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# Chest Wall Reconstruction with Porcine Acellular Dermal Matrix (Strattice) and Autologous Tissue Transfer for High Risk Patients with Chest Wall Tumors

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**Background:** Tenets of chest wall reconstruction for malignancy are well known; however, the insertion of synthetic prosthetic material to achieve rigid and nonrigid skeletal reconstruction could be associated with more complications including infection and extrusion especially in high risk patients. This includes fungating, infected tumors, previous radiotherapy, scars, smoking, diabetes, and morbid obesity. Bioprosthesis Acellular Dermal Matrix Strattice would be a substitute to resist infection and provide stable coverage.

**Methods:** A retrospective data analysis study on 8 high-risk patients who underwent chest wall resection and reconstruction with bioprosthesis (Strattice) with/without titanium plates in the period between 2012 and 2017 was performed. Patient's demographics, risk factors, clinico-pathological, radiological, operative details, adjuvant therapy, postoperative morbidity, and follow-up data were recorded.

**Results:** Sarcoma was the pathology in 7 and recurrent breast cancer in 1 with age range from 21 to 71 years (mean, 50) and preponderance of female patients (n = 5). Defects were located anterior/anterolateral with size ranging from 270 to 1,050 cm<sup>2</sup> (mean, 511). Reconstruction was performed using Strattice only in 4 patients, whereas in 4 it was combined with titanium plates. All patients required flap reconstruction (3 pedicled and 5 free). The follow-up ranged from 9 to 52 months (mean, 24.8). Minor complications occurred in 2 patients; however, good functional outcome was achieved in all.

**Conclusion:** Strattice would act as a safe alternative modality for chest wall reconstruction to resist infection in high-risk patients with extensive defects. It should be considered as a valuable tool in the armamentarium of chest wall reconstruction. (*Plast Reconstr Surg Glob Open* 2018;6:e1703; doi: 10.1097/GOX.0000000000001703; Published online 18 May 2018.)

## INTRODUCTION

Surgery is considered the corner stone for management of chest wall malignancy.<sup>1-3</sup> The wide resection resulting in large chest wall defects can lead to instability, interference with respiration with the potential increase in postopera-

tive morbidity and mortality.<sup>4,5</sup> The principles of chest wall reconstruction are well known, radical resection ensuring adequate tumor-free margins, maintenance of chest wall stability, lung functional, and acceptable cosmetic results.<sup>1,2</sup> However, the insertion of prosthetic material to achieve rigid and nonrigid skeletal reconstruction could be associated with more complications including surgical-site infection (SSI), wound dehiscence, and possible extrusion of prosthesis especially in high-risk patients.<sup>3,6,7</sup> Uni/multivariate analysis of predictors of complications has been previously discussed in the literature including general factors (diabetes, immune-suppression, morbid obesity, smoking) and local factors (previous radiotherapy and surgery, fungating and infected tumors).<sup>1,6,8</sup> Various materials have

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been reported to be used for either rigid or nonrigid chest wall reconstruction to achieve functional chest wall stability including mainly synthetic meshes [Polypropylene (Marlex) poly tetrafluoroethylene (Gore-Tex)], Methylmethacrylate sandwich mesh, Stratos osteosynthesis system comprising titanium bars and clips and other metallic substitutes.<sup>3,9–11</sup> On the other hand, there is sparse evidence reported in the literature mainly reported cases on the use of biological mesh (Permocal, Alloderm, and Strattice) both in clinical practice and animal model.<sup>7,12–17</sup> We report our case series experience using acellular dermal matrix (ADM) Strattice in patients at increased risks of synthetic prosthetic mesh-related complications due to mainly local factors including infected fungating and necrotic tumors, previous surgery and perioperative radiotherapy/chemotherapy and to a lesser extent general factors as smoking, morbid obesity, and diabetes.

## MATERIALS AND METHODS

Prospectively collected data from 8 consecutive cancer patients who underwent chest wall resection and reconstruction with Strattice (ADM) biological mesh in the period between January 2012 and April 2017 were reviewed. The indication to consider bioprosthetic material Strattice included high-risk patients having contamination of the operative field (fungating, infected, and necrotic tumors), previous local radiotherapy and surgery and/or associated general risk factors as diabetes, active smoking, chemotherapy, and morbid obesity. All patients were managed through a multidisciplinary approach including thoracic surgeon (M.K.) and reconstructive surgeon (H.K.) to optimize their perioperative treatment plan. This included preoperative preparation, pulmonary function test to assess fitness, radiological investigations including computed tomographic (CT) scan and/or angiography and magnetic resonance imaging to plan the surgical resection, possible skeletal and soft-tissue reconstructive options and oncological adjuvant therapy.

