## UNIVERSITY<sup>OF</sup> BIRMINGHAM University of Birmingham Research at Birmingham

### The relationship between transpiration and nutrient uptake in wheat changes under elevated atmospheric CO

Houshmandfar, Alireza; Fitzgerald, Glenn J.; O'Leary, Garry; Tausz-Posch, Sabine; Fletcher, Andrew; Tausz, Michael

DOI: 10.1111/ppl.12676

License: Other (please specify with Rights Statement)

Document Version Peer reviewed version

Citation for published version (Harvard):

Houshmandfar, A, Fitzgerald, GJ, O'Leary, G, Tausz-Posch, S, Fletcher, A & Tausz, M 2017, 'The relationship between transpiration and nutrient uptake in wheat changes under elevated atmospheric CO', *Physiologia Plantarum*. https://doi.org/10.1111/ppl.12676

Link to publication on Research at Birmingham portal

#### **Publisher Rights Statement:**

This is the peer reviewed version of the following article: Houshmandfar, A., Fitzgerald, G. J., O'Leary, G., Tausz-Posch, S., Fletcher, A. and Tausz, M. (2018), The relationship between transpiration and nutrient uptake in wheat changes under elevated atmospheric CO2. Physiol Plantarum. Accepted Author Manuscript. . doi:10.1111/ppl.12676, which has been published in final form at 10.1111/ppl.12676. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving.

#### **General rights**

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

•Users may freely distribute the URL that is used to identify this publication.

•Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.

•User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?) •Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

#### Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

# The relationship between transpiration and nutrient uptake in wheat changes under elevated atmospheric CO<sub>2</sub>

Alireza Houshmandfar<sup>1,2,\*</sup>, Glenn J. Fitzgerald<sup>3,2</sup>, Garry O'Leary<sup>3</sup>, Sabine Tausz-Posch<sup>2,4</sup>, Andrew Fletcher<sup>1</sup>, Michael Tausz<sup>5,6</sup>

- 1) CSIRO Agriculture and Food, Private Bag 5, P.O., Wembley, WA 6913, Australia
- 2) Faculty of Veterinary and Agricultural Sciences, The University of Melbourne, Creswick, Victoria 3363, Australia
- Agriculture Victoria, Victoria State Department of Economic Development, Jobs, Transport and Resources, Horsham 3401, Australia
- School of Biosciences, University of Birmingham, Edgbaston, Birmingham B15 2TT, United Kingdom
- 5) Birmingham Institute of Forest Research, The University of Birmingham, Edgbaston, Birmingham B15 2TT, United Kingdom
- Department of Ecosystem and Forest Sciences, The University of Melbourne, Creswick, Victoria 3363, Australia

<u>Corresponding author:</u> Alireza Houshmandfar, CSIRO Agriculture and Food, Private Bag 5, P.O., Wembley, WA 6913, Australia. Email: <u>alireza.houshmandfar@csiro.au</u> Faculty of Veterinary and Agricultural Sciences, University of Melbourne, 4 Water Street, Creswick, Victoria 3363, Australia. Email: alireza.houshmandfar@unimelb.edu.au

#### Abstract

The impact of elevated  $[CO_2]$  (e $[CO_2]$ ) on crops often includes a decrease in their nutrient concentrations where reduced transpiration-driven mass flow of nutrients has been suggested to play a role. We used two independent approaches, a Free-Air CO<sub>2</sub> Enrichment (FACE) experiment in the South Eastern wheat belt of Australia and a simulation study employing the Agricultural Production Systems Simulator (APSIM), to show that transpiration (mm) and nutrient uptake (g m<sup>-2</sup>) of nitrogen (N), potassium (K), sulfur (S), calcium (Ca), magnesium (Mg), and manganese (Mn) in wheat are correlated under  $e[CO_2]$ , but that nutrient uptake per unit water transpired is higher under  $e[CO_2]$  than under ambient  $[CO_2]$  ( $a[CO_2]$ ). This result suggests that transpiration-driven mass flow of nutrients contributes to decreases in nutrient concentrations under  $e[CO_2]$ , but cannot solely explain the overall decline.

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1111/ppl.12676

Keywords: APSIM, FACE, Jarvis model, nitrogen uptake, Triticum aestivum L.

**Abbreviations:** a[CO<sub>2</sub>], ambient [CO<sub>2</sub>]; CNC, critical nitrogen concentration; EC, electrical conductivity; e[CO<sub>2</sub>], elevated [CO<sub>2</sub>]; FACE, Free-Air CO<sub>2</sub> Enrichment; LAI, leaf area index; ns, not significant; PAR, photosynthetically active radiation; RUE, radiation use efficiency;  $g_s$ , stomatal conductance; TE, transpiration efficiency; VPD, vapour pressure deficit

#### Introduction

Anthropogenic activities such as fossil fuel consumption and deforestation have caused atmospheric  $[CO_2]$  to increase rapidly (Canadell et al., 2007) from 315 µmol mol<sup>-1</sup> in 1960 to approximately 400 µmol mol<sup>-1</sup> in 2015. Future  $[CO_2]$  is likely to reach 550 µmol mol<sup>-1</sup> or above by 2050 (IPCC, 2013). Plants respond to elevated  $[CO_2]$  (e $[CO_2]$ ) with increased photosynthesis (Ainsworth and Long, 2005; Nowak et al., 2004) and reduced stomatal conductance ( $g_s$ ) (Bunce, 2004; Leakey et al., 2006), and these responses are fundamental for all other CO<sub>2</sub>-driven effects on plants and their accompanying ecosystems (Ainsworth and Rogers, 2007; Long et al., 2004).

In C<sub>3</sub> crops, these changes improve water use efficiency and grain yield (Ainsworth et al., 2008; Leakey et al., 2012) but are likely to reduce nutrient concentrations in vegetative tissues, which, in turn, can result in lower nutrient concentrations in the grain (Conroy and Hocking, 1993; Hogy et al., 2009; Pleijel and Hogy, 2015). Concentrations of nutrients such as nitrogen (N), potassium (K), sulfur (S), calcium (Ca), magnesium (Mg), and manganese (Mn) have all been reported to decrease by up to 22% under e[CO<sub>2</sub>] (Loladze, 2002; Loladze, 2014; McGrath and Lobell, 2013).

Limited uptake of nutrients per unit root mass is one of the hypotheses suggested to explain the decline in nutrient concentration under e[CO<sub>2</sub>] (Taub and Wang, 2008). This reduced root uptake could result from changes in root systems, such as less efficient root system architecture or decreased uptake capacity per unit root length (BassiriRad et al., 2001; Pritchard and Rogers, 2000), or from reduced transpiration-driven mass flow of nutrients due to decreased  $g_s$  under e[CO<sub>2</sub>] (BassiriRad et al., 2001; Del Pozo et al., 2007; McGrath and Lobell, 2013).

The significance of transpirational mass flow for nutrient uptake is supported by a wealth of circumstantial data (Masle et al., 1992; Polley et al., 1999; Russell and Barber, 1960; Sellin et al., 2013). Transpiration rate may affect nutrient uptake both directly, through effects on the rate of radial transport of nutrients through the apoplasm (Marschner and Marschner, 2012), and indirectly, by influencing the supply of nutrients to the plasma membrane of root cells (Cramer and Hawkins, 2009; Tinker and Nye, 2000). Decreased transpiration rates may reduce rhizosphere nutrient depletion resulting from plant uptake, especially for solute nutrients (Kupper et al., 2012; Scholz et al., 2007).

Few attempts have been made to address the effect of  $e[CO_2]$  on the relationship between transpirational mass flow and nutrient uptake. McDonald et al. (2002) used a short duration (7-day period) pot experiment with  $e[CO_2]$  (approximately 1000 µmol mol<sup>-1</sup>) and high humidity under glasshouse conditions to report that transpiration rate and nitrate (NO<sub>3</sub><sup>-</sup>) uptake were positively related. A similar correlation was found by McGrath and Lobell (2013) in a meta-analysis that included experiments mostly conducted in open-top chambers, growth cabinets or glasshouse conditions. The authors pointed out that none of the experiments had measured transpirational mass flow and nutrient uptake simultaneously under  $e[CO_2]$ . Glasshouse or chamber conditions may also distort responses and lead to conclusions that are not immediately transferable to field conditions (Long et al., 2006; Long et al., 2004; Mcconnaughay et al., 1993; McLeod and Long, 1999). In addition, pot experiments can change the water dynamics necessary to evaluate the relationship between transpirational mass flow and nutrient uptake. Because of the small pot volume, nutrients are easily available to the roots, regardless of whether transpiration is reduced or not (McGrath and Lobell, 2013). Free-Air CO<sub>2</sub> enrichment (FACE) studies with plants rooted in the ground would likely give better estimates because roots have unrestricted access to soil volume, and water dynamics represent typical field conditions.

