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**Identification and structural characterisation of a partially arabinosylated
lipoarabinomannan variant isolated from a *Corynebacterium glutamicum ubiA-*
mutant**

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Running title: arabinosylated LAM variant from a *Corynebacterium glutamicum ubiA-* mutant

Abbreviations: Ac, acylated; Ara, arabinose; Araf, arabinofuranose; AG, arabinogalactan; BHI, brain heart infusion; DPA, decaprenyl-monophosphoryl-**D**-arabinose; DPM, decaprenyl-monophosphoryl-mannose; DPR, decaprenyl-monophosphoryl-**D**-ribose; DPPR, decaprenyl-monophosphoryl-5-phosphoribose; GC/MS, gas chromatography mass spectrometry; LAM, lipoarabinomannan; LM, lipomannan; LB, Luria-Bertani; Man, mannose; mAGP, mycolyl-arabinogalactan-peptidoglycan; NMR, nuclear magnetic resonance; p, pyranose; PI, phosphatidyl-*myo*-inositol; PIMs, phosphatidyl-*myo*-inositol mannosides; pRpp, 5-phosphoribofuranose pyrophosphate.

SUMMARY

Arabinan polysaccharide side-chains are present in both *Mycobacterium tuberculosis* and *Corynebacterium glutamicum* in the heteropolysaccharide arabinogalactan (AG), and in *M. tuberculosis* in the lipoglycan, lipoarabinomannan (LAM). Herein, we show by quantitative sugar and glycosyl linkage analysis that *C. glutamicum* possesses a much smaller LAM version, Cg-LAM, characterised by single *t*-Araf residues linked to the $\alpha(1\rightarrow6)$ -linked mannan backbone. MALDI-TOF MS showed an average molecular weight of 13 800-15 400 Da for Cg-LAM. The biosynthetic origin of Araf residues found in the extracytoplasmic arabinan domain of AG and LAM is well known to be provided by decaprenyl-monophosphoryl-**D**-arabinose (DPA). However, the characterisation of LAM in a *C. glutamicum::ubiA* mutant devoid of prenyltransferase activity and devoid of DPA-dependent arabinan deposition into AG, revealed partial formation of LAM, albeit with a slightly altered molecular weight. These data suggest that in addition to DPA utilisation as an Araf donor, alternative pathways exist in *Corynebacteriaceae* for Araf delivery, possibly *via* an unknown sugar nucleotide.

INTRODUCTION

The *Corynebacteriaceae* represent a distinct and unusual group within Gram-positive bacteria, with the most prominent members being the human pathogens *Mycobacterium tuberculosis* and *Mycobacterium leprae* (Bloom & Murray, 1992). In addition, non-pathogenic bacteria also belong to this taxon, such as *Corynebacterium glutamicum*, which is used in the industrial production of amino acids (Sahm *et al.*, 2000). These bacteria collectively belong to the same sub-order, and share a similar genome, cell envelope and corresponding cell wall biosynthetic enzymes (Dover *et al.*, 2004; Stackebrandt *et al.*, 1997).

The characteristic cell envelope of this distinct group of bacteria contain mycolic acids, arabinogalactan (AG) and peptidoglycan, which are covalently linked to each other to form the mycolyl-arabinogalactan-peptidoglycan (mAGP) complex (Besra *et al.*, 1995; Brennan & Nikaido, 1995; Brennan, 2003; Daffé *et al.*, 1990; Dmitriev *et al.*, 2000; Dover *et al.*, 2004; McNeil *et al.*, 1990; McNeil *et al.*, 1991). In addition, they also possess an array of cell wall associated glycolipids, such as phosphatidyl-*myo*-inositol (PI), phosphatidyl-*myo*-inositol mannosides (PIMs) and the lipoglycans, lipomannan (LM) and lipoarabinomannan (LAM) (Brennan & Ballou, 1967; Brennan & Ballou, 1968; Chatterjee *et al.*, 1991; Chatterjee *et al.*, 1993; Guerardel *et al.*, 2002; Hill & Ballou, 1966; Khoo *et al.*, 1995). As a result, arabinose is present in two polysaccharides with markedly different structures. Interestingly, the occurrence of arabinosyl-containing glycoconjugates in bacteria (Brennan & Nikaido, 1995), plants (Fincher *et al.*, 1983), and

protozoan parasites (Dobson *et al.*, 2003; Goswami *et al.*, 2006; Guha-Niyogi *et al.*, 2001; Previato *et al.*, 1982; Xavier Da Silveira *et al.*, 1998), and their absence in mammalian cells is well known. Moreover, *Corynebacteriaceae* arabinan is an extremely complex homopolymer of **D**-arabinofuranose (**D**-Araf) residues based on discrete structural motifs, however the complete structures and biosynthesis of these polymers is still to be established.

