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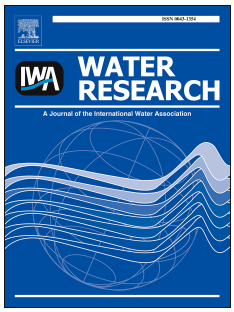
Accepted Manuscript



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PII: S0043-1354(16)30148-8

DOI: 10.1016/j.watres.2016.03.021

Reference: WR 11905

To appear in: *Water Research*

Received Date: 16 November 2015

Revised Date: 7 March 2016

Accepted Date: 8 March 2016

Please cite this article as: Carstea, E.M., Bridgeman, J., Baker, A., Reynolds, D.M., Fluorescence spectroscopy for wastewater monitoring: A review, *Water Research* (2016), doi: 10.1016/j.watres.2016.03.021.

Fluorescence spectroscopy for wastewater monitoring: a review

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Abstract: Wastewater quality is usually assessed using physical, chemical and microbiological tests, which are not suitable for online monitoring, provide unreliable results, or use hazardous chemicals. Hence, there is an urgent need to find a rapid and effective method for the evaluation of water quality in natural and engineered systems and for providing an early warning of pollution events. Fluorescence spectroscopy has been shown to be a valuable technique to characterize and monitor wastewater in surface waters for tracking sources of pollution, and in treatment works for process control and optimization. This paper reviews the current progress in

applying fluorescence to assess wastewater quality. Studies have shown that, in general, wastewater presents higher fluorescence intensity compared to natural waters for the components associated with peak T (living and dead cellular material and their exudates) and peak C (microbially reprocessed organic matter). Furthermore, peak T fluorescence is significantly reduced after the biological treatment process and peak C is almost completely removed after the chlorination and reverse osmosis stages. Thus, simple fluorometers with appropriate wavelength selectivity, particularly for peaks T and C could be used for online monitoring in wastewater treatment works. This review also shows that care should be taken in any attempt to identify wastewater pollution sources due to potential overlapping fluorophores. Correlations between fluorescence intensity and water quality parameters such as biochemical oxygen demand (BOD) and total organic carbon (TOC) have been developed and dilution of samples, typically up to x10, has been shown to be useful to limit inner filter effect. It has been concluded that the following research gaps need to be filled: lack of studies on the on-line application of fluorescence spectroscopy in wastewater treatment works and lack of data processing tools suitable for rapid correction and extraction of data contained in fluorescence excitation-emission matrices (EEMs) for real-time studies.

Key words: fluorescence spectroscopy, wastewater, organic matter, monitoring

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1 Introduction

Environmental monitoring is applied to determine the compliance with ambient and discharge standards and to identify areas with persistent issues for timely and effective remediation (Cahoon and Mallin 2013). Wastewater quality assessment is an essential part of environmental monitoring due to the high anthropogenic impact of treated and untreated discharges on water bodies (Suthar et al. 2010). There are two important aspects of wastewater quality monitoring: the first concerns the detection of pollution events for early warning and rapid remedial responses of water bodies, while the second aspect relates to wastewater treatment works where quality monitoring is required for process control and compliance with regulations at the effluent discharge point (Bourgeois et al. 2001, Michael et al. 2015, Rehman et al. 2015).

The quality of wastewater is generally assessed using physical, chemical and microbiological tests. Among these techniques, reliance is often placed on biological oxygen demand (BOD), chemical oxygen demand (COD) and total organic carbon (TOC) (Bourgeois et al. 2001, Bridgeman et al. 2013). However, these global parameters depend on expensive

or time-consuming methods, offering only snapshots of moments in time (Bourgeois et al. 2001, Chong et al. 2013, Yang et al. 2015a), which makes them unsuitable for online monitoring. Research conducted almost two decades ago (Ahmad and Reynolds 1995, Tartakovsky et al. 1996, Reynolds and Ahmad 1997, Ahmad and Reynolds 1999) has shown that fluorescence spectroscopy could be used for wastewater quality assessment as a tool for discharge detection in natural water systems and for process control in wastewater treatment plants (WwTPs). Fluorescence is the release of energy in the form of light when molecules or moieties, named fluorophores, are excited with a high-energy light source (Lakowicz 2006, Reynolds 2014). The technique has been suggested for its multiple advantages: it is fast, inexpensive, reagentless, requires little sample preparation, is highly sensitive and non-invasive (Reynolds 2003, Hudson et al. 2007, Cao et al. 2009, Henderson et al. 2009, Hambly et al. 2010, Murphy et al. 2011, Chong et al. 2013, Yang et al. 2015a). According to Reynolds (2002) fluorescence monitoring could provide rapid feedback, allowing dynamic, high spatial and temporal resolution studies.

In the past decades, more studies have proved the potential of fluorescence spectroscopy as a monitoring and detection tool in natural and engineered systems. This technique has been used successfully to characterize organic matter in seawater (Coble et al. 1990, Coble 1996, Conmy et al.

2004, Drozdowska 2007), freshwater (Baker 2001, McKnight et al. 2001, Spencer et al. 2007b, Carstea et al. 2009) or estuarine water (Huguet et al. 2009). Also, it has been used to monitor riverine organic matter and diesel pollution (Downing et al. 2009, Carstea et al. 2010), evaluate drinking water treatment processes (Bieroza et al. 2009, Cumberland et al. 2012, Shutova et al. 2014) or detect pesticides (Ferretto et al. 2014). Fluorescence spectroscopy has been used to assess the quality of raw sewage and effluents (Baker 2001, Boving et al. 2004, Pfeiffer et al. 2008), industrial (Santos et al. 2001, Borisover et al. 2011, Li et al. 2015), or farm (Baker 2002b, Old et al. 2012) discharges into natural systems. Moreover, recent studies on short and long-term fluorescence monitoring along the WwTPs process train have been undertaken, to determine the potential of the technique for treatment processes control (for example, (Murphy et al. 2011, Bridgeman et al. 2013, Cohen et al. 2014, Ou et al. 2014, Singh et al. 2015). Although considerable work has been done so far in this field, there are still issues with regard to the "matrix effects", as reviewed by Henderson et al. (2009), or with fouling (Reynolds 2002) that must be overcome to allow application of the technique in WwTPs.

Other reviews proved the potential of applying fluorescence spectroscopy to water quality monitoring (Hudson et al. 2007, Henderson et al. 2009, Fellman et al. 2010, Ishii

and Boyer 2012, Yang et al. 2015b). However, none of them focused only on wastewater, which requires a specific discussion due to its complexity in composition and impact on the environment. Moreover, a growing number of studies are published each year on the application of fluorescence spectroscopy to wastewater quality evaluation, proving its scientific and industrial importance. In this paper, we review the current progress in applying fluorescence spectroscopy to assess wastewater quality. The technique's capabilities as a detection and early warning tool of pollution with treated or raw wastewater from different sources are discussed. Also, its potential for process control in WwTPs is presented.

2 Fluorescence assessment of wastewater components

2.1 Organic matter fluorescence assessment

The most common methods of recording fluorescence spectra for wastewater are excitation – emission matrices (EEM) and synchronous fluorescence spectra (SFS). EEMs represent fluorescence contour maps, which comprise a series of repeated emission scans recorded in a range of excitation wavelengths (Coble 1996). SFS are obtained by scanning simultaneously both excitation and emission monochromators at a fixed wavelength interval between them (Patra and Mishra 2002, Reynolds 2003). For many years, since the mid-1970s, SFS were preferred as a multidimensional technique for the

179 analysis of complex solutions, because it provided better peak
180 resolution, compared to emission spectra, and faster recording
181 time than EEMs (Ryder 2005). However, the improvement of
182 instrumentation allowed researchers to obtain fast, high-
183 resolution EEM collection, which increased the method
184 popularity in the research community. In addition, EEMs offer
185 varied possibilities of data interpretation, from simple peak-
186 picking and Fluorescence Regional Integration to the more
187 complex Parallel Factor Analysis (PARAFAC) and Self-
188 Organizing Maps. Among these methods, peak-picking and
189 PARAFAC are the most popular in the research community
190 and therefore only these two methods will be discussed in the
191 following sections.

192 The peak-picking method is a very simple tool to identify
193 components based on their maximum intensity and
194 corresponding excitation and emission wavelength pairs (Coble
195 1996). An example of peak-picking analysis is shown in Figure
196 1 (a). According to Goldman et al. (2012), peak-picking is a
197 viable analysis technique and can be employed for the
198 development and use of a real-time tool and may be related to
199 custom sensors available today. However, its applicability may
200 be limited due to peak shifts, possible overlapping and
201 interferences between peaks (Yang et al. 2015b). Moreover, it
202 may lead to misleading observations by associating each peak

with a specific fluorophore, when two excitation wavelengths are seen at fluorescent components (Fig. 1).

PARAFAC is a mathematical tri-liniar model that deconvolutes EEMs into chemically meaningful components (Fig. 1b). It separates the contribution of different fluorophores without additional assumptions about their excitation and emission spectra (Cohen et al. 2014). A thorough description of PARAFAC method and components in wastewater is given by Yang et al. (2015b). PARAFAC has become common practice in water quality studies, over the past 10 years (Murphy et al. 2014). Yang et al. (2015b) proposed that PARAFAC be developed into a surrogate method for conventional water quality parameters, treatability of organic matter (OM) and performance of treatment processes. Yu et al. (2014) suggested that the PARAFAC tool, the EEMizer, developed by Bro and Vidal (2011), could be implemented to monitor on-line the WwTPs performance. The studies of Yu et al. (2015a) implied that PARAFAC is able to identify contamination events and can be used for early warning, but the component that indicates contamination must be spectrally different from the existing components, without major spectral overlap, which may undermine the online monitoring strategy. Similarly, Murphy et al. (2011) showed that at times PARAFAC had difficulties distinguishing between components, returning hybridized spectra. Also, in a comparison between chromatographic

fluorescence fingerprints and EEM-PARAFAC, Li et al. (2014) showed that the latter method could not reflect the variety of organic matter species with similar fluorescence, but different physico-chemical properties. In addition, PARAFAC is currently applied only as post-processing technique, making it unsuitable for continuous monitoring. Also, there is no consensus regarding the optimum model in terms of sample size and variability (Yu et al. 2015a).

All these techniques have been employed successfully to analyse OM from various natural to engineered sources. A thorough review on OM fluorescence is provided by Hudson et al. (2007) and Fellman et al. (2010). Crude sewage is a combination of domestic waste, industrial discharges, surface runoff and storm flow. Its composition varies depending on the age and type of sewerage, time of day, weather conditions and type of incoming sewer (Ahmad and Reynolds 1995, Hudson et al. 2007). Ellis (2004) showed that the general organic composition of wastewater is 50 % proteins, 14 % carbohydrates, 10 % fats and oils and trace amounts of priority pollutants and surfactants, which are present in detergents, soaps, shampoo and similar consumer products. More recently, Huang et al. (2010) found that fibres, proteins and sugars are the largest groups of OM in wastewaters, accounting for 20.64 %, 12.38 % and 10.65 %, respectively, of the total TOC. According to the researchers, food related substances are the

main source of OM in wastewaters (Huang et al. 2010). Using gas chromatography/mass spectrometry, Huang et al. (2010) detected 90 compounds from the groups of alkyls and aromatic hydrocarbons, alkenes, alcohols, organic acids, ketones, phenols, nitrogenous compounds, ethers, amines and esters. In addition, they found lipids, volatile fatty acids, humic acids, DNA + RNA, tannic acids and linear alkylbenzene sulfonates. Within the organic composition, there are numerous overlapping fluorophores that contribute to the EEMs (Aiken 2014). Due to the difficulty of assigning specific fluorophores to the peaks identified in EEMs, the fluorescence of wastewater will be discussed as two regions based on the classification provided by Li et al. (2014): the region Em < 380 nm is associated with fluorophores containing a limited number of aromatic rings and the indole moiety of free tryptophan whilst the region > 380 nm is associated with polycyclic aromatic fluorophores.

2.2 Region Em < 380 nm

Based on the peak-picking method, fluorescence in this region is represented by peak T ($\lambda_{excitation}$ / $\lambda_{emission}$ *~225 (~280) / ~350 nm*) and peak B ($\lambda_{excitation}$ / $\lambda_{emission}$ *~225 (~280) / ~305 nm*) (Fig. 1a). Peaks T and B have been observed in all studies that used the peak-picking method for EEM processing, irrespective of the wastewater source (Table SM1). These peaks have been associated with living and dead cellular

material and their exudates and indicate microbial activity (Bridgeman et al. 2013) and material derived from anthropogenic activities (Yu et al. 2014). In PARAFAC, the region Em < 380 nm is generally identified as components with 2 excitation wavelengths and 1 emission wavelength (Fig. 1b) in the same wavelength ranges as peaks T and B in the peak-picking method. These components are identified in both municipal and industrial wastewater samples; however, the component similar to peak T is more common in wastewater compared to other components in this region (Table SM2).