In this study, we used two independent approaches applied to the same wheat cultivars grown at the same site to investigate the potential relationship between transpiration and nutrient uptake. For the first approach, measurements in the Australian Grains Free-Air CO<sub>2</sub> enrichment (AGFACE) facility were used to simultaneously evaluate the nutrient uptake (N, K, S, Ca, Mg and Mn) and transpiration rates in wheat (Triticum aestivum L.) grown under ambient [CO<sub>2</sub>] (a[CO<sub>2</sub>], approximately 390 µmol mol<sup>-1</sup>) and  $e[CO_2]$  (approximately 550 µmol mol<sup>-1</sup>). During one growing season (2013) nutrient uptake was measured in detail and transpiration was estimated for the same period using the Jarvis-type (Jarvis, 1976) empirical  $g_s$  model. This model was parameterised from approximately 1500 leaf level  $g_s$ measurements on the same wheat cultivars at the AGFACE site (Houshmandfar et al., 2015a). For the second approach, nutrient uptake (for N only) and transpiration was simulated by the Agricultural Production Systems Simulator (APSIM) modeling framework over ten consecutive growing seasons from 2007 to 2016. APSIM is a well-tested agricultural model internationally recognised as a highly advanced simulator of cropping systems (Holzworth et al., 2014). The model simulates plant water uptake (approximately equal to transpiration (Taub, 2010)) and N uptake in wheat (APSIM-Wheat) for an adjustable duration of time, along with a wide range of other capabilities. APSIM-Wheat has been broadly evaluated in a range of experimental (Holzworth and Huth, 2011; Zhang et al., 2012) and farm conditions (Hochman et al., 2009), also under e[CO<sub>2</sub>] scenarios (Asseng et al., 2004; Reyenga et al., 1999), and in particular at the AGFACE site (O'Leary et al., 2015).

If the relationship between transpiration and nutrient uptake is identical under  $a[CO_2]$  and  $e[CO_2]$  (and explains a significant proportion of the data variance), then changes in transpiration rate will be sufficient to explain changes in crop nutrition under  $e[CO_2]$ .

Understanding the relative role of transpiration in lower nutrient concentration in crops grown under e[CO<sub>2</sub>] would improve our currently limited ability to predict responses for different crops and regions, and help guide breeding and agronomic strategies to adapt crops to higher [CO<sub>2</sub>].

#### Material and methods

#### Experimental setup and plant material

A field experiment using two cultivars of wheat (*Triticum aestivum* L. cv. Scout and cv. Yitpi) was conducted at the AGFACE site located in Horsham, Victoria, Australia (36°45′07″S, 142°06′52″E; 127 m above mean sea level) in the 2013 growing season. The soil type is a Vertosol clay with nondispersive and pedal surface (Isbell, 2002), approximately 35% clay at the top increasing to 60% at 1.4 m depth. The AGFACE is located in a semi-arid cropping area, which has a Mediterranean type climate but with cooler and drier winters (Hutchinson et al., 2005). Long-term average annual rainfall of the area is 435 mm, with 274 mm typically falling during the wheat growing season (June to November). Long-term mean growing season temperature is 16.5 °C (Australian Bureau of Meteorology). The soil had a pH of 8.2, EC (electrical conductivity) of 0.14  $\mu$ S cm<sup>-1</sup>, Mehlich 3 extractable (Mehlich, 1984) Ca of 6.4 (±0.5) g kg<sup>-1</sup>, K of 501.8 (±42.5) mg kg<sup>-1</sup>, Mg of 1.1 (±0.0) g kg<sup>-1</sup>, Mn of 0.1 (±0.0) mg kg<sup>-1</sup>, Fe of 120 (±9.2) mg kg<sup>-1</sup>, zinc (Zn) of 2.3 (±0.5) mg kg<sup>-1</sup>, copper (Cu) of 1.6 (±0.1) mg kg<sup>-1</sup>, Bray extractable (Bray and Kurtz, 1945) phosphate (PO4<sup>3-</sup>) of 19.2 (±2.2) mg kg<sup>-1</sup>, as well as AQ2 measured (Automated Discrete Analyzer, Seal Analytical Ltd, UK) nitrate (NO3<sup>-</sup>) and ammonium (NH4<sup>+</sup>) of 6.1 (±0.2) and 3.4 (±0.1) mg kg<sup>-1</sup>, respectively. These values are averages of 32 soil samples from 0 to 25 cm depth.

In 2013, the AGFACE was fully randomized in a complete block design with four replications, with eight octagonal plots (16 m diameter) of which four were  $a[CO_2]$  (approximately 390 µmol mol<sup>-1</sup>) and four  $e[CO]_2$  (approximately 550 µmol mol<sup>-1</sup>). Each cultivar was sown into two randomly allocated replicate subplots (1.5 m × 4 m), one each in opposing halves of the ring. Measurements from the two subplots were averaged for each replicate plot. Each  $e[CO_2]$  plot was encircled by horizontal CO<sub>2</sub>-release-tubes in an octagonal shape. The tubes were progressively raised as the crop grew so that the CO<sub>2</sub> was injected about 0.1 - 0.15 m above the canopy. A plot center [CO<sub>2</sub>] value of 550 µmol mol<sup>-1</sup> was targeted for the  $e[CO_2]$  treatment from sunrise to sunset starting from germination. Average plot central [CO<sub>2</sub>] were recorded every minute with an infrared gas analyzers (IRGA, SBA-4, PP Systems, MA, USA) located at the central part of each plot. Detailed engineering specifications, performance of the FACE system, and treatment descriptions are found in Mollah et al. (2011) and Fitzgerald et al. (2016).

#### Plant sampling and nutrient analysis

Leaf area index (LAI, one-sided green leaf area per m<sup>-2</sup> ground area) and nutrient uptake into aboveground biomass (g m<sup>-2</sup> ground area) were simultaneously measured at four different growing

stages from stem elongation until the end of anthesis: DC31, DC34, DC65 and DC69 (DC decimal code, according to Zadoks et al. (1974) cereal growth scale). Labelled areas of  $1.35 \times 0.5 \text{ m}^2$  were harvested at DC31 and DC65, and 30 to 50 random tillers sampled at DC34 and DC69 experimental unit. Plant densities were 414.4 (±15.3) and 462.5 (±13.2) tillers m<sup>-2</sup> under ambient and elevated [CO<sub>2</sub>], respectively. For each sampling time, entire aboveground biomass including leaves, stems, and spikes (DC 65 and DC 69) were harvested, then, after measuring leaf area with a calibrated leaf area meter (Li-3100, Li-Cor, NE, USA), dried for 72 h in a 70 °C of oven temperature. N concentration was measured using LECO Nitrogen Macro Determinator (TruMac, LECO Corporation, MI, USA). K, S, Ca, Mg, and Mn concentrations were measured using inductively coupled plasma atomic emission spectrometry (ICP-AES) (Applied Research Laboratories, 3580B, Switzerland) after digestion in concentrated nitric acid (HNO<sub>3</sub>) following the procedure described by Zarcinas et al. (1987). We studied N, K, S, Ca, Mg and Mn because these nutrients are considered to be transported to a significant extent by transpirational flow (solute transport) (Marschner and Marschner, 2012; Oliveira et al., 2010). Nutrient uptake into aboveground was calculated as nutrient concentration multiplied with dry weights expressed per m<sup>2</sup> ground area.

