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Potential Role of Adipose Tissue and its Hormones in Burns and Critically Ill Patients

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Abstract

Obesity has become a world-wide pandemic and is considered a major risk factor for various diseases. Despite this, recent intriguing clinical observations have been made suggesting that being overweight has some advantages. Overweight and some obese patients were reported to have significantly lower all-cause mortality, described as the 'obesity paradox'. This phenomenon resulted in increased research aimed at investigating the influence of adipose tissue on outcomes of various clinical states including critical illness. In this review, we summarise research findings on the effect burn injury and trauma-related critical illness have on adipose tissue and discuss potential mechanisms by which adipose tissue influences outcomes in burn and other critically ill patients. Burn injury and critical illness influence adipose tissue functionally and morphologically, with circulating levels of fat derived hormones, adipokines, altered in patients following injury and/or critical illness. As adipokines regulate a variety of processes including inflammation and metabolism, this disruption in the adipokine axis may explain the obesity paradox phenomenon observed in critically ill patients. We conclude that further research on the influence of individual adipokines on prognosis in burn and critically ill patients and the mechanisms involved is required to increase understanding of their therapeutic potential.

Highlights

- The Obesity Paradox has been reported in critically ill populations.
- Burn injury and critical illness affect adipose tissue morphologically and functionally.
- Adipokines exert anti- and pro-inflammatory effects influencing patient outcomes.
- Current scar reduction treatments utilising adipose tissue potentially mediate their effects through release of adipokines.

Keywords

Burns, Thermal Injury, Adipose Tissue, Fat, Hormones, Adipokines

Abbreviations

APACHE	Acute Physiological Assessment and Chronic Health Evaluation
CRP	C-Reactive Protein
FFA	Free Fatty Acids
HMG-CoA	Hydroxymethyl Glutaryl Coenzyme A
IL	Interleukin
RCT	Randomised Controlled Trial
SAPS	Simplified Acute Physiology Score
SOFA	Sequential Organ Failure Assessment

Introduction

Obesity is a complex multifactorial condition that affects over a third of the world's population [1]. With increasing prevalence of overweight and obese individuals [2, 3], obesity is being described as a global pandemic [4] as obesity greatly impacts the individual's health status and quality of life [5, 6] being a major risk factor for various pathologies including cancer, cardiovascular disease, diabetes and osteoarthritis [7].

In this context a recent and intriguing observation is that all-cause mortality is reported to be significantly lower in overweight and some obese patients [8]. This phenomenon, where outcomes are paradoxically better in overweight and obese patients compared to normal weight individuals, is described as the 'obesity paradox' and is the subject of increasing interest in scientific and medical communities [9-12]. The underlying mechanisms behind this phenomenon remain poorly understood and this is particularly the case in critically ill populations where the data on the obesity paradox are limited.

This review aims to summarise the observations suggesting the presence of the obesity paradox in critically ill patients, including burns and to discuss potential mechanisms that may explain the difference in outcomes between normal weight, overweight and obese patients, focusing primarily on hormones secreted by adipose tissue (adipokines).

Adipose Tissue in Burns and Critical Illness

Adipose tissue is one of the largest organs in the human body. Importantly, it is no longer deemed an inert tissue that serves the roles of thermal/mechanical insulation protecting internal organs from external stimuli (such as cold and shock) or as an energy storage modality. Since the discovery in 1994 of leptin, an adipokine or adipose-derived hormone capable of controlling body energy balance [13], adipose tissue is now recognised as endocrine organ able to influence metabolism and inflammatory status. As a result extensive research has been carried out investigating potential roles of adipokines in various clinical conditions including autoimmune and inflammatory disorders and connective tissue diseases [14], metabolic disorders [15, 16], cardiovascular and neurovascular diseases [17], and cancer [18, 19].

Despite increasing interest in adipose tissues' role in clinical pathologies, its role in the context of critical illness including burns and trauma remains to be fully elucidated. With thousands of critically ill patients admitted to intensive care units every year [20] some interesting observations have been made. Patients requiring prolonged critical care were reported to lose lean body mass while adipose tissue mass remained preserved or even increased [21, 22]. **Furthermore, although morbid obesity (BMI \geq 40 kg/m²) is an independent risk factor for mortality in critically ill patients [23], improved survival rates were observed among overweight (BMI 25-30 kg/m²) and obese (BMI 30-40 kg/m²) patients compared to normal BMI patients during critical illness [23-26].** These paradoxical findings have stimulated research in to the interplay between critical illness and adipose tissue and their influence on patient outcomes. Moreover the profound

inflammatory and metabolic response to burn and trauma related critical illness suggest a potential involvement for adipose tissue and adipokines.

Critical illness following injury is a multifactorial heterogeneous disorder characterised by an overwhelming pro-inflammatory response accompanied by a compensatory anti-inflammatory reaction and subsequent immunosuppression [27, 28]. This classical paradigm also applies to severe forms of critical illness such as burns, the pathology of which we have described previously [29, 30]. The human response to burn injury includes a so-called 'genomic storm'[31], consistent with simultaneous increased systemic inflammation, innate immune activation and anti-inflammatory response [32, 33], as well as suppression of adaptive immunity[31]. In addition, burn patients and others with severe critical illness suffer from a prolonged hypermetabolic, hypercatabolic response [33, 34].

The metabolic response following thermal injury is characterised as a two phase response: the 'ebb' phase within 48 hours where metabolism, cardiac output and oxygen consumption are reduced, followed by the 'flow' phase at approximately 120 hours post-injury where these parameters increase and plateau [35]. This metabolic response includes: peripheral lipolysis and free fatty acid (FFA) [36] oxidation leading to an acute, global and complex increase in FFA levels[37]; systemic induction of endoplasmic reticulum (ER) stress and unfolded protein response [38]; up to 6-fold increase in breakdown rates of skeletal muscle protein[39]; elevation in resting energy expenditure up to 140%[40] that can be prolonged [33].

Burns and other severe critical illnesses have been reported to influence adipose tissue morphologically and functionally. Saraf *et al* reported the impact of severe burn injury on subcutaneous white adipose tissue in children and observed significantly reduced adipocyte size, increased collagen deposition and cell mitochondria content, increased immune cells such as macrophages, as well as increased inflammatory cytokine production [41]. These morphological changes suggest "browning" of subcutaneous adipose tissue following thermal injury, a finding which was confirmed biochemically and functionally. Sidossis *et al* reported significantly increased mitochondrial density and mitochondrial respiratory capacity, as well as an 80-fold increase in the expression of uncoupling protein 1 (UCP1), a molecule abundantly observed in brown adipose tissue depots [42], in burn patients compared to healthy controls [43]. In addition, Patsouris *et al* reported similar findings including significantly increased mitochondrial mass and adipose tissue browning markers in burn patients [44]. This could be a compensatory mechanism since brown adipose tissue is known to induce thermogenesis, modulate energy expenditure and exert local tissue effects such as stimulating angiogenesis and influencing macrophage polarization [45]. Similar morphological and metabolic activity alterations of adipose tissue have been reported in critically ill patients[46, 47]. A functional aspect of adipose tissue is its endocrine role through the production of adipokines and these may mediate many of the effects seen in burns and critical illness.

Overview of Adipokines and their Biological Effects

There are approximately 600 identified hormones secreted by adipose tissue [48], providing a rich source of potential novel biomarkers and therapeutic targets for the management of various pathologies. In this review, we will focus on Adiponectin, Ghrelin, Leptin, Resistin and Visfatin as the best characterised adipokines.

Adiponectin is released exclusively from white adipose tissue[49], and is the most abundant adipose-specific adipokine, with expression in subcutaneous fat being greater than visceral fat [50]. Adiponectin has anti-inflammatory effects [51]. Ghrelin is an orexigenic hormone that is an endogenous ligand to growth hormone and was initially thought to be produced mainly by the stomach [52], but has subsequently been identified in other tissues including adipose tissue [53]. Ghrelin signaling is associated with adiposity, changes in fat distribution and mobilisation, independent of growth hormone and dietary intake[54, 55]. Leptin is primarily secreted by subcutaneous white adipose tissue, the amount of leptin secreted into the circulation is proportional to adipose tissue mass and nutritional status[51]. Leptin exhibits structural similarities to cytokines [56] and is pro-inflammatory [57]. Resistin is also a pro-inflammatory adipokine expressed by adipocytes and other tissues including skeletal muscle [58, 59]. Visfatin, also called pre-B-cell colony enhancing factor, is primarily secreted by adipocytes in visceral white adipose tissue and exhibits pro-inflammatory effects [60].

Relevant to this review, adipokines have been reported to influence skin and adipose tissue. Adiponectin and ghrelin have been observed to exert anti-inflammatory and anti-fibrotic effects on skin [61-63] and were reported to enhance wound healing rates [64, 65]. Similarly, leptin has been observed to enhance human epidermal keratinocyte and epithelial cell proliferation, differentiation and migration, as well as promote angiogenesis within dermal connective tissues [66]. However, leptin was also found to be overexpressed in hypertrophic and keloid scars [67]. This could be due to increased pro-inflammatory cytokine release associated with leptin, as seen in inflammatory skin conditions [68]. Visfatin has been reported to enhance chemokine and antimicrobial peptide production in human keratinocytes [69, 70], as well as exhibit anti-fibrotic properties[71].

Adiponectin and leptin have been reported to induce browning of adipose tissue [72, 73] and adiponectin promotes adipogenesis as well as increasing lipid accumulation and insulin responsiveness of adipocytes [74]. In contrast, leptin inhibits insulin-dependent glucose uptake and lipogenesis and reverses insulin-induced lipolysis [75]. Ghrelin stimulates adipogenesis and glucose uptake, as well as inhibiting lipolysis, apoptosis and autophagy of adipocytes [76, 77]. Resistin and visfatin enhance pro-inflammatory cytokine expression in adipose tissue including TNF- α and IL-6 [78, 79]. Similarly, resistin and visfatin induce insulin resistance in adipocytes [79, 80]. The influence of these adipokines is not limited to skin and adipose tissue. The beneficial and detrimental effects of these adipokines on various cell types and tissues are summarized in Figure 1 and the reader is also referred to recent reviews for further detail [17, 81-88].

Figure 1. Effects of adipokines on various tissues and organs

Adipokine changes in burns and critically ill patients

Several studies have demonstrated acute reductions in circulating adiponectin levels in critical illness and/or injury including burns, sepsis and trauma [89-95]. In addition, an inverse association was reported between serum adiponectin levels and severity of illness as measured by C-Reactive Protein (CRP), Simplified Acute Physiology Score (SAPS) II and Sequential Organ Failure Assessment (SOFA) scores [91, 92, 95]. Similar findings were observed in patients with acute pancreatitis, where adiponectin levels in the blood were negatively associated with severity of disease and incidence of tissue necrosis [93]. Furthermore, adiponectin levels progressively increase with patient recovery [91, 94]. Although the above findings indicate that decreased serum adiponectin levels may lead to poor outcomes, other research has reported different findings. Circulating adiponectin levels in severely ill patients did not correlate with inflammatory markers including Interleukin (IL) -6, IL-10 and Tumour Necrosis Factor (TNF)- α [89, 96, 97] and clinical scores including Acute Physiologic Assessment and Chronic Health Evaluation (APACHE) II score and SOFA [96, 98, 99]. Furthermore, higher blood adiponectin levels were associated with increased risk of mortality during critical illness [100-103].

Only two studies have investigated circulating ghrelin levels in critical illness. Wade *et al* reported significantly reduced ghrelin levels in severely burned patients correlating with metabolic/caloric needs. **No other associations with other parameters such as injury severity and inflammatory status were observed [89].** Santacruz *et al* also observed significantly reduced plasma ghrelin levels in critically ill patients but saw no correlations with feeding status [104].

