

Acute Myeloid Leukaemia

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Editorial

Acute Myeloid Leukaemia: New Targets and Therapies

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The most common acute hematological malignancy in adults is acute myeloid leukaemia (AML), accounting for more than 80% of cases in patients over 60 years of age [1]. AML is the second most common acute leukaemia in children, accounting for 15–20% of leukaemia cases [2,3]. Morphological classification of AML into eight sub-types (FAB M0–M7) based on the type of cell from which the leukemia developed, and on its degree of maturity, was in use until the end of the last century [4]. A new World Health Organization (WHO) classification introduced four main AML groups, based on cytogenetic abnormalities, and is important to the approach used to treat this disease [5]. In particular, acute promyelocytic leukaemia (APL), a distinct M3 subtype of AML, was once an incurable disease and the use of all-*trans* retinoic acid (ATRA)-based differentiation therapy and anthracycline-based chemotherapy now provides a cure rate above 80% (reviewed in [6]). However, aside from APL, around 90% of older patients (aged \geq 60 years) and more than half of young adult patients (aged $<$ 60 years) die from their disease [7], and AML accounts for the largest number of annual deaths due to leukaemias in the US [2]. The survival of older patients who are unable to tolerate chemotherapy is dismal, at only five to 10 months [8], and complete remission rates for relapsed patients are $<$ 25% [9,10].

The Fms-like tyrosine kinase 3 (FLT3), a class III receptor tyrosine kinase, and FLT3 ligand (FL) play an important role in the proliferation, differentiation, and survival of hematopoietic cells [11]. Mutated FLT3 is expressed in a subset of AML patients and confers a poor prognosis. The most common mutation, which occurs in about 25% of AML patients, is an internal tandem duplication (ITD) in the juxtamembrane region of FLT3, which causes ligand-independent dimerization and constitutive receptor activation [12]. Point mutations in the tyrosine kinase domain (TKD), FLT3-TKD, occur in approximately 7% of AMLs [13]. FLT3 tyrosine kinase inhibitors are well tolerated, as a monotherapy and with intensive chemotherapy, and second generation inhibitors have shown significant promise as a treatment for relapsed and refractory AML. The paper by Mooney and colleagues [14] and the review by Tsapogas and colleagues [15] advance the knowledge of FLT3 and FL. Mooney and colleagues have examined the expression of mRNA FLT3 (mRNA and cell surface protein) by hematopoietic stem cells (HSC) and various progenitor cells (HPC). A sub-population of short-term and long-term HSC express FLT3. As expected, expression by HPC was observed for these cells with lymphoid, granulocytic, and myelomonocytic potential. Regarding FLT3 expression by HSC, Tsapogas provides evidence to support an instructive role of FL signalling at early stages of hematopoiesis, in addition to a role in promoting cell survival and proliferation. The precise pattern of expression of FLT3 by HSC/HPC and the roles of FL in normal hematopoiesis are critical to a better understanding of AML subtypes, the process of disease progression, as well as for the development of therapeutic strategies to target FLT3-mutated AML.

Regarding other kinase inhibitors, T315, an inhibitor of integrin-linked kinase, has been shown to suppress the proliferation of breast and stomach cancer cells and chronic lymphocytic leukaemia cells.

Chiu and colleagues [16] report that this agent is cytotoxic against the AML cell lines HL-60 and THP-1 and primary leukaemia cells from AML patients. T315 also suppresses the growth of THP-1 xenografts. Chiu and colleagues describe aspects of the mechanism of action of T315, in driving apoptosis and autophagic cell death, and suggest further assessment of the use in treating AML and other leukaemias. However, there is the need to modify T315 to increase efficacy and reduce toxicity. Apoptosis can also be induced in human leukaemia cells by the curcumin analogue EF-24. In their study, Skoupa and colleagues [17] investigate the mechanism by which EF-24 induces cell death in K562 cells. They propose that EF-24 may be suitable as an anticancer agent, specifically in cases of drug resistance.

The oncogenic or chromosomal mutations that are present in a patient's AML cells at diagnosis are important to personalized treatment. The best known is a balanced translocation between chromosomes 15 and 17 [t(15;17)(q22;q21)] resulting in the formation of promyelocytic leukemia (PML) and retinoic acid receptor alpha (RAR α) fusion protein [18]. Leukaemic blasts which carry this mutation are susceptible to ATRA-induced cell differentiation. Watts and colleagues [19] report a salutary lesson from the treatment outcome of a case of AML. The case is characterised by a new t(4;15)(q31;q22) translocation, resulting in the expression of the TMEM154-RASGRF1 fusion protein. The patient who was treated with ATRA as a part of a clinical study died from rapid disease progression. An increase in the expression of RAR γ was observed upon treatment of the patient's cells *ex vivo* with ATRA, and they proliferated in response to ATRA and a RAR γ agonist. The disease progression could be related to an increase in RAR γ , which plays a role in hematopoietic stem cell self-renewal and proliferation. Furthermore, there are implications for the use of retinoid-based differentiation therapy for certain cases.