By examining the list of wastewater organic components (Dignac et al. 2000, Huang et al. 2010, Navalon et al. 2011), and the literature review of Aiken (2014), Stedmon and Cory (2014) and Baker et al. (2014), the following components were considered as contributors to the fluorescence in the region Em < 380 nm: phenols (for example cresols), indoles, mono and polyaromatic hydrocarbons, DNA, aromatic amino acids (phenylalanine, tyrosine), degradation products of lignin (lignin phenols, vanillic acid, syringic acid etc.). These compounds are derived from domestic waste, chemical, pharmaceutical, plastic, petrochemical, paper, leather or textile industries (del Olmo et al. 1996, Pokhrel and Viraraghavan 2004, He et al. 2007, Tchaikovskaya et al. 2007, Tertuliani et al. 2008). The potential contributing fluorophores to this region are presented in Table 1.

303

304 **2.3 Region Em > 380 nm**

305 The peak-picking method classifies this region as
306 follows: Peak A ($\lambda_{excitation}$ / $\lambda_{emission}$ *~225 / 400 - 500 nm)*, peak C
307 ($\lambda_{excitation}$ / $\lambda_{emission}$ *300 - 350 / 400 - 500 nm)* and peak M
308 ($\lambda_{excitation}$ / $\lambda_{emission}$ *310 - 320 / 380 – 420 nm)* (Fig. 1a). All
309 studies done so far on wastewater OM have identified peak C
310 and most studies found peak A (Table 1); however, peak M
311 was analysed only by Yu et al. (2014) at municipal wastewater.
312 Most of the studies that employed PARAFAC for EEM
313 analysis identified a maximum of 4 components associated and
314 microbially and terrestrially derived DOM (example of two
315 components in Fig 1b). However, Ishii and Boyer (2012) have
316 identified the PARAFAC components common in natural and
317 engineered water systems: Component 1 similar to peak A with
318 excitation in the region < *230 – 260 nm* and emission between
319 *400* and *500 nm*; Component 2 similar to peaks A + C found in
320 excitation region < *240 – 275 (339 – 420 nm)* and emission
321 within *434 – 520 nm*; and Component 3 similar to peak A + M
322 appearing in the excitation domain *<240 – 260 nm (295 – 380*
323 *nm)* and within the *374 – 450 nm* emission range. According to
324 Ishii and Boyer (2012), component 1 is found mostly in OM
325 sources dominated by terrestrial precursor material. Component
326 2 was defined as reduced quinone-like and was identified in
327 OM from a wide variety of aquatic systems, including those

dominated by terrestrial and microbial inputs. While, component 3 fluorophores were defined as oxidised quinone-like and were similar to those with terrestrial and marine precursors. Component 1 has not been reported in wastewater studies, but components 2 and 3 were seen at studies made on municipal and industrial wastewater (Table SM2). Additional components were observed in wastewater (Table SM2), but they vary depending on source.

As shown in Table 1, there are several fluorophores that could contribute to the fluorescence of region Em > 380 nm: lignins, PAHs, flavonoids, humic acids, quinones, aromatic ketones, fluorescent whitening agents (FWAs), pharmaceutically active compounds (Dignac et al. 2000, Huang et al. 2010, Aiken 2014, Baker et al. 2014, Stedmon and Cory 2014). Among these components, FWAs have been proposed as an indicator of human faecal contamination (Assaad et al. 2014), sewer misconnections (Chandler and Lerner 2015) and presence of landfill leachates (Graham et al. 2015). FWAs are highly soluble and poorly biodegraded, and therefore likely to pass through biological treatment in WwTPs (Kramer et al. 1996, Poiger et al. 1998, Assaad et al. 2014). Research has shown that these components can be detected with handheld fluorometers, which enhances the capability for in situ water monitoring (Hartel et al. 2007). Nevertheless, issues with detecting FWAs in waters have been reported: the fluorescence

of other peak C fluorophores overlap the peaks of FWAs, these components are easily photodegraded and DOM hinders the reaction of FWAs (Kramer et al. 1996, Baker 2002a, Hartel et al. 2007, Assaad et al. 2014). Solutions to overcome fluorescence overlap have been proposed, yet the other issues identified may limit the method's applicability in detecting sewage. The following solutions have been proposed: a) to use the photodegradation rate to separate FWAs from organic matter (Hartel et al. (2007); b) to take into account the differences in shape of the photodecay curve between FWAs and natural organic matter (Cao et al. (2009)); c) to use a baseline correction method to compare the differences in fluorescence intensity of FWA, between the regions *320 nm – 345 nm* and *345 nm – 360 nm*, with the same values for the water samples (Takahashi and Kawamura (2006)); and d) to apply three-way analysis of EEMs assisted by second-order chemometric analyses (Gholami et al. 2015). Discrimination between humic substances and FWAs was achieved by Boving et al. (2004), who analysed FWAs in solution with humic acid and tannic acid. FWAs were recorded at *344 nm* and *422 nm* emission wavelength, and *250 nm* excitation wavelength. The authors found that the second peak of the FWAs was separated from humic acids by 22 nm, but there was a 4 nm separation from tannic acid. Therefore, the $\lambda_{excitation}$ / $\lambda_{emission}$ = *250 / 422*

377 *nm* peak could be used for FWAs detection without
378 interference from humic acid.

379 As shown above, there are several fluorophores that
380 contribute to the < 380 nm > Em regions, but the list is not
381 exhaustive. More studies are needed to identify new fluorescent
382 components and especially those specific to source with the
383 highest contribution to EEMs. Since the regions exhibit the
384 fluorescence of xenobiotic compounds, both can be used for
385 wastewater quality assessment. In particular, peaks T and C,
386 and the PARAFAC analogous components, are present in all
387 wastewater studies (Tables SM1 and SM2) and may be applied
388 to the control of wastewater treatment processes. However, it
389 may be difficult to identify the source and type of sewage
390 pollution in receiving water bodies. In this sense, Baker et al.
391 (2014) advise caution and stress the importance of using a good
392 sampling framework combined with an appropriate
393 multivariate analysis of data for successful investigation of
394 water pollution.

395

396 **3 Correlation of the fluorescence peaks with BOD, COD**
397 **and TOC**

398 In order to assess the capability of fluorescence
399 spectroscopy to act as a monitoring tool it is important to
400 consider the correlations between fluorescence peaks and BOD,
401 COD and TOC, commonly used indicators of OM

concentration in natural waters and wastewater. As reviewed by Bourgeois et al. (2001) and (Jouanneau et al. 2014), BOD is a desirable measurement in treatment processes, it presents several disadvantages, which make this technique unsuitable for on-line monitoring and process control: it is slow to yield information, it is labour intensive, toxic substances affect bacteria, it may not reflect conditions in the treatment processes, it is insensitive and imprecise at low concentrations and has an uncertainty of 15-20% in the results. COD takes less time to give a result than BOD (2-4 h) and is not affected by toxic substances. However, it is still not suitable for on-line monitoring and process control due to the measuring time and because it requires hazardous chemicals. Also, COD is able to discriminate between biodegradable and biologically inert organic matter only in conjunction with BOD and not on its own (Bourgeois et al. 2001, Chen et al. 2014). TOC is very fast, as triplicates can be analyzed in minutes. However, it cannot differentiate between biodegradable and nonbiodegradable OM (Orhon et al. 2009). Also, conflicting results have been reported between different techniques of measuring TOC (Bourgeois et al. 2001).

Correlation between fluorescence and standard parameters revealed that peaks T and C relate to BOD, COD and TOC, as reviewed by (Henderson et al. 2009). Slightly better correlation with BOD is seen at peak T compared to peak

C. An exception to the above observation is found at the study of Wang et al. (2007) who obtained better correlation with the PARAFAC component exhibiting fluorescence in the peak C region, compared to the peak T component (Table 2). They observed the best correlation with BOD at the component similar to peak M (0.73). The researchers concluded that this component contributed the most to BOD for wastewater-impacted lakes. Nevertheless, these results highlight the complexity of the source and that there are potentially several fluorophores, which display fluorescence in the peak T/C regions. It also shows that both regions could contribute to BOD. The difference in correlation coefficients could also be determined by the low sample sizes in some studies, which might under or overestimate the relationship between fluorescence and BOD, COD and TOC (Table 2). Another cause of the difference could be the method used for data processing, as PARAFAC offers better separation of overlapping components compared to peak-picking.

Based on the correlation between BOD and peak T fluorescence, Hur and Kong (2008) tried to estimate, using SFS and first derivative spectra, the concentration of BOD of samples from urban rivers affected by treated sewage. They found that the relative fluorescence intensity, at *283 nm* to *245 nm* from SFS, is the optimum estimation index as it has the best positive correlation with BOD values (0.91). It has been

452 reported that the multiple regression method, using the light
453 scattering intensity at *633 nm* or turbidity, greatly enhances the
454 correlation between measured and predicted BOD values. Hur
455 and Kong (2008) also observed that filtered samples presented
456 enhanced correlation; however, Bridgeman et al. (2013)
457 reported slightly higher correlation coefficient between BOD
458 and fluorescence at unfiltered samples compared to filtered
459 with 0.45 or 0.2 µm. These differences could be site specific
460 and may depend on the sizes of OM components.

461 As reviewed by Baker et al. (2014), the correlation
462 between BOD and peak T fluorescence suggests a direct link
463 with microbiological activity in this region of fluorescence,
464 although the source of peak T fluorescence is generally
465 unknown. It was also implied that handheld instruments could
466 be used in the future to investigate the temporal variability of
467 BOD (Baker et al. 2014). Due to the relation with
468 microbiological activity, peak T fluorescence was suggested as
469 indicator of the presence / absence faecal coliforms (Sorensen
470 et al. 2015, Sorensen et al. 2016). Pfeiffer et al. (2008) obtained
471 excellent correlation (0.90 – 0.95) with faecal coliforms on
472 samples from a wastewater polluted river and (Tedetti et al.
473 2012) found a good correlation (0.78) between the PARAFAC
474 component and Escherichia Coli + enterococci on wastewater
475 impacted coastal water samples. More recently, (Baker et al.
476 2015) obtained a log correlation of 0.74 between fluorescence

and E. Coli measurements. These findings are encouraging, but more work should be done to explore the link between fluorescent components and faecal coliforms and its potential use in on-line monitoring applications. In a comparison with flow cytometer measurements, peak T intensity correlated with an increase of total live and dead bacteria numbers (Bridgeman et al. 2015). The researchers found that four bacteria isolated from a potable water tap sample showed different responses in the fluorescence signal, although the intensity of peak T fluorescence did not correlate with the bacteria counts. Nevertheless, peak T fluorescence could be used to assess the microbiological activity in a water system.

4 Fluorescence detection of wastewater pollution

Fluorescence spectroscopy has shown its capabilities as a real-time assessment tool for wastewater quality due to its advantages and correlation with standard parameters. This technique could be very effective in detecting raw wastewater contamination in water bodies. Also, the impact of wastewater effluents on natural waters could be evaluated, since effluent organic matter has different composition and characteristics from naturally occurring OM (Wang et al. 2015). Therefore it is important to look at the different types of wastewater for particular characteristics that may facilitate identification in the receiving water bodies.

4.1 Sources of wastewater

Studies published so far on fluorescence spectroscopy have focused on domestic, farm and industrial wastewater, which includes textile, pulp mill, coke or brewery industries. More studies are needed on wastewater from oil refineries, metal processing, fermentation factories, pharmaceutical industry, chemical plants, meatpacking and processing etc.

4.1.1 Domestic wastewater

Wastewater is the flow of water used by a community and includes household wastes, commercial and industrial waste stream flows, and stormwater (Drinan and Spellman 2012). Domestic wastewater contains the solid and liquid discharges of humans and animals, contributing with millions of bacteria, virus, and non-pathogenic and pathogenic organisms. It may also contain sanitary products, cleaners and detergents, trash, garbage and any other substances that are poured or flushed into the sewer system (Drinan and Spellman 2012). Public treatment facilities may also collect industrial effluents and thus chemicals, dyes, acids, alkalies, grit or detergents can be found in municipal wastewater (Drinan and Spellman 2012). Stormwater runoff, if collected by WwTPs, may bring into the system large amounts of sand, gravel, road-salt and other grit (Drinan and Spellman 2012).

As discussed in the previous sections, there are numerous compounds that may contribute to the fluorescence peaks. Generally, fluorescence spectra of untreated and treated domestic wastewater are characterized by intense peaks in the region Em < 380 nm, especially peak T, associated with high microbial abundance, and by significantly lower intensity peaks A and C fluorescence (Baker 2001, Hudson et al. 2007, Hur and Cho 2012, Bridgeman et al. 2013). In some studies, the fluorescence spectra of effluents showed a higher prevalence of peaks A and C, compared to peaks T and B (Ghervase et al. 2010a, Riopel et al. 2014). Among peaks, T and C seem to be present at most municipal wastewater samples (Tables SM1 and SM2) and may serve as indicators of wastewater contamination. Peak B is rarely analysed at wastewater EEMs due to the potential interferences from scattering; however, this fraction could indicate the proximity of the measurement point to the discharge point or freshness of the contamination. According to Pfeiffer et al. (2008), the fluorescence of both peak T and peak B decreases in intensity with increasing distance from the release point, but peak B is completely removed at longer distances, due to dilution or breakdown of the organic fraction. For peak B removal, seasonal shifts should also be taken into account as rainfall could contribute to dilution, sunlight irradiation could cause photodegradation or increase microbial uptake during summer (Meng et al. 2013).

