechanical setup and control method was used to achieve 360° sectional images via  
18 mechanical scanning of a high frequency (~30 MHz) ultrasound transducer. This overcomes the  
19 manufacture challenges on 3D printing of small parts, e.g. small gears, and improve the reliability of the  
20 device. It demonstrated that micro-ultrasound with a mechanical rotation scheme is feasible for assessing  
21 the GI tract by providing high-resolution ultrasound images. The proposed scheme in this study paves the  
22 way for the implementation of capsule ultrasound devices. This is further demonstrated by the high-  
23 resolution, cross-sectional *in vivo* imaging of the small intestine and oesophagus possible with this  
24 capsule.

1 The 10 mm diameter and 30 mm length of the prototype devices conformed to commercial video  
2 capsule endoscopes (VCE) dimensions, which are found to be acceptable for clinical use. It is possible to  
3 short the length but require more engineering work. The motor can be customized for at least 4mm shorter  
4 than current design. Although wireless capsules are desirable for the inspection of full GI tract, the  
5 tethered devices not only allow the research on the ultrasound imaging modality in the capsule form to be  
6 successful after insulating other challenges in the development of CE devices, such as power supply, data  
7 communication, location tracking and locomotion control (Basar et al. 2012; Ciuti et al. 2016), but also  
8 give extra benefits in the development stage by its capability of pull-back to the marked positions for  
9 repeated tests. Moreover, there is no easily suitable, commercially-available ICs for USCE onboard image  
10 processing.

11 Satisfactory acoustic coupling between the capsule and the mucosa of the lumen wall was achieved  
12 augmented by saline drips during the *in vivo* experiments. Good agreement of layer structure and sublayer  
13 thickness between *in vivo* images, and *ex vivo* images have been achieved, especially for the small  
14 intestine tissue. Ring down artifacts caused by the capsule shell can reduce the visibility of the mucosa  
15 layer, while the lumen tissue compresses against the capsule. This may indicate that careful design of the  
16 capsule shell will be one of the challenges to be overcome for further development of ultrasound capsule  
17 endoscopes.