In addition to ATRA, the differentiating agent 1,25-dihydroxyvitamin D (1,25D) has been reported to have beneficial effects in combination therapy for cancer. The paper by Janik and colleagues [20] examines the regulation of the vitamin D receptor (*VDR*) gene by 1,25D and ATRA in blood cells at early stages of their differentiation. ATRA, but not 1,25D, upregulates the expression of *VDR* in human early-stage blood cells. As to early-stage mouse cells, *VDR* is upregulated by 1,25D, but not by ATRA. Hence, *VDR* regulation in humans and mice is different, which is germane to testing combinations of agents for use in differentiation therapy. The findings also bring to attention that the level of expression of *VDR* protein is low in patients' AML blasts that do not respond to 1,25D. The level can be upregulated by ATRA treatment and this is relevant to the combined therapeutic use of ATRA and 1,25D.

There is an urgent need for new treatments for AML, which will include the need to identify new molecules to target. A number of recurrent mutations in AML involve genes concerned with regulating the epigenetic landscape [21]. Gain or loss of function of the gene encoding the EZH2 methyltransferase occurs in various malignancies. Sbirkov and colleagues [22] describe the use of affinity-purification mass spectrometry to identify new partners of EZH2 and potential new non-histone substrates. Of particular note is that EZH2 has a role in regulating translation, via interacting with RNA binding proteins and methylating eEF1A1. eEF1A1 is a component of protein synthesis, highly expressed in tumours, and therapeutic targeting is a possibility in AML. The review by Gbolahan and colleagues [23] focusses on the benefit of immunotherapeutic interventions to the long-term control of AML, including the use of hypomethylating agents. The results from early phase immunotherapeutic interventions, for example, the use of a monoclonal antibody to CD33 which is highly expressed on AML blasts, are encouraging. The interleukin-3 receptor α chain (CD123), Fc γ RI (CD64) and C-type lectin-like molecule 1 are also promising targets. The review considers the use of hypomethylating agents to increase the antigenicity of AML cells and their role as immunomodulatory drugs.

The articles by Player et al., Khan et al., and Almeida et al. describe the outcomes from clinical studies. Player and colleagues [24] examine azacitidine for front-line therapy of patients with AML, by reference to international phase 3 trial data and data from the Austrian Azacitidine registry. The authors report clinically-meaningful improvement in overall survival for patients treated with front-line azacitidine versus conventional regimens, and conclude that azacitidine appears efficacious as a front-line treatment for WHO-AML patients. Khan and colleagues [25] examine the clinical outcomes in patients with *RUNX1*-mutated AML. The response of older patients to treatment with

hypomethylating agents and survival were found to be independent of *RUNX1* status, leading the authors to conclude that future studies should focus on the prognostic value of *RUNX1* mutations relative to co-occurring mutations. Almeida and colleagues [26] examine the outcomes from the various types of treatment for acute erythroleukemia (AEL). AEL typically has a poor prognosis. From a comparison of patients treated with hypomethylating agents, front-line, and intensive chemotherapy, the results provide support to the use of hypomethylating agents to treat AEL. However, high-risk patients treated first-line with hypomethylating agents live just a few months longer (13.3 versus 7.5 for patients treated with intensive chemotherapy).

In summary, there is quite some way to go to extend the success in treating APL to other types of AML. As mentioned above, 90% of older patients die from their disease. In populations of developed countries, cancers are turning into prevalent diseases of the aged. According to epidemiological data, about 80% of cancers are diagnosed in patients older than 55, with the median age at diagnosis above 60 years [27]. This is also the case for AML for which the median age at diagnosis is 67 years [28]. Many older AML patients are not able to receive intensive chemotherapy. Hence, there is the need for new approaches to the treatment of AML, and other malignancies, in elderly patients. New therapeutic targets are important, as is consideration of differentiation therapy, including agents that modify the epigenetic status of cells, combined with gentler chemotherapy. A means of preventing AML relapse and holding in check for life will provide a better quality of life to elderly patients.