#### **Transpiration estimates**

Transpiration was estimated on a leaf area basis using the principles of Fick's law, and  $g_s$  computed with the Jarvis-type (Jarvis, 1976) empirical model previously parameterised for wheat using many measurements of the same cultivars at the AGFACE site (Houshmandfar et al., 2015a). The parameterised Jarvis-type model predicts  $g_s$  by multiplying maximum  $g_s$  (a[CO<sub>2</sub>]: 0.823 (±0.250),  $e[CO_2]$ : 0.529 (±0.127) mol m<sup>-2</sup> s<sup>-1</sup>) measured under optimum conditions with functions of the main microclimate and phenological variables including temperature, vapour pressure deficit (VPD), soil moisture content, time of day, leaf aging, and photosynthetically active radiation (PAR), parameterised separately for ambient and elevated [CO<sub>2</sub>] growing conditions (Houshmandfar et al., 2015a). Microclimatic variables were taken from a continuously logging (15-min intervals) meteorological station located in the AGFACE site, except for the 143<sup>rd</sup> day after germination when average daily weather observations were taken from the nearby (<20 km) meteorological station # 079028 (Australian Bureau of Meteorology). The soil water content was omitted because soil water was regarded as nonlimiting during the measurement period of this experiment (an exceptionally high rainfall growing season for the site, Table 1). VPD of air was calculated using temperature and relative humidity as described by Monteith and Unsworth (2013) and ranged from 0.005 to 3.993 kPa. Stomatal conductance was calculated on a 15-min average basis. The equation (transpiration =  $g_s \times VPD$ ) described by Hunt et al. (1985) was used to calculate the estimates for transpiration rates (Wright et al., 2012). Leaf-level transpiration was estimated separately for each replicate subplot from germination until the time of nutrient data collection, i.e. from germination until DC31, DC34, DC65 or DC69. Transpiration was then (approximately) scaled up to stand-level by multiplying leaf-level transpiration with LAI (Wright

et al., 2012). The air-temperature-based VPD was used as an approximation rather than leaf-temperature-based VPD (Wright et al., 2012). Transpiration was expressed as mm.

**Table 1** Average values for temperature and solar radiation, as well as rainfall amount from 2007 to2016.

	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016
	2007	2008	2005	2010	2011	2012	2013	2014	2015	2010
Annual average minimum air temperature (°C)	8.0	6.7	7.6	7.3	7.4	6.8	7.5	7.3	7.3	8.0
Annual average maximum air temperature (°C)	22.4	21.4	22.2	20.8	21.1	21.5	21.7	22.5	22.3	21.6
Annual rainfall (mm)	428.9	335.4	399.8	559.4	507.0	287.2	378.0	221.8	269.8	550.4
Rainfall during the growing season (mm)	164.0	166.2	266.6	268.6	215.4	174.8	320.8	115.0	125.2	326.4
Annual average solar radiation (MJ m <sup>-2</sup> )	16.3	15.6	16.3	18.2	15.3	16.0	15.8	16.2	16.1	15.8

#### **APSIM model**

APSIM-Wheat (v 7.8) (Holzworth et al., 2014) was used to simulate monthly transpiration and N uptake values in cv. Yitpi and cv. Scout from germination until approximately DC69 under a[CO<sub>2</sub>] (as per the ambient [CO<sub>2</sub>] in the growing season) and e[CO<sub>2</sub>] (550 µmol mol<sup>-1</sup>) in ten consecutive wheat seasons (2007-2016). Ambient [CO<sub>2</sub>] was set to 360 µmol mol<sup>-1</sup> for 2007 and 2008, 370 µmol mol<sup>-1</sup> for 2009 and 2010, 380 µmol mol<sup>-1</sup> for 2011 and 2012, 390 µmol mol<sup>-1</sup> for 2013 and 2014, and 400 µmol mol<sup>-1</sup> for 2015 and 2016, the approximate values corresponding to changes in the ambient [CO<sub>2</sub>] in the growing seasons. The model was run for ten years to broaden the range of the environmental conditions to test if the relationships remain unchanged by variations in the growing seasons. In APSIM-Wheat, e[CO<sub>2</sub>] impacts upon simulated growth and resource use via changes to radiation use efficiency (RUE), transpiration efficiency (TE) and the critical N concentration (CNC) (Reyenga et al., 1999). To capture CO<sub>2</sub> effects on RUE, and interactions with temperature, the model scales RUE using the ratio of the light-limited photosynthetic response at the e[CO<sub>2</sub>] to that at 350 µmol mol<sup>-1</sup>. The responses of TE and leaf CNC to e[CO<sub>2</sub>] are assumed to be linear with changes of +37 and -7%, respectively, for a doubling of [CO<sub>2</sub>] to 700 µmol mol<sup>-1</sup> (O'Leary et al., 2015). Actual transpiration is indirectly reduced under e[CO<sub>2</sub>] through the gain in TE.

The experimental site was defined by its soil type (Table 2), and daily weather conditions (solar radiation, maximum and minimum temperature and rainfall, Table 1) which were mostly taken from the continuously logging meteorological station located in the AGFACE site. When on-site measurements were unavailable, daily weather observations were taken from the nearby meteorological station *#* 079028, extracted from the SILO climate data archive (Jeffrey et al., 2001). Additional experimental site related characteristics are found in O'Leary et al. (2015). Cultivar-specific parameters used to define wheat growth in the APSIM simulation are listed in Table 3. No change to the species parameters and the module source code was made.

Depth	BD	Air dry	LL	DUL	SAT	OC	APSIM KL	APSIM XF
(cm)	(g cc <sup>-1</sup> )	(mm mm <sup>-1</sup> )	(%)	(day⁻¹)	(0-1)			
0-10	1.14	0.15	0.20	0.39	0.46	1.248	0.06	1.00
10-20	1.30	0.18	0.23	0.40	0.47	0.708	0.06	1.00
20-40	1.37	0.25	0.27	0.42	0.48	0.354	0.04	1.00
40-60	1.40	0.27	0.30	0.43	0.47	0.177	0.02	0.80
60-80	1.40	0.28	0.33	0.45	0.47	0.089	0.02	0.80
80-100	1.40	0.30	0.35	0.45	0.47	0.044	0.02	0.60
100-120	1.40	0.32	0.36	0.45	0.47	0.022	0.02	0.60
120-140	1.40	0.33	0.37	0.45	0.47	0.011	0.02	0.20
140-160	1.40	0.34	0.37	0.45	0.47	0.011	0.02	0.20
160-180	1.40	0.34	0.37	0.45	0.47	0.011	0.02	0.20

**Table 2** Soil profile data describing the bulk density (BD), air dry, crop lower limit (LL), drained upper limit (DUL), saturation (SAT), organic carbon (OC), and the water availability coefficients of KL and XF.

**Table 3** Key parameters used to define cultivar-specific settings in APSIM-Wheat. <sup>a</sup> The thermal time from emergence until end of juvenile is influenced by the number of cumulative vernalising days during the period. <sup>b</sup> The phase from end of juvenile until floral initiation is influenced by the photoperiod sensitivity.

Cultivar parameters	Yitpi	Scout
Sensitivity to vernalisation <sup>a</sup>	1.5	1.8
Sensitivity to photoperiod <sup>b</sup>	3.0	3.5
Kernel number per stem weight at the beginning of grain filling (g)	25	25
Potential daily grain filling rate (g grain <sup>-1</sup> day <sup>-1</sup> )	0.002	0.002
Grain growth rate from flowering to grain filling (g grain $^1$ day $^1$ )	0.001	0.001
Maximum grain size (g)	0.041	0.041
Thermal time from start grain filling to maturity (*C days)	545	550
Thermal time from floral initiation to flowing (*C days)	555	555
Thermal time needed in anthesis phase (°C days)	120	120
Thermal time needed from sowing to end of juvenile (°C days)	400	400
Maximum root depth (cm)	180	180

#### Statistical analyses

Analysis of variance for the effects of  $CO_2$ , cultivar, and their interaction on total transpiration and nutrient uptake into aboveground biomass were performed using split plot module ( $CO_2 \times$  cultivar treatment structure) separately for each growth stage with "agricolae" package (de Mendiburu and de Mendiburu, 2016) in R software (v 3.0.3) (R Core Team, 2000). No cultivar or cultivar by  $CO_2$ interaction effect was found to be statistically significant, therefore, we only report the  $CO_2$ -driven effects.