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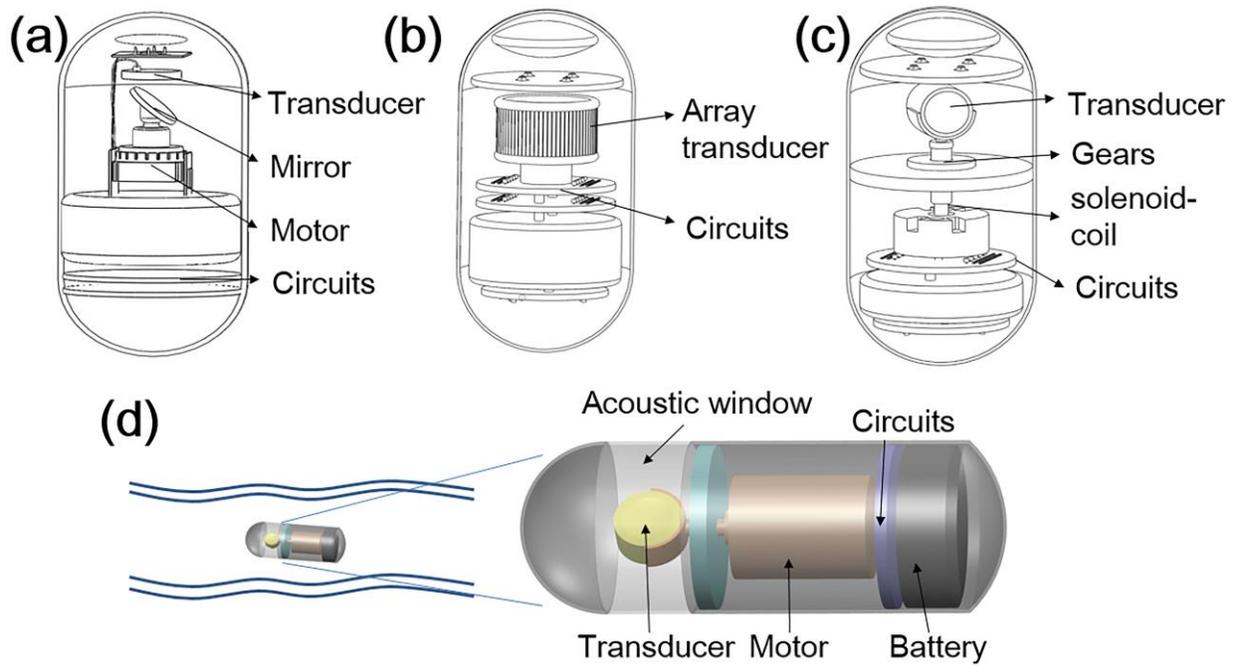
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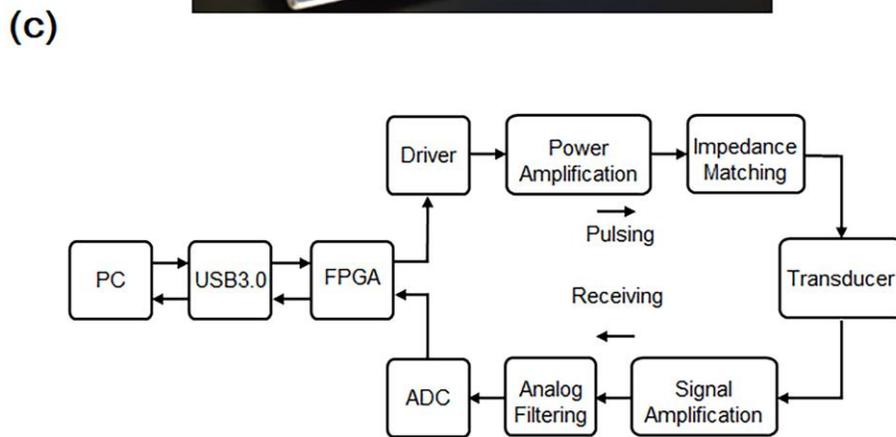