### Data Collection

A prospectively maintained database was used to identify the patients, and their records were assessed retrospectively. The demographic information and potential identifiable risks and comorbidities, previous history of malignancy, and treatment were collected. Tumor characteristics and details of adjuvant therapy including either chemotherapy or radiotherapy were also documented. The results of the radiological diagnostic workup included standard chest x-ray, CT scan, and magnetic resonance imaging to delineate the extent of the tumor in the chest wall, and involvement of adjacent surrounding structures was noted. The operative details identified the numbers of ribs and costochondral junctions along with other structures that were resected as part of the tumor extirpation and included intercostal muscles, chest wall muscles, lungs, diaphragm, and overlying soft tissue. The sizes of the skeletal and soft-tissue defect were recorded in all patients. Documentation of whether Strattice was used solely or in adjunct with titanium plates and rib clips (Strasbourg

Thoracic Osteosyntheses System; MedXpert GmbH, Heitersheim, Germany) to restore chest wall integrity was noted. In addition, the methods of soft-tissue coverage were also documented. The postoperative outcomes in terms of length of stay, complications particularly SSI, wound dehiscence, chest wall stability, and respiratory function were identified from the patient records.

### Operative Consideration

All patients included were considered for surgery after been reviewed by the multidisciplinary thoracic oncoplastic team in the sarcoma and breast multidisciplinary team meeting (MDT). The resection and reconstruction were performed as a 1-stage procedure in all patients with a 2 team approach. In the majority of cases (5/8), the 2 teams worked simultaneously when free flap reconstruction was performed, whereas in 2 (2 of 8), the thoracic surgeons initially performed the resection and restored the skeletal wall integrity; this was followed by harvesting and inset of the selected regional flap. On the other hand, in 1 patient, the reconstructive team harvested the selected regional flap first to facilitate tumor extirpation and also salvage the regional flap from being damaged through the conventional thoracic approach. Post tumor resection chest wall stability was achieved using firm Strattice ADM (30×30 or 15×15 cm), which was washed in vitro with 5 L of saline to remove any chemical preservatives, followed by fashioning of the sheet to the defect (onlay technique) and securing it to the surrounding rib edges with interrupted Ethibond 2/0 sutures. Titanium plates and rib clips (Stratos osteosynthesis MedExpert) were used in addition to the Strattice ADM in 4 patients due to the extent of the resection and the location of the tumor being anterolateral to maintain structural support and stability. The site, size of the defect, availability of tissues, and preference of the reconstructive surgeon determined the choice of the soft-tissue flap. Soft-tissue reconstruction was achieved utilizing loco-regional pedicled muscle flaps as latissimus dorsi with or without split-thickness graft. Free microvascular tissue was indicated in more extensive resection with large surface area or when loco-regional flaps deemed not feasible due to direct tumor involvement, vascular compromise, and previous radiotherapy or surgery. We were mindful of creating a suitable window in the Strattice sheet in the cases where the internal mammary pedicle was used as the recipient pedicle for the free flap to avoid any compromise of the vascular supply postmicrovascular anastomosis. As a routine, all surgical sites were copiously irrigated with pulse-jet lavage using 3 L of saline (0.9% NaCl) solution to mechanically remove any debris or loose diathermized tissue and to maintain moisture of the Strattice. A chest wall tube was placed in the thoracic cavity, whereas suction drains were inserted deep into the flap and subcutaneous tissue. The duration of the drainage depended on the clinical, chest x-ray, and biological follow-up. Intraoperative collagen implant impregnated with aminoglycoside antibiotic gentamicin (Collatamp, EUSA Pharma, Hempstead, UK) was inserted in the operative field before closure. All 8 patients were transferred to intensive therapy unit with 6 patients (6 of 8) extubated

at the end of surgery and 2 within 12 hours. Standard flap observation chart protocol was followed, including Doppler signal, skin color, temperature, and capillary refill. At induction, all patients received antibiotics intravenous therapy, determined by appropriate tissue penetrability with special reference to bone and joint infections for prosthetic device<sup>18</sup> and continued routinely for 48 hours postoperatively followed by oral therapy for the duration of time the drains remained in situ. All patients received structured physiotherapy and rehabilitation program for both chest and donor site depending on flap selection.

## RESULTS

The patient, tumor, and chest wall defect characteristics including surgical management and outcomes for the cases are summarized in Table 1. The risks for SSI included fungating, infected and necrotic tumors, previous local radiotherapy, and surgical scar along with general risk factors as diabetes, active smoking, chemotherapy, and morbid obesity. The tumor tissue was recurrent breast cancer in 1 and sarcoma in 7 and with age range from 21 to 71 years (mean, 50) and preponderance of female patients ( $n = 5$ ). Resultant defects were located anterior, lateral, and anterolateral with sizes ranging from 270 to 1,050 cm<sup>2</sup> (mean, 511). The average number of ribs resected with adjoining costochondral cartilage was between 3 and 10 (median, 4). In addition, partial sternectomy was performed in 1 patient and complete sternal resection in another one. Adequate clear surgical resection margins were achieved in all the patients. The resultant chest wall defect was repaired using Strattice in all patients. In 4 patients, the defects were large, crossing midline and required additional rigid reconstruction using titanium plates and rib clips (Stratos). The repaired defects were covered in all patients with soft-tissue flaps successfully achieved with free microvascular tissue transfer in 5 patients [muscle sparing transverse abdominus myocutaneous (MSTRAM) type I ( $n = 4$ ) and tensor fasciae latae (TFL;  $n = 1$ )], whereas in 3 pedicled latissimus dorsi muscle with split-thickness graft and musculocutaneous flap were performed. The recipient vessels for the microvascular anastomosis were internal mammary pedicle ( $n = 4$ ) and thoracodorsal pedicle ( $n = 1$ ; Figs 1–5; **see video, Supplemental Digital Content 1**, demonstrating the multidisciplinary 2-team simultaneous approach for en bloc resection of a fungating infected large chondrosarcoma of the anterior chest wall. This video is available in the “Related Videos” section of PRSGlobalOpen.com or at <http://links.lww.com/PRSGO/A722>).