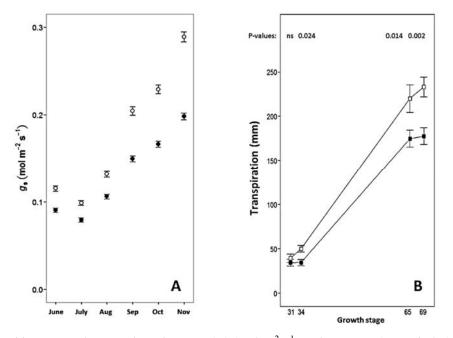
Simple regression analyses with R software were used to assess the relationship between transpiration and nutrient uptake under ambient and elevated [CO<sub>2</sub>]. Significance of differences between the fitted

coefficients (slopes and intercepts) of the regression lines under ambient and elevated [CO<sub>2</sub>] were evaluated using 95% confidence intervals calculated from the standard error (Diem and Seldrup, 1982). Similar to the analysis of variance, we did not find a significant difference between the two cultivars and therefore (for the regression analysis only) data for both cultivars were pooled, increasing the degrees of freedom (from 14 to 30). Graphs were produced using "ggplot2" package (Wickham, 2009) of R software.

#### Results

#### FACE experiment - estimated g<sub>s</sub> and transpiration

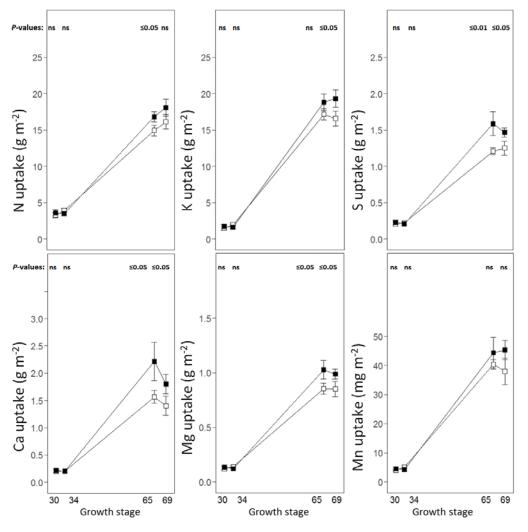
Lower mean  $g_s$  was estimated for plants grown under  $e[CO_2]$  than under  $a[CO_2]$  conditions (Fig. 1), and the difference was smaller earlier than later in the growing season. Elevated  $[CO_2]$  decreased  $g_s$  by a mean of 27% across all estimated values over the study period. This translated into a 31% (15 mm) lower total transpiration under  $e[CO_2]$  at DC34, 21% (45 mm) at DC65, and 24% (56 mm) at DC69 (Fig. 1). Total transpiration estimates, from germination until DC69, were 233.1 (±11.1) and 177.4 (±9.3) mm under ambient and elevated  $[CO_2]$ , respectively.



**Fig. 1** Monthly-averaged stomatal conductance  $(g_s) \pmod{m^{-2} s^{-1}}$  under  $a[CO_2]$  (open circles) and  $e[CO_2]$  (filled circles) estimated using a Jarvis-type model developed at this site (Houshmandfar et al., 2015a) (A). Cumulative transpiration estimates (±standard error) from germination until DC31, DC34, DC65 and DC69 in wheat grown under FACE conditions ( $a[CO_2]$ : open squares;  $e[CO_2]$ : filled squares) (B). Each data point represents a mean of 4 replicates × 2 cultivars (±standard error). Transpiration was estimated as  $g_s$  multiplied with VPD and LAI.

#### FACE experiment - nutrient uptake

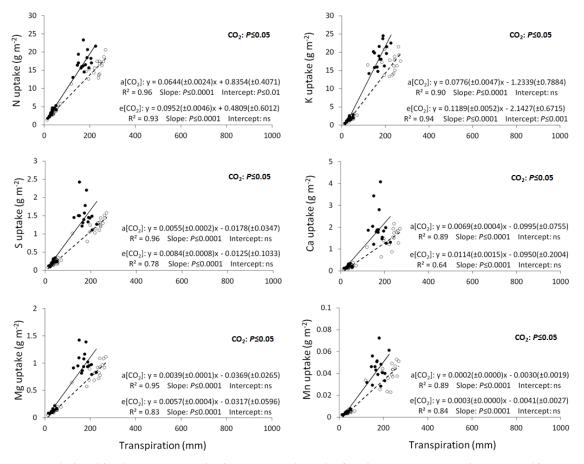
Total nutrient uptake into aboveground biomass was higher in plants grown under  $e[CO_2]$  than under  $a[CO_2]$  when measured at later stages of development, i.e. DC65 and DC69 (albeit not always significant) (Fig. 2). At DC31 and DC34 (early growth stages), this difference was not apparent (Fig. 2).



**Fig. 2** Nutrient uptake into aboveground biomass from germination until DC31, DC34, DC65 and DC69 in wheat grown under FACE conditions (a[CO<sub>2</sub>]: open squares; e[CO<sub>2</sub>]: filled squares). Each data point represents a mean of 4 replicates plot  $\times$  2 cultivars (±standard error).

#### FACE experiment - relationship between transpiration and nutrient uptake

Transpiration (mm) was positively correlated with uptake  $m^{-2}$  of N, K, S, Ca, Mg, and Mn under both ambient and elevated CO<sub>2</sub> conditions (Fig. 3). The coefficient of determinations ( $R^2$ ) were 0.96 and 0.93 for N, 0.90 and 0.94 for K, 0.96 and 0.78 for S, 0.89 and 0.64 for Ca, 0.95 and 0.83 for Mg, and 0.89 and 0.84 for Mn, under ambient and elevated CO<sub>2</sub>, respectively (Fig. 3). The slopes of the relationships were consistently steeper under  $e[CO_2]$  than under  $a[CO_2]$  and this difference was significant for all nutrients ( $P \le 0.05$ ). Nutrient uptake per unit water transpired was on average 50% higher under  $e[CO_2]$  than  $a[CO_2]$ .



**Fig. 3** Relationships between transpiration (mm) and uptake for nitrogen (N), potassium (K), sulfur (S), calcium (Ca), magnesium (Mg), and manganese (Mn) under  $a[CO_2]$  (dashed lines and open circles) and  $e[CO_2]$  (bold lines and filled circles). ±: standard error for the estimated parameters (slope and intercept of the linear relationship). ns: not significant. Each data point represents a replicate subplot (n=32).

#### **APSIM study – simulated transpiration and N uptake**

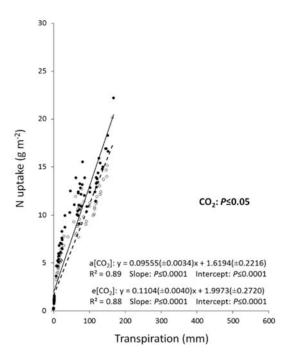
Differences between simulated values for transpiration under ambient and elevated  $[CO_2]$  were small in any of the studied years from 2007 to 2016 (Table 4). Total N uptake into aboveground biomass from germination until approximately DC69 was 20% higher in 2007, 24% in 2008, 21% in 2009, 17% in 2010, 18% in 2011, 21% in 2012, 20% in 2013, 16% in 2014, 13% in 2015, and 17% in 2016 in plants grown under  $e[CO_2]$  than under  $a[CO_2]$ , respectively (Table 4).

	-		,			
Simulated year	Transpiratio	on (mm)	Nitrogen uptake (g m <sup>-2</sup> )			
	a[CO <sub>2</sub> ]	e[CO <sub>2</sub> ]	a[CO <sub>2</sub> ]	e[CO <sub>2</sub> ]		
2007	119.1±0.9	121.5±0.7	11.2±0.0	13.5±0.1		
2008	113.0±2.1	117.0±1.7	10.3±0.4	12.8±0.4		
2009	144.6±4.8	148.4±3.0	14.4±0.7	17.5±0.6		
2010	115.2±2.0	118.2±0.9	10.7±0.6	12.6±1.2		
2011	126.5±1.9	128.7±1.4	13.2±0.1	15.6±0.2		
2012	117.9±2.7	122.2±2.3	11.2±0.6	13.6±0.7		
2013	139.6±9.0	145.0±6.9	13.0±0.9	15.7±0.8		
2014	91.3±0.4	92.5±0.3	9.1±0.1	10.6±0.2		
2015	101.0±0.7	102.1±0.6	11.1±0.1	12.6±0.2		
2016	139.8±7.4	142.1±5.5	13.5±1.1	15.8±1.0		

**Table 4** APSIM-simulated values of transpiration (mm) and N uptake (g  $m^{-2}$ ) from germination until approximately DC 69. ±: standard error for the difference between simulated values for the two studied cultivars (cv. Yitpi and cv. Scout).