Leptin levels in the blood have been reported to be elevated in critical illness [105-108]. Furthermore, leptin was positively associated with pro-inflammatory status of severely ill patients, as measured by CRP, IL-6, sTNFR1 and TNF- α [106, 108-111]. **Additionally, other studies have reported that serum-soluble leptin receptor (SLR) in patients correlated with inflammatory response and illness severity as measured by IL-6, lactate, procalcitonin and APACHE II score [112, 113].** Interestingly, elevated levels of leptin were observed in survivors of acute sepsis [105], while increased SLR levels in critically ill patients were associated with increased mortality [112]. However, other studies have reported different findings. Blood leptin levels in severely ill patients were similar or reduced compared to healthy volunteers [89, 94, 95, 109, 112, 114] and **no associations were found between circulating leptin levels and inflammatory status, illness severity, or mortality in critical illness [94, 95, 107, 109, 111, 112, 114].**

In contrast to the heterogeneity of results reported on the impact adiponectin, ghrelin and leptin on critical illness outcomes, the influence of resistin and visfatin on outcomes

of severely ill patients is consistent in the literature. Critically ill patients exhibit significantly elevated circulating levels of resistin [89, 95, 98, 99, 111, 115-120] and visfatin [111, 121-127]. Additionally, both resistin and visfatin significantly correlated with pro-inflammatory responses (including CRP, IL-6, IL-8 and TNF- α), and worse clinical severity scores (including APACHE II, Glasgow Coma score, multiple organ dysfunction score, SAPS II and SOFA)[89, 95, 98, 99, 111, 115-119, 121-127]. Furthermore, high resistin and visfatin levels in blood were associated with poor outcomes including mortality [116, 117, 122-126].

A systematic review examining the evidence for adipokines having an influence on critical care patients has been published recently [128]. It concludes that although strong observations were reported indicating the influence of adipokines on the prognosis of critical illness, additional larger studies that incorporate more diverse cohorts (such as age, gender, BMI, ethnic groups and different pathologies) is required to better understand the relationship between adipokines and critical illness. This is essential in order to validate the potential clinical value and utility of adipokines as diagnostic and/or prognostic biomarkers, as well their potential as therapeutic targets in critical illness including burn and trauma. Furthermore, studies to date have investigated the association of adipokines with critical illness in the acute setting only. This focus on the acute setting has further limited the translation of adipokines in clinical settings. Importantly, since medical care advancements have improved survival rates after critical trauma [129-131], greater emphasis is now placed on the prevention and treatment of potentially debilitating long-term sequelae experienced by survivors of severe illness including chronic critical illness [132-134], prolonged pathophysiological responses[33] and scarring [135].

Conclusions

Several studies have reported changes in the serum levels of specific adipokines and their role in the regulation of a range of biological responses to injury including inflammation, metabolic dysregulation and wound healing is emerging. However a robust characterisation of the impact of such changes in individual adipokines on patient outcomes, especially in burns patients, is lacking. Large clinical and scientific studies are required to establish the mechanisms by which adipose tissue may influence patient outcomes and translate the research into clinical practice to improve short and long-term outcomes of burn and critically ill patients.

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References

- [1] Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*. 2014;384:766-81.
- [2] Trends in adult body-mass index in 200 countries from 1975 to 2014: a pooled analysis of 1698 population-based measurement studies with 19.2 million participants. *Lancet*. 2016;387:1377-96.
- [3] Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: a pooled analysis of 2416 population-based measurement studies in 128.9 million children, adolescents, and adults. *Lancet*. 2017;390:2627-42.
- [4] Swinburn BA, Sacks G, Hall KD, McPherson K, Finegood DT, Moodie ML, et al. The global obesity pandemic: shaped by global drivers and local environments. *Lancet*. 2011;378:804-14.
- [5] Visscher TL, Seidell JC. The public health impact of obesity. *Annual review of public health*. 2001;22:355-75.
- [6] Taylor VH, Forhan M, Vigod SN, McIntyre RS, Morrison KM. The impact of obesity on quality of life. *Best practice & research Clinical endocrinology & metabolism*. 2013;27:139-46.
- [7] Seidell JC, Halberstadt J. The global burden of obesity and the challenges of prevention. *Annals of nutrition & metabolism*. 2015;66 Suppl 2:7-12.
- [8] Flegal KM, Kit BK, Orpana H, Graubard BI. Association of all-cause mortality with overweight and obesity using standard body mass index categories: a systematic review and meta-analysis. *Jama*. 2013;309:71-82.
- [9] Braun N, Gomes F, Schutz P. "The obesity paradox" in disease--is the protective effect of obesity true? *Swiss medical weekly*. 2015;145:w14265.
- [10] Park J, Ahmadi SF, Streja E, Molnar MZ, Flegal KM, Gillen D, et al. Obesity paradox in end-stage kidney disease patients. *Progress in cardiovascular diseases*. 2014;56:415-25.
- [11] Lavie CJ, McAuley PA, Church TS, Milani RV, Blair SN. Obesity and cardiovascular diseases: implications regarding fitness, fatness, and severity in the obesity paradox. *Journal of the American College of Cardiology*. 2014;63:1345-54.
- [12] Valentijn TM, Galal W, Tjeertes EK, Hoeks SE, Verhagen HJ, Stolker RJ. The obesity paradox in the surgical population. *The surgeon : journal of the Royal Colleges of Surgeons of Edinburgh and Ireland*. 2013;11:169-76.
- [13] Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature*. 1994;372:425-32.
- [14] Fietta P, Delsante G. Focus on adipokines. *Theoretical biology forum*. 2013;106:103-29.
- [15] Jialal I, Devaraj S. Subcutaneous adipose tissue biology in metabolic syndrome. *Hormone molecular biology and clinical investigation*. 2018;33.
- [16] Nicholson T, Church C, Baker DJ, Jones SW. The role of adipokines in skeletal muscle inflammation and insulin sensitivity. *Journal of inflammation (London, England)*. 2018;15:9.

- [17] Opatrilova R, Caprnda M, Kubatka P, Valentova V, Uramova S, Nosal V, et al. Adipokines in neurovascular diseases. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. 2018;98:424-32.
- [18] Lengyel E, Makowski L, DiGiovanni J, Kolonin MG. Cancer as a Matter of Fat: The Crosstalk between Adipose Tissue and Tumors. *Trends in cancer*. 2018;4:374-84.
- [19] Morris EV, Edwards CM. Adipokines, adiposity, and bone marrow adipocytes: Dangerous accomplices in multiple myeloma. *J Cell Physiol*. 2018.
- [20] Mullins PM, Goyal M, Pines JM. National growth in intensive care unit admissions from emergency departments in the United States from 2002 to 2009. *Academic emergency medicine : official journal of the Society for Academic Emergency Medicine*. 2013;20:479-86.
- [21] Hart DW, Wolf SE, Herndon DN, Chinkes DL, Lal SO, Obeng MK, et al. Energy expenditure and caloric balance after burn: increased feeding leads to fat rather than lean mass accretion. *Ann Surg*. 2002;235:152-61.
- [22] Plank LD, Connolly AB, Hill GL. Sequential changes in the metabolic response in severely septic patients during the first 23 days after the onset of peritonitis. *Ann Surg*. 1998;228:146-58.
- [23] Nasraway SA, Jr., Albert M, Donnelly AM, Ruthazer R, Shikora SA, Saltzman E. Morbid obesity is an independent determinant of death among surgical critically ill patients. *Crit Care Med*. 2006;34:964-70; quiz 71.
- [24] Fonarow GC, Srikanthan P, Costanzo MR, Cintron GB, Lopatin M. An obesity paradox in acute heart failure: analysis of body mass index and inhospital mortality for 108,927 patients in the Acute Decompensated Heart Failure National Registry. *American heart journal*. 2007;153:74-81.
- [25] Peake SL, Moran JL, Ghelani DR, Lloyd AJ, Walker MJ. The effect of obesity on 12-month survival following admission to intensive care: a prospective study. *Crit Care Med*. 2006;34:2929-39.
- [26] Trivedi V, Jean RE, Genese F, Fuhrmann KA, Saini AK, Mangulabnan VD, et al. Impact of Obesity on Outcomes in a Multiethnic Cohort of Medical Intensive Care Unit Patients. *Journal of intensive care medicine*. 2018;33:97-103.
- [27] Greathouse KC, Hall MW. Critical Illness-Induced Immune Suppression: Current State of the Science. *American journal of critical care : an official publication, American Association of Critical-Care Nurses*. 2016;25:85-92.
- [28] Vanzant EL, Lopez CM, Ozrazgat-Baslanti T, Ungaro R, Davis R, Cuenca AG, et al. Persistent inflammation, immunosuppression, and catabolism syndrome after severe blunt trauma. *The journal of trauma and acute care surgery*. 2014;76:21-9; discussion 9-30.
- [29] Al-Tarrah K, Hewison M, Moiemien N, Lord JM. Vitamin D status and its influence on outcomes following major burn injury and critical illness. *Burns & trauma*. 2018;6:11.
- [30] Al-Tarrah K, Moiemien N, Lord JM. The influence of sex steroid hormones on the response to trauma and burn injury. *Burns & trauma*. 2017;5:29.
- [31] Xiao W, Mindrinos MN, Seok J, Cuschieri J, Cuenca AG, Gao H, et al. A genomic storm in critically injured humans. *J Exp Med*. 2011;208:2581-90.
- [32] Hampson P, Dinsdale RJ, Wearn CM, Bamford AL, Bishop JRB, Hazeldine J, et al. Neutrophil Dysfunction, Immature Granulocytes, and Cell-free DNA are Early

- Biomarkers of Sepsis in Burn-injured Patients: A Prospective Observational Cohort Study. *Ann Surg.* 2017;265:1241-9.
- [33] Jeschke MG, Gauglitz GG, Kulp GA, Finnerty CC, Williams FN, Kraft R, et al. Long-term persistence of the pathophysiologic response to severe burn injury. *PLoS one.* 2011;6:e21245.
- [34] De Cosmi V, Milani GP, Mazzocchi A, D'Oria V, Silano M, Calderini E, et al. The Metabolic Response to Stress and Infection in Critically Ill Children: The Opportunity of an Individualized Approach. *Nutrients.* 2017;9.
- [35] Wolfe RR. Review: acute versus chronic response to burn injury. *Circ Shock.* 1981;8:105-15.
- [36] Otterbein LR, Cosio C, Graceffa P, Dominguez R. Crystal structures of the vitamin D-binding protein and its complex with actin: structural basis of the actin-scavenger system. *Proc Natl Acad Sci U S A.* 2002;99:8003-8.
- [37] Qi P, Abdullahi A, Stanojic M, Patsouris D, Jeschke MG. Lipidomic analysis enables prediction of clinical outcomes in burn patients. *Scientific reports.* 2016;6:38707.
- [38] Jeschke MG, Finnerty CC, Herndon DN, Song J, Boehning D, Tompkins RG, et al. Severe Injury Is Associated With Insulin Resistance, Endoplasmic Reticulum Stress Response, and Unfolded Protein Response. *Ann Surg.* 2012;255:370-8.
- [39] Chao T, Herndon DN, Porter C, Chondronikola M, Chaidemenou A, Abdelrahman DR, et al. Skeletal Muscle Protein Breakdown Remains Elevated in Pediatric Burn Survivors up to One-Year Post-Injury. *Shock.* 2015;44:397-401.
- [40] Jeschke MG, Chinkes DL, Finnerty CC, Kulp G, Suman OE, Norbury WB, et al. Pathophysiologic response to severe burn injury. *Ann Surg.* 2008;248:387-401.
- [41] Saraf MK, Herndon DN, Porter C, Toliver-Kinsky T, Radhakrishnan R, Chao T, et al. Morphological Changes in Subcutaneous White Adipose Tissue After Severe Burn Injury. *Journal of burn care & research : official publication of the American Burn Association.* 2016;37:e96-103.
- [42] Kalinovich AV, de Jong JM, Cannon B, Nedergaard J. UCP1 in adipose tissues: two steps to full browning. *Biochimie.* 2017;134:127-37.
- [43] Sidossis LS, Porter C, Saraf MK, Borsheim E, Radhakrishnan RS, Chao T, et al. Browning of Subcutaneous White Adipose Tissue in Humans after Severe Adrenergic Stress. *Cell metabolism.* 2015;22:219-27.
- [44] Patsouris D. Burn Induces Browning of the Subcutaneous White Adipose Tissue in Mice and Humans. 2015;13:1538-44.
- [45] Wang GX, Zhao XY, Lin JD. The brown fat secretome: metabolic functions beyond thermogenesis. *Trends in endocrinology and metabolism: TEM.* 2015;26:231-7.
- [46] Langouche L, Perre SV, Thiessen S, Gunst J, Hermans G, D'Hoore A, et al. Alterations in adipose tissue during critical illness: An adaptive and protective response? *American journal of respiratory and critical care medicine.* 2010;182:507-16.
- [47] Marques MB, Langouche L. Endocrine, metabolic, and morphologic alterations of adipose tissue during critical illness. *Crit Care Med.* 2013;41:317-25.
- [48] Lehr S, Hartwig S, Sell H. Adipokines: a treasure trove for the discovery of biomarkers for metabolic disorders. *Proteomics Clinical applications.* 2012;6:91-101.