For LAM biosynthesis we initially proposed the following biosynthetic pathway PI→PIM→LM→LAM (Besra *et al.*, 1997) in *M. tuberculosis*, which is now largely supported by biochemical and genetic evidence (Gurcha *et al.*, 2002; Kordulakova *et al.*, 2002; Kremer *et al.*, 2002; Schaeffer *et al.*, 1999). PimA catalyses the addition of Man_p provided by GDP-Mannose to the 2-position of the *myo*-inositol of PI to form PIM₁ (Kordulakova *et al.*, 2002), whereas PimB might be responsible for the addition of a second Man_p to the 6-position to yield Ac₁PIM₂ (Schaeffer *et al.*, 1999; Tatituri *et al.*, 2007). PimC has been demonstrated to allow further mannosylation to Ac₁PIM₃ (Kremer *et al.*, 2002) and more recently, PimE has been shown to be involved in the biosynthesis of Ac₁PIM₅ (Morita *et al.*, 2006). It has been proposed that PIM₄ is the direct precursor of LM, characterised by a linear $\alpha(1\rightarrow6)$ -linked mannan backbone linked with $\alpha(1\rightarrow2)$ mannopyranose side chains generated through a mannosyltransferase encoded by Rv2181 (Kaur *et al.*, 2006). LM is then further glycosylated with arabinan to produce LAM, and finally ‘mannose-capped’ to produce ManLAM, a process initiated by the capping enzyme encoded by Rv1635c (Dinadayala *et al.*, 2006). For the addition of arabinose residues into mycobacterial LAM, EmbC is required (Berg *et al.*, 2005; Shi *et al.*, 2006;

Zhang *et al.*, 2003), whereas AftA, EmbA and EmbB perform arabinan polymerisation in AG (Alderwick *et al.*, 2006b; Belanger *et al.*, 1996; Escuyer *et al.*, 2001). In contrast, *C. glutamicum* possesses a singular Emb, and an AftA orthologue, which are used in AG biosynthesis (Alderwick *et al.*, 2005; Alderwick *et al.*, 2006b; Seidel *et al.*, 2007). It is interesting to note that the arabinan domains of AG and LAM utilise several different *Araf* linkages, which suggests that additional arabinofuranosyltransferases must be required to form AG and LAM, and still remain to be identified in *Corynebacteriaceae*.

It has been shown that the activated *Araf* sugar donor in *Corynebacteriaceae* is decaprenyl-monophosphoryl-**D**-arabinose (DPA) (Wolucka *et al.*, 1994). It was also proposed that decaprenyl-monophosphoryl-**D**-ribose (DPR) could be an additional precursor involved in arabinan synthesis, *via* a 2'-epimerase (Wolucka & de Hoffmann, 1995). Recently, it was shown that 5-phosphoribofuranose pyrophosphate (pRpp) is converted to decaprenyl-monophosphoryl-5-phosphoribose (DPPR) *via* a 5-phospho- α -**D**-ribose-1-diphosphate-decaprenyl-phosphate 5-phosphoribosyltransferase, (UbiA; Rv3806c and NCgl2781 in *M. tuberculosis* and *C. glutamicum*, respectively) (Huang *et al.*, 2005), which is then dephosphorylated to form DPR. DPR is then enzymatically oxidised to form a keto-intermediate, DPX, followed by an enzymatic reduction to DPA (Mikusova *et al.*, 2005). Additionally, disruption of *ubiA* in *C. glutamicum*, the first enzyme involved in the biosynthesis of DPA, resulted in a complete loss of cell wall arabinan (Alderwick *et al.*, 2006a). To date, no sugar nucleotides of *Araf* have been identified. The key discovery that *Araf* residues in AG arise from DPA, also raised doubts whether mycobacteria neither produce nor require *Araf* sugar nucleotides. Herein, we

now clearly show that an *ubiA* disrupted mutant of *C. glutamicum* still produces a modified LAM variant (Cg-LAM), which is arabinosylated even in the absence of the sugar donor DPA (Alderwick *et al.*, 2006a). Taken together, the data suggests that alternative pathways may exist in *Corynebacteriaceae*, independent of DPA utilisation, specifically involved in arabinosylation events in Cg-LAM biosynthesis.

METHODS

Strains, construction of plasmids and culture conditions. *C. glutamicum* ATCC 13032 (the wild type strain, and referred for the remainder of the text as *C. glutamicum*) and *Escherichia coli* DH5 α mc^r were grown in Luria-Bertani (LB) broth (Difco) at 30 °C and 37 °C, respectively. The *C. glutamicum::ubiA* mutant was grown on complex medium brain heart infusion (BHI) (Difco). Kanamycin and ampicillin were used at a concentration of 50 $\mu\text{g ml}^{-1}$. The vector used for inactivation of *C. glutamicum::ubiA* was pCg::*ubiA*. It contained an internal *ubiA*-fragment of 321 bp amplified with the primer pairs ATC TTC AAC CAG CGC ACG ATC and AAT ATC GAT CAC TGG CAT GTG, which was ligated into the SmaI site of the non-replicative vector pK18mob to yield pCg::*ubiA*. To enable chromosomal inactivation of *ubiA*, pCg::*ubiA* was introduced into *C. glutamicum* by electroporation (Alderwick *et al.*, 2005).