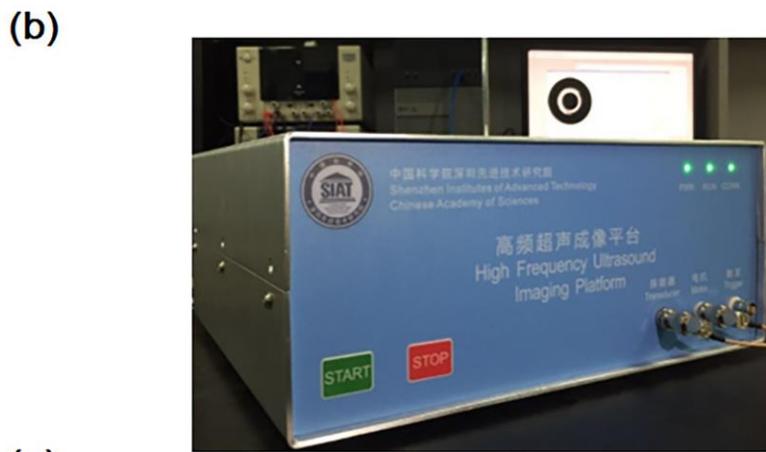
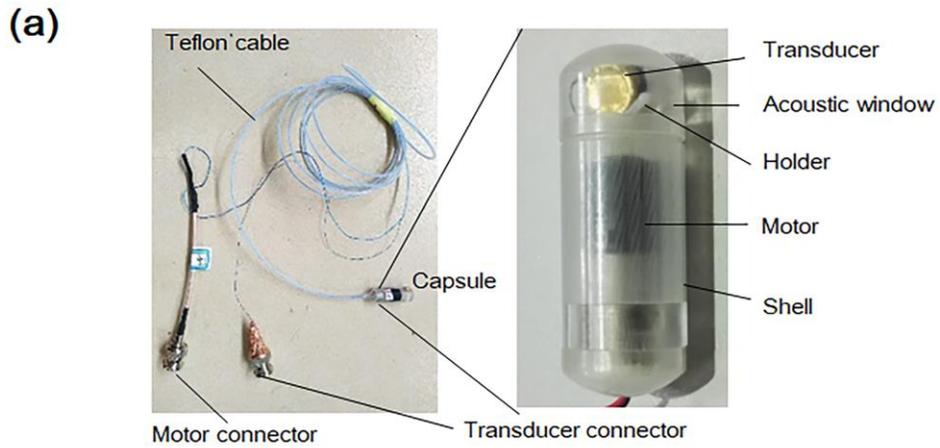
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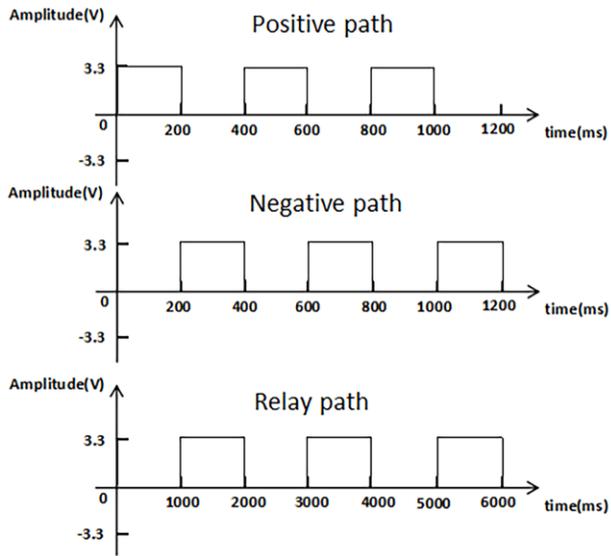
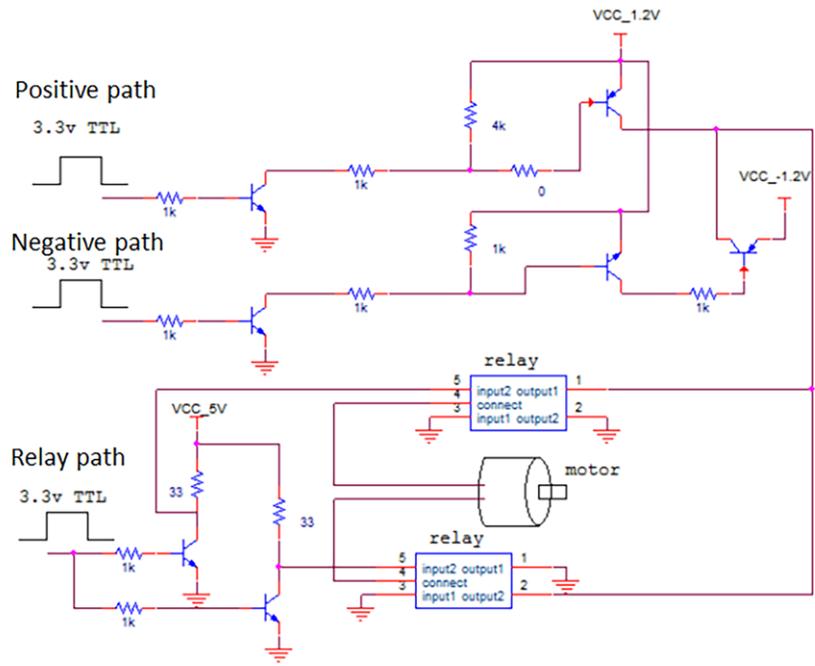