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References

1. Pollyea, D.; Kohrt, H.; Medeiros, B. Acute myeloid leukaemia in the elderly: A review. *Br. J. Haematol.* **2011**, *152*, 524–542. [[CrossRef](#)] [[PubMed](#)]
2. O'Donnell, M.; Tallman, M.; Abboud, C.; Altman, J.; Appelbaum, F.; Arber, D.; Attar, E.; Borate, U.; Coutre, S.; Damon, L.; et al. National Comprehensive Cancer, N., Acute myeloid leukemia, version 2.2013. *J. Natl. Compr. Cancer Netw.* **2013**, *11*, 1047–1055. [[CrossRef](#)]
3. Rubnitz, J. How I treat pediatric acute myeloid leukemia. *Blood* **2012**, *119*, 5980–5988. [[CrossRef](#)] [[PubMed](#)]
4. Bennett, J.; Catovsky, D.; Daniel, M.; Flandrin, G.; Galton, D.; Gralnick, H.; Sultan, C. Proposals for the classification of the acute leukaemias. French-American-British (FAB) co-operative group. *Br. J. Haematol.* **1976**, *33*, 451–458. [[CrossRef](#)] [[PubMed](#)]
5. Vardiman, J.; Harris, N.; Brunning, R. The World Health Organization (WHO) classification of the myeloid neoplasms. *Blood* **2002**, *100*, 2292–2302. [[CrossRef](#)] [[PubMed](#)]
6. Lo-Coco, F.; Cicconi, L.; Breccia, M. Current standard treatment of adult acute promyelocytic leukaemia. *Br. J. Haematol.* **2016**, *172*, 841–854. [[CrossRef](#)] [[PubMed](#)]
7. Ferrara, F.; Schiffer, C. Acute myeloid leukaemia in adults. *Lancet* **2013**, *381*, 484–495. [[CrossRef](#)]
8. Dohner, H.; Weisdorf, D.; Bloomfield, C. Acute Myeloid Leukemia. *N. Engl. J. Med.* **2015**, *373*, 1136–1152. [[CrossRef](#)] [[PubMed](#)]
9. Leopold, L.; Willemze, R. The treatment of acute myeloid leukemia in first relapse: A comprehensive review of the literature. *Leuk. Lymphoma* **2002**, *43*, 1715–1727. [[CrossRef](#)] [[PubMed](#)]
10. Greenberg, P.; Lee, S.; Advani, R.; Tallman, M.; Sikic, B.; Letendre, L.; Dugan, K.; Lum, B.; Chin, D.; Dewald, G.; et al. Mitoxantrone, etoposide, and cytarabine with or without valspodar in patients with relapsed or refractory acute myeloid leukemia and high-risk myelodysplastic syndrome: A phase III trial (E2995). *J. Clin. Oncol.* **2004**, *22*, 1078–1086. [[CrossRef](#)] [[PubMed](#)]
11. Rosnet, O.; Schiff, C.; Pébusque, M.; Marchetto, S.; Tonnelle, C.; Toiron, Y.; Birg, F.; Birnbaum, D. Human FLT3/FLK2 gene: CDNA cloning and expression in hematopoietic cells. *Blood* **1993**, *82*, 1110–1119. [[PubMed](#)]