552 From the myriad of fluorophores, FWAs may display
553 distinctive features in the EEMs for municipal wastewater
554 samples (Bridgeman et al. 2013). However, this fraction is not
555 specific to domestic wastewater, as it has been detected at
556 paper mill effluents (Baker 2002a, Ciputra et al. 2010,
557 Bassandeh et al. 2013) or landfill leachates (Graham et al.
558 2015). Therefore, peaks T and C seem to be the best tools of
559 monitoring domestic wastewater quality.

560 In addition to fluorescence intensity increase, it has been
561 shown that discharge of domestic sewage may change the
562 properties of OM from the receiving water bodies. For
563 example, Xue et al. (2011) found that sewage effluents change
564 the capacity of OM to form disinfection by-products and
565 decrease its sensitivity to UV light. Also, changes in
566 aromaticity and hydrophobicity of OM have been reported.
567 These OM characteristics have been assessed after discharge,
568 using the emission wavelength of peak C. In two studies
569 undertaken by Goldman et al. (2012) on OM wastewater
570 effluent and by Ghervase et al. (2010b) on untreated sewage
571 discharge, it was found that the fluorescence signal of the two
572 types of samples presented lower peak C emission wavelength,
573 indicating lower aromaticity compared to natural OM. While,
574 Spencer et al. (2007a) reported higher aromaticity of the OM
575 from an estuarine sample with anthropogenic impact from
576 domestic wastewater effluents, compared to the estuarine OM.

577 Goldman et al. (2012) found that the mixture of effluent and
578 river waters produce midrange values and, therefore, a potential
579 increase in aromaticity with distance from discharge could be
580 expected. In marine environments, fluorescence measurements
581 on wastewater discharges showed great complexity of the
582 mixing properties. Petrenko et al. (1997) observed 4 layers in
583 the seawater column, 2 layers being affected by sewage
584 representing the "old" and "new" plume waters and 2 layers
585 unaffected by effluent. According to the researchers, the release
586 of wastewater increased 2 fold to the concentration of
587 ammonium, silicate and phosphate in sewage affected plumes
588 and could stimulate the growth of phytoplankton. Baker and
589 Inverarity (2004) also found an increase in nitrate and
590 phosphate concentrations downstream of discharge into urban
591 rivers.
592
593 ***4.1.2 Animal wastewater***
594 Animal wastes represent an important source of water
595 pollution, through the release of untreated wastewater or
596 surface runoff from farms. This type of wastewater produces
597 BOD values that are 1 to 3 times higher than sewage BOD
598 (Baker 2002b). Most meat processing units treat the wastewater
599 prior to release, however animal wastewater varies temporally
600 in composition, requiring continuous monitoring for effective
601 detection and removal of pollutants. Relatively few studies

have looked at the potential of using fluorescence spectroscopy to monitor the quality of animal wastewater. However, data gathered so far can help define particular characteristics of animal wastewater OM. The fluorescence of animal wastewater is generally dominated by the region Em < 380 nm. In particular, peak T fluorescence seems to be common to all samples, as it has been detected at farmyard runoff (Old et al. 2012), pig and cattle slurry, silage liquor, sheep barn waste (Baker 2002b), poultry processing unit (Ghervase et al. 2010b) and cattle slaughter house (Louvet et al. 2013). The researchers also observed a low peak C fluorescence relative to peak T. Baker (2002b) calculated the ratio between the fluorescence intensity of these two peaks and found that peak T intensity was 2 to 25 times higher than that of peak C, the highest ratio being obtained for silage liquor, while the lowest was seen at the sheep barn waste. A similar peak T/C ratio was obtained by Old et al. (2012) at farmyard runoff samples. The ratio of peaks T and C fluorescence intensity shows that farm waste pollution events could leave a signature in river waters (Baker 2002b) and confirm the potential of using fluorescence as a low cost and rapid technique for tracing animal derived pollutants (Old et al. 2012). Interestingly, pig and cattle slurry presented peak B fluorescence at a similar intensity to that of peak T. Peak B was also detected at poultry wastewater (Ghervase et al. 2010b), having even higher fluorescence than that of peak T.

Ghervase et al. (2010b) suggested using the ratio of peak T and peak B to detect poultry wastewater pollution in rivers. However, this ratio applicability could be limited only to certain types of animal wastewaters.

Cattle slaughterhouse wastewater may contain albumin and haemoglobin that would contribute to the Em < 380 nm fluorescence region (Louvet et al. 2013). Also, bovine serum albumin may contribute to the fluorescence region of Em > 380 nm. Louvet et al. (2013) found another fluorescence peak that could belong to metalloporphyrins ($\lambda_{excitation}$ / $\lambda_{emission}$ = *400 - 440 nm / 450 - 510 nm)*. These components are attributed to red blood, which is a major pollutant in slaughterhouse wastewater. Again, the ratio of peaks T and C fluorescence intensity was found to be an effective indicator of biodegradation of slaughter house wastewater (Louvet et al. 2013). Nevertheless, the composition of animal derived pollutants is highly variable in time and depends on the animal species, physiological state and diet (Baker 2002b, Louvet et al. 2013). Therefore, more studies are needed to better understand the properties of OM from animal derived wastewater and set clear characteristics for enhanced detection of pollution events.

4.1.3 Industrial sources of wastewater

Industrial wastewater is primarily derived from the manufacturing and processing of chemicals, textiles, wood,

pulp mill or paper. The composition of effluents varies depending on the raw materials used, the type of process and the efficiency of material removal (Sánchez Rojas and Bosch Ojeda 2005). Studies on continuous monitoring and evaluation of industrial wastewater using fluorescence spectroscopy are scarce, limiting identification of particular features of wastewater fluorescence spectra. Few studies focussed on wastewater from petrochemical, chemical and biochemical industry (Borisover et al. 2011), brewery (Janhom et al. 2009, Janhom et al. 2011), textile (Li et al. 2015), pulp mill and paper processing (Baker 2002a, Ciputra et al. 2010, Cawley et al. 2012, Bassandeh et al. 2013) computer components manufacturing (Cohen et al. 2014) and coke industry (Ou et al. 2014). In one short-term monitoring study, Yang et al. (2015a) analysed and compared the fluorescence spectra of samples from the effluents of 57 facilities belonging to 12 industrial categories (non-alcoholic drinks, electronic devices, food, leather and fur, meat, organic chemicals, pulp and paper, petrochemical, resin and plastic, steel, steam-power and textile dyeing) aiming to evaluate the potential of fluorescence spectroscopy to identify wastewater sources. The researchers were able to characterise and differentiate industrial effluents using cluster analysis, EEM-PARAFAC and FT-IR. Components from both < 380 nm > regions were observed, but no component dominated over all samples. For instance, the

peak T component presented the highest fluorescence intensity at leather and fur wastewater, while peak C components dominated the EEMs of food wastewater samples. Therefore, Yang et al. (2015a) concluded that, without additional analyses it may be difficult to identify an industrial source with fluorescence spectroscopy. However, Borisover et al. (2011) observed a bathochromic shift of the peak T component induced by polarity and composition of local environment. They studied samples collected from rivers impacted by industrial effluents of oil refineries, petroleum and chemical and biochemical plants. The researchers recommended using this component as fluorescent tracer of non-specific industrial pollution.

Studies that evaluated wastewater samples from particular industries have identified specific fluorophores. For example, at pulp mill wastewater effluents, Cawley et al. (2012) found a component that was attributed to lignosulfonic acid or to a mixture of fluorophores from the many lignin degradation products. However, the authors highlighted that this component may exhibit different emission maxima depending on variations in the actual chemical moieties present in each sample. A similar component was found by Bassandeh et al. (2013) at samples collected from the biologically treated effluent of a newsprint mill and the authors attributed it to lignins or chemicals involved in the paper making process.

Cawley et al. (2012) and Bassandeh et al. (2013) both identified distinctive PARAFAC peaks for the lignin derived components. However, Santos et al. (2001) observed very intense peaks and additional shoulders at the peak C for samples collected from rivers downstream of pulp mill effluent discharge. Also, compared to samples upstream, the researchers detected an additional peak at $\lambda_{excitation}$ / $\lambda_{emission}$ *~290 / ~340 nm*, which coincides with the peak T fluorescence. Baker (2002a) suggested that peak T fluorescence results from the lignin and sugars produced by the pulping process, which are likely to be rich in aromatic proteins. This component correlated with TOC (r=0.62, N=18), indicating that peak T fluorescence was a significant contributor to the TOC at paper mill effluents, as this correlation was not seen at the river samples. In addition to lignin derived components, Baker (2002a) identified a peak associated with FWAs, which are commonly used in papers. The differences in results, found by these studies, could be attributed to variations in chemical moieties or to the fact that Cawley et al. (2012) and Bassandeh et al. (2013) used PARAFAC for data processing to provide better separation between lignin and other peak T or peak C fluorophores.

A distinctive feature was also detected at textile industry effluents by Li et al. (2015), who found a triple excitation component with emission wavelength at *460 nm*. They considered this feature as specific to textile-derived

components, because most fluorophores in region Em > 380 nm present dual excitation peaks at emission wavelength between *400* and *500 nm*. The triple excitation peaks were associated with 1-amino-2-naphtol structure, based on a spectral comparison with the standard solution and were suggested to be used as specific indicators in textile effluents. Li et al. (2015) also found that for peak T fluorescence there were much more species with varying emission wavelengths, which could relate to azo dyes as these substances emit similar fluorescence in this region.

As shown in section 2.2 and Table 1, peak B fluorescence could represent phenol-like matter, hydrocarbons or cresols as found by Ou et al. (2014) at coke wastewater samples. In addition to peak B and peak C fluorophores, Ou et al. (2014) identified a component associated with heterocyclic components and polycyclic aromatic hydrocabons (PAHs), such as fluoranthene or naphtol. PAHs were also detected by Cohen et al. (2014) at samples collected from a WwTPs that receives 50% of its crude wastewater from a computer component factory. Based on spectral similarities, Cohen et al. (2014) suggested that this component contains a pyrene-like moiety.

While for textile, pulp mill or coke wastewater, distinctive components have been identified, brewery wastewater has been shown to contain only the typical peaks T,

752 A and C (Janhom et al. 2009, Janhom et al. 2011), generated by
753 the cleaning and washing of raw materials. They also showed
754 that the fluorescence of brewery wastewater samples belonged
755 primarily to hydrophobic acids and hydrophilic bases OM
756 fractions.
757

758 **4.2 Wastewater tracking in aquatic systems**

759 Discrimination between sources using fluorescence
760 spectroscopy may be challenging since domestic wastewater
761 can be mixed with industrial effluents and agricultural runoffs
762 (Andersen et al. 2014). Industrial wastewater could also contain
763 domestic discharges from the toilets and kitchens within
764 factories (Reynolds and Ahmad 1995). Moreover, organic
765 pollutants like optical brighteners, PAHs or lignins have
766 widespread application and thus can be found in any type of
767 wastewater.

768 In particular for industrial wastewater it may be more
769 difficult to separate sources due to the varied composition of
770 the solution. The release of industrial effluents in water bodies
771 may lead to the production of fluorescent fractions formed of a
772 mixture of proteinaceous and non-proteinaceous substances,
773 which generates a bathchromic shift in the typical peak T
774 fluorescence emission wavelength. According to Borisover et
775 al. (2011) this component may be used as a tracer of non-
776 specific industrial pollution. However, various industrial

777 wastewaters produce high quantities of particular fluorophores
778 like PAHs or heterocyclic compounds, differentiating them
779 from domestic wastewater. As shown by Cohen et al. (2014)
780 the pyrene-like components separated the wastewater with 50%
781 industrial input from the more domestic wastewater sources.
782 Also, the devices, developed by Tedetti et al. (2013) and Puiu
783 et al. (2015), that separate PAHs from other peak T
784 fluorophores, hold great promise in detecting both domestic
785 and industrial sources of pollution. Additionally, chemical
786 separation can be undertaken by the use of time resolved laser
787 induced fluorescence, which is capable to identify components
788 based on their lifetimes. PAHs have a relatively long
789 fluorescence lifetimes and great quantum efficiency, which
790 help at distinguishing PAHs from the OM background
791 (McGowin 2005).

792 However, the question remains as to how to differentiate
793 between wastewater from domestic, animal farms and industry
794 sources, which are characterized by intense Em < 380 nm
795 region. Domestic wastewater contains PAHs (Huang et al.
796 2010), which have a distinctive fluorescence signal; however,
797 the quantities could be too low in comparison to other
798 fluorophores and therefore the fluorescence of PAHs could be
799 exceeded by other compounds.

800 Component distinction can also be undertaken by
801 PARAFAC, which may be able to separate overlapping

components or identify specific pollutant indicators (Cohen et al. 2014, Yang et al. 2015b). However, in case of low concentrated pollutants, such as detergents, peak picking has been shown to be more effective than PARAFAC (Mostofa et al. 2010). Therefore, a combination of these techniques could better provide a thorough view of the sample composition and OM interaction with pollutants. Fluorescence spectroscopy could be used as an early warning system in case of accidental pollution and could serve as a quick method in initial identification of the source of wastewater, before more complex and expensive analyses would be employed.