- [49] Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem*. 1995;270:26746-9.
- [50] Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the Release of Adipokines by Adipose Tissue, Adipose Tissue Matrix, and Adipocytes from Visceral and Subcutaneous Abdominal Adipose Tissues of Obese Humans. *Endocrinology*. 2004;145:2273-82.
- [51] Ahima RS. Metabolic actions of adipocyte hormones: focus on adiponectin. *Obesity* (Silver Spring, Md). 2006;14 Suppl 1:9s-15s.
- [52] Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature*. 1999;402:656-60.
- [53] Baatar D, Patel K, Taub DD. The effects of ghrelin on inflammation and the immune system. *Molecular and cellular endocrinology*. 2011;340:44-58.
- [54] Sangiao-Alvarellos S, Vazquez MJ, Varela L, Nogueiras R, Saha AK, Cordido F, et al. Central ghrelin regulates peripheral lipid metabolism in a growth hormone-independent fashion. *Endocrinology*. 2009;150:4562-74.
- [55] Al Massadi O, Lopez M, Tschop M, Dieguez C, Nogueiras R. Current Understanding of the Hypothalamic Ghrelin Pathways Inducing Appetite and Adiposity. *Trends in neurosciences*. 2017;40:167-80.
- [56] Leal VdO, Mafrá D. Adipokines in obesity. *Clinica Chimica Acta*. 2013;419:87-94.
- [57] Fernandez-Riejos P, Najib S, Santos-Alvarez J, Martin-Romero C, Perez-Perez A, Gonzalez-Yanes C, et al. Role of leptin in the activation of immune cells. *Mediators of inflammation*. 2010;2010:568343.
- [58] Stepan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, et al. The hormone resistin links obesity to diabetes. *Nature*. 2001;409:307-12.
- [59] Jamaluddin MS, Weakley SM, Yao Q, Chen C. Resistin: functional roles and therapeutic considerations for cardiovascular disease. *British Journal of Pharmacology*. 2012;165:622-32.
- [60] Sun Z, Lei H, Zhang Z. Pre-B cell colony enhancing factor (PBEF), a cytokine with multiple physiological functions. *Cytokine & growth factor reviews*. 2013;24:433-42.
- [61] Lakota K, Wei J, Carns M, Hinchcliff M, Lee J, Whitfield ML, et al. Levels of adiponectin, a marker for PPAR-gamma activity, correlate with skin fibrosis in systemic sclerosis: potential utility as biomarker? *Arthritis research & therapy*. 2012;14:R102.
- [62] Shibata S, Tada Y, Hau CS, Mitsui A, Kamata M, Asano Y, et al. Adiponectin regulates psoriasiform skin inflammation by suppressing IL-17 production from gammadelta-T cells. *Nature communications*. 2015;6:7687.
- [63] Koca SS, Ozgen M, Sarikaya M, Dagli F, Ustundag B, Isik A. Ghrelin prevents the development of dermal fibrosis in bleomycin-induced scleroderma. *Clinical and experimental dermatology*. 2014;39:176-81.
- [64] Liu C, Huang J, Li H, Yang Z, Zeng Y, Liu J, et al. Ghrelin accelerates wound healing through GHS-R1a-mediated MAPK-NF- κ B/GR signaling pathways in combined radiation and burn injury in rats. *Scientific reports*. 2016;6.
- [65] Shibata S, Tada Y, Asano Y, Hau CS, Kato T, Saeki H, et al. Adiponectin regulates cutaneous wound healing by promoting keratinocyte proliferation and migration via the ERK signaling pathway. *J Immunol*. 2012;189:3231-41.

- [66] Tadokoro S, Ide S, Tokuyama R, Umeki H, Tatehara S, Kataoka S, et al. Leptin Promotes Wound Healing in the Skin. *PloS one*. 2015;10.
- [67] Seleit I, Bakry OA, Samaka RM, Tawfik AS. Immunohistochemical Evaluation of Leptin Expression in Wound Healing: A Clue to Exuberant Scar Formation. *Applied immunohistochemistry & molecular morphology : AIMM*. 2016;24:296-306.
- [68] Johnston A, Arnadottir S, Gudjonsson JE, Aphale A, Sigmarsdottir AA, Gunnarsson SI, et al. Obesity in psoriasis: Leptin and resistin as mediators of cutaneous inflammation. *The British journal of dermatology*. 2008;159:342-50.
- [69] Hau CS, Kanda N, Noda S, Tatsuta A, Kamata M, Shibata S, et al. Visfatin enhances the production of cathelicidin antimicrobial peptide, human beta-defensin-2, human beta-defensin-3, and S100A7 in human keratinocytes and their orthologs in murine imiquimod-induced psoriatic skin. *Am J Pathol*. 2013;182:1705-17.
- [70] Kanda N, Hau CS, Tada Y, Tatsuta A, Sato S, Watanabe S. Visfatin enhances CXCL8, CXCL10, and CCL20 production in human keratinocytes. *Endocrinology*. 2011;152:3155-64.
- [71] Masui Y, Asano Y, Shibata S, Noda S, Akamata K, Aozasa N, et al. A possible contribution of visfatin to the resolution of skin sclerosis in patients with diffuse cutaneous systemic sclerosis via a direct anti-fibrotic effect on dermal fibroblasts and Th1 polarization of the immune response. *Rheumatology (Oxford, England)*. 2013;52:1239-44.
- [72] Hui X, Gu P, Zhang J, Nie T, Pan Y, Wu D, et al. Adiponectin Enhances Cold-Induced Browning of Subcutaneous Adipose Tissue via Promoting M2 Macrophage Proliferation. *Cell metabolism*. 2015;22:279-90.
- [73] Dodd G, Descherf S, Loh K, Simonds SE, Wiede F, Balland E, et al. Leptin and insulin act on POMC neurons to promote the browning of white fat. *Cell*. 2015;160:88-104.
- [74] Fu Y, Luo N, Klein RL, Garvey WT. Adiponectin promotes adipocyte differentiation, insulin sensitivity, and lipid accumulation. *Journal of lipid research*. 2005;46:1369-79.
- [75] Harris RB. Direct and indirect effects of leptin on adipocyte metabolism. *Biochim Biophys Acta*. 2014;1842:414-23.
- [76] Rodriguez A. Novel molecular aspects of ghrelin and leptin in the control of adipobiology and the cardiovascular system. *Obesity facts*. 2014;7:82-95.
- [77] Rodriguez A, Gomez-Ambrosi J, Catalan V, Rotellar F, Valenti V, Silva C, et al. The ghrelin O-acyltransferase-ghrelin system reduces TNF-alpha-induced apoptosis and autophagy in human visceral adipocytes. *Diabetologia*. 2012;55:3038-50.
- [78] Lehrke M, Reilly MP, Millington SC, Iqbal N, Rader DJ, Lazar MA. An inflammatory cascade leading to hyperresistinemia in humans. *PLoS medicine*. 2004;1:e45.
- [79] Chang YC, Chang TJ, Lee WJ, Chuang LM. The relationship of visfatin/pre-B-cell colony-enhancing factor/nicotinamide phosphoribosyltransferase in adipose tissue with inflammation, insulin resistance, and plasma lipids. *Metabolism: clinical and experimental*. 2010;59:93-9.
- [80] Fu Y, Luo L, Luo N, Garvey WT. Proinflammatory cytokine production and insulin sensitivity regulated by overexpression of resistin in 3T3-L1 adipocytes. *Nutrition & metabolism*. 2006;3:28.

- [81] Shibata R, Ouchi N, Ohashi K, Murohara T. The role of adipokines in cardiovascular disease. *Journal of cardiology*. 2017;70:329-34.
- [82] Romacho T, Elsen M, Rohrborn D, Eckel J. Adipose tissue and its role in organ crosstalk. *Acta physiologica (Oxford, England)*. 2014;210:733-53.
- [83] Li F, Li Y, Duan Y, Hu CA, Tang Y, Yin Y. Myokines and adipokines: Involvement in the crosstalk between skeletal muscle and adipose tissue. *Cytokine & growth factor reviews*. 2017;33:73-82.
- [84] Zhu Q, Scherer PE. Immunologic and endocrine functions of adipose tissue: implications for kidney disease. *Nature reviews Nephrology*. 2018;14:105-20.
- [85] Feakins RM. Obesity and metabolic syndrome: pathological effects on the gastrointestinal tract. *Histopathology*. 2016;68:630-40.
- [86] Hawkes CP, Mostoufi-Moab S. Fat-bone interaction within the bone marrow milieu: Impact on hematopoiesis and systemic energy metabolism. *Bone*. 2018.
- [87] Francisco V, Perez T, Pino J, Lopez V, Franco E, Alonso A, et al. Biomechanics, obesity, and osteoarthritis. The role of adipokines: When the levee breaks. *Journal of orthopaedic research : official publication of the Orthopaedic Research Society*. 2018;36:594-604.
- [88] Maurizi G, Della Guardia L, Maurizi A, Poloni A. Adipocytes properties and crosstalk with immune system in obesity-related inflammation. *J Cell Physiol*. 2018;233:88-97.
- [89] Wade CE, Mora AG, Shields BA, Pidcoke HF, Baer LA, Chung KK, et al. Signals from fat after injury: plasma adipokines and ghrelin concentrations in the severely burned. *Cytokine*. 2013;61:78-83.
- [90] Jernas M, Olsson B, Sjöholm K, Sjogren A, Rudemo M, Nellgard B, et al. Changes in adipose tissue gene expression and plasma levels of adipokines and acute-phase proteins in patients with critical illness. *Metabolism: clinical and experimental*. 2009;58:102-8.
- [91] Welters ID, Bing C, Ding C, Leuwer M, Hall AM. Circulating anti-inflammatory adipokines High Molecular Weight Adiponectin and Zinc- α 2-glycoprotein (ZAG) are inhibited in early sepsis, but increase with clinical recovery: a pilot study. *BMC anesthesiology*. 2014;14.
- [92] Venkatesh B, Hickman I, Nisbet J, Cohen J, Prins J. Changes in serum adiponectin concentrations in critical illness: a preliminary investigation. *Critical care (London, England)*. 2009;13:R105.
- [93] Sharma A, Muddana V, Lamb J, Greer J, Papachristou GI, Whitcomb DC. Low serum adiponectin levels are associated with systemic organ failure in acute pancreatitis. *Pancreas*. 2009;38:907-12.
- [94] Langouche L, Vander Perre S, Frystyk J, Flyvbjerg A, Hansen TK, Van den Berghe G. Adiponectin, retinol-binding protein 4, and leptin in protracted critical illness of pulmonary origin. *Critical care (London, England)*. 2009;13:R112.
- [95] Hillenbrand A, Knippschild U, Weiss M, Schrezenmeier H, Henne-Bruns D, Huber-Lang M, et al. Sepsis induced changes of adipokines and cytokines - septic patients compared to morbidly obese patients. *BMC surgery*. 2010;10:26.