Extraction and purification of lipoglycans. Purification procedures were adapted from protocols established for the extraction and purification of mycobacterial lipoglycans (Nigou *et al.*, 1999). Briefly, 10 g of cells grown to and absorbance at 600 nm of 1.0, were delipidated at 60 °C by using 500 ml CHCl₃/CH₃OH (1:1, v/v). The delipidated cells were resuspended in deionized water and disrupted by probe sonication (MSE Soniprep 150, 12 micron amplitude, 60 s ON, 90 s OFF for 10 cycles, on ice) and the cell debris refluxed 5 times with 100 ml of C₂H₅OH/H₂O (1:1, v/v) at 68 °C, for 12 h intervals. The cell debris was removed by centrifugation and the supernatant containing lipoglycans, neutral glycans and proteins dried. The dried supernatant was then treated with a hot 80

% (w/w) phenol-H₂O, biphasic wash at 70 °C for 1 h, followed by several protease treatments and the lipoglycan fraction recovered following extensive dialysis against deionized water (Nigou *et al.*, 1999).

The crude lipoglycan extract was resuspended in buffer A (50 mM ammonium acetate and 15 % propan-1-ol) and subjected to Octyl-Sepharose CL-4B hydrophobic interaction chromatography (2.5 cm x 50 cm) as previously reported (Leopold & Fischer, 1993). The column was initially washed with 4 column volumes of buffer A to ensure removal of neutral glycans followed by an increasing gradient of propan-1-ol ranging from 25-65 % keeping the concentration of ammonium acetate constant. The eluates were collected and extensively dialysed against deionized water and concentrated to approximately 1 ml and precipitated using 5 ml of C₂H₅OH and the sample freeze-dried using a Savant SpeedVac. The freeze-dried sample containing the retained material from the hydrophobic interaction column was then re-suspended in buffer B (0.2 M NaCl, 0.25 % sodium deoxycholate (w/v), 1 mM EDTA and 10 mM Tris-HCl, pH 8) to a final concentration of 200 mg ml⁻¹. The sample was gently mixed and left to incubate for 48 h at room temperature. The sample was then loaded onto a Sephacryl S-200 column (2.5 cm x 50 cm) previously equilibrated with buffer B. The sample was washed with 400 ml of buffer C (0.2 M NaCl, 1 mM EDTA and 10 mM Tris-HCl, pH 8) at a flow rate of 3 ml h⁻¹, collecting 1.5 ml fractions using a Bio-Rad auto-sampler. The fractions were monitored by SDS-PAGE using either a silver stain utilising periodic acid and silver nitrate (Hunter *et al.*, 1986) or a Pro-Q emerald glycoprotein stain (Invitrogen) and individual fractions pooled and dialysed extensively against buffer C for 72 h with frequent changes. The

samples were further dialysed against deionized water for 48 h with frequent changes of deionized water, lyophilised and stored at - 20°C.

Glycosyl composition and linkage analysis of lipoglycans by alditol acetates.

Lipoglycans from wild type *C. glutamicum* and the *C. glutamicum::ubiA* mutant were hydrolysed using 2 M trifluoroacetic acid (TFA), reduced with NaBD₄, and the resultant alditols were per-*O*-acetylated and examined by Gas Chromatography (GC) (Tatituri *et al.*, 2007). Lipoglycans (2 mgs) were per-*O*-methylated using dimethyl sulfinyl carbanion as described previously (Alderwick *et al.*, 2005; Besra *et al.*, 1995; Daffé *et al.*, 1990). In this procedure, lipoglycan samples (2 mgs) were resuspended in 0.5 ml dimethyl sulphoxide (anhydrous) and 100 µl of 4.8 M dimethyl sulfinyl carbanion. The reaction mixture was stirred for 1 h and then iodomethane (50 µl) was slowly added and the suspension stirred for 1 h, this process was repeated for a total of three times. The reaction mixture was then diluted with an equal volume of water, dialysed against deionized water and dried. The resulting retentate was applied to a pre-equilibrated C₁₈ Sep-Pak cartridge and initially washed with 10 ml water, 10 ml 20 % acetonitrile, and the per-*O*-methylated lipoglycan eluted with 2 ml acetonitrile and 2 ml ethanol. After drying the combined organic eluate under nitrogen, the per-*O*-methylated lipoglycan was hydrolysed using 250 µl of 2 M TFA at 120 °C for 2 h. The resulting hydrolysate was reduced with NaBD₄, per-*O*-acetylated and examined by Gas Chromatography/Mass Spectrometry (GC/MS) (Tatituri *et al.*, 2007). Per-*O*-methylation of lipoglycans for Matrix-assisted Laser Desorption Ionisation-Mass Spectrometry (MALDI-MS) analysis was performed as described previously (Dell *et al.*, 1993). Briefly, 1 ml of a dimethyl

sulfoxide/NaOH slurry was added followed by 0.5 ml of iodomethane. The reaction mixture was vigorously shaken for 10 min at room temperature and the reaction quenched with 1 ml of water. Per-*O*-methylated samples were then extracted into chloroform (1 ml) and washed several times with water before drying under a stream of nitrogen.