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Figure 1. The schemes towards for ultrasound capsule. (a) A mirror was employed for ultrasound reflection; (b) Circular array transducer-based scheme; (c) A solenoid coil was used to rotate the transducer through gears; (d) A micro-motor was employed to rotate the transducer back and forth for circular imaging.



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2 Figure 2. (a) Tethered  $\mu$ USCE device, (b) high-frequency ultrasound imaging and motor control  
3 system, (c) a process diagram of the ultrasound imaging process.  
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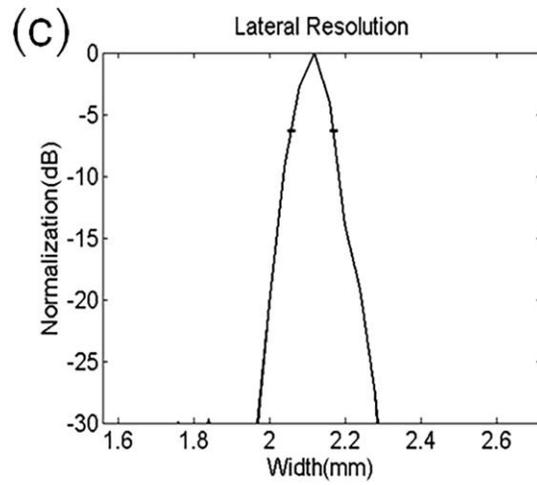
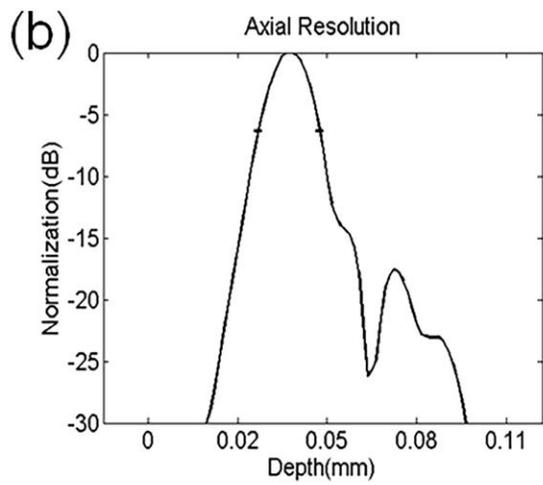
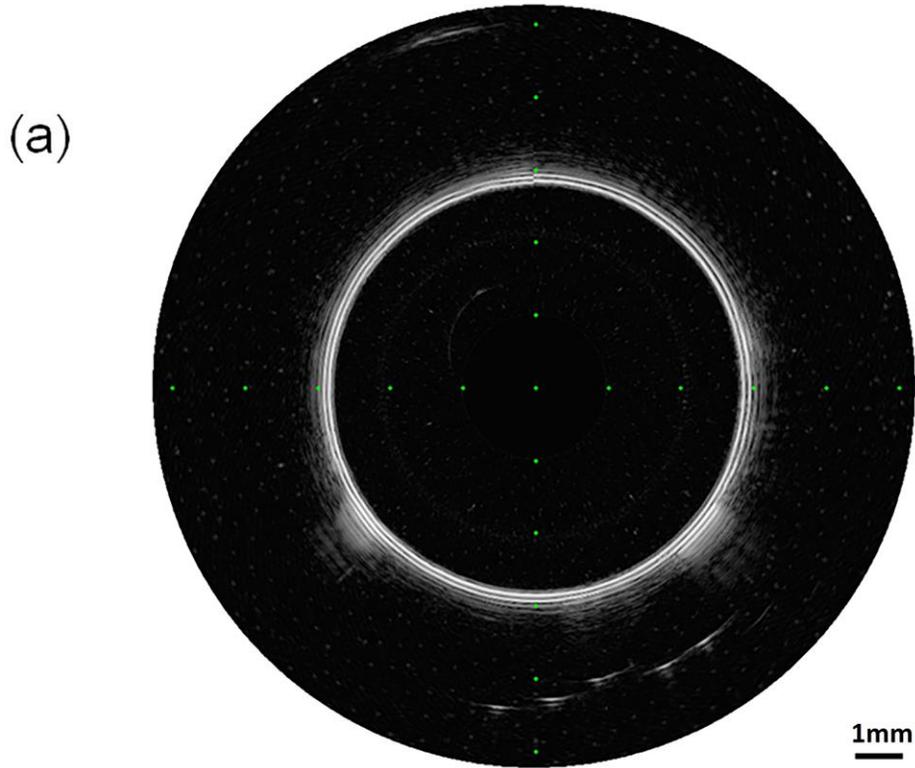


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Figure 3. Micromotor control circuit diagram with input time sequences.