5 Control of wastewater treatment processes using fluorescence spectroscopy

Two decades ago, the studies of Reynolds and Ahmad (1995) and Tartakovsky et al. (1996) demonstrated the potential of using fluorescence spectroscopy for both off- and on-line monitoring in wastewater treatment. Recent studies have suggested that this technique could be applied to process control and optimization (Bridgeman et al. 2013). With increasingly stringent regulation it will be more difficult to control treatment efficiency with current techniques, (BOD, COD and TOC), which are expensive, time-consuming and unreliable (Bridgeman et al. 2013, Rehman et al. 2015). More pressure is put on WwTPs when other environmental

827 implications, such as energy and chemical consumption or
828 greenhouse gases emissions are considered (Wang et al. 2015).
829 Fluorescence spectroscopy offers a robust technique available
830 for a rapid and low cost estimation of effluent quality.
831 However, studies on fluorescence monitoring of WwTPs
832 processes are scarce and only one long-term study at 5
833 municipal WwTPs has been achieved (Cohen et al. 2014).
834 Also, only one real-time monitoring study has been published
835 on two recycled water systems (Singh et al. 2015). According
836 to Reynolds (2002), WwTPs are hostile environments, making
837 continuous and dynamic monitoring of wastewater quality
838 difficult due to problems associated with fouling. This would
839 require regular cleaning, which is time consuming. In addition,
840 the fluorescence signal could be affected by pH, IFE,
841 temperature and metal ions, requiring subsequent corrections.
842 However, recent development of devices, already on market,
843 show great promise since they convert the on-line peak T
844 fluorescence signal into BOD equivalent values, using an
845 internal calibration factor or a multispectral approach
846 (ChelseaInstruments 2015, ModernWater 2015,
847 ZAPSTechnologies 2015). This type of instruments could
848 provide an immediate estimation of changes in wastewater
849 quality, displaying capabilities of effective process control.
850

5.1 Monitoring of fluorescent OM

Fluorescence real-time monitoring of wastewater quality is difficult to implement due to multiple potential factors that may interfere with the signal. The only real-time monitoring study was undertaken by (Galinha et al. 2011a) on a pilot scale membrane bioreactor system to predict performance parameters. EEMs were recorded for 10 months and processed with multivariate techniques. They concluded that although fluorescence was able to describe total COD for influent and effluent, it could not accurately predict other performance parameters and hence, fluorescence cannot totally replace conventional monitoring of membrane bioreactors (Galinha et al. 2011a). Nevertheless, real-time monitoring studies at full-scale WwTPs should be undertaken in order to assess the feasibility of the method and the issues that can arise from its implementation. The studies done on the monitoring of surface waters identified major issues and offered solutions, which could be used to build a strategy for wastewater on-line monitoring. The issues reported so far include: biofilm formation, temperature, turbidity, inner filter effect, calibration procedure, presence of quenching elements. Most of these problems are thoroughly reviewed by Henderson et al. (2009). Therefore, only the recent studies will be discussed. Before the study of Carstea et al. (2010) no long-term, real-time monitoring experiments were done due to fouling issues.

Carstea et al. (2010) showed that over a period of 11 days of continuous EEM recordings on an urban river, biofilm formation on the water extraction system had no influence on the fluorescence signal. However, higher rates of biofilm formation are expected in wastewater, compared to surface water, due to the large quantities of extracellular polymeric substances that enhance cell adhesion to solid surfaces (Tsuneda et al. 2003).

Regarding temperature, Chen et al. (2015) tested a newly developed, portable laser induced fluorescence system, for its monitoring capabilities, on estuarine water and found that temperature changes affected the fluorescence results. Yamashita et al. (2015) and Khamis et al. (2015) also reported the impact of temperature on the fluorescence of OM, at monitoring studies on open ocean and urban river. Carstea et al. (2014) have shown that peak T fluorescence suffers more thermal quenching at samples with higher urban anthropogenic impact compared to natural sources. Therefore, temperature could have a major impact on OM fluorescence from wastewater. However, a temperature-compensating tool has been proposed and tested by Watras et al. (2011). Khamis et al. (2015) also proposed a compensating tool for turbidity, which can have a great impact on the fluorescence signal when large particles are present. It is yet to be tested on wastewater samples.

The inner filter effect (IFE) is another major issue at wastewater samples. The IFE is the apparent decrease in the emitted fluorescence intensity or a distortion of the band-shape resulting from the absorption of the excited and emitted radiation (Henderson et al. 2009). Kothawala et al. (2013) found that the best correction tool for the IFE is the absorbance based approach, proposed by Lakowicz (2006). This approach can be applied to samples with absorbance values of up to 1.5 cm^{-1}; at samples above this value a dilution of 2x is recommended (Kothawala et al. 2013). However, the study of Kothawala et al. (2013) was undertaken on lake water samples and it is not known if these rules apply to wastewater monitoring. As seen in Tables SM1 and SM2, for the wastewater evaluation studies there are two preferred methods for reducing the IFE: dilution and post-measurement mathematical correction. A dilution factor of 10 was used in some studies, while in others the samples were diluted until a specific absorbance value was achieved. Most studies report the absorbance values at wavelengths within the excitation region of peak T. In specific studies, no dilution was used to analyse samples as this procedure in not applied to on-line measurements (for example, (Baker and Inverarity 2004, Louvet et al. 2013, Li et al. 2014). However, IFE could be a serious issue for monitoring studies, as this factor might lead to an underestimation of the degree of pollution and poor

prediction of BOD, COD or TOC. In this case, dilutions to a certain absorbance value (< 0.05 cm^{-1}, as used in most studies, Tables SM1 and SM2) or post-measurement IFE correction are recommended. However, other solutions should be found to counteract IFE, as the use of UV absorbance measurements, in addition to fluorescence spectroscopy, reduces the practicality of the method for on-line monitoring.

In addition, Yamashita et al. (2015) proposed fluorescence sensors calibration for dark blanks and/or sensitivity. Solutions of L-tryptophan (Tedetti et al. 2013, Khamis et al. 2015, Sorensen et al. 2015) and quinine sulphate (Conmy et al. 2004, Chen et al. 2015, Yamashita et al. 2015) are generally used as calibration standards for the two fluorescence regions. However, Khamis et al. (2015) mention that uncalibrated systems may be used if qualitative data is needed.

Finally regarding the presence of quenching components, Wang et al. (2014) have proved that the presence of humic-like components could reduce the fluorescence of peak T in effluent organic matter. However, even more complex interactions could occur in wastewater samples. Galinha et al. (2011b) found that the addition of bovine serum albumin to domestic wastewater samples determined a decrease with 31-58 % of peak T fluorescence. They concluded that the complexity of interferences on the fluorescence signal might not allow the

simple and direct quantitative measurement of specific fluorophores in complex biological systems, such as wastewater. Also, in a study aiming to identify the contribution of extracellular polymeric substances to dye removal, Wei et al. (2015) showed that methylene blue has a substantial quenching effect on peaks T and C fluorescence. Several studies (Baker 2001, 2002b, Spencer et al. 2007a, Xue et al. 2011) have stressed that, although peak T is dominant in fluorescence spectra of wastewater, it is very likely that sewage generates high quantities of other components, which may significantly impact peak T fluorescence. Nevertheless, a study conducted by Zhou et al. (2015) on a drinking water source contaminated with domestic wastewater, showed that all peaks were sensitive to pollutant concentration, especially peak T, which could be used as an early warning tool for contamination. Moreover, Goldman et al. (2012) were able to predict the percentage of municipal wastewater in rivers with 80 % confidence, by the use of multivariate linear regression and the fluorescence of both peak T and peak C. They recommended applying this model to develop in situ instruments, inform monitoring progress and develop additional water quality indicators. Also, Hur and Cho (2012) recommended the use of absorbance values at 220 nm and 254 nm, and PARAFAC components similar to peaks T and C, as estimation indices for BOD and COD in wastewater effluent contaminated river.

5.2 Monitoring of treatment processes with fluorescence spectroscopy

Typical wastewater treatment begins with a series of physical operations (pre-treatment and primary treatment), such as screening and sedimentation to remove the floating and settleable solids. These steps are followed by biological processes, which are used to convert the finely divided and dissolved OM from wastewater into flocculant settleable biological solids (Tchobanoglous et al. 1991). Biological processes include the suspended growth activated sludge process, anaerobic/anoxic/oxic, sequencing batch reactor, membrane reactor, trickling filter, etc. Activated sludge is the most common process, involving the entrainment of air for microbial degradation of OM. In the final steps of the biological treatment, the sludge flocs are separated from the treated effluent, through sedimentation, before the effluent is discharged to a water body. In some WwTPs, additional treatment processes (tertiary and quaternary), such as filtration, chlorination, UV disinfection or reverse osmosis are adopted after the biological treatment and subsequent sedimentation (Yang et al. 2015b).

Few studies have focused, so far, on wastewater quality monitoring in treatment works, using fluorescence spectroscopy, to understand the behavior of OM along the

process train, the removal of components and the potential of applying fluorescence as a control tool. Among these studies, some looked into the treatment of specific domestic/industrial wastewater (Janhom et al. 2009, Janhom et al. 2011, Zhu et al. 2011, Yu et al. 2013), the removal and behavior of refractory OM in treatment works (Hur et al. 2011), characterization of reverse osmosis permeates (Singh et al. 2009, Singh et al. 2012, 2015) or compared fluorescence EEM-PARAFAC and HPLC/HPSEC techniques (Li et al. 2014). Fluorescence monitoring of wastewater quality was performed at time frames spanning from 1 month to 20 months, by collecting samples from the inlet and outlet (Reynolds 2002, Riopel et al. 2014) or along different treatment steps (Singh et al. 2009, Hambly et al. 2010, Murphy et al. 2011, Singh et al. 2012, Bridgeman et al. 2013, Cohen et al. 2014, Ou et al. 2014, Singh et al. 2015). The longest monitoring study was undertaken by Cohen et al. (2014), who analyzed the wastewater quality from municipal treatment plants during 20 months. _ENREF_23_ENREF_126Most of the monitoring studies involved WwTPs that employed activated sludge, as biological treatment process. Nevertheless, a few long-term and short-term monitoring studies have proven the capacity of fluorescence to evaluate the treatment performance in plants that used trickling filters (Bridgeman et al. 2013), anaerobic/anoxic/oxic (Yu et al. 2014), a novel anoxic/aerobic/aerobic system (Ou et al. 2014) or other

advanced biological treatments, such as phase isolated ditches, bio-Denipho process, sequencing batch reactors (Hur et al. 2011). Hur et al. (2011) found no difference in OM fluorescence characteristics between conventional and advanced biological treatment, while Bridgeman et al. (2013) were able to show, using fluorescence spectroscopy, that activated sludge was more effective than trickling filters, in removing the organic fraction. Variations in the fluorescence signal among WwTPs were also observed by Murphy et al. (2011). Nevertheless, the general consensus is that the behavior of certain fluorescence peaks can be followed along treatment plants to test performance. Cohen et al. (2014) suggested using both peak T and peak C components as indicators of total microbial activity in wastewater. Therefore, varied instrumentation available on market or under development (Bridgeman et al. 2015) that measure both components may be applied to monitor treatment efficiency.

5.3 Removal of fluorescence components along the treatment plant processes

Studies have shown that the OM, especially in the region Em < 380 nm is significantly removed after the biological treatment process (Fig. 2). This is to be expected since the biological treatment removes biodegradable material (Cohen et al. 2014). Riopel et al. (2014) reported a 60% reduction in the

peak T fluorescence. Within the Em < 380 nm region, peak T component experiences a different degree of removal compared to peak B component. Yu et al. (2013) found that peak T fluorescence decreases with 60 % in the anaerobic/anoxic zone, almost 40 % in the oxic zone and 5% in the final clarification process, whilst peak B fluorescence is reduced by 55%, almost 100% and 0% in the respective zones. Yu et al. (2014) reported slightly higher reduction percentages for peak B in the anaerobic/anoxic/oxic system. They also observed that peak T remained relatively consistent in the treatment process (41 - 48 %), but peak B decreased dramatically (33 - 7 %). However, Murphy et al. (2011) and Janhom et al. (2009) found a poor removal of peak B fluorescence. Janhom et al. (2009) stated that peak B substances are not considered refractory and suggested that these substances could be related to some humic-bound proteinaceous constituents, which may be biologically resistant. Nevertheless, Cohen et al. (2014) advises caution when comparing the sensitivity of fluorescent components to wastewater treatment due to possible multiple differences in the treatment system. In addition to the biological treatment, Cohen et al. (2014) found that soil-aquifer treatment causes a further significant decrease in the concentration of the OM fluorescing in the Em < 380 nm region. Murphy et al. (2011) and Hambly et al. (2010) also observed that chlorination generated a high removal rate of the

peak T fraction at recycled treatment plants.