GC and GC/MS analysis. Analysis of partially per-*O*-methylated, per-*O*-acetylated alditol acetate sugar derivatives was performed on a CE Instruments ThermoQuest Trace GC 2000. Samples were injected in the split mode. The column used was a DB225 (Supelco). The oven was programmed to hold at an isothermal temperature of 275 °C for a run time of 15 min. GC/MS was carried out on a BPX5 column (Supelco) and a Finnigan Polaris/GCQ Plus™, as described previously (Besra *et al.*, 1995; Daffé *et al.*, 1990). GC/MS analysis of alditol acetate sugar derivatives was performed on a Perkin Elmer Clarus 500 Instrument. Samples were injected in the splitless mode. The column used was a RTX-5 (30 m x 0.25 mm internal diameter, Restek Corp.). Initial temperature was set at 60 °C then ramped to 300 °C at 8 °C /min.

NMR Spectroscopy. NMR spectra of lipoglycans were recorded on a Bruker DMX-500 equipped, with a double resonance (1H/X)-BBI z-gradient probe head. All samples were exchanged in D₂O (D, 99.97% from Euriso-top, Saint-Aubin, France), with intermediate lyophilisation, and then dissolved in 0.5 ml D₂O and analysed at 313K. The ¹H and ¹³C NMR chemical shifts were referenced relative to internal acetone at 2.225 and 34.00 ppm, respectively. All the details concerning NMR sequences used and experimental

procedures were described in previous studies (Gilleron *et al.*, 1999; Gilleron *et al.*, 2000; Nigou *et al.*, 1999).

MALDI-MS Analysis. MALDI-MS was performed using a PerSeptive Biosystems Voyager DE[®] STR mass spectrometer. Native lipoglycans were dissolved in methanol/H₂O 50:50 and 1 µl of the sample was loaded onto a metal plate. After evaporation 1 µl of the matrix 2,5-dihydroxybenzoic acid was added on the spot. Samples were analysed using the linear negative mode. Per-*O*-methylated samples were dissolved in methanol/H₂O (80:20, v/v) and analysed in the linear positive mode using 2,5-dihydroxybenzoic acid as matrix. Sequazyme peptide mass standards were used as external calibrants (Applied Biosystems, Foster City, CA).

RESULTS

Disruption of *Cg-ubiA*. The *ubiA* gene product was shown in prior work to synthesise DPPR, which is converted to DPA, thus supplying the substrate for the “priming” arabinosyltransferase AftA (Alderwick *et al.*, 2006b). These initial Araf residues “prime” the galactan backbone for further attachment of $\alpha(1\rightarrow5)$ linked Araf residues. These reactions require the arabinofuranosyltransferase activities of Mt-EmbA and Mt-EmbB, or Cg-Emb, respectively, and also the utilisation of DPA (Alderwick *et al.*, 2005; Escuyer *et al.*, 2001; Seidel *et al.*, 2007), to eventually result in mature AG. In light of our present studies demonstrating the occurrence of Cg-LAM we inactivated the mycobacterial orthologue of *C. glutamicum*, NCgl2781, by transforming the wild type to kanamycin resistance conferred by the vector borne *aph* gene product of pCg::*ubiA* with a view of generating a clear *C. glutamicum* LM-only phenotype. The vector, as previously described (Alderwick *et al.*, 2005), was integrated into the chromosomal *ubiA* gene, thus disrupting *ubiA*, as confirmed by two independent PCR analyses with two different primer pairs (Alderwick *et al.*, 2005). As expected, the resulting strain *C. glutamicum*::*ubiA* exhibited a strong reduced growth (Alderwick *et al.*, 2006a).

Purification of lipoglycans from *C. glutamicum* and *C. glutamicum*::*ubiA*. Analysis of the crude phenol-extracted lipoglycans from *C. glutamicum* (Fig. 1A) and *C. glutamicum*::*ubiA* (Fig. 1B) visualised on SDS-PAGE revealed two closely migrating lipoglycans. A two-step purification protocol was performed to fractionate the crude lipoglycans. In the first step, the neutral glycans and nucleic acids were eliminated by using

an octyl-Sepharose CL-4B column (Amersham Biosciences). In the second step, the lipoglycans were individually purified using a Sephacryl S-200 column (Amersham Biosciences). Fractions containing the lipoglycans were monitored by SDS-PAGE stained with either silver nitrate or Pro-Q emerald glycoprotein stains and pooled accordingly to afford lower (1) and upper (2) lipoglycans (Fig. 1A and B).