#### **APSIM study - relationship between transpiration and N uptake**

The simulated transpiration and N uptake were positively correlated under ambient and elevated  $CO_2$  conditions (Fig. 4). The slope of the relationship under  $e[CO_2]$  was steeper than under  $a[CO_2]$ : N uptake per unit water transpired was approximately 15% higher under  $e[CO_2]$  than  $a[CO_2]$  ( $P \le 0.05$ ) (Fig. 4).



**Fig. 4** Relationship between simulated transpiration (mm) and nitrogen (N) uptake (expressed as g m<sup>-2</sup> ground area) under a[CO<sub>2</sub>] (dashed line and open circles) and e[CO<sub>2</sub>] (bold line and filled circles), simulated monthly average values using APSIM modeling frame work (2007- 2016).  $\pm$ : standard error for the estimated parameters (slope and intercept of the linear relationship). Difference between for the

slopes of the relationships under ambient and elevated  $CO_2$  was significant at  $P \le 0.05$ . The difference was not significant for intercepts (n=100).

#### Discussion

The Jarvis-type empirical model has been widely used for estimating  $g_s$  in transpiration studies (Jarvis, 1976; Stewart, 1988; Wright et al., 2012). We employed the Jarvis model to allow estimation from repeated measurement days to the whole growing season. The Jarvis model was parameterised previously for wheat under ambient and elevated CO<sub>2</sub> growing conditions at the AGFACE site, employing the same set of cultivars (cv. Yitpi and cv. Scout) (Houshmandfar et al., 2015a). Stomatal conductance was on average 27% lower under elevated than under ambient CO<sub>2</sub>. This is in agreement with CO<sub>2</sub>-driven decreases of between 18 to 30% reported by earlier FACE trials measuring *in situ*  $g_s$  in wheat, depending on intraspecific variations and environmental conditions under which plants were grown or  $g_s$  measurements were collected (Garcia et al., 1998; Houshmandfar et al., 2016; McGrath and Lobell, 2013; Wall et al., 2000).

The difference between estimated  $g_s$  under ambient and elevated CO<sub>2</sub> was smaller earlier than later in the growing season. It has been shown in previous papers that wheat grown under e[CO<sub>2</sub>] can have increased stomatal sensitivity to environmental factors (Bunce, 2004; Houshmandfar et al., 2015a), in particular response functions to temperature and VPD in a Jarvis model changed (Houshmandfar et al. 2015a). Temperature and VPD are likely to have higher values during the late than early growing season, which can explain the increasing difference in  $g_s$ .

The simple approach used to upscale from leaf-level  $g_s$  to an estimate of canopy transpiration (multiplying  $g_s$  with VPD and LAI; Wright et al 2012) ignores within-canopy differences in stomatal responses and micro-environment as well as potential boundary layer effects, and is therefore likely to overestimate transpiration. We have no independent transpiration to further evaluate our estimates directly, but a study on soybean in FACE showed that stand-level evapotranspiration scaled well with  $g_s$  measured on upper canopy leaves, implying strong coupling between canopy and atmosphere and consequently high boundary layer conductance (Bernacchi et al., 2007). Since soybean canopies are denser than dryland wheat, and the AGFACE site sees average wind speeds, it is a reasonable assumption that the wheat canopy in our study was at least as well coupled to the atmosphere and strong bias by boundary layer effects (as e.g. in dense forest canopies; Kauwe et al. (2013)), is unlikely. Calculating TE for biomass production with our transpiration estimates resulted in about 4 g dry aboveground biomass L<sup>-1</sup> transpiration (using anthesis biomass data for ambient CO<sub>2</sub> reported in Houshmandfar et al. (2016)). Given that 2013 was a very high rainfall season at the site, this compares well to corresponding values between 4-6 g L<sup>-1</sup> determined at anthesis for a number of Australian wheat cultivars under non-limiting water conditions (Fig. 4 in Fletcher and Chenu (2015)).

Uncertainties in upscaling aside, our focus was on the comparison between ambient and elevated  $CO_2$  grown wheat, and the estimated 23% decrease for stand-level transpiration under e[ $CO_2$ ] compared to

 $a[CO_2]$  is in good agreement with reports in the literature that suggest up to 22% lower stand-level transpiration in wheat grown under FACE, depending on growing conditions (Hunsaker et al., 1996; Hunsaker et al., 2000; Leakey et al., 2009; Tausz Posch et al., 2013). The soybean FACE study cited above also showed that differences in transpiration between ambient and elevated CO<sub>2</sub> were governed by differences in  $g_s$ , but not by changes in LAI or canopy structure (Bernacchi et al., 2007). Increasing [CO<sub>2</sub>] has been shown to cause partial stomatal closure, which reduces transpiration per unit of leaf (e.g. Wall et al. (2000)), and in the above mentioned reports, per unit ground area. Conversely, CO<sub>2</sub> stimulation of growth can result in larger plants with higher LAI, which would tend to increase stand-level transpiration (Kimball et al., 1995; Rosenberg et al., 1990), so that the actual change depends mainly on the relative magnitude of effects on leaf level transpiration and LAI. Some studies found that the leaf area effect may dominate, especially in dry soils where e[CO<sub>2</sub>] grown wheat could produce greater LAI (Samarakoon et al., 1995).

APSIM simulated whole-season water use and wheat yield of three previous seasons at AGFACE well, but results for intermediate growth stages, as in this study, were less accurate (O'Leary et al., 2015). Discrepancies of transpiration assessments by different methods warrant further investigation, but for the purpose of this study the differences between ambient and elevated CO<sub>2</sub> were more important. In contrast to the transpiration estimates extrapolated from leaf level measurements, our APSIM simulations suggested no considerable differences between the transpiration rates in plants grown under ambient and elevated  $[CO_2]$ . This result is in line with O'Leary et al. (2015) who tested various modeling frameworks including APSIM-Wheat with AGFACE data under various environmental conditions (e.g. differential time of sowing and watering regimes) in 2007, 2008, and 2009 growing seasons. Their results suggested that APSIM overestimated stimulation of LAI by e[CO<sub>2</sub>], especially during early growing stages, with the result that although measured water use during the growing season was reduced by  $e[CO_2]$ , the simulation by APSIM-Wheat showed no such reduction (O'Leary et al., 2015). For the purpose of this paper the important question was whether the simulation by APSIM indicates a change for the transpiration-nutrient uptake relationship. An overestimation of transpiration under  $e[CO_2]$ would only give false negative, but not false positive results, that is, the disparity between the relationships under ambient and elevated [CO<sub>2</sub>] may be underestimated.

Total nutrient uptake into aboveground biomass was higher in plants grown under  $e[CO_2]$  than under  $a[CO_2]$  (albeit not always significant) in the AGFACE experiment. This result was consistent with APSIM output where higher N uptake into aboveground biomass was simulated under elevated than under ambient  $[CO_2]$  in all the simulated years with an acceptable absolute error for 2013. Tissue concentration of nutrients was mostly, albeit not always significantly, lower in plants grown under  $e[CO_2]$  (e.g. meta-analyses by Loladze (2002), Loladze (2014), and McGrath and Lobell (2013)) but plants take up more nutrients on an area basis because of increased biomass production (Adam et al., 2000; Brooks et al., 2000; O'Leary et al., 2015; Wechsung et al., 1995). In this particular growing season (2013) nutrient concentrations, for the most part, were not significantly lower (data not shown), but

over multiple years the same tendency towards lower concentrations was observed at this site (e.g. Walker et al. (2016), Buchner et al. (2015), Fernando et al. (2014), and Panozzo et al. (2014)).