Compared to peaks T and B components, peaks A and C are removed to a lower extent in the first stages of the treatment works (Fig. 2). Riopel et al. (2014) reported a reduction in the peak C component of 28 % and an increase in peak M with 4 % from influent to effluent. Cohen et al. (2014) found that one component in the Em > 380 nm region, sensitive to microbial activity, was removed, while other two components could not be removed by the biological treatment. Yu et al. (2013) observed a reduction in peak C - like component below 10 %. Later, Yu et al. (2014) showed that one component in the region Em > 380 nm increases from 6 % in the primary treatment to 19 % after the biological treatment. An increase in the fluorescence of this component was observed by Ou et al. (2014) in anoxic and aerobic treatments. Poor degradation of these components was also reported by Janhom et al. (2011) at an activated sludge treatment process. Yu et al. (2015b) found that with increasing retention times at sequencing batch reactor the peak C components increase in the soluble microbial products. These products are generated by substrate utilization or biomass decay and cell lysis, and are regarded as autochthonous matter. Cohen et al. (2014) and Riopel et al. (2014) suggest that these fluorescent components are either potentially produced during the process or are recalcitrant to decomposition. Riopel et al. (2014) mention that large

molecules degrade into smaller molecules that have a fulvic-like behavior, based on the polyphenol postulate of humic susbtances formation. They explain that due to the high microbial activity in WwTPs, the secreted exocellular enzymes will oxidize the polyphenols into quinones. The quinones will agglomerate with metabolites like amino acids or peptides, leading to the formation of humic polymers, which could be fulvic acids because they are smaller in size. Another explanation for the poor removal of these components is provided by Hur et al. (2011) who studied the fate of refractory OM in WwTPs. Refractory OM is not easily removed by the biological treatment process due to its recalcitrant nature. Moreover, Hur et al. (2011) showed that in most WwTPs, the percentage distribution of refractory OM increases in the effluents.

Tertiary and quaternary treatment stages are responsible for removing most of the fraction that fluoresces in the region Em > 380 nm (Fig. 2). Hambly et al. (2010) observed that chlorination generated a higher reduction in peak C compared to previous treatment steps. Singh et al. (2012) found a minimum of 97 % removal of peak C fluorophores after the reverse osmosis process. Murphy et al. (2011) also reported almost complete removal of components following reverse osmosis treatment step.

Removal of fluorescent compounds, like FWAs and PAHs, was also analysed. Bridgeman et al. (2013) found FWAs only in crude wastewater and not after other treatment steps, concluding that this fluorescent fraction associates with particulate matter, which is removed by the primary treatment stage. In addition, Tavares et al. (2008) stated that subsequent disinfection processes may further remove FWAs from wastewater. According to Hayashi et al. (2002), up to 80 % of FWAs are removed after the biological treatment, and thus these compounds could be used as molecular markers of less effective treatment processes. Ou et al. (2014) found that, for coke wastewater, the novel anoxic/aerobic/aerobic system successfully removed PAHs. While, Cohen et al. (2014) observed no reduction in the pyrene-like component along the treatment steps.

In most monitoring studies, other changes in the fluorescence spectra with regard to peak shape and position were observed. However, the findings regarding peak position are not consistent across studies, potentially due to differences in the treatment process or source of wastewater. For example, Zhu et al. (2011) observed that peak C presented a blue shift of 5 nm for the excitation wavelength and of 21 nm for the emission wavelength, from influent to effluent, at membrane bioreactor treated supermarket wastewater. Hur et al. (2011) reported a 20 nm excitation wavelength red shift between

influent and effluent, at refractory OM from municipal wastewater. Yet, Riopel et al. (2014), using PARAFAC, found no change in the peak C position or shape between sample locations. Riopel et al. (2014) observed that the PARAFAC component similar to peak T was elongated to longer wavelengths at influent samples compared to effluent. They attributed this elongation to the free or bound nature of the components. In the study of Zhu et al. (2011), peak T fluorescence displayed a red shift of 5 nm in the emission wavelength, from influent to effluent (Zhu et al. 2011). According to Zhu et al. (2011), the red shift is associated with the presence of carbonyl containing substances, hydroxyl, alkoxyl, amino groups and carboxyl constituents, while a blue shift is linked to a decomposition of condensed aromatic moieties and the break-up of the large molecules into small molecules.

5.4 Fluorescence control and optimisation of treatment processes

Increasingly stringent regulation has put major pressure on water utilities to find new technologies and implement control concepts that would improve the overall performance of WwTPs (Rehman et al. 2015). As discussed in previous sections, fluorescence spectroscopy has the potential to be used as a highly effective monitoring technique of treatment quality.

This could be achieved through the use of peak T fluorescence, which could replace the out-dated and inaccurate BOD (Bridgeman et al. 2013). Consequently, fluorescence spectroscopy could provide the WwTPs with the optimum tool for real-time control and remediation of plant performance failures (Chong et al. 2013).

Additionally, Bridgeman et al. (2013) and Ahmad and Reynolds (1995) suggested that fluorescence could improve the process control in activated sludge process. The bacteria and microorganisms that form the activated sludge are fed with wastewater containing organic waste. In order to sustain the biological activities into the activated sludge process for BOD reduction, air is pumped into the tanks to provide sufficient quantities of dissolved oxygen. Aeration is one of the most energy intensive operations from the WwTPs, almost 65 % of energy being consumed for the activated sludge process (Rehman et al. 2015). Water utilities often over aerate to ensure meeting discharge regulations (Bridgeman et al. 2013). It is estimated that, by monitoring OM in WwTPs, 40 % of the energy costs could be saved (Ahmad and Reynolds 1995). Thus, fluorescence may be used to optimize process control in treatment works and eliminate the unnecessary costs associated with overtreatment (Bridgeman et al. 2013).

Promising results regarding online monitoring and process control were obtained by Singh et al. (2015), who

published the first real-time study on two municipal recycled treatment plants. The researchers used a peak C sensor to prove the robustness of the technique in detecting reverse osmosis membrane fouling and integrity. They showed that the sensor was sufficiently sensitive to detect subtle differences between membrane permeates and identify underperformance issues. Also, no indication of fouling on probe and no deviation of probe performance were observed, during the experimental period. This study demonstrated the potential of using fluorescence for treatment process assessment and control.

6 Conclusions and future considerations

The use of real-time fluorescence could lead to a positive change in the water industry, as operators would be able to start immediate remedial actions in case of accidental pollution events, cut costs associated with complex analytical approaches and comply with discharge regulation. Wastewater treatment processes reduce peak T fluorescence primarily by biological treatment, and peak C through chlorination and reverse osmosis. There are several simple probes or fluorometers available on market that measure these two components or more complex systems that convert the peak T fluorescence signal into BOD values.

However, in case of monitoring surface waters contaminated with wastewater, the use of simple fluorometers

may not be the best solution to identify the exact source and take the appropriate remedial actions. Several fluorophores, with varied origins, were shown to contribute to peaks T and C, hindering the identification of the source of wastewater pollution in natural water systems. Single or double wavelength instruments could only be used as a time and cost effective first measure for early warning.

Implementation of fluorescence instrumentation for on-line monitoring is relatively slow due to several factors, such as high quantities of suspended solids, temperature, fouling etc. In order to counteract these issues, dilution of samples is recommended: to a factor of 10 or to an absorbance value of < 0.05 cm^{-1}, in the peak T absorbance region. However, wastewaters are highly variable in concentration and composition and therefore a general dilution factor may not be recommended. In addition, post-measurement mathematical correction could be applied to reduce the impact produced by external factors.

Acknowledgements

The work of E.M. Carstea was supported by the European Commission Framework Programme 7, through a Marie Curie Intra-European Fellowship (PIEF-GA-2012-329962).

References

Ahmad, S.R. and Reynolds, D.M. (1995) Synchronous fluorescence spectroscopy of wastewater and some potential constituents. Water Res 29(6), 1599-1602.

Ahmad, S.R. and Reynolds, D.M. (1999) Monitoring of water quality using fluorescence technique: prospect of on-line process control. Water Res 33(9), 2069-2074.

Aiken, G.R. (2014) Aquatic Organic Matter Fluorescence. Coble, P.G., Lead, J., Baker, A., Reynolds, D.M. and Spencer, R.G.M. (eds), pp. 35-75, Cambridge University Press, NY, USA.

Andersen, C.B., Lewis, G., Hart, M. and Pugh, J. (2014) The Impact of Wastewater Treatment Effluent on the Biogeochemistry of the Enoree River, South Carolina, During Drought Conditions. Water, Air, & Soil Pollution 225(5), 1-21.

Assaad, A., Pontvianne, S. and Pons, M.N. (2014) Photodegradation-based detection of fluorescent whitening agents in a mountain river. Chemosphere 100, 27-33.

Baker, A. (2001) Fluorescence Excitation−Emission Matrix Characterization of Some Sewage-Impacted Rivers. Environ Sci Technol 35(5), 948-953.

Baker, A. (2002a) Fluorescence Excitation−Emission Matrix Characterization of River Waters Impacted by a Tissue Mill Effluent. Environ Sci Technol 36(7), 1377-1382.

Baker, A. (2002b) Fluorescence properties of some farm wastes: implications for water quality monitoring. Water Res 36(1), 189-195.

Baker, A. and Inverarity, R. (2004) Protein-like fluorescence intensity as a possible tool for determining river water quality. Hydrological Processes 18(15), 2927-2945.

Baker, A., Andersen, M.S., Marjo, C.E., Zainuddin, N.S., Rutlidge, H., Graham, P.W. and Henderson, R.K. (2014) Investigation of Pollution in Rivers and Groundwater by Fluorescence. 1-14.

Baker, A., Cumberland, S.A., Bradley, C., Buckley, C. and Bridgeman, J. (2015) To what extent can portable fluorescence spectroscopy be used in the real-time assessment of microbial water quality? Sci Total Environ 532, 14-19.

Bassandeh, M., Antony, A., Le-Clech, P., Richardson, D. and Leslie, G. (2013) Evaluation of ion exchange resins for the removal of dissolved organic matter from biologically treated paper mill effluent. Chemosphere 90(4), 1461-1469.

Bieroza, M., Baker, A. and Bridgeman, J. (2009) Exploratory analysis of excitation-emission matrix fluorescence spectra with self-organizing maps as a basis for determination of organic matter removal efficiency at water treatment works. Journal of Geophysical Research: Biogeosciences 114(G4), G00F07.

Borisover, M., Laor, Y., Saadi, I., Lado, M. and Bukhanovsky, N. (2011) Tracing Organic Footprints from Industrial Effluent Discharge in Recalcitrant Riverine Chromophoric Dissolved Organic Matter. Water, Air, & Soil Pollution 222(1-4), 255-269.

Bourgeois, W., Burgess, J.E. and Stuetz, R.M. (2001) On-line monitoring of wastewater quality: a review. Journal of Chemical Technology & Biotechnology 76(4), 337-348.

1303 Boving, T., Meritt, D. and Boothroyd, J. (2004) Fingerprinting
1304 sources of bacterial input into small residential watersheds: fate of
1305 fluorescent whitening agents. Environmental Geology 46(2).
1306 Bridgeman, J., Baker, A., Carliell-Marquet, C. and Carstea, E.
1307 (2013) Determination of changes in wastewater quality through a
1308 treatment works using fluorescence spectroscopy. Environ Technol
1309 34(21-24), 3069-3077.
1310 Bridgeman, J., Baker, A., Brown, D. and Boxall, J.B. (2015)
1311 Portable LED fluorescence instrumentation for the rapid assessment
1312 of potable water quality. Science of The Total Environment 524-
1313 525C, 338-346.
1314 Bro, R. and Vidal, M. (2011) EEMizer: Automated modeling
1315 of fluorescence EEM data. Chemometrics and Intelligent Laboratory
1316 Systems 106(1), 86-92.
1317 Cahoon, L.B. and Mallin, M.A. (2013) Monitoring Water
1318 Quality. Ahuja, S. (ed), pp. 149-169, Elsevier, Amsterdam.
1319 Cao, Y., Griffith, J.F. and Weisberg, S.B. (2009) Evaluation of
1320 optical brightener photodecay characteristics for detection of human
1321 fecal contamination. Water Res 43(8), 2273-2279.
1322 Carstea, E.M., Baker, A., Pavelescu, G. and Boomer, I. (2009)
1323 Continuous fluorescence assessment of organic matter variability on
1324 the Bournbrook River, Birmingham, UK. Hydrological Processes
1325 23(13), 1937-1946.
1326 Carstea, E.M., Baker, A., Bieroza, M. and Reynolds, D. (2010)
1327 Continuous fluorescence excitation–emission matrix monitoring of
1328 river organic matter. Water Res 44(18), 5356-5366.
1329 Carstea, E.M., Baker, A., Bieroza, M., Reynolds, D.M. and
1330 Bridgeman, J. (2014) Characterisation of dissolved organic matter
1331 fluorescence properties by PARAFAC analysis and thermal
1332 quenching. Water Res 61(0), 152-161.
1333 Cawley, K.M., Butler, K.D., Aiken, G.R., Larsen, L.G.,
1334 Huntington, T.G. and McKnight, D.M. (2012) Identifying fluorescent
1335 pulp mill effluent in the Gulf of Maine and its watershed. Marine
1336 Pollution Bulletin 64(8), 1678-1687.
1337 Chandler, D.M. and Lerner, D.N. (2015) A low cost method to
1338 detect polluted surface water outfalls and misconnected drainage.
1339 Water and Environment Journal 29(2), 202-206.
1340 ChelseaInstruments (2015) UviLux BOD Indicator, Chelsea
1341 Instruments,
1342 http://www.chelsea.co.uk/marine/fluorometers/uvilux-bod-
1343 indicator.
1344 Chen, B., Wu, H. and Li, S.F.Y. (2014) Development of
1345 variable pathlength UV–vis spectroscopy combined with partial-
1346 least-squares regression for wastewater chemical oxygen demand
1347 (COD) monitoring. Talanta 120, 325-330.
1348 Chen, P., Pan, D., Mao, Z. and Tao, B. (2015) Detection of
1349 water quality parameters in Hangzhou Bay using a portable laser
1350 fluorometer. Marine Pollution Bulletin 93(1-2), 163-171.
1351 Chong, S., Aziz, A. and Harun, S. (2013) Fibre Optic Sensors
1352 for Selected Wastewater Characteristics. Sensors (Basel) 13(7),
1353 8640-8668.
1354 Ciputra, S., Antony, A., Phillips, R., Richardson, D. and
1355 Leslie, G. (2010) Comparison of treatment options for removal of
1356 recalcitrant dissolved organic matter from paper mill effluent.
1357 Chemosphere 81(1), 86-91.