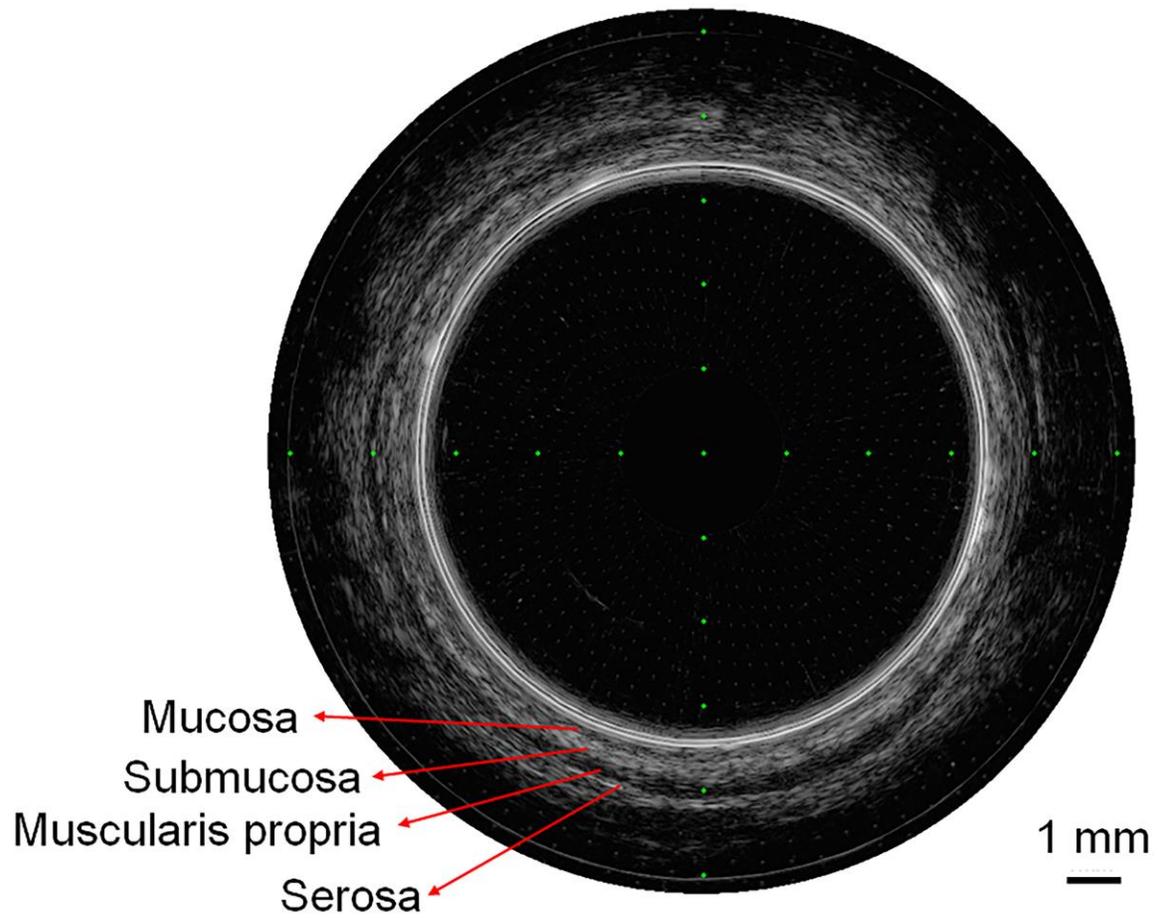
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2 Figure 4. (a) An ultrasound B-scan image of the wire phantom, with (b) axial and (c) lateral resolutions.