Two patients experienced minor complications, 1 developed wound breakdown and superficial SSI 10 months postoperatively after receiving radiotherapy. This required minor surgical debridement with secondary closure. The same patient developed local recurrence and distant metastases (month 12 postoperatively) and was referred to the palliative team. The other patient developed superficial minor distal flap necrosis, which required debridement and flap readvancement day 14 postoperatively; in neither patients, the Strattice was exposed. During the follow-up period of 9–52 months (mean, 24.8), there was

no local recurrence or distant metastases in any of the other patients with clinically good chest wall function. No readmissions were encountered within the 30 days from discharge. One patient died at 14 months due to systemic disease progression.

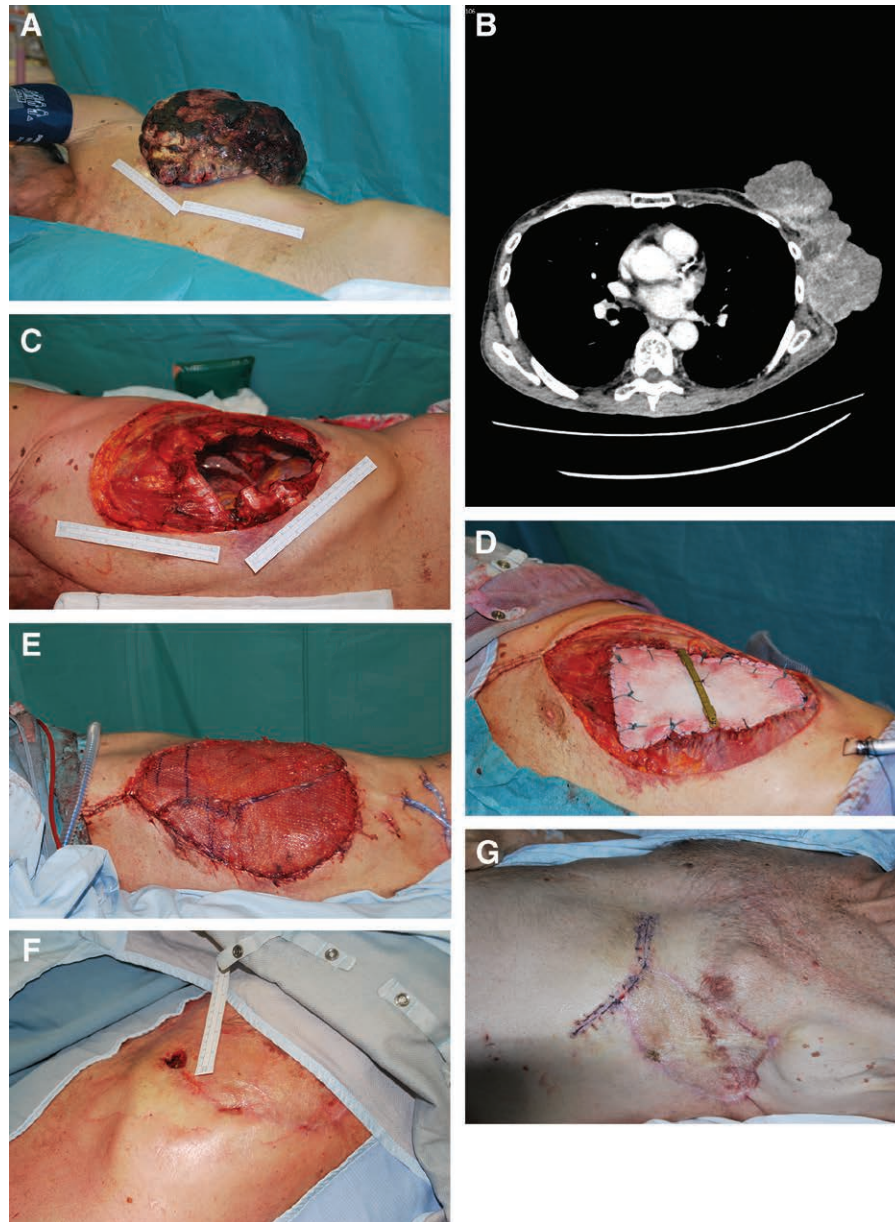
## DISCUSSION

The synthetic prosthesis mesh is used in the majority of chest wall reconstruction following resection of tumors. However, there is an increased risk of SSI, particularly if used in compromised surgical fields including fungating infected necrotic tumors, previously local radiotherapy and surgical scarring and in patients with general risk factors as diabetes, active smoking, chemotherapy, and morbid obesity.<sup>7</sup> Furthermore, if infection and/or exposure of mesh ensue in the event of being exposed due to dehiscence or breakdown of overlying skin, surgical removal is often required. Observation from previously published reports demonstrated the relatively high infection rate (6–22%) in chest wall reconstruction in noncontaminated defects with up to 42% requirement of removal of synthetic mesh.<sup>6</sup> In this series, of extensive chest wall defects (mean, 511 cm<sup>2</sup>) in compromised surgical fields, Strattice ADM was used to substitute synthetic mesh to resist infection. The general consensus accepted for chest wall skeletal reconstruction that we also applied in our series is that any defect less than 5 cm in size in any location, and those up to 10 cm in size posteriorly do not need rigid reconstruction for functional reasons. On the other hand, posterior defects in proximity to the tip of the scapula and larger lesions mostly anterior and lateral are likely to produce paradoxical chest wall motion therefore requires rigid reconstruction.<sup>1,6,10,11</sup> Four patients in our series required additional structural support with rigid skeletal reconstruction using titanium plates (Stratos) due to the extent and location of the defects in the anterolateral chest wall region to maintain chest wall integrity. To date, there are few case reports demonstrating the use of Strattice in chest wall reconstruction.<sup>14,16,17</sup> Alternatively, other bioprosthetic meshes as Alloderm and Permacol in both human and animal models have shown to be efficacious in similar scenarios.<sup>7,12,13</sup> The advantages of biological mesh materials over synthetic material have been reported, which includes superior handling properties, reduced visceral adhesions, resistance to infection, better integration into the autologous tissue due to host cell infiltration, and revascularization while maintaining structural integrity.