Coble, P.G., Green, S.A., Blough, N.V. and Gagosian, R.B. (1990) Characterization of dissolved organic matter in the Black Sea by fluorescence spectroscopy. Nature 348(6300), 432-435.

Coble, P.G. (1996) Characterization of marine and terrestrial DOM in seawater using excitation-emission matrix spectroscopy. Marine Chemistry 51(4), 325-346.

Cohen, E., Levy, G.J. and Borisover, M. (2014) Fluorescent components of organic matter in wastewater: Efficacy and selectivity of the water treatment. Water Res 55(0), 323-334.

Conmy, R.N., Coble, P.G. and Castillo, C.E.D. (2004) Calibration and performance of a new in situ multi-channel fluorometer for measurement of colored dissolved organic matter in the ocean. Continental Shelf Research 24(3), 431-442.

Cumberland, S., Bridgeman, J., Baker, A., Sterling, M. and Ward, D. (2012) Fluorescence spectroscopy as a tool for determining microbial quality in potable water applications. Environ Technol 33(4-6), 687-693.

del Olmo, M., Díez, C., Molina, A., de Orbe, I. and Vílchez, J.L. (1996) Resolution of phenol, o-cresol, m-cresol and p-cresol mixtures by excitation fluorescence using partial least-squares (PLS) multivariate calibration. Analytica Chimica Acta 335(1–2), 23-33.

Dignac, M.F., Ginestet, P., Rybacki, D., Bruchet, A., Urbain, V. and Scribe, P. (2000) Fate of wastewater organic pollution during activated sludge treatment: nature of residual organic matter. Water Res 34(17), 4185-4194.

Downing, B.D., Boss, E., Bergamaschi, B.A., Fleck, J.A., Lionberger, M.A., Ganju, N.K., Schoellhamer, D.H. and Fujii, R. (2009) Quantifying fluxes and characterizing compositional changes of dissolved organic matter in aquatic systems in situ using combined acoustic and optical measurements. Limnology and Oceanography: Methods 7(1), 119-131.

Drinan, J.E. and Spellman, F. (2012) Water and Wastewater Treatment: A Guide for the Nonengineering Professional, CRC Press, FL, USA.

Drozdowska, V. (2007) The lidar investigation of the upper water layer fluorescence spectra of the Baltic Sea. The European Physical Journal Special Topics 144(1), 141-145.

Ellis, T.G. (2004) Encyclopedia of Life Support System, Eolss Publishers, Oxford, UK

Fellman, J.B., Hood, E. and Spencer, R.G.M. (2010) Fluorescence spectroscopy opens a new windows into dissolved organic matter dynamics in freshwater ecosystems: A review. Limnology and Oceanography 55(6), 2452-2462.

Ferretto, N., Tedetti, M., Guigue, C., Mounier, S., Redon, R. and Goutx, M. (2014) Identification and quantification of known polycyclic aromatic hydrocarbons and pesticides in complex mixtures using fluorescence excitation-emission matrices and parallel factor analysis. Chemosphere 107, 344-353.

Galinha, C.F., Carvalho, G., Portugal, C.A.M., Guglielmi, G., Oliveira, R., Crespo, J.G. and Reis, M.A.M. (2011a) Real-time monitoring of membrane bioreactors with 2D-fluorescence data and statistically based models. Water Science and Technology 63(7), 1381-1388.

Galinha, C.F., Carvalho, G., Portugal, C.A.M., Guglielmi, G., Reis, M.A.M. and Crespo, J.G. (2011b) Two-dimensional

1413 fluorescence as a fingerprinting tool for monitoring wastewater
1414 treatment systems. Journal of Chemical Technology &
1415 Biotechnology 86(7), 985-992.
1416 Ghervase, L., Carstea, E.M., Pavelescu, G., Borisova, E. and
1417 Daskalova, A. (2010a) Fluorescence evaluation of anthropogenic
1418 influence on rivers crossing Sofia Romanian Reports in Physics
1419 62(1), 193-201.
1420 Ghervase, L., Carstea, E.M., Pavelescu, G. and Savastru, D.
1421 (2010b) Laser Induced Fluorescence efficiency in water quality
1422 assessment Romanian Reports in Physics 62(3), 652-659.
1423 Gholami, A., Masoum, S., Mohsenikia, A. and Abbasi, S.
1424 (2015) Chemometrics-assisted excitation-emission fluorescence
1425 analytical data for rapid and selective determination of optical
1426 brighteners in the presence of uncalibrated interferences.
1427 Spectrochim Acta A Mol Biomol Spectrosc 153, 108-117.
1428 Goldman, J.H., Rounds, S.A. and Needoba, J.A. (2012)
1429 Applications of fluorescence spectroscopy for predicting percent
1430 wastewater in an urban stream. Environ Sci Technol 46(8), 4374-
1431 4381.
1432 Graham, P.W., Baker, A., Andersen, M.S. and Acworth, I.
1433 (2015) Field Measurement of Fluorescent Dissolved Organic
1434 Material as a Means of Early Detection of Leachate Plumes. Water,
1435 Air, & Soil Pollution 226(7).
1436 Hambly, A.C., Henderson, R.K., Storey, M.V., Baker, A.,
1437 Stuetz, R.M. and Khan, S.J. (2010) Fluorescence monitoring at a
1438 recycled water treatment plant and associated dual distribution
1439 system--implications for cross-connection detection. Water Res
1440 44(18), 5323-5333.
1441 Hartel, P.G., Hagedorn, C., McDonald, J.L., Fisher, J.A.,
1442 Saluta, M.A., Dickerson, J.W., Jr., Gentit, L.C., Smith, S.L.,
1443 Mantripragada, N.S., Ritter, K.J. and Belcher, C.N. (2007) Exposing
1444 water samples to ultraviolet light improves fluorometry for detecting
1445 human fecal contamination. Water Res 41(16), 3629-3642.
1446 Hayashi, Y., Managaki, S. and Takada, H. (2002) Fluorescent
1447 Whitening Agents in Tokyo Bay and Adjacent Rivers: Their
1448 Application as Anthropogenic Molecular Markers in Coastal
1449 Environments. Environ Sci Technol 36(16), 3556-3563.
1450 He, Q., Yao, K., Sun, D. and Shi, B. (2007) Biodegradability
1451 of tannin-containing wastewater from leather industry.
1452 Biodegradation 18(4), 465-472.
1453 Henderson, R.K., Baker, A., Murphy, K.R., Hambly, A.,
1454 Stuetz, R.M. and Khan, S.J. (2009) Fluorescence as a potential
1455 monitoring tool for recycled water systems: a review. Water Res
1456 43(4), 863-881.
1457 Huang, M.-h., Li, Y.-m. and Gu, G.-w. (2010) Chemical
1458 composition of organic matters in domestic wastewater. Desalination
1459 262(1–3), 36-42.
1460 Hudson, N., Baker, A. and Reynolds, D. (2007) Fluorescence
1461 analysis of dissolved organic matter in natural, waste and polluted
1462 waters—a review. River Research and Applications 23(6), 631-649.
1463 Huguet, A., Vacher, L., Relexans, S., Saubusse, S., Froidefond,
1464 J.M. and Parlanti, E. (2009) Properties of fluorescent dissolved
1465 organic matter in the Gironde Estuary. Organic Geochemistry 40(6),
1466 706-719.

1467 Hur, J. and Kong, D.S. (2008) Use of synchronous
1468 fluorescence spectra to estimate biochemical oxygen demand (BOD)
1469 of urban rivers affected by treated sewage. Environ Technol 29(4),
1470 435-444.
1471 Hur, J., Lee, T.-H. and Lee, B.-M. (2011) Estimating the
1472 removal efficiency of refractory dissolved organic matter in
1473 wastewater treatment plants using a fluorescence technique. Environ
1474 Technol 32(16), 1843-1850.
1475 Hur, J. and Cho, J. (2012) Prediction of BOD, COD, and Total
1476 Nitrogen Concentrations in a Typical Urban River Using a
1477 Fluorescence Excitation-Emission Matrix with PARAFAC and UV
1478 Absorption Indices. Sensors (Basel) 12(1), 972-986.
1479 Ishii, S.K.L. and Boyer, T.H. (2012) Behavior of Reoccurring
1480 PARAFAC Components in Fluorescent Dissolved Organic Matter in
1481 Natural and Engineered Systems: A Critical Review. Environ Sci
1482 Technol 46(4), 2006-2017.
1483 Janhom, T., Wattanachira, S. and Pavasant, P. (2009)
1484 Characterization of brewery wastewater with spectrofluorometry
1485 analysis. J Environ Manage 90(2), 1184-1190.
1486 Janhom, T., Pavasant, P. and Wattanachira, S. (2011) Profiling
1487 and monitoring of DOM in brewery wastewater and treated
1488 wastewater. Environ Monit Assess 176(1-4), 403-418.
1489 Jouanneau, S., Recoules, L., Durand, M.J., Boukabache, A.,
1490 Picot, V., Primault, Y., Lakel, A., Sengelin, M., Barillon, B. and
1491 Thouand, G. (2014) Methods for assessing biochemical oxygen
1492 demand (BOD): A review. Water Res 49, 62-82.
1493 Khamis, K., Sorensen, J.P.R., Bradley, C., Hannah, D.M.,
1494 Lapworth, D.J. and Stevens, R. (2015) In situ tryptophan-like
1495 fluorometers: assessing turbidity and temperature effects for
1496 freshwater applications. Environmental Science: Processes &
1497 Impacts.
1498 Kothawala, D.N., Murphy, K.R., Stedmon, C.A.,
1499 Weyhenmeyer, G.A. and Tranvik, L.J. (2013) Inner filter correction
1500 of dissolved organic matter fluorescence. Limnology and
1501 Oceanography: Methods 11(12), 616-630.
1502 Kramer, J.B., Canonica, S., Hoigné, J. and Kaschig, J. (1996)
1503 Degradation of Fluorescent Whitening Agents in Sunlit Natural
1504 Waters. Environ Sci Technol 30(7), 2227-2234.
1505 Lakowicz, J. (2006) Principles of Fluorescence Spectroscopy,
1506 Springer US, NY, USA.
1507 Li, W., Xu, Z., Wu, Q., Li, Y., Shuang, C. and Li, A. (2015)
1508 Characterization of fluorescent-dissolved organic matter and
1509 identification of specific fluorophores in textile effluents.
1510 Environmental Science and Pollution Research 22(6), 4183-4189.
1511 Li, W.T., Chen, S.Y., Xu, Z.X., Li, Y., Shuang, C.D. and Li,
1512 A.M. (2014) Characterization of dissolved organic matter in
1513 municipal wastewater using fluorescence PARAFAC analysis and
1514 chromatography multi-excitation/emission scan: a comparative study.
1515 Environ Sci Technol 48(5), 2603-2609.
1516 Louvet, J.N., Homeky, B., Casellas, M., Pons, M.N. and
1517 Dagot, C. (2013) Monitoring of slaughterhouse wastewater
1518 biodegradation in a SBR using fluorescence and UV-Visible
1519 absorbance. Chemosphere 91(5), 648-655.
1520 McGowin, A.E. (2005) Chromatographic Science Series.
1521 Nollet, L.M.L. (ed), p. 599, CRC Press, Boca Raton, FL.

McKnight, D.M., Boyer, E.W., Westerhoff, P.K., Doran, P.T., Kulbe, T. and Andersen, D.T. (2001) Spectrofluorometric characterization of dissolved organic matter for indication of precursor organic material and aromaticity. Limnology and Oceanography 46(1), 38-48.

Meng, F., Huang, G., Yang, X., Li, Z., Li, J., Cao, J., Wang, Z. and Sun, L. (2013) Identifying the sources and fate of anthropogenically impacted dissolved organic matter (DOM) in urbanized rivers. Water Res 47(14), 5027-5039.

Michael, I., Michael, C., Duan, X., He, X., Dionysiou, D.D., Mills, M.A. and Fatta-Kassinos, D. (2015) Dissolved effluent organic matter: Characteristics and potential implications in wastewater treatment and reuse applications. Water Res 77, 213-248.