The nutrient uptake per unit water transpired was higher under  $e[CO_2]$  than under  $a[CO_2]$ . Although we have not measured xylem nutrient concentrations in this study directly, our results indicate that on average nutrient concentrations in the transpiration stream would be greater under  $e[CO_2]$ , assuming nutrients are transported in the transpiration stream. This is in apparent contrast with our earlier work reporting  $e[CO_2]$  decreases both transpiration flow and concentrations of nutrients in the xylem sap of wheat (Houshmandfar et al., 2015b). In that study, Ca and Mg concentrations in the xylem were evaluated at anthesis stage (DC 60 and DC 69). That result was also supported by Li et al. (2016), whose measurements taken ten days after DC 65 also demonstrated that xylem sap concentrations of Ca, Mg, and K were lower in plants grown under  $e[CO_2]$  than under  $a[CO_2]$ . It is important to note that those results may be specific to the phenological stage when the measurements were done - around or just after anthesis, which corresponds to end of the phenological period investigated in this present paper. Our results here imply that such lower nutrient concentrations in the xylem stream at the later stage have been offset during earlier growth, possibly up to anthesis, to arrive at an overall greater uptake per unit transpiration.

There are a number of possible mechanisms to modify the effect of e[CO<sub>2</sub>] on nutrient uptake during phenological development. It is of vital importance for plant adaptation to respond flexibly to changes in environmental conditions (Forde and Lorenzo, 2001; Robinson, 1994). Although reports about root growth under e[CO<sub>2</sub>] are highly variable in details (e.g. Burkart et al. (2004) and Pacholski et al. (2015)), root biomass is generally stimulated by e[CO<sub>2</sub>] in line with shoot biomass. Earlier in the season, this may allow good access to soil nutrients and high uptake rates. Nutrient concentrations in soil and plant tissues act as signals continuously modifying lateral and seminal root formation (Lopez-Bucio et al., 2003; Morgan and Connolly, 2013). There are some suggestions that e[CO<sub>2</sub>] stimulates lateral root growth preferentially over the elongation of primary roots, leading to highly branched, shallower root system architecture (Burkart et al. (2004) and Pacholski et al. (2015)). Such changes can make the root systems less efficient in soil exploration and thus nutrient uptake (Berntson, 1994; Pritchard and Rogers, 2000; Taub and Wang, 2008), and this may only become manifest towards the end of the season when exploration of deeper soil layers becomes more important.

Transpiration and nutrient uptake were strongly correlated. This supports the literature, e.g. as reviewed in Taub and Wang (2008), suggesting that limitations to the transpiration-driven mass flow rate of nutrients due to decreased  $g_s$  is a contributing factor to the a decrease in nutrient concentration under e[CO<sub>2</sub>]. If this reduction resulting from decreased transpirational flow was the only mechanism, the nutrient uptake per unit water transpired should not have been different under ambient and elevated CO<sub>2</sub> conditions, and in the graphs (Fig. 3 and 4) measurements from e[CO<sub>2</sub>] and a[CO<sub>2</sub>] grown plants should be part of the same relationship. Because in our data the nutrient uptake per unit water transpired was consistently higher under e[CO<sub>2</sub>] than a[CO<sub>2</sub>], we conclude that mechanisms other than mass flow of nutrients are involved. On average, these mechanisms increase nutrient uptake into the transpiration stream, and may therefore to some extent mitigate the decrease in transpiration. Such mechanisms could be associated with changes in root system architecture or function (Berntson, 1994; Pritchard and Rogers, 2000; Taub and Wang, 2008).

#### Conclusions

We used a FACE experiment and the APSIM modelling framework with wheat to investigate potential changes of the relationship between nutrient uptake and transpiration rates under  $e[CO_2]$  to test whether limited transpiration-driven mass flow of nutrients can explain nutrient decline under  $e[CO_2]$ . Our results suggest that transpiration and nutrient uptake of N, K, S, Ca, Mg and Mn are correlated under  $e[CO_2]$ , but that on average across the active growing season nutrient uptake per unit water transpired is higher in plants grown under  $e[CO_2]$  than  $a[CO_2]$ . We therefore concluded that limited transpiration-driven mass flow of nutrients to decreases in nutrient concentrations under  $e[CO_2]$ , but cannot solely account for the overall, more complex relationship between plant nutrition and  $e[CO_2]$ .

#### Acknowledgements

The AGFACE facility is jointly run by the University of Melbourne and the Victorian Department of Economic Development, Jobs, Transport and Resources (Agriculture Victoria) and received funding support from the Australian Commonwealth Department of Agriculture, the Grain Research and Development Corporation (GRDC), and the Australian Research Council (ARC). We gratefully acknowledge the contribution of the AGFACE field team lead by Russel Argall for agronomic trial management, Mahabubur Mollah for operating the CO<sub>2</sub> enrichment technology, Najib Ahmadi for laboratory management, and Chao Chen for her help on the APSIM simulations. AH received a Melbourne International Research Scholarship.

#### References

- Adam NR, Wall GW, Kimball BA, Pinter PJ, LaMorte RL, Hunsaker DJ, Adamsen FJ, Thompson T, Matthias AD, Leavitt SW and Webber AN (2000) Acclimation response of spring wheat in a free-air CO2 enrichment (FACE) atmosphere with variable soil nitrogen regimes. 1. Leaf position and phenology determine acclimation response. *Photosynth Res* 66:65-77.
- Ainsworth EA, Beier C, Calfapietra C, Ceulemans R, Durand-Tardif M, Farquhar GD, Godbold DL, Hendrey GR, Hickler T, Kaduk J, Karnosky DF, Kimball BA, Koerner C, Koornneef M, Lafarge T, Leakey ADB, Lewin KF, Long SP, Manderscheid R, Mcneil DL, Mies TA, Miglietta F, Morgan JA, Nagy J, Norby RJ, Norton RM, Percy KE, Rogers A, Soussana JF, Stitt M, Weigel HJ and White JW (2008) Next generation of elevated [CO2] experiments with crops: a critical investment for feeding the future world. *Plant Cell Environ* **31**:1317-1324.
- Ainsworth EA and Long SP (2005) What have we learned from 15 years of free-air CO2 enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy. *New Phytol* **165**:351-371.
- Ainsworth EA and Rogers A (2007) The response of photosynthesis and stomatal conductance to rising [CO2]: mechanisms and environmental interactions. *Plant Cell Environ* **30**:258-270.

- Asseng S, Jamieson P, Kimball B, Pinter P, Sayre K, Bowden J and Howden S (2004) Simulated wheat growth affected by rising temperature, increased water deficit and elevated atmospheric CO 2. *Field Crops Research* **85**:85-102.
- BassiriRad H, Gutschick VP and Lussenhop J (2001) Root system adjustments: regulation of plant nutrient uptake and growth responses to elevated CO2. *Oecologia* **126**:305-320.
- Bernacchi CJ, Kimball BA, Quarles DR, Long SP and Ort DR (2007) Decreases in stomatal conductance of soybean under open-air elevation of [CO2] are closely coupled with decreases in ecosystem evapotranspiration. *Plant Physiology* **143**:134-144.
- Berntson GM (1994) Modeling Root Architecture Are There Tradeoffs between Efficiency and Potential of Resource Acquisition. *New Phytol* **127**:483-493.
- Bray RH and Kurtz LT (1945) Determination of Total, Organic, and Available Forms of Phosphorus in Soils. *Soil Sci* **59**:39-45.
- Brooks TJ, Wall GW, Pinter PJ, Kimball BA, LaMorte RL, Leavitt SW, Matthias AD, Adamsen FJ, Hunsaker DJ and Webber AN (2000) Acclimation response of spring wheat in a free-air CO2 enrichment (FACE) atmosphere with variable soil nitrogen regimes. 3. Canopy architecture and gas exchange. *Photosynth Res* **66**:97-108.
- Buchner P, Tausz M, Ford R, Leo A, Fitzgerald GJ, Hawkesford MJ and Tausz-Posch S (2015) Expression patterns of C-and N-metabolism related genes in wheat are changed during senescence under elevated CO 2 in dry-land agriculture. *Plant Science* **236**:239-249.
- Bunce JA (2004) Carbon dioxide effects on stomatal responses to the environment and water use by crops under field conditions. *Oecologia* **140**:1-10.
- Burkart S, Manderscheid R and Weigel H-J (2004) Interactive effects of elevated atmospheric CO 2 concentrations and plant available soil water content on canopy evapotranspiration and conductance of spring wheat. *European Journal of Agronomy* **21**:401-417.
- Canadell JG, Le Quere C, Raupach MR, Field CB, Buitenhuis ET, Ciais P, Conway TJ, Gillett NP, Houghton RA and Marland G (2007) Contributions to accelerating atmospheric CO(2) growth from economic activity, carbon intensity, and efficiency of natural sinks. *P Natl Acad Sci USA* **104**:18866-18870.
- Conroy J and Hocking P (1993) Nitrogen Nutrition of C-3 Plants at Elevated Atmospheric Co2 Concentrations. *Physiol Plantarum* **89**:570-576.
- Cramer MD and Hawkins HJ (2009) A physiological mechanism for the formation of root casts. *Palaeogeogr Palaeocl* **274**:125-133.
- de Mendiburu F and de Mendiburu MF (2016) Package 'agricolae'. p^pp.
- Del Pozo A, Perez P, Gutierrez D, Alonso A, Morcuende R and Martinez-Carrasco R (2007) Gas exchange acclimation to elevated CO2 in upper-sunlit and lower-shaded canopy leaves in relation to nitrogen acquisition and partitioning in wheat grown in field chambers. *Environ Exp Bot* **59**:371-380.
- Diem K and Seldrup J (1982) Geigy Scientific Tables. Vol. 2. Introduction to Statistics. Statistical Tables. Mathematical Formulae. *Geigy Scientific Tables Vol 2 Introduction to Statistics Statistical Tables Mathematical Formulae*.
- Fernando N, Panozzo J, Tausz M, Norton RM, Fitzgerald GJ, Myers S, Nicolas ME and Seneweera S (2014) Intra-specific variation of wheat grain quality in response to elevated [CO 2] at two sowing times under rain-fed and irrigation treatments. *Journal of Cereal Science* 59:137-144.
- Fitzgerald GJ, Tausz M, O'Leary G, Mollah MR, Tausz-Posch S, Seneweera S, Mock I, Löw M, Partington DL and McNeil D (2016) Elevated atmospheric [CO2] can dramatically increase wheat yields in semi-arid environments and buffer against heat waves. *Global change biology*.
- Fletcher A and Chenu K (2015) Change in biomass partitioning and transpiration efficiency in Australian wheat varieties over the last decades, in *17th Australian Agronomy Conference*, Australian Society of Agronomy. p^pp 394-397.
- Forde B and Lorenzo H (2001) The nutritional control of root development. *Plant Soil* 232:51-68.