<sup>7,13</sup> In addition, bioprosthesis has better tolerance to exposure due to tissue breakdown, hence can be managed conservatively rather than surgical extirpation.<sup>7,13,19</sup> Despite the ability of both Alloderm and Strattice to become revascularized and resist infection, previous case reports have highlighted the superiority of porcine-derived over human-derived biological mesh including less elasticity to avoid stretch, been harvested in larger and consistent sheets achieving coverage of larger defects with a single sheet.<sup>15,17,20</sup> This was in concordance with our results, as 1 sheet was used per patient notwithstanding that the defect sizes reconstructed in this series

**Table 1. The Patient, Tumor, and Chest Wall Defect Characteristics Including Surgical Management and Outcomes**

	Pathology	Risk Factors	Resection	Defect Size	Skeletal Reconstruction	Soft-tissue Reconstruction	Follow-up	Survival
Case 1 (53 y); male	Fungating chondrosarcoma anterior chest wall	Fungation Infected Smoker	Bilateral (whole anterior chest wall) Total sternectomy + 5 ribs bilaterally including costochondral junctions	1,050 cm <sup>2</sup>	Strattice ADM + STRATOS bars	Free TFL flap	48 mo No recurrence	Alive
Case 2 (71 y); female	Radiation-induced sarcoma Previous wide local excision for breast cancer	Previous chronic deep abscess Previous radiation Smoker Diabetic Fungation Infection smoker	Unilateral (right) 5 ribs + costochondral junctions + completion mastectomy (R0 resection)	900 cm <sup>2</sup>	Strattice ADM	Free MSTRAM	12 mo No recurrence	Alive
Case 3 (54 y); male	High-grade infected fungating spindle sarcoma chest wall		Unilateral anterolateral (left) 4 ribs (R0 resection)	500 cm <sup>2</sup>	Strattice ADM + STRATOS bars	Lattissimus dorsi muscle flap + split-thickness graft	14 mo Recurrence 12 mo	Died 14 mo with disease progression
Case 4 (62 y); female	Radiation-induced sarcoma Previous breast cancer	Previous radiation Morbid obesity	Unilateral (Left) 5 ribs Pleura + lung segmentectomy (R0 resection)	300 cm <sup>2</sup>	Strattice ADM	Free MSTRAM	22 mo No recurrence	Alive
Case 5 (40 y); female	Recurrent nonmetastatic breast cancer involving chest wall	Previous radiation Long-term chemotherapy	Unilateral anterior (Right) 3 ribs + costochondral cartilages (R0 resection)	375 cm <sup>2</sup>	Strattice ADM	Free MSTRAM	5 mo No recurrence	Alive
Case 6 (51 y); female	Recurrent synovial sarcoma	Previous surgery Previous radiation Smoker Diabetic Previous radiation	Unilateral anterolateral (right) 5 ribs (R0 resection)	400 cm <sup>2</sup>	Strattice ADM + STRATOS	Free MSTRAM	36 mo No recurrence	Alive
Case 7 (48 y); female	Radiation-induced sarcoma Previous breast cancer	Previous radiation	Unilateral anterior (Right) 3 ribs (R0 resection)	270 cm <sup>2</sup>	Strattice ADM	Lattissimus dorsi musculocutaneous flap	28 mo No recurrence	Alive
Case 8 (21 y)	Ewing sarcoma	Opportunistic infections (pneumonia + perianal abscess) 15 Cycles (neoadjuvant chemotherapy) proton beam radiotherapy	Unilateral anterolateral with posterior extension (right)	300 cm <sup>2</sup>	Strattice ADM + STRATOS	Lattissimus dorsi muscle flap	3 mo No recurrence	Alive