- [96] Robinson K, Jones M, Ordonez J, Grice J, Davidson B, Prins J, et al. Random measurements of adiponectin and IL-6 may not be indicative of the 24-h profile in critically ill patients. *Clinical endocrinology*. 2013;79:892-8.
- [97] Koch A, Sanson E, Voigt S, Helm A, Trautwein C, Tacke F. Serum adiponectin upon admission to the intensive care unit may predict mortality in critically ill patients. *Journal of critical care*. 2011;26:166-74.
- [98] Yu P, Wang S, Qiu Z, Bai B, Zhao Z, Hao Y, et al. Efficacy of resistin and leptin in predicting persistent organ failure in patients with acute pancreatitis. *Pancreatology : official journal of the International Association of Pancreatology (IAP)* [et al]. 2016;16:952-7.
- [99] Vassiliadi DA, Tzanela M, Kotanidou A, Orfanos SE, Nikitas N, Armaganidis A, et al. Serial changes in adiponectin and resistin in critically ill patients with sepsis: associations with sepsis phase, severity, and circulating cytokine levels. *Journal of critical care*. 2012;27:400-9.
- [100] Karampela I, Kandri E, Antonakos G, Vogiatzakis E, Christodoulatos GS, Nikolaidou A, et al. Kinetics of circulating fetuin-A may predict mortality independently from adiponectin, high molecular weight adiponectin and prognostic factors in critically ill patients with sepsis: A prospective study. *Journal of critical care*. 2017;41:78-85.
- [101] Walkey AJ, Rice TW, Konter J, Ouchi N, Shibata R, Walsh K, et al. Plasma adiponectin and mortality in critically ill subjects with acute respiratory failure. *Crit Care Med*. 2010;38:2329-34.
- [102] Walkey AJ, Demissie S, Shah D, Romero F, Puklin L, Summer RS. Plasma Adiponectin, clinical factors, and patient outcomes during the acute respiratory distress syndrome. *PloS one*. 2014;9:e108561.
- [103] Palakshappa JA, Anderson BJ, Reilly JP, Shashaty MG, Ueno R, Wu Q, et al. Low Plasma Levels of Adiponectin Do Not Explain Acute Respiratory Distress Syndrome Risk: a Prospective Cohort Study of Patients with Severe Sepsis. *Critical care (London, England)*. 2016;20:71.
- [104] Santacruz CA, Quintairos A, Righy C, Crippa IA, Couto L, Jr., Imbault V, et al. Is There a Role for Enterohormones in the Gastroparesis of Critically Ill Patients? *Crit Care Med*. 2017;45:1696-701.
- [105] Bornstein SR, Licinio J, Tauchnitz R, Engelmann L, Negrao AB, Gold P, et al. Plasma leptin levels are increased in survivors of acute sepsis: associated loss of diurnal rhythm, in cortisol and leptin secretion. *The Journal of clinical endocrinology and metabolism*. 1998;83:280-3.
- [106] Arnalich F, Lopez J, Codoceo R, Jimenez M, Madero R, Montiel C. Relationship of plasma leptin to plasma cytokines and human survival in sepsis and septic shock. *J Infect Dis*. 1999;180:908-11.
- [107] Tzanela M, Orfanos SE, Tsirantonaki M, Kotanidou A, Sotiropoulou C, Christophoraki M, et al. Leptin alterations in the course of sepsis in humans. *In vivo (Athens, Greece)*. 2006;20:565-70.
- [108] Kythreotis P, Kokkini A, Avgeropoulou S, Hadjioannou A, Anastasakou E, Rasidakis A, et al. Plasma leptin and insulin-like growth factor I levels during acute exacerbations of chronic obstructive pulmonary disease. *BMC pulmonary medicine*. 2009;9:11.

- [109] Papathanassoglou ED, Moynihan JA, Ackerman MH, Mantzoros CS. Serum leptin levels are higher but are not independently associated with severity or mortality in the multiple organ dysfunction/systemic inflammatory response syndrome: a matched case control and a longitudinal study. *Clinical endocrinology*. 2001;54:225-33.
- [110] Yousef AA, Amr YM, Suliman GA. The diagnostic value of serum leptin monitoring and its correlation with tumor necrosis factor-alpha in critically ill patients: a prospective observational study. *Critical care (London, England)*. 2010;14:R33.
- [111] Schaffler A, Landfried K, Volk M, Furst A, Buchler C, Scholmerich J, et al. Potential of adipocytokines in predicting peripancreatic necrosis and severity in acute pancreatitis: pilot study. *Journal of gastroenterology and hepatology*. 2007;22:326-34.
- [112] Koch A, Weiskirchen R, Zimmermann HW, Sanson E, Trautwein C, Tacke F. Relevance of serum leptin and leptin-receptor concentrations in critically ill patients. *Mediators of inflammation*. 2010;2010.
- [113] Shapiro NI, Khankin EV, Van Meurs M, Shih SC, Lu S, Yano M, et al. Leptin exacerbates sepsis-mediated morbidity and mortality. *J Immunol*. 2010;185:517-24.
- [114] Quasim T, McMillan DC, Wallace AM, Kinsella J. The relationship between leptin concentrations, the systemic inflammatory response and illness severity in surgical patients admitted to ITU. *Clinical nutrition (Edinburgh, Scotland)*. 2004;23:233-8.
- [115] Sunden-Cullberg J, Nystrom T, Lee ML, Mullins GE, Tokics L, Andersson J, et al. Pronounced elevation of resistin correlates with severity of disease in severe sepsis and septic shock. *Crit Care Med*. 2007;35:1536-42.
- [116] Koch A, Gressner OA, Sanson E, Tacke F, Trautwein C. Serum resistin levels in critically ill patients are associated with inflammation, organ dysfunction and metabolism and may predict survival of non-septic patients. *Critical care (London, England)*. 2009;13:R95.
- [117] Dong XQ, Yang SB, Zhu FL, Lv QW, Zhang GH, Huang HB. Resistin is associated with mortality in patients with traumatic brain injury. *Critical care (London, England)*. 2010;14:R190.
- [118] Macdonald SP, Stone SF, Neil CL, van Eeden PE, Fatovich DM, Arendts G, et al. Sustained elevation of resistin, NGAL and IL-8 are associated with severe sepsis/septic shock in the emergency department. *PloS one*. 2014;9:e110678.
- [119] Schaffler A, Hamer O, Dickopf J, Goetz A, Landfried K, Voelk M, et al. Admission resistin levels predict peripancreatic necrosis and clinical severity in acute pancreatitis. *The American journal of gastroenterology*. 2010;105:2474-84.
- [120] Duffy SL, Lagrone L, Herndon DN, Mileski WJ. Resistin and postburn insulin dysfunction. *J Trauma*. 2009;66:250-4.
- [121] Lu LF, Yang SS, Wang CP, Hung WC, Yu TH, Chiu CA, et al. Elevated visfatin/pre-B-cell colony-enhancing factor plasma concentration in ischemic stroke. *Journal of stroke and cerebrovascular diseases : the official journal of National Stroke Association*. 2009;18:354-9.
- [122] Lee KA, Gong MN. Pre-B-cell colony-enhancing factor and its clinical correlates with acute lung injury and sepsis. *Chest*. 2011;140:382-90.
- [123] Chen J, Weng J-F, Hong W-C, Luo L-F, Yu W, Luo S-D. Change in plasma visfatin level after severe traumatic brain injury. *Peptides*. 2012;38:8-12.

- [124] Yin CG, Jiang L, Tang B, Zhang H, Qian Q, Niu GZ. Prognostic significance of plasma visfatin levels in patients with ischemic stroke. *Peptides*. 2013;42:101-4.
- [125] Huang Q, Dai WM, Jie YQ, Yu GF, Fan XF, Wu A. High concentrations of visfatin in the peripheral blood of patients with acute basal ganglia hemorrhage are associated with poor outcome. *Peptides*. 2013;39:55-8.
- [126] Lee K, Huh JW, Lim CM, Koh Y, Hong SB. Clinical role of serum pre-B cell colony-enhancing factor in ventilated patients with sepsis and acute respiratory distress syndrome. *Scandinavian journal of infectious diseases*. 2013;45:760-5.
- [127] Schaffler A, Hamer OW, Dickopf J, Goetz A, Landfried K, Voelk M, et al. Admission visfatin levels predict pancreatic and peripancreatic necrosis in acute pancreatitis and correlate with clinical severity. *The American journal of gastroenterology*. 2011;106:957-67.
- [128] Hajri T, Gharib M, Kaul S, Karpel MS, Jr. Association between adipokines and critical illness outcomes. *The journal of trauma and acute care surgery*. 2017;83:507-19.
- [129] Jackson PC, Hardwicke J, Bamford A, Nightingale P, Wilson Y, Papini R, et al. Revised estimates of mortality from the Birmingham Burn Centre, 2001-2010: a continuing analysis over 65 years. *Ann Surg*. 2014;259:979-84.
- [130] Wearn C, Hardwicke J, Kitsios A, Siddons V, Nightingale P, Moiemmen N. Outcomes of burns in the elderly: revised estimates from the Birmingham Burn Centre. *Burns*. 2015;41:1161-8.
- [131] Zimmerman JE, Kramer AA, Knaus WA. Changes in hospital mortality for United States intensive care unit admissions from 1988 to 2012. *Critical care (London, England)*. 2013;17:R81.
- [132] Loss SH, Nunes DSL, Franzosi OS, Salazar GS, Teixeira C, Vieira SRR. Chronic critical illness: are we saving patients or creating victims? *Revista Brasileira de Terapia Intensiva*. 2017;29:87-95.
- [133] Kahn JM, Le T, Angus DC, Cox CE, Hough CL, White DB, et al. The epidemiology of chronic critical illness in the United States*. *Crit Care Med*. 2015;43:282-7.
- [134] Marchioni A, Fantini R, Antenora F, Clini E, Fabbri L. Chronic critical illness: the price of survival. *European journal of clinical investigation*. 2015;45:1341-9.
- [135] Lee KC, Dretzke J, Grover L, Logan A, Moiemmen N. A systematic review of objective burn scar measurements. *Burns & trauma*. 2016;4:14.

Highlights

- The Obesity Paradox has been reported in critically ill populations.
- Burn injury and critical illness affect adipose tissue morphologically and functionally.
- Adipokines exert anti- and pro-inflammatory effects influencing patient outcomes.
- Current validated medical treatments utilize adipose tissue and potentially adipokines.