- Garcia RL, Long SP, Wall GW, Osborne CP, Kimball BA, Nie GY, Pinter PJ, Lamorte RL and Wechsung F (1998) Photosynthesis and conductance of spring-wheat leaves: field response to continuous free-air atmospheric CO2 enrichment. *Plant Cell Environ* **21**:659-669.
- Hochman Z, Holzworth D and Hunt J (2009) Potential to improve on-farm wheat yield and WUE in Australia. *Crop and Pasture Science* **60**:708-716.
- Hogy P, Wieser H, Kohler P, Schwadorf K, Breuer J, Franzaring J, Muntifering R and Fangmeier A (2009)
   Effects of elevated CO2 on grain yield and quality of wheat: results from a 3-year free-air CO2 enrichment experiment. *Plant Biology* **11**:60-69.
- Holzworth DP and Huth NI (2011) Simple software processes and tests improve the reliability and usefulness of a model. *Environmental modelling & software* **26**:510-516.
- Holzworth DP, Huth NI, Zurcher EJ, Herrmann NI, McLean G, Chenu K, van Oosterom EJ, Snow V, Murphy C and Moore AD (2014) APSIM–evolution towards a new generation of agricultural systems simulation. *Environmental Modelling & Software* **62**:327-350.
- Houshmandfar A, Fitzgerald GJ, Armstrong R, Macabuhay AA and Tausz M (2015a) Modelling stomatal conductance of wheat: An assessment of response relationships under elevated CO 2. *Agricultural and Forest Meteorology* **214**:117-123.
- Houshmandfar A, Fitzgerald GJ, Macabuhay AA, Armstrong R, Tausz-Posch S, Löw M and Tausz M (2016) Trade-offs between water-use related traits, yield components and mineral nutrition of wheat under Free-Air CO 2 Enrichment (FACE). *European Journal of Agronomy* **76**:66-74.
- Houshmandfar A, Fitzgerald GJ and Tausz M (2015b) Elevated CO 2 decreases both transpiration flow and concentrations of Ca and Mg in the xylem sap of wheat. *Journal of plant physiology* **174**:157-160.
- Hunsaker D, Kimball B, Pinter Jr P, LaMorte R and Wall G (1996) Carbon dioxide enrichment and irrigation effects on wheat evapotranspiration and water use efficiency. *Transactions of the* ASAE **39**:1345-1355.
- Hunsaker D, Kimball B, Pinter P, Wall G, LaMorte R, Adamsen F, Leavitt S, Thompson T, Matthias A and Brooks T (2000) CO 2 enrichment and soil nitrogen effects on wheat evapotranspiration and water use efficiency. *Agricultural and Forest Meteorology* **104**:85-105.
- Hunt ER, Weber JA and Gates DM (1985) Effects of Nitrate Application on Amaranthus-Powellii Wats
   .3. Optimal Allocation of Leaf Nitrogen for Photosynthesis and Stomatal Conductance. *Plant Physiol* **79**:619-624.
- Hutchinson MF, McIntyre S, Hobbs RJ, Stein JL, Garnett S and Kinloch J (2005) Integrating a global agroclimatic classification with bioregional boundaries in Australia. *Global Ecol Biogeogr* **14**:197-212.
- IPCC (2013) Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change, (Stocker TF, D. Qin, G.-K. Plattner, M. Tignor, S.K. Allen, J. Boschung, A. Nauels, Y. Xia, V. Bex and P.M. Midgley ed), NY, USA. p^pp 1535.
- Isbell RF (2002) The Australian Soil Classification. CSIRO Publishing Collingwood, Australia.
- Jarvis PG (1976) Interpretation of Variations in Leaf Water Potential and Stomatal Conductance Found in Canopies in Field. *Philos T Roy Soc B* **273**:593-610.
- Jeffrey SJ, Carter JO, Moodie KB and Beswick AR (2001) Using spatial interpolation to construct a comprehensive archive of Australian climate data. *Environmental Modelling & Software* **16**:309-330.
- Kauwe MG, Medlyn BE, Zaehle S, Walker AP, Dietze MC, Hickler T, Jain AK, Luo Y, Parton WJ and Prentice IC (2013) Forest water use and water use efficiency at elevated CO2: a model-data intercomparison at two contrasting temperate forest FACE sites. *Global Change Biology* **19**:1759-1779.
- Kimball BA, Pinter PJ, Garcia RL, LaMORTE RL, Wall GW, Hunsaker DJ, Wechsung G and Wechsung F (1995) Productivity and water use of wheat under free-air CO2 enrichment. *Global Change Biology* 1:429-442.