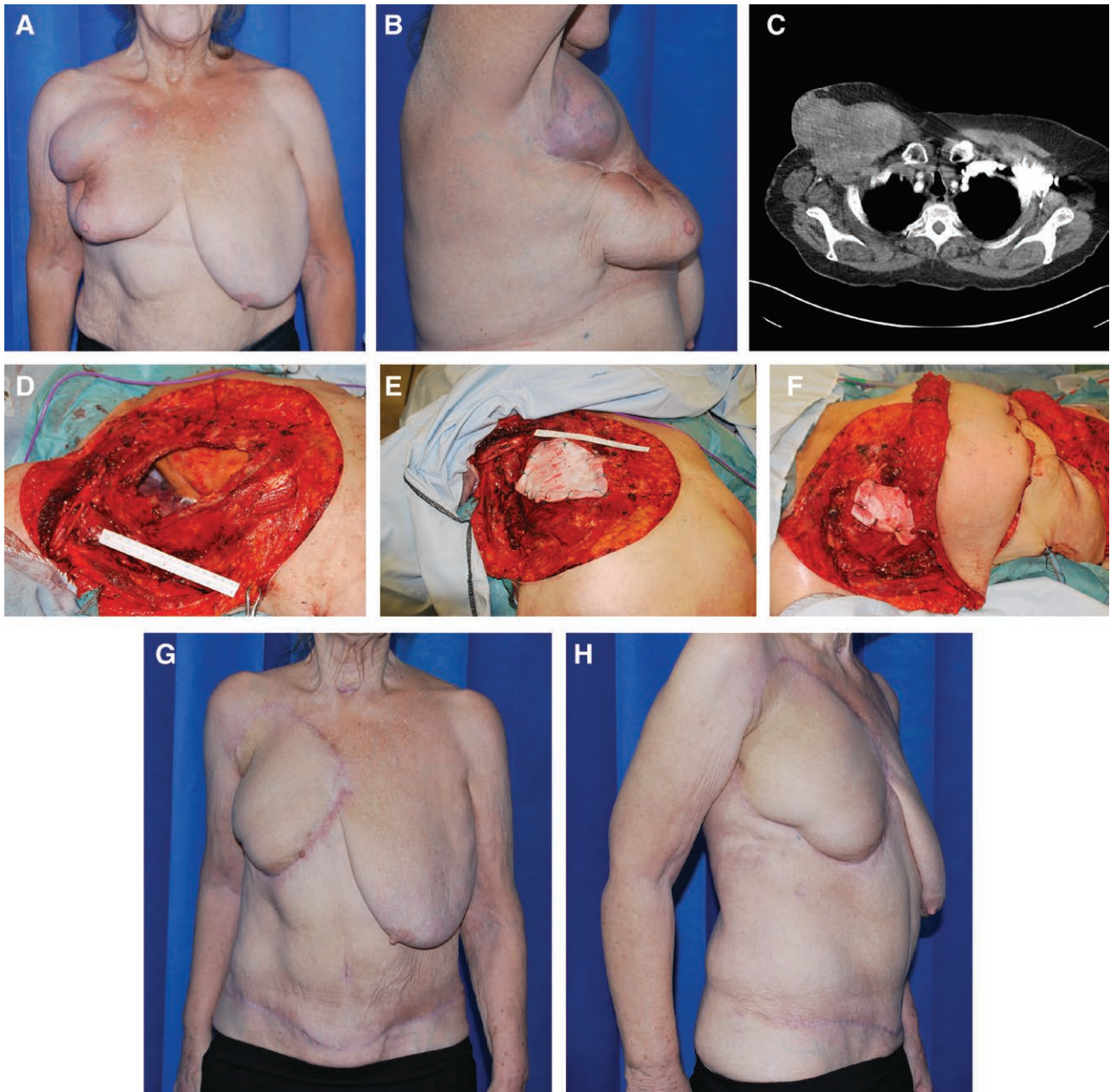




**Fig. 1.** A, Large undifferentiated fungating necrotic soft-tissue sarcoma left chest wall in a 53-year-old smoker male patient. B, CT scan defining the extent of the lesion with involvement of underlying ribs and intercostals space necessitating resection of ribs to achieve negative surgical margins; C, Intraoperative photograph post en bloc tumor resection including 4 ribs demonstrating the skeletal defect exposing underlying thoracic viscera. D, Intraoperative photograph showing reconstruction of the skeletal defect using Strattice ADM used with STRATOS bar. E, Intraoperative photograph showing soft-tissue coverage achieved with latissimus dorsi muscle flap and meshed skin graft. F, 10 Months postoperative photograph showing 1×2 cm distal wound dehiscence postradiotherapy. G, Postoperative photograph 3 weeks postdebridement and secondary closure of the dehiscence area. Note the full maturation of the graft.