General structural features of lipoglycans from *C. glutamicum* and *C.*

***glutamicum::ubiA*.** The lower lipoglycan (1) from both *C. glutamicum* and *C. glutamicum::ubiA* were similar and exhibited the basic components of a structure related to mycobacterial LM (now termed Cg-LM) and contained solely mannopyranose (Manp). Per-*O*-methylation analysis of Cg-LM from both strains indicated the presence of *t*-Manp, 2-Manp, 6-Manp and 2,6-Manp residues (Tatituri *et al.*, 2007). GC analysis of the total acid hydrolysed upper lipoglycan (2) from *C. glutamicum* contained arabinose and mannose in a ratio (23:77) and was related to mycobacterial LAM (Fig. 2A). In addition, per-*O*-methylation analysis of wild type Cg-LAM indicated the presence of *t*-Araf, *t*-Manp, 2-Manp, 6-Manp and 2,6-Manp residues (Fig. 3A). Interestingly, Cg-LAM of the wild-type strain was previously shown to be composed of a PI anchor linked to an $\alpha(1\rightarrow6)$ Manp backbone substituted at most of the *O*-2 positions by the structural motifs *t*-Araf, *t*-Manp, *t*-Araf(1 \rightarrow 2)-Manp and *t*-Manp(1 \rightarrow 2)-Manp units (Fig. 4E) (Tatituri *et al.*, 2007). Accordingly, the 1D ^1H -NMR anomeric region (Fig. 4A) exhibited a complex pattern of overlapping resonances corresponding to Araf and Manp units. The different spin systems were characterised by ^1H - ^{13}C HMQC NMR (Fig. 4B) and anomeric resonances were attributed as follows: $\delta\text{H}_1\text{C}_1$ 5.20/112.2 (I₁) to two overlapping *t*-Araf units (see below) and

5.13/112.0 (II₁) to a third *t*-Araf unit, 5.06/105.2 (III₁) to *t*-Manp units, 5.12/101.4 (IV₁), 5.07/101.7 (V₁) and 5.04/101.9 (VI₁) to 2,6-Manp units, 5.06/105.2 (VII₁) to 6-Manp units and 5.00/104.9 (VIII₁) to 2-Manp units.

Our previous experiments demonstrated that the *C. glutamicum::ubiA* mutant failed to synthesise DPA (Alderwick *et al.*, 2006a) and possessed an arabinan deficient cell wall phenotype (Alderwick *et al.*, 2005). As a consequence, we anticipated that the Cg-LAM from the mutant would be devoid of arabinose. Surprisingly, glycosyl compositional analysis of Cg-LAM from the *C. glutamicum::ubiA* mutant contained arabinose, albeit substantially reduced, and mannose in a ratio (4.5:95.5) (Fig. 2B). In addition, per-*O*-methylation analysis of Cg-LAM from the mutant indicated the presence of *t*-Araf, *t*-Manp, 2-Manp, 6-Manp and 2,6-Manp, although, the relative abundance of *t*-Araf residues was reduced (Fig. 3B). In agreement with the glycosyl compositional and linkage analysis data indicating a reduced amount of *t*-Araf units, the 1D ¹H anomeric region of Cg-LAM from the *glutamicum::ubiA* mutant (Fig. 4C) was simpler than that observed for Cg-LAM from wild type *C. glutamicum* (Fig. 4A). Resonance I₁ corresponding to a *t*-α-Araf unit was dramatically reduced, suggesting two overlapping *t*-Araf signals. This was further confirmed by ¹H-¹³C HMQC analysis (Fig. 4D). Indeed, the relative intensity of the correlation at δH₁C₁ 5.20/112.2 (I₁) was much weaker and correlation at δH₁C₁ 5.13/112.0 (II₁) of the third *t*-Araf residue was absent.

The native Cg-LAMs from wild type *C. glutamicum* and the *C. glutamicum::ubiA* mutant were analysed by MALDI TOF MS. MALDI spectra were performed in the linear

negative mode. The mass spectra show broad unresolved molecular ion envelopes due to the heterogeneity of the Cg-LAMs and the relatively low resolution of this type of mass spectrometry. The native Cg-LAM from wild type *C. glutamicum* shows an average molecular weight of around 15 400 Da (Fig. 5A). The native Cg-LAM from the *C. glutamicum::ubiA* mutant shows a somewhat lower average molecular weight in the region of 13 900 Da (Fig. 5B). Samples of Cg-LAM from wild type *C. glutamicum* and *C. glutamicum::ubiA* were subjected to per-*O*-methylation and analysed by MALDI TOF MS (data not shown). Per-*O*-methylation replaces fatty acyl substituents with methyl groups as well as methylating all the free hydroxyl groups. The average molecular weight of the derivatised Cg-LAM from wild type *C. glutamicum* was observed around 15 600 Da, whilst the molecular weight of the corresponding Cg-LAM from *C. glutamicum::ubiA* was observed around 15 200 Da.