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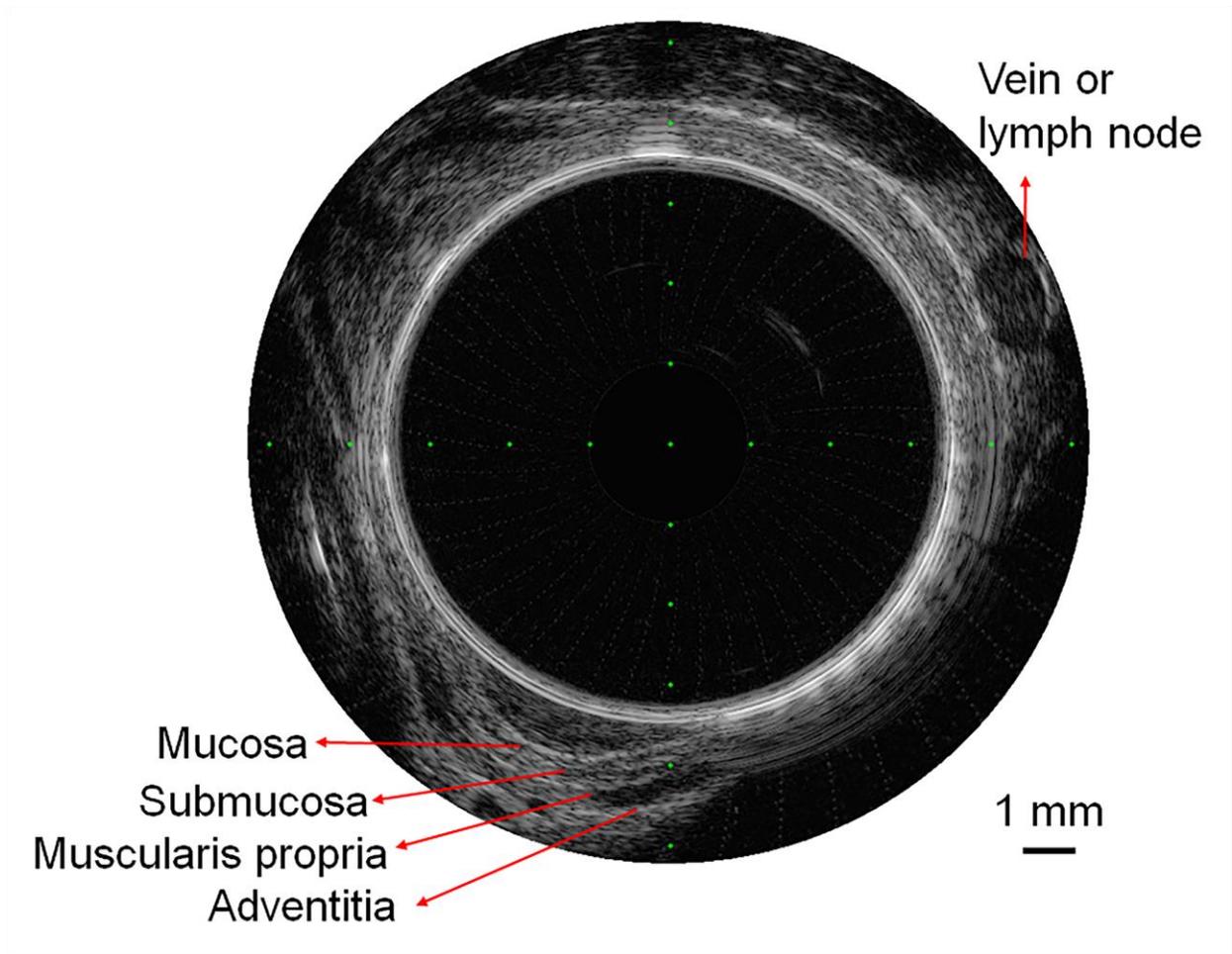


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2 Figure 5. A post-processed cross-sectional ultrasound B-scan transmurial image of a pig small intestine.

3 Cardinal histological layers have been labelled from the superficial luminal layer (i.e. Mucosa) to the  
4 deeper layer (Serosa).

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2 Figure 6. A post-processed cross-sectional ultrasound B-scan transmural image of a pig oesophagus.

3 Cardinal histological layers have been labelled from the superficial luminal layer (i.e. Mucosa) to the

4 deeper layer (Adventitia).

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