Potential Role of Adipose Tissue and its Hormones in Burns and Critically Ill Patients

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Abstract

Obesity has become a world-wide pandemic and is considered a major risk factor for various diseases. Despite this, recent intriguing clinical observations have been made suggesting that being overweight has some advantages. Overweight and some obese patients were reported to have significantly lower all-cause mortality, described as the 'obesity paradox'. This phenomenon resulted in increased research aimed at investigating the influence of adipose tissue on outcomes of various clinical states including critical illness. In this review, we summarise research findings on the effect burn injury and trauma-related critical illness have on adipose tissue and discuss potential mechanisms by which adipose tissue influences outcomes in burn and other critically ill patients. Burn injury and critical illness influence adipose tissue functionally and morphologically, with circulating levels of fat derived hormones, adipokines, altered in patients following injury and/or critical illness. As adipokines regulate a variety of processes including inflammation and metabolism, this disruption in the adipokine axis may explain the obesity paradox phenomenon observed in critically ill patients. We conclude that further research on the influence of individual adipokines on prognosis in burn and critically ill patients and the mechanisms involved is required to increase understanding of their therapeutic potential.

Highlights

- The Obesity Paradox has been reported in critically ill populations.
- Burn injury and critical illness affect adipose tissue morphologically and functionally.
- Adipokines exert anti- and pro-inflammatory effects influencing patient outcomes.
- Current scar reduction treatments utilising adipose tissue potentially mediate their effects through release of adipokines.

Keywords

Burns, Thermal Injury, Adipose Tissue, Fat, Hormones, Adipokines

Abbreviations

APACHE	Acute Physiological Assessment and Chronic Health Evaluation
CRP	C-Reactive Protein
FFA	Free Fatty Acids
HMG-CoA	Hydroxymethyl Glutaryl Coenzyme A
IL	Interleukin
RCT	Randomised Controlled Trial
SAPS	Simplified Acute Physiology Score
SOFA	Sequential Organ Failure Assessment

Introduction

Obesity is a complex multifactorial condition that affects over a third of the world's population [1]. With increasing prevalence of overweight and obese individuals [2, 3], obesity is being described as a global pandemic [4] as obesity greatly impacts the individual's health status and quality of life [5, 6] being a major risk factor for various pathologies including cancer, cardiovascular disease, diabetes and osteoarthritis [7].

In this context a recent and intriguing observation is that all-cause mortality is reported to be significantly lower in overweight and some obese patients [8]. This phenomenon, where outcomes are paradoxically better in overweight and obese patients compared to normal weight individuals, is described as the 'obesity paradox' and is the subject of increasing interest in scientific and medical communities [9-12]. The underlying mechanisms behind this phenomenon remain poorly understood and this is particularly the case in critically ill populations where the data on the obesity paradox are limited.

This review aims to summarise the observations suggesting the presence of the obesity paradox in critically ill patients, including burns and to discuss potential mechanisms that may explain the difference in outcomes between normal weight, overweight and obese patients, focusing primarily on hormones secreted by adipose tissue (adipokines).

Adipose Tissue in Burns and Critical Illness

Adipose tissue is one of the largest organs in the human body. Importantly, it is no longer deemed an inert tissue that serves the roles of thermal/mechanical insulation protecting internal organs from external stimuli (such as cold and shock) or as an energy storage modality. Since the discovery in 1994 of leptin, an adipokine or adipose-derived hormone capable of controlling body energy balance [13], adipose tissue is now recognised as endocrine organ able to influence metabolism and inflammatory status. As a result extensive research has been carried out investigating potential roles of adipokines in various clinical conditions including autoimmune and inflammatory disorders and connective tissue diseases [14], metabolic disorders [15, 16], cardiovascular and neurovascular diseases [17], and cancer [18, 19].

Despite increasing interest in adipose tissues' role in clinical pathologies, its role in the context of critical illness including burns and trauma remains to be fully elucidated. With thousands of critically ill patients admitted to intensive care units every year [20] some interesting observations have been made. Patients requiring prolonged critical care were reported to lose lean body mass while adipose tissue mass remained preserved or even increased [21, 22]. Furthermore, although morbid obesity (BMI ≥ 40 kg/m²) is an independent risk factor for mortality in critically ill patients [23], improved survival rates were observed among overweight (BMI 25-30 kg/m²) and obese (BMI 30-40 kg/m²) patients compared to normal BMI patients during critical illness [23-26]. These paradoxical findings have stimulated research in to the interplay between critical illness and adipose tissue and their influence on patient outcomes. Moreover the profound

inflammatory and metabolic response to burn and trauma related critical illness suggest a potential involvement for adipose tissue and adipokines.

Critical illness following injury is a multifactorial heterogeneous disorder characterised by an overwhelming pro-inflammatory response accompanied by a compensatory anti-inflammatory reaction and subsequent immunosuppression [27, 28]. This classical paradigm also applies to severe forms of critical illness such as burns, the pathology of which we have described previously [29, 30]. The human response to burn injury includes a so-called 'genomic storm'[31], consistent with simultaneous increased systemic inflammation, innate immune activation and anti-inflammatory response [32, 33], as well as suppression of adaptive immunity[31]. In addition, burn patients and others with severe critical illness suffer from a prolonged hypermetabolic, hypercatabolic response [33, 34].

The metabolic response following thermal injury is characterised as a two phase response: the 'ebb' phase within 48 hours where metabolism, cardiac output and oxygen consumption are reduced, followed by the 'flow' phase at approximately 120 hours post-injury where these parameters increase and plateau [35]. This metabolic response includes: peripheral lipolysis and free fatty acid (FFA) [36] oxidation leading to an acute, global and complex increase in FFA levels[37]; systemic induction of endoplasmic reticulum (ER) stress and unfolded protein response [38]; up to 6-fold increase in breakdown rates of skeletal muscle protein[39]; elevation in resting energy expenditure up to 140%[40] that can be prolonged [33].

Burns and other severe critical illnesses have been reported to influence adipose tissue morphologically and functionally. Saraf *et al* reported the impact of severe burn injury on subcutaneous white adipose tissue in children and observed significantly reduced adipocyte size, increased collagen deposition and cell mitochondria content, increased immune cells such as macrophages, as well as increased inflammatory cytokine production [41]. These morphological changes suggest "browning" of subcutaneous adipose tissue following thermal injury, a finding which was confirmed biochemically and functionally. Sidossis *et al* reported significantly increased mitochondrial density and mitochondrial respiratory capacity, as well as an 80-fold increase in the expression of uncoupling protein 1 (UCP1), a molecule abundantly observed in brown adipose tissue depots [42], in burn patients compared to healthy controls [43]. In addition, Patsouris *et al* reported similar findings including significantly increased mitochondrial mass and adipose tissue browning markers in burn patients [44]. This could be a compensatory mechanism since brown adipose tissue is known to induce thermogenesis, modulate energy expenditure and exert local tissue effects such as stimulating angiogenesis and influencing macrophage polarization [45]. Similar morphological and metabolic activity alterations of adipose tissue have been reported in critically ill patients[46, 47]. A functional aspect of adipose tissue is its endocrine role through the production of adipokines and these may mediate many of the effects seen in burns and critical illness.

Overview of Adipokines and their Biological Effects

There are approximately 600 identified hormones secreted by adipose tissue [48], providing a rich source of potential novel biomarkers and therapeutic targets for the management of various pathologies. In this review, we will focus on Adiponectin, Ghrelin, Leptin, Resistin and Visfatin as the best characterised adipokines.

Adiponectin is released exclusively from white adipose tissue[49], and is the most abundant adipose-specific adipokine, with expression in subcutaneous fat being greater than visceral fat [50]. Adiponectin has anti-inflammatory effects [51]. Ghrelin is an orexigenic hormone that is an endogenous ligand to growth hormone and was initially thought to be produced mainly by the stomach [52], but has subsequently been identified in other tissues including adipose tissue [53]. Ghrelin signaling is associated with adiposity, changes in fat distribution and mobilisation, independent of growth hormone and dietary intake[54, 55]. Leptin is primarily secreted by subcutaneous white adipose tissue, the amount of leptin secreted into the circulation is proportional to adipose tissue mass and nutritional status[51]. Leptin exhibits structural similarities to cytokines [56] and is pro-inflammatory [57]. Resistin is also a pro-inflammatory adipokine expressed by adipocytes and other tissues including skeletal muscle [58, 59]. Visfatin, also called pre-B-cell colony enhancing factor, is primarily secreted by adipocytes in visceral white adipose tissue and exhibits pro-inflammatory effects [60].

Relevant to this review, adipokines have been reported to influence skin and adipose tissue. Adiponectin and ghrelin have been observed to exert anti-inflammatory and anti-fibrotic effects on skin [61-63] and were reported to enhance wound healing rates [64, 65]. Similarly, leptin has been observed to enhance human epidermal keratinocyte and epithelial cell proliferation, differentiation and migration, as well as promote angiogenesis within dermal connective tissues [66]. However, leptin was also found to be overexpressed in hypertrophic and keloid scars [67]. This could be due to increased pro-inflammatory cytokine release associated with leptin, as seen in inflammatory skin conditions [68]. Visfatin has been reported to enhance chemokine and antimicrobial peptide production in human keratinocytes [69, 70], as well as exhibit anti-fibrotic properties[71].

Adiponectin and leptin have been reported to induce browning of adipose tissue [72, 73] and adiponectin promotes adipogenesis as well as increasing lipid accumulation and insulin responsiveness of adipocytes [74]. In contrast, leptin inhibits insulin-dependent glucose uptake and lipogenesis and reverses insulin-induced lipolysis [75]. Ghrelin stimulates adipogenesis and glucose uptake, as well as inhibiting lipolysis, apoptosis and autophagy of adipocytes [76, 77]. Resistin and visfatin enhance pro-inflammatory cytokine expression in adipose tissue including TNF- α and IL-6 [78, 79]. Similarly, resistin and visfatin induce insulin resistance in adipocytes [79, 80]. The influence of these adipokines is not limited to skin and adipose tissue. The beneficial and detrimental effects of these adipokines on various cell types and tissues are summarized in Figure 1 and the reader is also referred to recent reviews for further detail [17, 81-88].

Figure 1. Effects of adipokines on various tissues and organs

Adipokine changes in burns and critically ill patients

Several studies have demonstrated acute reductions in circulating adiponectin levels in critical illness and/or injury including burns, sepsis and trauma [89-95]. In addition, an inverse association was reported between serum adiponectin levels and severity of illness as measured by C-Reactive Protein (CRP), Simplified Acute Physiology Score (SAPS) II and Sequential Organ Failure Assessment (SOFA) scores [91, 92, 95]. Similar findings were observed in patients with acute pancreatitis, where adiponectin levels in the blood were negatively associated with severity of disease and incidence of tissue necrosis [93]. Furthermore, adiponectin levels progressively increase with patient recovery [91, 94]. Although the above findings indicate that decreased serum adiponectin levels may lead to poor outcomes, other research has reported different findings. Circulating adiponectin levels in severely ill patients did not correlate with inflammatory markers including Interleukin (IL) -6, IL-10 and Tumour Necrosis Factor (TNF)- α [89, 96, 97] and clinical scores including Acute Physiologic Assessment and Chronic Health Evaluation (APACHE) II score and SOFA [96, 98, 99]. Furthermore, higher blood adiponectin levels were associated with increased risk of mortality during critical illness [100-103].

Only two studies have investigated circulating ghrelin levels in critical illness. Wade *et al* reported significantly reduced ghrelin levels in severely burned patients correlating with metabolic/caloric needs. No other associations with other parameters such as injury severity and inflammatory status were observed [89]. Santacruz *et al* also observed significantly reduced plasma ghrelin levels in critically ill patients but saw no correlations with feeding status [104].