Altogether, these data indicate that the *C. glutamicum::ubiA* mutant produces a slightly truncated Cg-LAM with a severe reduction in *t*-Araf content but maintaining and possibly extending its mannan core. In addition, the presence of UbiA and the biosynthesis of DPA, appears essential for the higher arabinosylation of Cg-LAM. Moreover, these findings suggest that arabinose deposition in the *C. glutamicum::ubiA* mutant is a result of incorporation of *t*-Araf through another arabinose substrate, and one which is not DPA dependent.

DISCUSSION

Understanding the biosynthesis of mycobacterial arabinan is paramount to identifying potential new drug targets for the treatment of tuberculosis. However, as discussed earlier a complete understanding of these pathways is far from clear. In this respect, *C. glutamicum* is a useful tool in understanding mycobacterial cell wall biosynthesis, since this organism possesses the core structural elements in *Corynebacterineae* with few gene duplications, and deletion of orthologous genes which is often lethal in mycobacterial species is possible (Alderwick *et al.*, 2005; Alderwick *et al.*, 2006b; Gande *et al.*, 2004). Specifically, the deletion of the DPA requiring arabinosyltransferase Emb in *C. glutamicum* resulted in a strongly abrogated arabinan domain of AG (Alderwick *et al.*, 2005), and deletion of AftA also disabled attachment of the remaining singular *t*-Araf residues to galactan (Alderwick *et al.*, 2006b). Furthermore, the loss of DPA synthesis in a *C. glutamicum::ubiA* mutant also resulted in the full loss of the arabinan domain in mature AG (Alderwick *et al.*, 2005). In contrast, Cg-LM and Cg-LAM isolated from the corresponding *C. glutamicum* Δemb and $\Delta aftA$ mutants were similar to the wild type glycans (data not shown). An absence of Araf residues was expected in Cg-LAM of the *C. glutamicum::ubiA* mutant. Strikingly, a total absence of arabinan was not observed (Fig. 2), thus requiring an alternative source and mechanism of addition of this sugar which may be attributed to the presence of an uncharacterised nucleotide sugar donor in *Corynebacterineae*.

In contrast to mycobacteria, parasites do synthesise and utilise a sugar nucleotide to provide an activated arabinose sugar donor. Schneider *et al.* (1994) characterised GDP- α -**D-Arap**, the precursor of **D-Ara** in *Leishmania major* lipophosphoglycan. In trypanosomatids, this sugar nucleotide is believed to be synthesised in a two step process through the combined activities of an Ara-1-kinase and GDP-Ara pyrophosphorylase, whereby arabinose is phosphorylated to form arabinose 1-phosphate and then activated to the nucleotide level by GDP-**D-Arap** pyrophosphorylase (Mengeling & Turco, 1999; Schneider *et al.*, 1995). Compared with the many genes encoding glycosyltransferases in prokaryotic and eukaryotic systems, very little is known about those that encode arabinopyranosyltransferases. Dobson *et al.* (2003) identified two genes (*SCA1/2*) encoding arabinosyltransferases mediating scAra capping, and recently it was demonstrated by heterologous expression that *Leishmania SCA1* encodes an arabinopyranosyltransferase (Goswami *et al.*, 2006).

Glycosyltransferases belonging to one of the families, GT-A and GT-B, or both, may be responsible for the initial addition of *t-Araf* residues on to Cg-LAM in the absence of DPA. These enzymes may be present on the cytoplasmic side of the membrane and probably aid in the transfer of *t-Araf* residues from a putative sugar nucleotide donor. These results could explain the presence of *t-Araf* in Cg-LAM, even though DPA biosynthesis has been completely abrogated in the *C. glutamicum::ubiA* mutant (Alderwick *et al.*, 2005). The mechanism as to how these *Araf* residues are added is not clear. One possible explanation is that *Araf* residues from a nucleotide precursor are added in a step-wise fashion to Cg-LM on the cytoplasmic side to give motif-A proximal

to PI, and then this complex polysaccharide is then probably flipped on to the periplasmic side. It is thought that the flipping mechanism is a prerequisite for arabinosylation to occur outside of the membrane, where the arabinan and mannan domains are formed through specific GT-C glycosyltransferases, by the addition of either *t-Araf* and *t-Manp* residues, from DPA or decaprenyl-monophosphoryl-**D**-mannose (DPM) to give motif-B, C and D (Fig. 4E and Fig. 6) (Berg *et al.*, 2007; Seidel *et al.*, 2007; Tatituri *et al.*, 2007). The biosynthetic origin of mycobacterial *Araf* residues is generally a poorly understood area. To date, no sugar nucleotides of *Araf* have been identified, and as a result, a number of theories regarding the generation of arabinan in mycobacteria have been put forward (Huang *et al.*, 2005; Mikusova *et al.*, 2005; Scherman *et al.*, 1995; Scherman *et al.*, 1996). Depending on the organisation of the arabinan chains, it is speculated that five or six arabinofuranosyltransferases are required for arabinan biosynthesis in mycobacteria. In this regard, it is interesting to note that an *M. smegmatis embC* mutant (Escuyer *et al.*, 2001; Zhang *et al.*, 2003) was found to be devoid of LAM but from the chemical analysis, the resulting LM precursor, possessed 2-3 *Araf* residues. The possibility exists that these residues are initially added *via* a non-DPA dependent glycosyltransferase and requires further investigation.