ModernWater (2015) BODCheck Modern Water, http://www.modernwater.com/assets/downloads/Factsheets/MW_Factsheet_BODChek_highres.pdf.

Mostofa, K.G., Wu, F., Liu, C.-Q., Fang, W., Yuan, J., Ying, W., Wen, L. and Yi, M. (2010) Characterization of Nanming River (southwestern China) sewerage-impacted pollution using an excitation-emission matrix and PARAFAC. Limnology 11(3), 217-231.

Murphy, K.R., Hambly, A., Singh, S., Henderson, R.K., Baker, A., Stuetz, R. and Khan, S.J. (2011) Organic matter fluorescence in municipal water recycling schemes: toward a unified PARAFAC model. Environ Sci Technol 45(7), 2909-2916.

Murphy, K.R., Stedmon, C.A., Wenig, P. and Bro, R. (2014) OpenFluor- an online spectral library of auto-fluorescence by organic compounds in the environment. Analytical Methods 6(3), 658-661.

Navalon, S., Alvaro, M. and Garcia, H. (2011) Analysis of organic compounds in an urban wastewater treatment plant effluent. Environ Technol 32(3), 295-306.

Old, G.H., Naden, P.S., Granger, S.J., Bilotta, G.S., Brazier, R.E., Macleod, C.J.A., Krueger, T., Bol, R., Hawkins, J.M.B., Haygarth, P. and Freer, J. (2012) A novel application of natural fluorescence to understand the sources and transport pathways of pollutants from livestock farming in small headwater catchments. Science of The Total Environment 417–418(0), 169-182.

Orhon, D., Babuna, F.G. and Karahan, O. (2009) Industrial Wastewater Treatment by Activated Sludge, IWA Publishing, London.

Ou, H.S., Wei, C.H., Mo, C.H., Wu, H.Z., Ren, Y. and Feng, C.H. (2014) Novel insights into anoxic/aerobic(1)/aerobic(2) biological fluidized-bed system for coke wastewater treatment by fluorescence excitation-emission matrix spectra coupled with parallel factor analysis. Chemosphere 113, 158-164.

Patra, D. and Mishra, A.K. (2002) Recent developments in multi-component synchronous fluorescence scan analysis. TrAC Trends in Analytical Chemistry 21(12), 787-798.

Petrenko, A.A., Jones, B.H., Dickey, T.D., LeHaitre, M. and Moore, C. (1997) Effects of a sewage plume on the biology, optical characteristics, and particle size distributions of coastal waters. Journal of Geophysical Research 102(C11), 25061.

Pfeiffer, E., Pavelescu, G., Baker, A., Roman, C., Ioja, C. and Savastru, D. (2008) Pollution analysis on the Arges River using

fluorescence spectroscopy. Journal of optoelectronics and advanced materials 10(6), 1489-1494.

Poiger, T., Field, J.A., Field, T.M., Siegrist, H. and Giger, W. (1998) Behavior of fluorescent whitening agents during sewage treatment. Water Res 32(6), 1939-1947.

Pokhrel, D. and Viraraghavan, T. (2004) Treatment of pulp and paper mill wastewater--a review. Sci Total Environ 333(1-3), 37-58.

Rehman, U., Vesvikar, M., Maere, T., Guo, L., Vanrolleghem, P.A. and Nopens, I. (2015) Effect of sensor location on controller performance in a wastewater treatment plant. Water Science & Technology 71(5), 700-708.

Reynolds, D. (2014) Aquatic Organic Matter Fluorescence. Coble, P.G., Lead, J., Baker, A., Reynolds, D.M. and Spencer, R.G.M. (eds), pp. 3-35, Cambridge University Press, NY, USA.

Reynolds, D.M. and Ahmad, S.R. (1995) The effect of metal ions on the fluorescence of sewage wastewater. Water Res 29(9), 2214-2216.

Reynolds, D.M. and Ahmad, S.R. (1997) Rapid and direct determination of wastewater BOD values using a fluorescence technique. Water Res 31(8), 2012-2018.

Reynolds, D.M. (2002) The differentiation of biodegradable and non-biodegradable dissolved organic matter in wastewaters using fluorescence spectroscopy. Journal of Chemical Technology & Biotechnology 77(8), 965-972.

Reynolds, D.M. (2003) Rapid and direct determination of tryptophan in water using synchronous fluorescence spectroscopy. Water Res 37(13), 3055-3060.

Riopel, R., Caron, F. and Siemann, S. (2014) Fluorescence Characterization of Natural Organic Matter at a Northern Ontario Wastewater Treatment Plant. Water, Air, & Soil Pollution 225(9), 1-17.

Ryder, A. (2005) Reviews in Fluorescence 2005. Geddes, C. and Lakowicz, J. (eds), pp. 169-198, Springer US.

Sánchez Rojas, F. and Bosch Ojeda, C. (2005) Effluent analysis in analytical chemistry: an overview. Anal Bioanal Chem 382(4), 978-991.

Santos, E.B.H., Filipe, O.M.S., Duarte, R.M.B.O., Pinto, H. and Duarte, A.C. (2001) Fluorescence as a Tool for Tracing the Organic Contamination from Pulp Mill Effluents in Surface Waters. Acta hydrochimica et hydrobiologica 28(7), 364-371.

Shutova, Y., Baker, A., Bridgeman, J. and Henderson, R.K. (2014) Spectroscopic characterisation of dissolved organic matter changes in drinking water treatment: From PARAFAC analysis to online monitoring wavelengths. Water Res 54, 159-169.

Singh, S., Henderson, R.K., Baker, A., Stuetz, R.M. and Khan, S.J. (2009) Distinguishing Stage 1 and 2 reverse osmosis permeates using fluorescence spectroscopy. Water Sci Technol 60(8), 2017-2023.

Singh, S., Henderson, R.K., Baker, A., Stuetz, R.M. and Khan, S.J. (2012) Characterisation of reverse osmosis permeates from municipal recycled water systems using fluorescence spectroscopy: Implications for integrity monitoring. Journal of Membrane Science 421-422, 180-189.

1630 Singh, S., Henderson, R.K., Baker, A., Stuetz, R.M. and Khan,
1631 S.J. (2015) Online fluorescence monitoring of RO fouling and
1632 integrity: analysis of two contrasting recycled water schemes.
1633 Environ. Sci.: Water Res. Technol. 1(5), 689-698.
1634 Sorensen, J.P., Lapworth, D.J., Marchant, B.P., Nkhuwa, D.C.,
1635 Pedley, S., Stuart, M.E., Bell, R.A., Chirwa, M., Kabika, J., Liemisa,
1636 M. and Chibesa, M. (2015) In-situ tryptophan-like fluorescence: A
1637 real-time indicator of faecal contamination in drinking water
1638 supplies. Water Res 81, 38-46.
1639 Sorensen, J.P.R., Sadhu, A., Sampath, G., Sugden, S., Dutta
1640 Gupta, S., Lapworth, D.J., Marchant, B.P. and Pedley, S. (2016) Are
1641 sanitation interventions a threat to drinking water supplies in rural
1642 India? An application of tryptophan-like fluorescence. Water Res 88,
1643 923-932.
1644 Spencer, R.G., Baker, A., Ahad, J.M., Cowie, G.L.,
1645 Ganeshram, R., Upstill-Goddard, R.C. and Uher, G. (2007a)
1646 Discriminatory classification of natural and anthropogenic waters in
1647 two U.K. estuaries. Science of The Total Environment 373(1), 305-
1648 323.
1649 Spencer, R.G.M., Pellerin, B.A., Bergamaschi, B.A., Downing,
1650 B.D., Kraus, T.E.C., Smart, D.R., Dahlgren, R.A. and Hernes, P.J.
1651 (2007b) Diurnal variability in riverine dissolved organic matter
1652 composition determined byin situ optical measurement in the San
1653 Joaquin River (California, USA). Hydrological Processes 21(23),
1654 3181-3189.
1655 Stedmon, C.A. and Cory, R.M. (2014) Aquatic Organic Matter
1656 Fluorescence. Coble, P.G., Lead, J., Baker, A., Reynolds, D.M. and
1657 Spencer, R.G.M. (eds), pp. 278-303, Cambridge University Press,
1658 NY, USA.
1659 Suthar, S., Sharma, J., Chabukdhara, M. and Nema, A. (2010)
1660 Water quality assessment of river Hindon at Ghaziabad, India:
1661 impact of industrial and urban wastewater. Environ Monit Assess
1662 165(1-4), 103-112.
1663 Takahashi, M. and Kawamura, K. (2006) Simple Measurement
1664 of 4,4'-bis(2-sulfostyryl)-biphenyl in River Water by Fluorescence
1665 Analysis and Its Application as an Indicator of Domestic Wastewater
1666 Contamination. Water, Air, and Soil Pollution 180(1-4), 39-49.
1667 Tartakovsky, B., Lishman, L.A. and Legge, R.L. (1996)
1668 Application of multi-wavelength fluorometry for monitoring
1669 wastewater treatment process dynamics. Water Res 30(12), 2941-
1670 2948.
1671 Tavares, M.E., Spivey, M.I.H., McIver, M.R. and Mallin, M.A.
1672 (2008) Testing for optical brighteners and fecal bacteria to detect
1673 sewage leaks in tidal creeks. Journal of the North Carolina Academy
1674 of Science 124(3), 91-97.
1675 Tchaikovskaya, O., Sokolova, I., Svetlichnyi, V., Karetnikova,
1676 E., Fedorova, E. and Kudryasheva, N. (2007) Fluorescence and
1677 bioluminescence analysis of sequential UV-biological degradation of
1678 p-cresol in water. Luminescence 22(1), 29-34.
1679 Tchobanoglous, G., Burton, F.L., Metcalf and Eddy (1991)
1680 Wastewater engineering : treatment, disposal, and reuse, McGraw-
1681 Hill, New York.
1682 Tedetti, M., Longhitano, R., Garcia, N., Guigue, C., Ferretto,
1683 N. and Goutx, M. (2012) Fluorescence properties of dissolved
1684 organic matter in coastal Mediterranean waters influenced by a

1685 municipal sewage effluent (Bay of Marseilles, France).
1686 Environmental Chemistry 9(5), 438-449.
1687 Tedetti, M., Joffre, P. and Goutx, M. (2013) Development of a
1688 field-portable fluorometer based on deep ultraviolet LEDs for the
1689 detection of phenanthrene- and tryptophan-like compounds in natural
1690 waters. Sensors and Actuators B: Chemical 182(0), 416-423.
1691 Tertuliani, J.S., Alvarez, D.A., Furlong, E.T., Meyer, M.T.,
1692 Zaugg, S.D. and Koltun, G.F. (2008) Occurrence of organic
1693 wastewater compounds in the Tinkers Creek watershed and two other
1694 tributaries to the Cuyahoga River, Northeast Ohio, p. 60, U.S.
1695 Geological Survey.
1696 Tsuneda, S., Aikawa, H., Hayashi, H., Yuasa, A. and Hirata,
1697 A. (2003) Extracellular polymeric substances responsible for
1698 bacterial adhesion onto solid surface. FEMS Microbiology Letters
1699 223(2), 287-292.
1700 Wang, X.-H., Wang, X., Huppes, G., Heijungs, R. and Ren,
1701 N.-Q. (2015) Environmental implications of increasingly stringent
1702 sewage discharge standards in municipal wastewater treatment
1703 plants: case study of a cool area of China. Journal of Cleaner
1704 Production 94, 278-283.
1705 Wang, Z., Cao, J. and Meng, F. (2014) Interactions between
1706 protein-like and humic-like components in dissolved organic matter
1707 revealed by fluorescence quenching. Water Res 68C, 404-413.
1708 Wang, Z.-g., Liu, W.-q., Zhao, N.-j., Li, H.-b., Zhang, Y.-j.,
1709 Si-Ma, W.-c. and Liu, J.-g. (2007) Composition analysis of colored
1710 dissolved organic matter in Taihu Lake based on three dimension
1711 excitation-emission fluorescence matrix and PARAFAC model, and
1712 the potential application in water quality monitoring. Journal of
1713 Environmental Sciences 19(7), 787-791.
1714 Watras, C.J., Hanson, P.C., Stacy, T.L., Morrison, K.M.,
1715 Mather, J., Hu, Y.H. and Milewski, P. (2011) A temperature
1716 compensation method for CDOM fluorescence sensors in freshwater.
1717 Limnology and Oceanography: Methods 9(7), 296-301.
1718 Wei, D., Wang, B., Ngo, H.H., Guo, W., Han, F., Wang, X.,
1719 Du, B. and Wei, Q. (2015) Role of extracellular polymeric
1720 substances in biosorption of dye wastewater using aerobic granular
1721 sludge. Bioresour Technol 185, 14-20.
1722 Xue, S., Zhao, Q., Ma, X., Li, F., Wang, J. and Wei, L. (2011)
1723 Comparison of dissolved organic matter fractions in a secondary
1724 effluent and a natural water. Environ Monit Assess 180(1-4), 371-
1725 383.
1726 Yamashita, Y., Lu, C.J., Ogawa, H., Nishioka, J., Obata, H.
1727 and Saito, H. (2015) Application of an in situ fluorometer to
1728 determine the distribution of fluorescent organic matter in the open
1729 ocean. Marine Chemistry 177, Part 2, 298-305.
1730 Yang, L., Han, D.H., Lee, B.-M. and Hur, J. (2015a)
1731 Characterizing treated wastewaters of different industries using
1732 clustered fluorescence EEM–PARAFAC and FT-IR spectroscopy:
1733 Implications for downstream impact and source identification.
1734 Chemosphere 127(0), 222-228.
1735 Yang, L., Hur, J. and Zhuang, W. (2015b) Occurrence and
1736 behaviors of fluorescence EEM-PARAFAC components in drinking
1737 water and wastewater treatment systems and their applications: a
1738 review. Environmental Science and Pollution Research, 1-11.