were relatively larger from previously reported cases. The relatively longer follow-up (mean, 24.8 months) when compared with previous reports illustrated the durability of the functional properties of a remodeled mature biologic scaffold in terms of strength and stretch. During the follow-up period, all patients showed satisfactory functional outcome assessed clinically and with pulmo-

nary function tests complimented with equally cosmetic satisfactory cosmetic outcomes. Hedgire et al.<sup>21</sup> reported on significant enhancement of Alloderm Biologic Mesh Spacer in the abdomen and pelvis detected on follow-up multi detector CT scan through progressive rise in Hounsfield unit values. This has been attributed to the possible revascularization of the ADM as supported by other

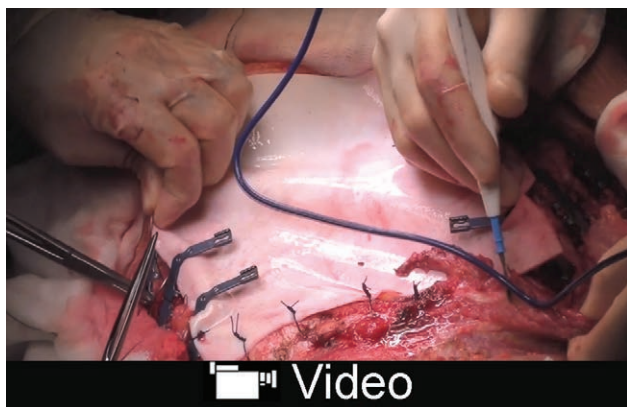


**Fig. 2.** A and B, Preoperative photograph showing radiation induced sarcoma of right upper chest wall in a 71-year-old female patient post breast cancer treatment. High risk factors included smoking, diabetes, previous radiotherapy, and previous chronic deep abscess (front-lateral view). C, CT scan defining the extent of the lesion with involvement of underlying ribs and intercostals space necessitating resection of ribs to achieve negative surgical margins. D, Intraoperative photograph post en bloc tumor resection including 3 ribs demonstrating the extent of skeletal and soft-tissue resection exposing underlying thoracic viscera, axillary neurovascular bundle and brachial plexus. E, Intraoperative photograph showing reconstruction of the skeletal defect using nonrigid reconstruction with Stratice ADM only. F, Intraoperative photograph postmicrovascular anastomosis of free MSTRAM type I using the internal mammary vascular pedicle as recipient vessels to provide soft-tissue coverage of the chest wall defect. G and H, 12 Months postoperative photograph showing complete survival of the flap with primary healing and restoration of chest wall anatomy (front-lateral view).

reports.<sup>22</sup> In our study, this was also observed in the CT scan follow-up performed postoperatively in 4 patients, which revealed well-enhanced scar tissue denoting revascularization and maturation providing stable coverage. Arguably other synthetic mesh materials would have been used; however, there would be an increased risk of infection of these synthetic meshes within this compromised

surgical field and their intolerance for contamination and/or cutaneous exposure, which has been highlighted in other studies.<sup>7,8,14</sup> Eventually, this would lead to detrimental consequences as a result of the requirement of mesh removal, which has been also highlighted in previous studies.<sup>6,7,14</sup> In his case report, Brunbjerg et al.<sup>14</sup> reported the use of Stratice successfully in chest wall reconstruc-





**Video Graphic 1.** See video, Supplement Digital Content 1 demonstrating the multidisciplinary 2-team simultaneous approach for en-bloc resection of a fungating infected large chondrosarcoma of the anterior chest wall. Strattice ADM and titanium plates (Stratos) were used to restore the skeletal chest wall integrity while soft-tissue reconstruction was achieved using free TFL flap. The resection included the sternum with the adjacent 7 costochondral cartilages and ribs on each side with exposure of the underlying thoracic viscera. Restoration of the chest wall integrity was subsequently achieved with Strattice ADM and titanium plates (Stratos). The free TFL flap provided soft coverage using the internal mammary vascular pedicle as the recipient vessels. The postoperative photographs demonstrate the complete survival of the flap primary wound healing and restoration of the chest wall integrity, whereas the postoperative CT scan reveals the maturation of the Strattice ADM providing stable chest wall reconstruction. This video is available in the "Related Videos" section of PRSGlobalOpen.com or at <http://links.lww.com/PRSGO/A722>.