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FIGURE LEGENDS

Figure 1. Lipoglycan profiles of wild type *C. glutamicum* (A) and *C. glutamicum::ubiA* (B). Lipoglycans were analysed using SDS-PAGE and visualised using the Pro-Q emerald glycoprotein stain (Invitrogen) specific for carbohydrates. Individual lipoglycans (**1** and **2**) were purified as described in the methods section. Abbreviations: C: crude lipoglycan; M: molecular weight marker (kDa).

Figure 2. Glycosyl compositional analysis of lipoglycan-2 from wild type *C. glutamicum* (A) and *C. glutamicum::ubiA* (B). Samples of individually purified lipoglycans (**2**) were hydrolysed with 2M TFA, reduced and per-*O*-acetylated. Alditol acetates were subjected to GC analysis. Peak area integration shows the arabinose content is 23 % in wild type *C. glutamicum* (Panel A) and 4.5% in the *C. glutamicum::ubiA* mutant (Panel B). Abbreviations: Ara, arabinose; Man, mannose.

Figure 3. Glycosyl linkage analysis of per-*O*-methylated lipoglycan-2 from *C. glutamicum* (A) and *C. glutamicum::ubiA* (B). Lipoglycan-2 from *C. glutamicum* and *C. glutamicum::ubiA* were per-*O*-methylated, hydrolysed using 2M TFA, reduced, and per-*O*-acetylated. The resulting partially per-*O*-methylated and per-*O*-acetylated glycosyl derivatives were analysed by GC/MS as described previously (Alderwick *et al.*, 2005).

Figure 4. NMR characterisation of wild-type Cg-LAM (A, B), Cg-LAM from the UbiA mutant (C, D) and a structural representation of Cg-LAM (E). 1D ¹H (A,C)

and 2D ^1H - ^{13}C HMQC (B,D) NMR spectra of Cg-LAMs in D_2O at 313K. Expanded regions (δ ^1H : 4.85-5.30) (A,C) and (δ ^1H : 4.85-5.30, δ ^{13}C : 100-114) (B,D) are shown. Glycosyl residues are labelled in Roman numerals and their carbons and protons in Arabic. I, II, *t*- α -Araf; III, *t*- α -Manp; IV, V, VI, 2,6- α -Manp; VII, 6- α -Manp; VIII, 2- α -Manp. Structural representation of Cg-LAM (Tatituri *et al.*, 2007). Cg-LAM contains an (1 \rightarrow 6)-Manp backbone almost completely substituted by *t*-Araf, *t*-Manp, *t*-Manp(1 \rightarrow 2)-Manp, and *t*-Araf(1 \rightarrow 2)-Manp units. X, either a *t*-Araf or a *t*-Manp unit (E).

Figure 5. MALDI-TOF MS spectra of Cg-LAM from wild type *C. glutamicum* (A) and *C. glutamicum::ubiA* (B). MALDI spectra were performed in the linear negative mode with delayed extraction using 2,5-dihydrobenzoic acid as matrix.

Figure 6. Proposed biosynthetic pathway of Cg-LAM. The addition of the first Araf unit is thought to occur on the cytoplasmic side of the membrane by a GT-A/B glycosyltransferase (Liu & Mushegian, 2003). After transportation across the membrane by an unknown “flippase” enzyme, further elaboration of the lipoglycan then occurs through the addition of mannose and arabinose units catalysed by several GT-C glycosyltransferases utilising DPM and DPA, respectively (Berg *et al.*, 2007; Seidel *et al.*, 2007; Tatituri *et al.*, 2007).