1739 Yu, H., Song, Y., Tu, X., Du, E., Liu, R. and Peng, J. (2013)
1740 Assessing removal efficiency of dissolved organic matter in
1741 wastewater treatment using fluorescence excitation emission matrices
1742 with parallel factor analysis and second derivative synchronous
1743 fluorescence. Bioresour Technol 144, 595-601.
1744 Yu, H., Song, Y., Liu, R., Pan, H., Xiang, L. and Qian, F.
1745 (2014) Identifying changes in dissolved organic matter content and
1746 characteristics by fluorescence spectroscopy coupled with self-
1747 organizing map and classification and regression tree analysis during
1748 wastewater treatment. Chemosphere 113, 79-86.
1749 Yu, H., Liang, H., Qu, F., Han, Z.-s., Shao, S., Chang, H. and
1750 Li, G. (2015a) Impact of dataset diversity on accuracy and sensitivity
1751 of parallel factor analysis model of dissolved organic matter
1752 fluorescence excitation-emission matrix. Scientific Reports 5, 10207.
1753 Yu, H., Qu, F., Sun, L., Liang, H., Han, Z., Chang, H., Shao, S.
1754 and Li, G. (2015b) Relationship between soluble microbial products
1755 (SMP) and effluent organic matter (EfOM): Characterized by
1756 fluorescence excitation emission matrix coupled with parallel factor
1757 analysis. Chemosphere 121, 101-109.
1758 ZAPSTechnologies (2015) LiquID Station, ZAPS
1759 Technologies, http://www.zapstechnologies.com/the-liquid-
1760 station/.
1761 Zhou, Y., Jeppesen, E., Zhang, Y., Shi, K., Liu, X. and Zhu, G.
1762 (2015) Dissolved organic matter fluorescence at wavelength 275/342
1763 nm as a key indicator for detection of point-source contamination in a
1764 large Chinese drinking water lake. Chemosphere 144, 503-509.
1765 Zhu, X., Wang, Z. and Wu, Z. (2011) Characterization of
1766 membrane foulants in a full-scale membrane bioreactor for
1767 supermarket wastewater treatment. Process Biochemistry 46(4),
1768 1001-1009.

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Figure 1. Main techniques of processing fluorescence EEMs. Examples of a) peaks identified with the peak picking method, and b) components identified with PARAFAC, for samples of water systems impacted by domestic wastewater.

Figure 2. Removal of fluorescent components during treatment; the removal percentages represent collective values from several studies (Tchobanoglous and Burton 1991, Reynolds 2002, Hambly et al. 2010, Janhom et al. 2011, Murphy et al. 2011, Singh et al. 2012, Cohen et al. 2014, Ou et al. 2014, Riopel et al. 2014, Yu et al. 2014) and unpublished data. Blue arrow – low decrease, Orange arrow – moderate removal, red arrow – high removal.

Table 1. Fluorophores contributing to regions Em < 380 nm >.

Potential fluorophores	Component	Region	Peak position (nm)	Reference	Potential sources in Ww
Lignins	Lignin phenols	Em < 380 nm	~ 245 (295) / 302	Walker et al. (2009)	Partially degraded food waste, undigested dietary fibre, toilet paper etc. Wastewater of paper and pulp industry (Pokhrel and Viraraghavan 2004) fibres from food (Huang et al. 2010)
			270-290 / 300-350	(Hernes et al. 2009)	
	Vanilic acid		/ 326	(Stedmon and Cory 2014)	
	Syringic acid		/ 338	(Stedmon and Cory 2014)	
	Breakdown products	Em > 380 nm	230-275 (300-390) / 400-520	(Baker 2002b, Ciputra et al. 2010, Osburn and Stedmon 2011, Cawley et al. 2012, Bassandeh et al. 2013)	Paper mill effluents (Baker 2002b, Ciputra et al. 2010, Cawley et al. 2012, Bassandeh et al. 2013)
Aromatic hydrocarbon	Toluene	Em < 380 nm	266 / 300 - 400	Persichetti et al. (2013)	Municipal Ww (Huang et al. 2010, Mrowiec 2014); Ww with petrol derivatives (Mehdizadeh et al. 2011)
Phenols	Cresols		210-285 / 290-310	del Olmo et al. (1996)	Pharmaceutical, fossil fuel or pesticide industries (Tchaikovskaya et al. 2007); Domestic Ww from disinfectants (Tertuliani et al. 2008)
Aromatic amino acids	Tyrosine		275 / 304	Lakowicz (2006)	Proteins and peptides (Lakowicz 2006); Domestic Ww(Burleson et al. 1980, Dignac et al. 2000, Huang et al. 2010)
	Tryptophan		295 / 353	Lakowicz (2006)	Proteins and peptides (Lakowicz 2006); Livestock Ww (Choi et al. 2013)
Indole			230 / 330-350	Determann et al. (1998)	Municipal Ww (Dignac et al. 2000, Tertuliani et al. 2008, Huang et al. 2010); Coal tar, oil shale, personal care products, pesticides and pharmaceuticals (Gu and Berry 1991, Tertuliani et al. 2008, Aiken 2014)
DNA			267 / 327	Vayá et al. (2010)	Proteins (Lakowicz 2006); Municipal Ww (Huang et al. 2010)
Polyaromatic hydrocarbons		Em < 380 nm	Short UV	Baker et al. (2014)	Municipal Ww (Guo et al. 2010, Huang et al. 2010); Landfill leachate (Baker and Curry 2004)
	Phenanthrene, anthracene, pyrene, fluoranthene, benzo[a]pyrene	Em > 380 nm	220-300 / 370-430	(Schwarz and Wasik 1976, Patra and Mishra 2001, Yang et al. 2016)	Industrial Ww (Cohen et al. 2014, Ou et al. 2014); Municipal Ww (Huang et al. 2010)
Quinones		Em > 380 nm			Microbes, fungi, plants (Aiken 2014); Activated sludge (Hu et al. 2000)
Flavonoids					Plants (Aiken 2014); food (Egert and Rimbach 2011); olive oil mill Ww (Leouifoudi et al. 2014)
Humic acids			220-320 (400-	IHSS (2015)	

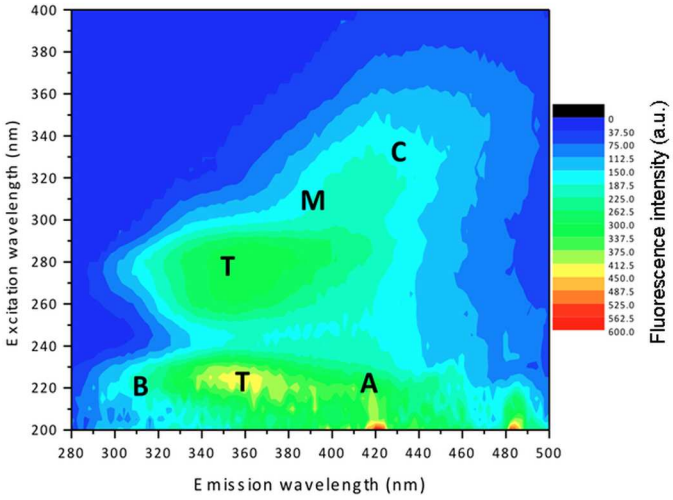
			500) / 400-550		Municipal Ww (Huang et al. 2010)
Pharmaceutically active compounds	Carbamazepine		308 / 410 (in 2 mol L^{-1} HCl, and 20 min irradiation time)	Hurtado-Sanchez Mdel et al. (2015)	Faeces, urine (Zhang et al. 2008)
	Fluorquinolone		290 / 500		
	Piroxican		294 / 372 (in media with pH < 2)		
Fluorescent whitening agents			360-365 / 400 - 440	(Takahashi and Kawamura 2006, Tavares et al. 2008)	Laundry detergents, sanitary products, toilet paper and tissues; Papermaking industry (Takahashi and Kawamura 2006, Assaad et al. 2014)

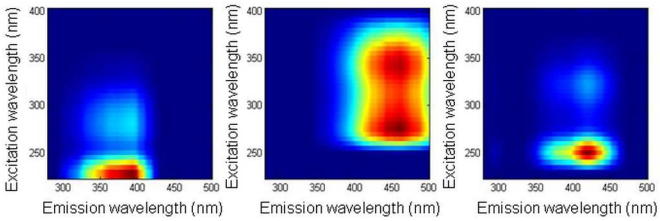
Ww – wastewater

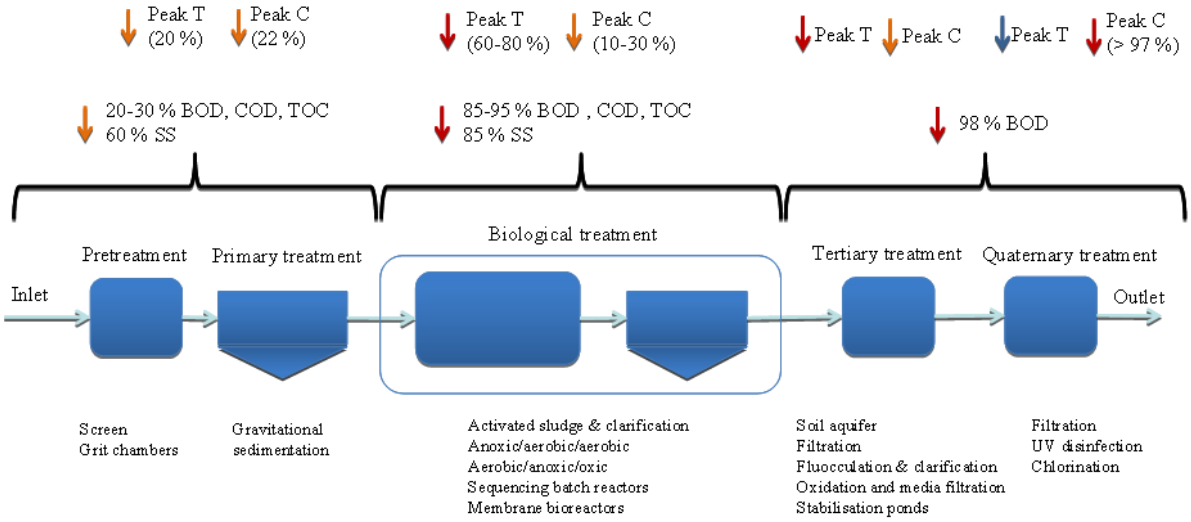
Table 2. Correlation coefficients for peaks T and C (or PARAFAC analogous components) with BOD, COD and TOC.

Reference	Samples	Sample size	Sample pH	Analysis temperature	Fluorescence Peak	BOD	COD	TOC
Reynolds and Ahmad (1997)	Raw, settled and treated Ww	129	N/A	Room temperature	280 / 340	0.94-0.97	N/A	N/A
Ahmad and Reynolds (1999)	Raw Ww	25	3 - 7	10-80^0 C	248 / 350	0.97	N/A	N/A
Reynolds (2002)	Raw Ww	56	6.8 ±0.4	26 ±10^0 C	280 / 350	0.93	0.94	0.93
Baker and Inverarity (2004)	Ww effluents and effluent impacted rivers	434	N/A	N/A	220 / 350	0.85	N/A	N/A
Wang et al. (2007)	Ww impacted lake	26	N/A	Room temperature	294 / 320	0.54	0.16	N/A
					360 / 425	0.65	0.03	N/A
Hudson et al. (2008)	Ww effluents	141	N/A	20^0 C	280 / 350	0.71	N/A	0.77
					300-370 / 400-500-	0.34	N/A	0.75
Bridgeman et al. (2013)	Domestic Ww, raw and treated	48	N/A	20^0 C	275-285 / 340-360	0.92	0.56	N/A
					320-355 / 410-470	0.88	0.78	N/A
Cohen et al. (2014)	Domestic and industrial Ww, raw and treated	25-34	7.8 – 8.5	Room temperature	<240 (275) / 346	0.82	0.82-0.99	0.85-0.99
					<240 (305) / 422	0.72	0.91	0.99
Ou et al. (2014)	Industrial Ww, raw and treated	120	7 - 9	Room temperature	280 / 320	N/A	0.92	N/A

Ww – wastewater; N/A – not available







- Several fluorophores contribute to common peaks hindering pollution source tracking
- Previous on-line studies may help build a strategy for wastewater analysis
- Dilution of samples, typically up to x10, useful to limit inner filter effect
- Calibration may not be needed for qualitative data
- Research gaps: online application of fluorescence and rapid data processing tools