Leptin levels in the blood have been reported to be elevated in critical illness [105-108]. Furthermore, leptin was positively associated with pro-inflammatory status of severely ill patients, as measured by CRP, IL-6, sTNFR1 and TNF- α [106, 108-111]. Additionally, other studies have reported that serum-soluble leptin receptor (SLR) in patients correlated with inflammatory response and illness severity as measured by IL-6, lactate, procalcitonin and APACHE II score [112, 113]. Interestingly, elevated levels of leptin were observed in survivors of acute sepsis [105], while increased SLR levels in critically ill patients were associated with increased mortality [112]. However, other studies have reported different findings. Blood leptin levels in severely ill patients were similar or reduced compared to healthy volunteers [89, 94, 95, 109, 112, 114] and no associations were found between circulating leptin levels and inflammatory status, illness severity, or mortality in critical illness [94, 95, 107, 109, 111, 112, 114].

In contrast to the heterogeneity of results reported on the impact adiponectin, ghrelin and leptin on critical illness outcomes, the influence of resistin and visfatin on outcomes

of severely ill patients is consistent in the literature. Critically ill patients exhibit significantly elevated circulating levels of resistin [89, 95, 98, 99, 111, 115-120] and visfatin [111, 121-127]. Additionally, both resistin and visfatin significantly correlated with pro-inflammatory responses (including CRP, IL-6, IL-8 and TNF- α), and worse clinical severity scores (including APACHE II, Glasgow Coma score, multiple organ dysfunction score, SAPS II and SOFA)[89, 95, 98, 99, 111, 115-119, 121-127]. Furthermore, high resistin and visfatin levels in blood were associated with poor outcomes including mortality [116, 117, 122-126].

A systematic review examining the evidence for adipokines having an influence on critical care patients has been published recently [128]. It concludes that although strong observations were reported indicating the influence of adipokines on the prognosis of critical illness, additional larger studies that incorporate more diverse cohorts (such as age, gender, BMI, ethnic groups and different pathologies) is required to better understand the relationship between adipokines and critical illness. This is essential in order to validate the potential clinical value and utility of adipokines as diagnostic and/or prognostic biomarkers, as well their potential as therapeutic targets in critical illness including burn and trauma. Furthermore, studies to date have investigated the association of adipokines with critical illness in the acute setting only. This focus on the acute setting has further limited the translation of adipokines in clinical settings. Importantly, since medical care advancements have improved survival rates after critical trauma [129-131], greater emphasis is now placed on the prevention and treatment of potentially debilitating long-term sequelae experienced by survivors of severe illness including chronic critical illness [132-134], prolonged pathophysiological responses[33] and scarring [135].

Conclusions

Several studies have reported changes in the serum levels of specific adipokines and their role in the regulation of a range of biological responses to injury including inflammation, metabolic dysregulation and wound healing is emerging. However a robust characterisation of the impact of such changes in individual adipokines on patient outcomes, especially in burns patients, is lacking. Large clinical and scientific studies are required to establish the mechanisms by which adipose tissue may influence patient outcomes and translate the research into clinical practice to improve short and long-term outcomes of burn and critically ill patients.

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References

- [1] Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980-2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*. 2014;384:766-81.
- [2] Trends in adult body-mass index in 200 countries from 1975 to 2014: a pooled analysis of 1698 population-based measurement studies with 19.2 million participants. *Lancet*. 2016;387:1377-96.
- [3] Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: a pooled analysis of 2416 population-based measurement studies in 128.9 million children, adolescents, and adults. *Lancet*. 2017;390:2627-42.
- [4] Swinburn BA, Sacks G, Hall KD, McPherson K, Finegood DT, Moodie ML, et al. The global obesity pandemic: shaped by global drivers and local environments. *Lancet*. 2011;378:804-14.
- [5] Visscher TL, Seidell JC. The public health impact of obesity. *Annual review of public health*. 2001;22:355-75.
- [6] Taylor VH, Forhan M, Vigod SN, McIntyre RS, Morrison KM. The impact of obesity on quality of life. *Best practice & research Clinical endocrinology & metabolism*. 2013;27:139-46.
- [7] Seidell JC, Halberstadt J. The global burden of obesity and the challenges of prevention. *Annals of nutrition & metabolism*. 2015;66 Suppl 2:7-12.
- [8] Flegal KM, Kit BK, Orpana H, Graubard BI. Association of all-cause mortality with overweight and obesity using standard body mass index categories: a systematic review and meta-analysis. *Jama*. 2013;309:71-82.
- [9] Braun N, Gomes F, Schutz P. "The obesity paradox" in disease--is the protective effect of obesity true? *Swiss medical weekly*. 2015;145:w14265.
- [10] Park J, Ahmadi SF, Streja E, Molnar MZ, Flegal KM, Gillen D, et al. Obesity paradox in end-stage kidney disease patients. *Progress in cardiovascular diseases*. 2014;56:415-25.
- [11] Lavie CJ, McAuley PA, Church TS, Milani RV, Blair SN. Obesity and cardiovascular diseases: implications regarding fitness, fatness, and severity in the obesity paradox. *Journal of the American College of Cardiology*. 2014;63:1345-54.
- [12] Valentijn TM, Galal W, Tjeertes EK, Hoeks SE, Verhagen HJ, Stolker RJ. The obesity paradox in the surgical population. *The surgeon : journal of the Royal Colleges of Surgeons of Edinburgh and Ireland*. 2013;11:169-76.
- [13] Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature*. 1994;372:425-32.
- [14] Fietta P, Delsante G. Focus on adipokines. *Theoretical biology forum*. 2013;106:103-29.
- [15] Jialal I, Devaraj S. Subcutaneous adipose tissue biology in metabolic syndrome. *Hormone molecular biology and clinical investigation*. 2018;33.
- [16] Nicholson T, Church C, Baker DJ, Jones SW. The role of adipokines in skeletal muscle inflammation and insulin sensitivity. *Journal of inflammation (London, England)*. 2018;15:9.

- [17] Opatrilova R, Caprnda M, Kubatka P, Valentova V, Uramova S, Nosal V, et al. Adipokines in neurovascular diseases. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. 2018;98:424-32.
- [18] Lengyel E, Makowski L, DiGiovanni J, Kolonin MG. Cancer as a Matter of Fat: The Crosstalk between Adipose Tissue and Tumors. *Trends in cancer*. 2018;4:374-84.
- [19] Morris EV, Edwards CM. Adipokines, adiposity, and bone marrow adipocytes: Dangerous accomplices in multiple myeloma. *J Cell Physiol*. 2018.
- [20] Mullins PM, Goyal M, Pines JM. National growth in intensive care unit admissions from emergency departments in the United States from 2002 to 2009. *Academic emergency medicine : official journal of the Society for Academic Emergency Medicine*. 2013;20:479-86.
- [21] Hart DW, Wolf SE, Herndon DN, Chinkes DL, Lal SO, Obeng MK, et al. Energy expenditure and caloric balance after burn: increased feeding leads to fat rather than lean mass accretion. *Ann Surg*. 2002;235:152-61.
- [22] Plank LD, Connolly AB, Hill GL. Sequential changes in the metabolic response in severely septic patients during the first 23 days after the onset of peritonitis. *Ann Surg*. 1998;228:146-58.
- [23] Nasraway SA, Jr., Albert M, Donnelly AM, Ruthazer R, Shikora SA, Saltzman E. Morbid obesity is an independent determinant of death among surgical critically ill patients. *Crit Care Med*. 2006;34:964-70; quiz 71.
- [24] Fonarow GC, Srikanthan P, Costanzo MR, Cintron GB, Lopatin M. An obesity paradox in acute heart failure: analysis of body mass index and inhospital mortality for 108,927 patients in the Acute Decompensated Heart Failure National Registry. *American heart journal*. 2007;153:74-81.
- [25] Peake SL, Moran JL, Ghelani DR, Lloyd AJ, Walker MJ. The effect of obesity on 12-month survival following admission to intensive care: a prospective study. *Crit Care Med*. 2006;34:2929-39.
- [26] Trivedi V, Jean RE, Genese F, Fuhrmann KA, Saini AK, Mangulabnan VD, et al. Impact of Obesity on Outcomes in a Multiethnic Cohort of Medical Intensive Care Unit Patients. *Journal of intensive care medicine*. 2018;33:97-103.
- [27] Greathouse KC, Hall MW. Critical Illness-Induced Immune Suppression: Current State of the Science. *American journal of critical care : an official publication, American Association of Critical-Care Nurses*. 2016;25:85-92.
- [28] Vanzant EL, Lopez CM, Ozrazgat-Baslanti T, Ungaro R, Davis R, Cuenca AG, et al. Persistent inflammation, immunosuppression, and catabolism syndrome after severe blunt trauma. *The journal of trauma and acute care surgery*. 2014;76:21-9; discussion 9-30.
- [29] Al-Tarrach K, Hewison M, Moiemien N, Lord JM. Vitamin D status and its influence on outcomes following major burn injury and critical illness. *Burns & trauma*. 2018;6:11.
- [30] Al-Tarrach K, Moiemien N, Lord JM. The influence of sex steroid hormones on the response to trauma and burn injury. *Burns & trauma*. 2017;5:29.
- [31] Xiao W, Mindrinis MN, Seok J, Cuschieri J, Cuenca AG, Gao H, et al. A genomic storm in critically injured humans. *J Exp Med*. 2011;208:2581-90.
- [32] Hampson P, Dinsdale RJ, Wearn CM, Bamford AL, Bishop JRB, Hazeldine J, et al. Neutrophil Dysfunction, Immature Granulocytes, and Cell-free DNA are Early

- Biomarkers of Sepsis in Burn-injured Patients: A Prospective Observational Cohort Study. *Ann Surg.* 2017;265:1241-9.
- [33] Jeschke MG, Gauglitz GG, Kulp GA, Finnerty CC, Williams FN, Kraft R, et al. Long-term persistence of the pathophysiologic response to severe burn injury. *PloS one.* 2011;6:e21245.
- [34] De Cosmi V, Milani GP, Mazzocchi A, D'Oria V, Silano M, Calderini E, et al. The Metabolic Response to Stress and Infection in Critically Ill Children: The Opportunity of an Individualized Approach. *Nutrients.* 2017;9.
- [35] Wolfe RR. Review: acute versus chronic response to burn injury. *Circ Shock.* 1981;8:105-15.
- [36] Otterbein LR, Cosio C, Graceffa P, Dominguez R. Crystal structures of the vitamin D-binding protein and its complex with actin: structural basis of the actin-scavenger system. *Proc Natl Acad Sci U S A.* 2002;99:8003-8.
- [37] Qi P, Abdullahi A, Stanojic M, Patsouris D, Jeschke MG. Lipidomic analysis enables prediction of clinical outcomes in burn patients. *Scientific reports.* 2016;6:38707.
- [38] Jeschke MG, Finnerty CC, Herndon DN, Song J, Boehning D, Tompkins RG, et al. Severe Injury Is Associated With Insulin Resistance, Endoplasmic Reticulum Stress Response, and Unfolded Protein Response. *Ann Surg.* 2012;255:370-8.
- [39] Chao T, Herndon DN, Porter C, Chondronikola M, Chaidemenou A, Abdelrahman DR, et al. Skeletal Muscle Protein Breakdown Remains Elevated in Pediatric Burn Survivors up to One-Year Post-Injury. *Shock.* 2015;44:397-401.
- [40] Jeschke MG, Chinkes DL, Finnerty CC, Kulp G, Suman OE, Norbury WB, et al. Pathophysiologic response to severe burn injury. *Ann Surg.* 2008;248:387-401.
- [41] Saraf MK, Herndon DN, Porter C, Toliver-Kinsky T, Radhakrishnan R, Chao T, et al. Morphological Changes in Subcutaneous White Adipose Tissue After Severe Burn Injury. *Journal of burn care & research : official publication of the American Burn Association.* 2016;37:e96-103.
- [42] Kalinovich AV, de Jong JM, Cannon B, Nedergaard J. UCP1 in adipose tissues: two steps to full browning. *Biochimie.* 2017;134:127-37.
- [43] Sidossis LS, Porter C, Saraf MK, Borsheim E, Radhakrishnan RS, Chao T, et al. Browning of Subcutaneous White Adipose Tissue in Humans after Severe Adrenergic Stress. *Cell metabolism.* 2015;22:219-27.
- [44] Patsouris D. Burn Induces Browning of the Subcutaneous White Adipose Tissue in Mice and Humans. 2015;13:1538-44.
- [45] Wang GX, Zhao XY, Lin JD. The brown fat secretome: metabolic functions beyond thermogenesis. *Trends in endocrinology and metabolism: TEM.* 2015;26:231-7.
- [46] Langouche L, Perre SV, Thiessen S, Gunst J, Hermans G, D'Hoore A, et al. Alterations in adipose tissue during critical illness: An adaptive and protective response? *American journal of respiratory and critical care medicine.* 2010;182:507-16.
- [47] Marques MB, Langouche L. Endocrine, metabolic, and morphologic alterations of adipose tissue during critical illness. *Crit Care Med.* 2013;41:317-25.
- [48] Lehr S, Hartwig S, Sell H. Adipokines: a treasure trove for the discovery of biomarkers for metabolic disorders. *Proteomics Clinical applications.* 2012;6:91-101.