tion in a patient with multiple recurrent breast cancer, chemotherapy, and radiotherapy requiring multiple chest wall resection and reconstruction with previous failure and extrusion of the polypropylene mesh. We previously reported the value of multidisciplinary thoracic oncoplastic approach in the reduction of infection (2.8%) in the presence of synthetic prosthetic material in noncontaminated chest wall defects.<sup>3</sup> Adopting the same approach in this relatively small cohort of high-risk patients, the pertinence of bioprosthetic mesh is seen to be associated with decreased incidence of SSI, less extrusion rate without any compromise of the chest wall stability that has also been supported by other studies.<sup>6,7,14</sup> Observation from previously published reports demonstrated the importance of the proximate collaboration between several disciplines in managing these patients; however, it underscores the occurrence of the wound morbidity even in the presence of soft-tissue coverage for rigid reconstruction ranging from 9% to 25%,<sup>1,8</sup> whereas in other series, this relationship was not highlighted.<sup>9,23</sup> The recruitment of well-vascularized soft tissue is paramount to provide coverage of the prosthesis, prevent infection, seal the pleural space, protect underlying viscera, obliterate dead space, contribute to structure integrity and healing, and enhance aesthetic outcome.<sup>1–3,7,24,25</sup> A wide variety of flaps could be used including loco-regional or free flaps depending on the site, size, and availability of tissues from donor sites and the available expertise.<sup>3,7,26</sup> Soft-tissue coverage was performed

in all patients in this series due to the extent of the cutaneous defects and to provide protection for the underlying Strattice and titanium plates. The majority of the patients required free flap including MSTRAM and TFL (5 of 8; 62.5%) when loco-regional flaps deemed not feasible, extent of the resection and due to the logistics of positioning the patients to allow thoracic and reconstructive surgeons working simultaneously. The efficacy of the xenogeneic nature of biological mesh has been questionable in previous reports<sup>7,13,27,28</sup>; however, with the rapidly increasing use of porcine-derived ADM in several indications including breast, abdominal, head and neck, dural reconstruction and static reconstruction postfacial nerve paralysis over the last decade has demonstrated its advantages.<sup>8,29–33</sup>

The current literature is largely limited to case studies, and to the authors best knowledge, this would be the largest series from a single institute to be reported with long-term follow-up addressing the decreased incidence of SSI and durability of chest wall reconstruction with the use of Strattice ADM in high risk patients. However, the limitation for this study is the relatively low number of patients from a single institute mainly due to the rarity of these presentations, the lack of available control group for direct comparison, and the requirement of potential objective analysis comparing pre- and postsurgery chest wall biomechanics. This study is driven to evaluate the indications and functional outcomes in patients undergoing chest wall reconstruction with bioprosthesis Strattice ADM, which would act as a safe alternative modality for chest wall reconstruction to resist infection in high risk patients with extensive defects. It should be considered as a valuable tool in the armamentarium of chest wall reconstruction.

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## REFERENCES

1. Gonfiotti A, Santini PF, Campanacci D, et al. Malignant primary chest-wall tumours: techniques of reconstruction and survival. *Eur J Cardiothorac Surg*. 2010;38:39–45.
2. Mansour KA, Thourani VH, Losken A, et al. Chest wall resections and reconstruction: a 25-year experience. *Ann Thorac Surg*. 2002;73:1720–1725; discussion 1725.
3. Khalil HH, Malahias MN, Balasubramanian B, et al. Multidisciplinary oncoplastic approach reduces infection in chest wall resection and reconstruction for malignant chest wall tumors. *Plast Reconstr Surg Glob Open*. 2016;4:e809.
4. Berthet JP, Wihlm JM, Canaud L, et al. The combination of polytetrafluoroethylene mesh and titanium rib implants: an in-