- Kupper P, Rohula G, Saksing L, Sellin A, Lohmus K, Ostonen I, Helmisaari HS and Sober A (2012) Does soil nutrient availability influence night-time water flux of aspen saplings? *Environ Exp Bot* 82:37-42.
- Leakey ADB, Ainsworth EA, Bernacchi CJ, Rogers A, Long SP and Ort DR (2009) Elevated CO2 effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. J Exp Bot **60**:2859-2876.
- Leakey ADB, Bernacchi CJ, Ort DR and Long SP (2006) Long-term growth of soybean at elevated [CO2] does not cause acclimation of stomatal conductance under fully open-air conditions. *Plant Cell Environ* **29**:1794-1800.
- Leakey ADB, Bishop KA and Ainsworth EA (2012) A multi-biome gap in understanding of crop and ecosystem responses to elevated CO2. *Curr Opin Plant Biol* **15**:228-236.
- Li X, Jiang D and Liu F (2016) Soil warming enhances the hidden shift of elemental stoichiometry by elevated CO2 in wheat. *Sci Rep-Uk* **6**.
- Loladze I (2002) Rising atmospheric CO 2 and human nutrition: toward globally imbalanced plant stoichiometry? *Trends in Ecology & Evolution* **17**:457-461.
- Loladze I (2014) Hidden shift of the ionome of plants exposed to elevated CO2 depletes minerals at the base of human nutrition. *Elife* **3**:e02245.
- Long SP, Ainsworth EA, Leakey ADB, Nosberger J and Ort DR (2006) Food for thought: Lower-thanexpected crop yield stimulation with rising CO2 concentrations. *Science* **312**:1918-1921.
- Long SP, Ainsworth EA, Rogers A and Ort DR (2004) Rising atmospheric carbon dioxide: Plants face the future. *Annu Rev Plant Biol* **55**:591-628.
- Lopez-Bucio J, Cruz-Ramirez A and Herrera-Estrella L (2003) The role of nutrient availability in regulating root architecture. *Curr Opin Plant Biol* **6**:280-287.
- Marschner H and Marschner P (2012) *Marschner's mineral nutrition of higher plants*. Academic Press, London ;.
- Masle J, Farquhar GD and Wong SC (1992) Transpiration Ratio and Plant Mineral-Content Are Related among Genotypes of a Range of Species. *Aust J Plant Physiol* **19**:709-721.
- Mcconnaughay KDM, Berntson GM and Bazzaz FA (1993) Limitations to Co2-Induced Growth Enhancement in Pot Studies. *Oecologia* **94**:550-557.
- McDonald EP, Erickson JE and Kruger EL (2002) Can decreased transpiration limit plant nitrogen acquisition in elevated CO2? *Funct Plant Biol* **29**:1115-1120.
- McGrath JM and Lobell DB (2013) Reduction of transpiration and altered nutrient allocation contribute to nutrient decline of crops grown in elevated CO2 concentrations. *Plant, Cell & Environment* **36**:697-705.
- McLeod AR and Long SP (1999) Free-air carbon dioxide enrichment (FACE) in global change research: A review. *Adv Ecol Res* **28**:1-56.
- Mehlich A (1984) Mehlich-3 Soil Test Extractant a Modification of Mehlich-2 Extractant. *Commun Soil Sci Plan* **15**:1409-1416.
- Mollah M, Partington D and Fitzgerald G (2011) Understand distribution of carbon dioxide to interpret crop growth data: Australian grains free-air carbon dioxide enrichment experiment. *Crop Pasture Sci* **62**:883-891.
- Monteith JL and Unsworth MH (2013) Principles of Environmental Physics: Plants, Animals, and the Atmosphere, 4th Edition. *Principles of Environmental Physics: Plants, Animals, and the Atmosphere, 4th Edition*:1-401.
- Morgan J and Connolly E (2013) Plant-soil interactions: nutrient uptake. Nat Educ Knowl 4:2.
- Nowak RS, Ellsworth DS and Smith SD (2004) Functional responses of plants to elevated atmospheric CO2 do photosynthetic and productivity data from FACE experiments support early predictions? *New Phytol* **162**:253-280.
- O'Leary GJ, Christy B, Nuttall J, Huth N, Cammarano D, Stöckle C, Basso B, Shcherbak I, Fitzgerald G and Luo Q (2015) Response of wheat growth, grain yield and water use to elevated CO2 under

a free-air CO2 enrichment (FACE) experiment and modelling in a semi-arid environment. *Global change biology* **21**:2670-2686.

- Pacholski A, Manderscheid R and Weigel H-J (2015) Effects of free air CO 2 enrichment on root growth of barley, sugar beet and wheat grown in a rotation under different nitrogen supply. *European Journal of Agronomy* **63**:36-46.
- Panozzo J, Walker C, Partington D, Neumann N, Tausz M, Seneweera S and Fitzgerald G (2014) Elevated carbon dioxide changes grain protein concentration and composition and compromises baking quality. A FACE study. *Journal of Cereal Science* **60**:461-470.
- Pleijel H and Hogy P (2015) CO2 dose-response functions for wheat grain, protein and mineral yield based on FACE and open-top chamber experiments. *Environmental pollution* **198**:70-77.
- Polley HW, Johnson HB, Tischler CR and Torbert HA (1999) Links between transpiration and plant nitrogen: Variation with atmospheric CO2 concentration and nitrogen availability. *Int J Plant Sci* **160**:535-542.
- Pritchard SG and Rogers HH (2000) Spatial and temporal deployment of crop roots in CO(2)-enriched environments. *New Phytol* **147**:55-71.
- R Core Team R (2000) R language definition. *Vienna, Austria: R foundation for statistical computing*.
- Reyenga PJ, Howden SM, Meinke H and McKeon GM (1999) Modelling global change impacts on wheat cropping in south-east Queensland, Australia. *Environmental modelling & software* **14**:297-306.
- Robinson D (1994) The Responses of Plants to Nonuniform Supplies of Nutrients. *New Phytol* **127**:635-674.
- Rosenberg NJ, Kimball B, Martin P and Cooper C (1990) From climate and CO2 enrichment to evapotranspiration. *Climate change and US water resources*:151-175.
- Russell RS and Barber DA (1960) The Relationship between Salt Uptake and the Absorption of Water by Intact Plants. *Annu Rev Plant Phys* **11**:127-140.
- Samarakoon A, Muller W and Gifford R (1995) Transpiration and leaf area under elevated CO2: effects of soil water status and genotype in wheat. *Functional Plant Biology* **22**:33-44.
- Scholz FG, Bucci SJ, Goldstein G, Meinzer FC, Franco AC and Miralles-Wilhelm F (2007) Removal of nutrient limitations by long-term fertilization decreases nocturnal water loss in savanna trees. *Tree Physiol* 27:551-559.
- Sellin A, Tullus A, Niglas A, Ounapuu E, Karusion A and Lohmus K (2013) Humidity-driven changes in growth rate, photosynthetic capacity, hydraulic properties and other functional traits in silver birch (Betula pendula). *Ecol Res* 28:523-535.
- Stewart JB (1988) Modeling Surface Conductance of Pine Forest. Agr Forest Meteorol 43:19-35.
- Taub D (2010) Effects of rising atmospheric concentrations of carbon dioxide on plants. *Nature Education Knowledge* **3**:21.
- Taub DR and Wang XZ (2008) Why are Nitrogen Concentrations in Plant Tissues Lower under Elevated CO2? A Critical Examination of the Hypotheses. *J Integr Plant Biol* **50**:1365-1374.
- Tausz-Posch S, Norton RM, Seneweera S, Fitzgerald GJ and Tausz M (2013) Will intra-specific differences in transpiration efficiency in wheat be maintained in a high CO2 world? A FACE study. *Physiologia Plantarum* **148**:232-245.
- Tinker PB and Nye PH (2000) Solute Movement in the Rhizosphere. Oxford University Press, USA.
- Walker C, Armstrong R, Panozzo J and Fitzgerald G (2016) Can Nitrogen Management maintain Grain Protein Content of wheat under elevated CO2? A FACE study.
- Wall GW, Adam NR, Brooks TJ, Kimball BA, Pinter PJ, LaMorte RL, Adamsen FJ, Hunsaker DJ, Wechsung G, Wechsung F, Grossman-Clarke S, Leavitt SW, Matthias AD and Webber AN (2000)
   Acclimation response of spring wheat in a free-air CO2 enrichment (FACE) atmosphere with variable soil nitrogen regimes. 2. Net assimilation and stomatal conductance of leaves. *Photosynth Res* 66:79-95.

- Wechsung G, Wechsung F, Wall GW, Adamsen FJ, Kimball BA, Garcia RL, Pinter PJ and Kartschall T (1995) Biomass and growth rate of a spring wheat root system grown in free-air CO2 enrichment (FACE) and ample soil moisture. *J Biogeogr* **22**:623-634.
- Wickham H (2009) ggplot2: Elegant Graphics for Data Analysis. Springer Publishing Company, Incorporated.
- Wright TE, Tausz M, Kasel S, Volkova L, Merchant A and Bennett LT (2012) Edge type affects leaf-level water relations and estimated transpiration of Eucalyptus arenacea. *Tree Physiol* **32**:280-293.
- Zadoks JC, Chang TT and Konzak CF (1974) Decimal Code for Growth Stages of Cereals. Weed Res 14:415-421.
- Zarcinas BA, Cartwright B and Spouncer LR (1987) Nitric-Acid Digestion and Multielement Analysis of Plant-Material by Inductively Coupled Plasma Spectrometry. *Commun Soil Sci Plan* **18**:131-146.
- Zhang Y, Feng L, Wang E, Wang J and Li B (2012) Evaluation of the APSIM-Wheat model in terms of different cultivars, management regimes and environmental conditions. *Canadian Journal of Plant Science* **92**:937-949.