- [49] Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem*. 1995;270:26746-9.
- [50] Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the Release of Adipokines by Adipose Tissue, Adipose Tissue Matrix, and Adipocytes from Visceral and Subcutaneous Abdominal Adipose Tissues of Obese Humans. *Endocrinology*. 2004;145:2273-82.
- [51] Ahima RS. Metabolic actions of adipocyte hormones: focus on adiponectin. *Obesity* (Silver Spring, Md). 2006;14 Suppl 1:9s-15s.
- [52] Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature*. 1999;402:656-60.
- [53] Baatar D, Patel K, Taub DD. The effects of ghrelin on inflammation and the immune system. *Molecular and cellular endocrinology*. 2011;340:44-58.
- [54] Sangiao-Alvarellos S, Vazquez MJ, Varela L, Nogueiras R, Saha AK, Cordido F, et al. Central ghrelin regulates peripheral lipid metabolism in a growth hormone-independent fashion. *Endocrinology*. 2009;150:4562-74.
- [55] Al Massadi O, Lopez M, Tschop M, Dieguez C, Nogueiras R. Current Understanding of the Hypothalamic Ghrelin Pathways Inducing Appetite and Adiposity. *Trends in neurosciences*. 2017;40:167-80.
- [56] Leal VdO, Mafrá D. Adipokines in obesity. *Clinica Chimica Acta*. 2013;419:87-94.
- [57] Fernandez-Riejos P, Najib S, Santos-Alvarez J, Martin-Romero C, Perez-Perez A, Gonzalez-Yanes C, et al. Role of leptin in the activation of immune cells. *Mediators of inflammation*. 2010;2010:568343.
- [58] Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, et al. The hormone resistin links obesity to diabetes. *Nature*. 2001;409:307-12.
- [59] Jamaluddin MS, Weakley SM, Yao Q, Chen C. Resistin: functional roles and therapeutic considerations for cardiovascular disease. *British Journal of Pharmacology*. 2012;165:622-32.
- [60] Sun Z, Lei H, Zhang Z. Pre-B cell colony enhancing factor (PBEF), a cytokine with multiple physiological functions. *Cytokine & growth factor reviews*. 2013;24:433-42.
- [61] Lakota K, Wei J, Carns M, Hinchcliff M, Lee J, Whitfield ML, et al. Levels of adiponectin, a marker for PPAR-gamma activity, correlate with skin fibrosis in systemic sclerosis: potential utility as biomarker? *Arthritis research & therapy*. 2012;14:R102.
- [62] Shibata S, Tada Y, Hau CS, Mitsui A, Kamata M, Asano Y, et al. Adiponectin regulates psoriasiform skin inflammation by suppressing IL-17 production from gammadelta-T cells. *Nature communications*. 2015;6:7687.
- [63] Koca SS, Ozgen M, Sarikaya M, Dagli F, Ustundag B, Isik A. Ghrelin prevents the development of dermal fibrosis in bleomycin-induced scleroderma. *Clinical and experimental dermatology*. 2014;39:176-81.
- [64] Liu C, Huang J, Li H, Yang Z, Zeng Y, Liu J, et al. Ghrelin accelerates wound healing through GHS-R1a-mediated MAPK-NF- κ B/GR signaling pathways in combined radiation and burn injury in rats. *Scientific reports*. 2016;6.
- [65] Shibata S, Tada Y, Asano Y, Hau CS, Kato T, Saeki H, et al. Adiponectin regulates cutaneous wound healing by promoting keratinocyte proliferation and migration via the ERK signaling pathway. *J Immunol*. 2012;189:3231-41.

- [66] Tadokoro S, Ide S, Tokuyama R, Umeki H, Tatehara S, Kataoka S, et al. Leptin Promotes Wound Healing in the Skin. *PloS one*. 2015;10.
- [67] Seleit I, Bakry OA, Samaka RM, Tawfik AS. Immunohistochemical Evaluation of Leptin Expression in Wound Healing: A Clue to Exuberant Scar Formation. *Applied immunohistochemistry & molecular morphology : AIMM*. 2016;24:296-306.
- [68] Johnston A, Arnadottir S, Gudjonsson JE, Aphale A, Sigmarsdottir AA, Gunnarsson SI, et al. Obesity in psoriasis: Leptin and resistin as mediators of cutaneous inflammation. *The British journal of dermatology*. 2008;159:342-50.
- [69] Hau CS, Kanda N, Noda S, Tatsuta A, Kamata M, Shibata S, et al. Visfatin enhances the production of cathelicidin antimicrobial peptide, human beta-defensin-2, human beta-defensin-3, and S100A7 in human keratinocytes and their orthologs in murine imiquimod-induced psoriatic skin. *Am J Pathol*. 2013;182:1705-17.
- [70] Kanda N, Hau CS, Tada Y, Tatsuta A, Sato S, Watanabe S. Visfatin enhances CXCL8, CXCL10, and CCL20 production in human keratinocytes. *Endocrinology*. 2011;152:3155-64.
- [71] Masui Y, Asano Y, Shibata S, Noda S, Akamata K, Aozasa N, et al. A possible contribution of visfatin to the resolution of skin sclerosis in patients with diffuse cutaneous systemic sclerosis via a direct anti-fibrotic effect on dermal fibroblasts and Th1 polarization of the immune response. *Rheumatology (Oxford, England)*. 2013;52:1239-44.
- [72] Hui X, Gu P, Zhang J, Nie T, Pan Y, Wu D, et al. Adiponectin Enhances Cold-Induced Browning of Subcutaneous Adipose Tissue via Promoting M2 Macrophage Proliferation. *Cell metabolism*. 2015;22:279-90.
- [73] Dodd G, Descherf S, Loh K, Simonds SE, Wiede F, Balland E, et al. Leptin and insulin act on POMC neurons to promote the browning of white fat. *Cell*. 2015;160:88-104.
- [74] Fu Y, Luo N, Klein RL, Garvey WT. Adiponectin promotes adipocyte differentiation, insulin sensitivity, and lipid accumulation. *Journal of lipid research*. 2005;46:1369-79.
- [75] Harris RB. Direct and indirect effects of leptin on adipocyte metabolism. *Biochim Biophys Acta*. 2014;1842:414-23.
- [76] Rodriguez A. Novel molecular aspects of ghrelin and leptin in the control of adipobiology and the cardiovascular system. *Obesity facts*. 2014;7:82-95.
- [77] Rodriguez A, Gomez-Ambrosi J, Catalan V, Rotellar F, Valenti V, Silva C, et al. The ghrelin O-acyltransferase-ghrelin system reduces TNF-alpha-induced apoptosis and autophagy in human visceral adipocytes. *Diabetologia*. 2012;55:3038-50.
- [78] Lehrke M, Reilly MP, Millington SC, Iqbal N, Rader DJ, Lazar MA. An inflammatory cascade leading to hyperresistinemia in humans. *PLoS medicine*. 2004;1:e45.
- [79] Chang YC, Chang TJ, Lee WJ, Chuang LM. The relationship of visfatin/pre-B-cell colony-enhancing factor/nicotinamide phosphoribosyltransferase in adipose tissue with inflammation, insulin resistance, and plasma lipids. *Metabolism: clinical and experimental*. 2010;59:93-9.
- [80] Fu Y, Luo L, Luo N, Garvey WT. Proinflammatory cytokine production and insulin sensitivity regulated by overexpression of resistin in 3T3-L1 adipocytes. *Nutrition & metabolism*. 2006;3:28.

- [81] Shibata R, Ouchi N, Ohashi K, Murohara T. The role of adipokines in cardiovascular disease. *Journal of cardiology*. 2017;70:329-34.
- [82] Romacho T, Elsen M, Rohrborn D, Eckel J. Adipose tissue and its role in organ crosstalk. *Acta physiologica (Oxford, England)*. 2014;210:733-53.
- [83] Li F, Li Y, Duan Y, Hu CA, Tang Y, Yin Y. Myokines and adipokines: Involvement in the crosstalk between skeletal muscle and adipose tissue. *Cytokine & growth factor reviews*. 2017;33:73-82.
- [84] Zhu Q, Scherer PE. Immunologic and endocrine functions of adipose tissue: implications for kidney disease. *Nature reviews Nephrology*. 2018;14:105-20.
- [85] Feakins RM. Obesity and metabolic syndrome: pathological effects on the gastrointestinal tract. *Histopathology*. 2016;68:630-40.
- [86] Hawkes CP, Mostoufi-Moab S. Fat-bone interaction within the bone marrow milieu: Impact on hematopoiesis and systemic energy metabolism. *Bone*. 2018.
- [87] Francisco V, Perez T, Pino J, Lopez V, Franco E, Alonso A, et al. Biomechanics, obesity, and osteoarthritis. The role of adipokines: When the levee breaks. *Journal of orthopaedic research : official publication of the Orthopaedic Research Society*. 2018;36:594-604.
- [88] Maurizi G, Della Guardia L, Maurizi A, Poloni A. Adipocytes properties and crosstalk with immune system in obesity-related inflammation. *J Cell Physiol*. 2018;233:88-97.
- [89] Wade CE, Mora AG, Shields BA, Pidcoke HF, Baer LA, Chung KK, et al. Signals from fat after injury: plasma adipokines and ghrelin concentrations in the severely burned. *Cytokine*. 2013;61:78-83.
- [90] Jernas M, Olsson B, Sjöholm K, Sjogren A, Rudemo M, Nellgard B, et al. Changes in adipose tissue gene expression and plasma levels of adipokines and acute-phase proteins in patients with critical illness. *Metabolism: clinical and experimental*. 2009;58:102-8.
- [91] Welters ID, Bing C, Ding C, Leuwer M, Hall AM. Circulating anti-inflammatory adipokines High Molecular Weight Adiponectin and Zinc- α 2-glycoprotein (ZAG) are inhibited in early sepsis, but increase with clinical recovery: a pilot study. *BMC anesthesiology*. 2014;14.
- [92] Venkatesh B, Hickman I, Nisbet J, Cohen J, Prins J. Changes in serum adiponectin concentrations in critical illness: a preliminary investigation. *Critical care (London, England)*. 2009;13:R105.
- [93] Sharma A, Muddana V, Lamb J, Greer J, Papachristou GI, Whitcomb DC. Low serum adiponectin levels are associated with systemic organ failure in acute pancreatitis. *Pancreas*. 2009;38:907-12.
- [94] Langouche L, Vander Perre S, Frystyk J, Flyvbjerg A, Hansen TK, Van den Berghe G. Adiponectin, retinol-binding protein 4, and leptin in protracted critical illness of pulmonary origin. *Critical care (London, England)*. 2009;13:R112.
- [95] Hillenbrand A, Knippschild U, Weiss M, Schrezenmeier H, Henne-Bruns D, Huber-Lang M, et al. Sepsis induced changes of adipokines and cytokines - septic patients compared to morbidly obese patients. *BMC surgery*. 2010;10:26.