- novative process for reconstructing large full thickness chest wall defects. *Eur J Cardiothorac Surg*. 2012;42:444–453.
5. Berthet JP, Solovei L, Tiffet O, et al. Chest-wall reconstruction in case of infection of the operative site: is there any interest in titanium rib osteosynthesis? *Eur J Cardiothorac Surg*. 2013;44:866–874.
6. Weyant MJ, Bains MS, Venkatraman E, et al. Results of chest wall resection and reconstruction with and without rigid prosthesis. *Ann Thorac Surg*. 2006;81:279–285.
7. Butler CE, Langstein HN, Kronowitz SJ. Pelvic, abdominal, and chest wall reconstruction with AlloDerm in patients at increased risk for mesh-related complications. *Plast Reconstr Surg*. 2005;116:1263–1275; discussion 1276.
8. Basta MN, Fischer JP, Lotano VE, et al. The thoracoplastic approach to chest wall reconstruction: preliminary results of a multidisciplinary approach to minimize morbidity. *Plast Reconstr Surg*. 2014;134:959e–967e.
9. Arnold PG, Pairolero PC. Chest-wall reconstruction: an account of 500 consecutive patients. *Plast Reconstr Surg*. 1996;98:804–810.
10. Losken A, Thourani VH, Carlson GW, et al. A reconstructive algorithm for plastic surgery following extensive chest wall resection. *Br J Plast Surg*. 2004;57:295–302.
11. Deschamps C, Tirnaksiz BM, Darbandi R, et al. Early and long-term results of prosthetic chest wall reconstruction. *J Thorac Cardiovasc Surg*. 1999;117:588–591; discussion 591.
12. Mirzabeigi MN, Moore JH Jr, Tuma GA. The use of Permacol® for chest wall reconstruction in a case of desmoid tumour resection. *J Plast Reconstr Aesthet Surg*. 2011;64:406–408.
13. Holton LH 3rd, Chung T, Silverman RP, et al. Comparison of acellular dermal matrix and synthetic mesh for lateral chest wall reconstruction in a rabbit model. *Plast Reconstr Surg*. 2007;119:1238–1246.
14. Brunbjerg ME, Juhl AA, Damsgaard TE. Chest wall reconstruction with acellular dermal matrix (Strattice™) and a TRAM flap. *Acta Oncol*. 2013;52:1052–1054.
15. Cothren CC, Gallego K, Anderson ED, et al. Chest wall reconstruction with acellular dermal matrix (AlloDerm) and a latissimus muscle flap. *Plast Reconstr Surg*. 2004;114:1015–1017.
16. Huston TL, Taback B, Rohde CH. Chest wall reconstruction with porcine acellular dermal matrix (strattice) and a latissimus myocutaneous flap. *Am Surg*. 2011;77:e115–e116.
17. Kaplan KM, Chopra K, Feiner J, et al. Chest wall reconstruction with strattice in an immunosuppressed patient. *Eplasty*. 2011;11:e46.
18. Recommendations for bone and joint prosthetic device infections in clinical practice (prosthesis, implants, osteosynthesis). *Societe de Pathologie infectieuse de langue francaise. Med Mal. Infect*. 2010;40:185–211.
19. Butler CE, Prieto VG. Reduction of adhesions with composite AlloDerm/polypropylene mesh implants for abdominal wall reconstruction. *Plast Reconstr Surg*. 2004;114:464–473.
20. Nahabedian MY. Does AlloDerm stretch? *Plast Reconstr Surg*. 2007;120:1276–1280.
21. Hedgire SS, Elmi A, Kambadakone AR, et al. MDCT imaging of Alloderm biologic mesh spacers in the abdomen and pelvis—preliminary experience. *Clin Imaging*. 2014;38:279–282.
22. Nahabedian MY. AlloDerm performance in the setting of prosthetic breast surgery, infection, and irradiation. *Plast Reconstr Surg*. 2009;124:1743–1753.
23. McKenna RJ Jr, Mountain CF, McMurtrey MJ, et al. Current techniques for chest wall reconstruction: expanded possibilities for treatment. *Ann Thorac Surg*. 1988;46:508–512.
24. Chapelier AR, Missana MC, Couturaud B, et al. Sternal resection and reconstruction for primary malignant tumors. *Ann Thorac Surg*. 2004;77:1001–1006; discussion 1006.
25. Rohrich RJ, Lowe JB, Hackney FL, et al. An algorithm for abdominal wall reconstruction. *Plast Reconstr Surg*. 2000;105:202–216; quiz 217.
26. Shahzad F, Wong KY, Maraka J, et al. Reconstruction of chest wall chondrosarcoma with an anterolateral thigh free flap: an illustration of decision-making in chest wall reconstruction. *Int J Surg Case Rep*. 2013;4:669–674.
27. Warren WL, Medary MB, Dureza CD, et al. Dural repair using acellular human dermis: experience with 200 cases: technique assessment. *Neurosurgery*. 2000;46:1391–1396.
28. Sandor M, Xu H, Connor J, et al. Host response to implanted porcine-derived biologic materials in a primate model of abdominal wall repair. *Tissue Eng Part A*. 2008;14:2021–2031.
29. Glasberg SB, Light D. AlloDerm and Strattice in breast reconstruction: a comparison and techniques for optimizing outcomes. *Plast Reconstr Surg*. 2012;129:1223–1233.
30. Kocak E, Nagel TW, Hulsen JH 3rd, et al. Biologic matrices in oncologic breast reconstruction after mastectomy. *Expert Rev Med Devices*. 2014;11:65–75.
31. Carruthers CA, Dearth CL, Reing JE, et al. Histologic characterization of acellular dermal matrices in a porcine model of tissue expander breast reconstruction. *Tissue Eng Part A*. 2015;21:35–44.
32. Melman LI, Jenkins ED, Hamilton NA, et al. Early biocompatibility of crosslinked and non-crosslinked biologic meshes in a porcine model of ventral hernia repair. *Hernia*. 2011;15:157–164.
33. Skovsted Yde S, Brunbjerg ME, Gudmundsdottir G, et al. Dural repair using porcine ADM: two cases and a literature review *Case Reports Plast Surg Hand Surg*. 2017;4:5–8.