12. Yokota, S.; Kiyoi, H.; Nakao, M.; Iwai, T.; Misawa, S.; Okuda, T.; Sonoda, Y.; Abe, T.; Kahsima, K.; Matsuo, Y.; et al. Internal tandem duplication of the FLT3 gene is preferentially seen in acute myeloid leukemia and myelodysplastic syndrome among various hematological malignancies. A study on a large series of patients and cell lines. *Leukemia* **1997**, *11*, 1605–1609. [[CrossRef](#)] [[PubMed](#)]
13. Callens, C.; Chevret, S.; Cayuela, J.; Cassinat, B.; Raffoux, E.; de Botton, S.; Thomas, X.; Guerc, I.A.; Fegueux, N.; Pigneux, A.; et al. Prognostic implication of FLT3 and Ras gene mutations in patients with acute promyelocytic leukemia (APL): A retrospective study from the European APL Group. *Leukemia* **2005**, *19*, 1153–1160. [[CrossRef](#)] [[PubMed](#)]
14. Mooney, C.; Cunningham, A.; Tsapogas, P.; Toellner, K.-M.; Brown, G. Selective Expression of Flt3 within the mouse hematopoietic stem cell compartment. *Int. J. Mol. Sci.* **2017**, *18*, 1037. [[CrossRef](#)] [[PubMed](#)]
15. Tsapogas, P.; Mooney, C.; Brown, G.; Rolink, A. The cytokine Flt3-ligand in normal and malignant hematopoiesis. *Int. J. Mol. Sci.* **2017**, *18*, 1115. [[CrossRef](#)] [[PubMed](#)]
16. Chiu, C.-F.; Weng, J.-R.; Jadhav, A.; Wu, C.-Y.; Sargeant, A.; Bai, L.-Y. T315 decreases acute myeloid leukemia cell viability through a combination of apoptosis induction and autophagic cell death. *Int. J. Mol. Sci.* **2016**, *17*, 1337. [[CrossRef](#)] [[PubMed](#)]
17. Skoupa, N.; Dolezel, P.; Ruzickova, E.; Mlejnek, P. Apoptosis induced by curcumin analogue EF-24 is neither mediated by oxidative stress related mechanism nor affected by expression of main drug transporters ABCB1 and ABCG2 in human leukemia cells. *Int. J. Mol. Sci.* **2017**, *18*, 2289. [[CrossRef](#)] [[PubMed](#)]
18. Kakizuka, A.; Miller, W.J.; Umesonon, K.; Warrell, R.J.; Frankel, S.; Murty, V.; Dmitrovsky, E.; Evans, R. Chromosomal translocation t(15;17) in human acute promyelocytic leukemia fuses RAR alpha with a novel putative transcription factor, PML. *Cell* **1991**, *66*, 663–674. [[CrossRef](#)]
19. Watts, J.; Perez, A.; Pereira, L.; Fan, Y.-S.; Brown, G.; Vega, F.; Petrie, K.; Swords, R.; Zelent, A. A case of AML characterized by a novel t(4;15)(q31;q22) translocation that confers a growth-stimulatory response to retinoid-based therapy. *Int. J. Mol. Sci.* **2017**, *18*, 1492. [[CrossRef](#)] [[PubMed](#)]
20. Janik, S.; Nowak, U.; Łaszkiwicz, A.; Satyr, A.; Majkowski, M.; Marchwicka, A.; Śniezewski, Ł.; Berkowska, K.; Gabryś, M.; Cebrat, M.; et al. Diverse REGulation of vitamin d receptor gene expression by 1,25-dihydroxyvitamin D and atra in murine and human blood cells at early stages of their differentiation. *Int. J. Mol. Sci.* **2017**, *18*, 1323. [[CrossRef](#)] [[PubMed](#)]
21. Conway O'Brien, E.; Prideaux, S.; Chevassut, T. The epigenetic landscape of acute myeloid leukemia. *Adv. Hematol.* **2014**, *2014*, 103175. [[CrossRef](#)] [[PubMed](#)]
22. Sbirkov, Y.; Kwok, C.; Bhamra, A.; Thompson, A.; Gil, V.; Zelent, A.; Petrie, K. Semi-quantitative mass spectrometry in AML cells identifies new non-genomic targets of the EZH2 methyltransferase. *Int. J. Mol. Sci.* **2017**, *18*, 1440. [[CrossRef](#)] [[PubMed](#)]
23. Gbolahan, O.; Zeidan, A.; Stahl, M.; Abu Zaid, M.; Farag, S.; Paczesny, S.; Konig, H. Immunotherapeutic concepts to target acute myeloid leukemia: Focusin on the role of monoclonal antibodies, hypomethylating agents and the leukemic microenvironment. *Int. J. Mol. Sci.* **2017**, *18*, 1660. [[CrossRef](#)] [[PubMed](#)]
24. Pleyer, L.; Döhner, H.; Dombret, H.; Seymour, J.; Schuh, A.; Beach, C.; Swern, A.; Burgstaller, S.; Stauder, R.; Girschikofsky, M.; et al. Azacitidine for front-line therapy of patients with AML: Reproducible efficacy established by direct comparison of international phase 3 trial data with registry data from the Austrian Azacitidine Registry of the AGMT study group. *Int. J. Mol. Sci.* **2017**, *18*, 415. [[CrossRef](#)] [[PubMed](#)]
25. Khan, M.; Cortes, J.; Kadia, T.; Naqvi, K.; Brandt, M.; Pierce, S.; Patel, K.; Borthakur, G.; Ravandi, F.; Konopleva, M.; et al. Clinical outcomes and co-occurring mutations in patients with RUNX1-mutated acute myeloid leukemia. *Int. J. Mol. Sci.* **2017**, *18*, 1618. [[CrossRef](#)] [[PubMed](#)]
26. Almeida, A.; Prebet, T.; Itzykson, R.; Ramos, F.; Al-Ali, H.; Shammo, J.; Pinto, R.; Maurillo, L.; Wetzel, J.; Musto, P.; et al. Clinical outcomes of 217 patients with acute erythroleukemia according to treatment type and line: A retrospective multinational study. *Int. J. Mol. Sci.* **2017**, *18*, 837. [[CrossRef](#)] [[PubMed](#)]
27. Marosi, C.; Köller, M. Challenge of cancer in the elderly. *ESMO Open* **2016**, *1*. [[CrossRef](#)] [[PubMed](#)]
28. Almeida, A.; Ramos, F. Acute myeloid leukemia in the older adults. *Leuk. Res. Rep.* **2016**, *6*, 1–7. [[CrossRef](#)] [[PubMed](#)]