- [96] Robinson K, Jones M, Ordonez J, Grice J, Davidson B, Prins J, et al. Random measurements of adiponectin and IL-6 may not be indicative of the 24-h profile in critically ill patients. *Clinical endocrinology*. 2013;79:892-8.
- [97] Koch A, Sanson E, Voigt S, Helm A, Trautwein C, Tacke F. Serum adiponectin upon admission to the intensive care unit may predict mortality in critically ill patients. *Journal of critical care*. 2011;26:166-74.
- [98] Yu P, Wang S, Qiu Z, Bai B, Zhao Z, Hao Y, et al. Efficacy of resistin and leptin in predicting persistent organ failure in patients with acute pancreatitis. *Pancreatology : official journal of the International Association of Pancreatology (IAP)* [et al]. 2016;16:952-7.
- [99] Vassiliadi DA, Tzanela M, Kotanidou A, Orfanos SE, Nikitas N, Armaganidis A, et al. Serial changes in adiponectin and resistin in critically ill patients with sepsis: associations with sepsis phase, severity, and circulating cytokine levels. *Journal of critical care*. 2012;27:400-9.
- [100] Karampela I, Kandri E, Antonakos G, Vogiatzakis E, Christodoulatos GS, Nikolaidou A, et al. Kinetics of circulating fetuin-A may predict mortality independently from adiponectin, high molecular weight adiponectin and prognostic factors in critically ill patients with sepsis: A prospective study. *Journal of critical care*. 2017;41:78-85.
- [101] Walkey AJ, Rice TW, Konter J, Ouchi N, Shibata R, Walsh K, et al. Plasma adiponectin and mortality in critically ill subjects with acute respiratory failure. *Crit Care Med*. 2010;38:2329-34.
- [102] Walkey AJ, Demissie S, Shah D, Romero F, Puklin L, Summer RS. Plasma Adiponectin, clinical factors, and patient outcomes during the acute respiratory distress syndrome. *PloS one*. 2014;9:e108561.
- [103] Palakshappa JA, Anderson BJ, Reilly JP, Shashaty MG, Ueno R, Wu Q, et al. Low Plasma Levels of Adiponectin Do Not Explain Acute Respiratory Distress Syndrome Risk: a Prospective Cohort Study of Patients with Severe Sepsis. *Critical care (London, England)*. 2016;20:71.
- [104] Santacruz CA, Quintairos A, Righy C, Crippa IA, Couto L, Jr., Imbault V, et al. Is There a Role for Enterohormones in the Gastroparesis of Critically Ill Patients? *Crit Care Med*. 2017;45:1696-701.
- [105] Bornstein SR, Licinio J, Tauchnitz R, Engelmann L, Negrao AB, Gold P, et al. Plasma leptin levels are increased in survivors of acute sepsis: associated loss of diurnal rhythm, in cortisol and leptin secretion. *The Journal of clinical endocrinology and metabolism*. 1998;83:280-3.
- [106] Arnalich F, Lopez J, Codoceo R, Jimenez M, Madero R, Montiel C. Relationship of plasma leptin to plasma cytokines and human survival in sepsis and septic shock. *J Infect Dis*. 1999;180:908-11.
- [107] Tzanela M, Orfanos SE, Tsirantonaki M, Kotanidou A, Sotiropoulou C, Christophoraki M, et al. Leptin alterations in the course of sepsis in humans. *In vivo (Athens, Greece)*. 2006;20:565-70.
- [108] Kythreotis P, Kokkini A, Avgeropoulou S, Hadjioannou A, Anastasakou E, Rasidakis A, et al. Plasma leptin and insulin-like growth factor I levels during acute exacerbations of chronic obstructive pulmonary disease. *BMC pulmonary medicine*. 2009;9:11.

- [109] Papathanassoglou ED, Moynihan JA, Ackerman MH, Mantzoros CS. Serum leptin levels are higher but are not independently associated with severity or mortality in the multiple organ dysfunction/systemic inflammatory response syndrome: a matched case control and a longitudinal study. *Clinical endocrinology*. 2001;54:225-33.
- [110] Yousef AA, Amr YM, Suliman GA. The diagnostic value of serum leptin monitoring and its correlation with tumor necrosis factor-alpha in critically ill patients: a prospective observational study. *Critical care (London, England)*. 2010;14:R33.
- [111] Schaffler A, Landfried K, Volk M, Furst A, Buchler C, Scholmerich J, et al. Potential of adipocytokines in predicting peripancreatic necrosis and severity in acute pancreatitis: pilot study. *Journal of gastroenterology and hepatology*. 2007;22:326-34.
- [112] Koch A, Weiskirchen R, Zimmermann HW, Sanson E, Trautwein C, Tacke F. Relevance of serum leptin and leptin-receptor concentrations in critically ill patients. *Mediators of inflammation*. 2010;2010.
- [113] Shapiro NI, Khankin EV, Van Meurs M, Shih SC, Lu S, Yano M, et al. Leptin exacerbates sepsis-mediated morbidity and mortality. *J Immunol*. 2010;185:517-24.
- [114] Quasim T, McMillan DC, Wallace AM, Kinsella J. The relationship between leptin concentrations, the systemic inflammatory response and illness severity in surgical patients admitted to ITU. *Clinical nutrition (Edinburgh, Scotland)*. 2004;23:233-8.
- [115] Sunden-Cullberg J, Nystrom T, Lee ML, Mullins GE, Tokics L, Andersson J, et al. Pronounced elevation of resistin correlates with severity of disease in severe sepsis and septic shock. *Crit Care Med*. 2007;35:1536-42.
- [116] Koch A, Gressner OA, Sanson E, Tacke F, Trautwein C. Serum resistin levels in critically ill patients are associated with inflammation, organ dysfunction and metabolism and may predict survival of non-septic patients. *Critical care (London, England)*. 2009;13:R95.
- [117] Dong XQ, Yang SB, Zhu FL, Lv QW, Zhang GH, Huang HB. Resistin is associated with mortality in patients with traumatic brain injury. *Critical care (London, England)*. 2010;14:R190.
- [118] Macdonald SP, Stone SF, Neil CL, van Eeden PE, Fatovich DM, Arendts G, et al. Sustained elevation of resistin, NGAL and IL-8 are associated with severe sepsis/septic shock in the emergency department. *PloS one*. 2014;9:e110678.
- [119] Schaffler A, Hamer O, Dickopf J, Goetz A, Landfried K, Voelk M, et al. Admission resistin levels predict peripancreatic necrosis and clinical severity in acute pancreatitis. *The American journal of gastroenterology*. 2010;105:2474-84.
- [120] Duffy SL, Lagrone L, Herndon DN, Mileski WJ. Resistin and postburn insulin dysfunction. *J Trauma*. 2009;66:250-4.
- [121] Lu LF, Yang SS, Wang CP, Hung WC, Yu TH, Chiu CA, et al. Elevated visfatin/pre-B-cell colony-enhancing factor plasma concentration in ischemic stroke. *Journal of stroke and cerebrovascular diseases : the official journal of National Stroke Association*. 2009;18:354-9.
- [122] Lee KA, Gong MN. Pre-B-cell colony-enhancing factor and its clinical correlates with acute lung injury and sepsis. *Chest*. 2011;140:382-90.
- [123] Chen J, Weng J-F, Hong W-C, Luo L-F, Yu W, Luo S-D. Change in plasma visfatin level after severe traumatic brain injury. *Peptides*. 2012;38:8-12.

- [124] Yin CG, Jiang L, Tang B, Zhang H, Qian Q, Niu GZ. Prognostic significance of plasma visfatin levels in patients with ischemic stroke. *Peptides*. 2013;42:101-4.
- [125] Huang Q, Dai WM, Jie YQ, Yu GF, Fan XF, Wu A. High concentrations of visfatin in the peripheral blood of patients with acute basal ganglia hemorrhage are associated with poor outcome. *Peptides*. 2013;39:55-8.
- [126] Lee K, Huh JW, Lim CM, Koh Y, Hong SB. Clinical role of serum pre-B cell colony-enhancing factor in ventilated patients with sepsis and acute respiratory distress syndrome. *Scandinavian journal of infectious diseases*. 2013;45:760-5.
- [127] Schaffler A, Hamer OW, Dickopf J, Goetz A, Landfried K, Voelk M, et al. Admission visfatin levels predict pancreatic and peripancreatic necrosis in acute pancreatitis and correlate with clinical severity. *The American journal of gastroenterology*. 2011;106:957-67.
- [128] Hajri T, Gharib M, Kaul S, Karpel MS, Jr. Association between adipokines and critical illness outcomes. *The journal of trauma and acute care surgery*. 2017;83:507-19.
- [129] Jackson PC, Hardwicke J, Bamford A, Nightingale P, Wilson Y, Papini R, et al. Revised estimates of mortality from the Birmingham Burn Centre, 2001-2010: a continuing analysis over 65 years. *Ann Surg*. 2014;259:979-84.
- [130] Wearn C, Hardwicke J, Kitsios A, Siddons V, Nightingale P, Moiemmen N. Outcomes of burns in the elderly: revised estimates from the Birmingham Burn Centre. *Burns*. 2015;41:1161-8.
- [131] Zimmerman JE, Kramer AA, Knaus WA. Changes in hospital mortality for United States intensive care unit admissions from 1988 to 2012. *Critical care (London, England)*. 2013;17:R81.
- [132] Loss SH, Nunes DSL, Franzosi OS, Salazar GS, Teixeira C, Vieira SRR. Chronic critical illness: are we saving patients or creating victims? *Revista Brasileira de Terapia Intensiva*. 2017;29:87-95.
- [133] Kahn JM, Le T, Angus DC, Cox CE, Hough CL, White DB, et al. The epidemiology of chronic critical illness in the United States*. *Crit Care Med*. 2015;43:282-7.
- [134] Marchioni A, Fantini R, Antenora F, Clini E, Fabbri L. Chronic critical illness: the price of survival. *European journal of clinical investigation*. 2015;45:1341-9.
- [135] Lee KC, Dretzke J, Grover L, Logan A, Moiemmen N. A systematic review of objective burn scar measurements. *Burns & trauma*. 2016;4:14.



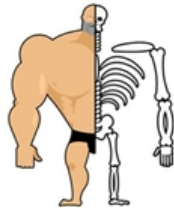
Cardiovascular System

- Influences myocardial and vascular inflammation, remodeling and fibrosis
- Affects myocardial metabolism and function
- Modulates atherosclerosis and thrombotic activity



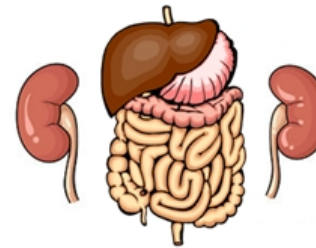
Nervous System

- Influences neuronal responsiveness
- Affects neuroendocrine mechanisms associated with feeding and energy expenditure
- Modulates neuro-inflammation



Musculoskeletal System

- Influences glucose uptake and insulin sensitivity
- Modulates protein and fatty acid metabolism
- Affects lipid deposition in muscle



Gastrointestinal and Renal System

- Modulates gastrointestinal and renal inflammation
- Influences cellular apoptosis
- Affects renal metabolism and function

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Declaration of interest

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