Figure 1

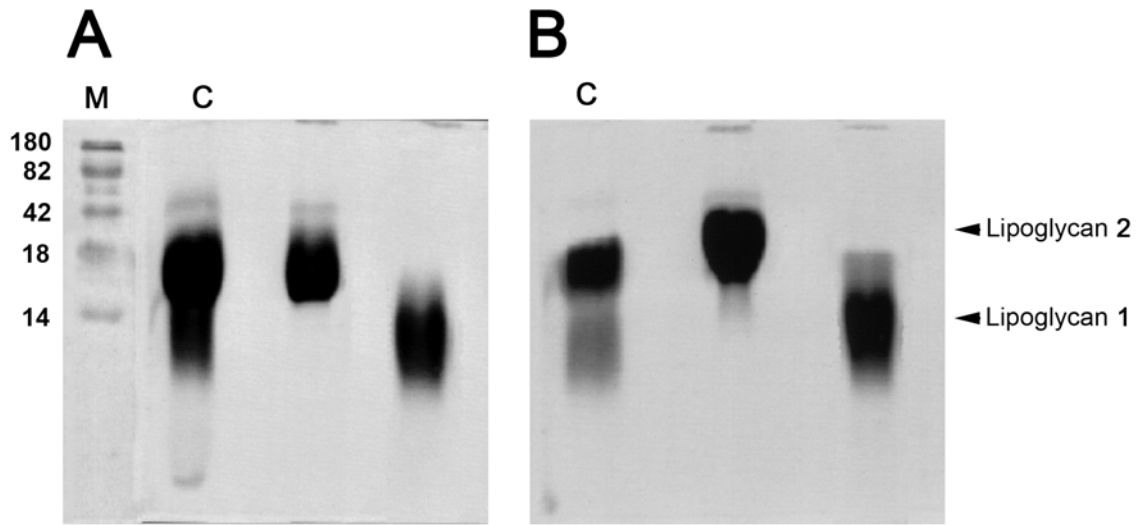


Figure 2

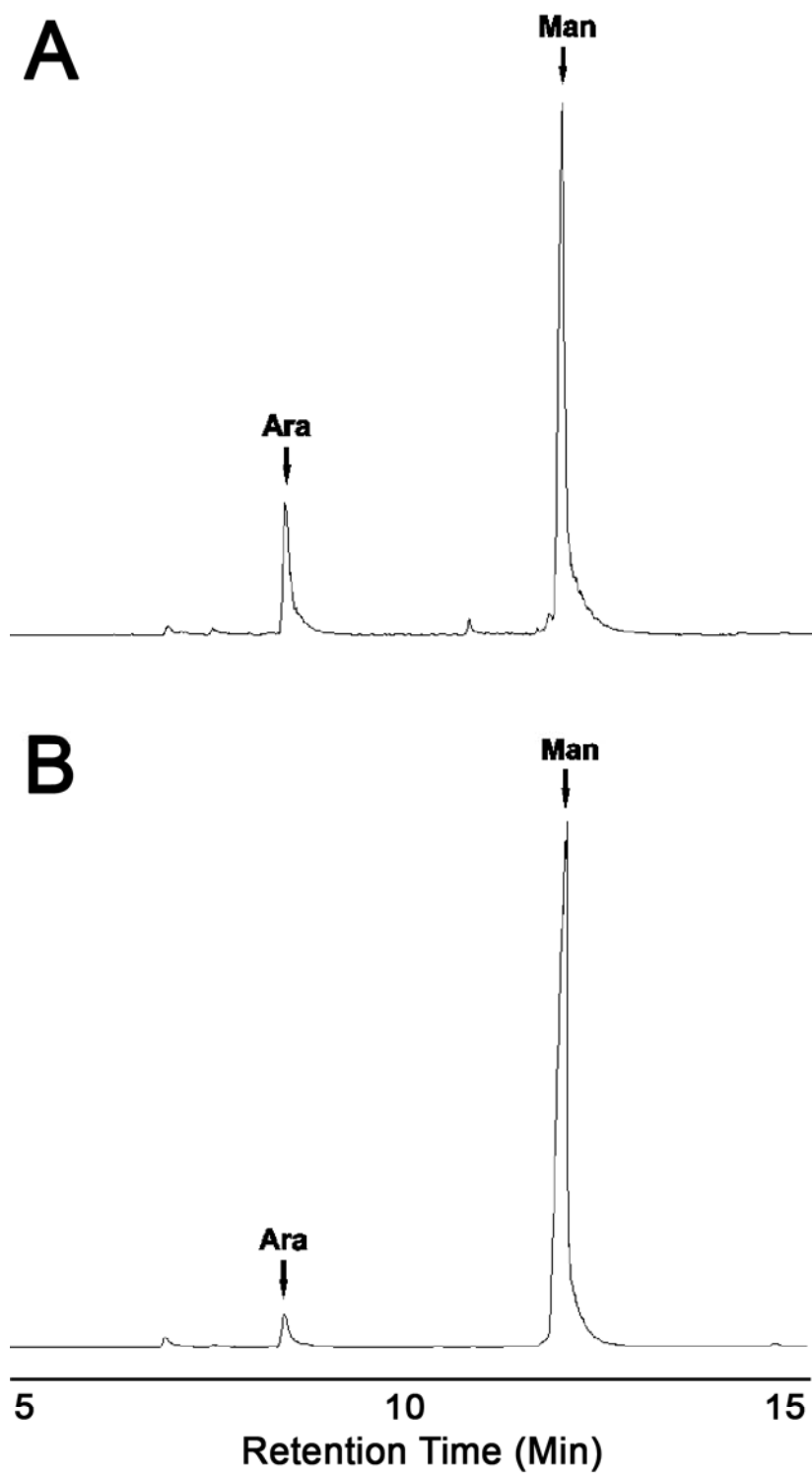


Figure 3

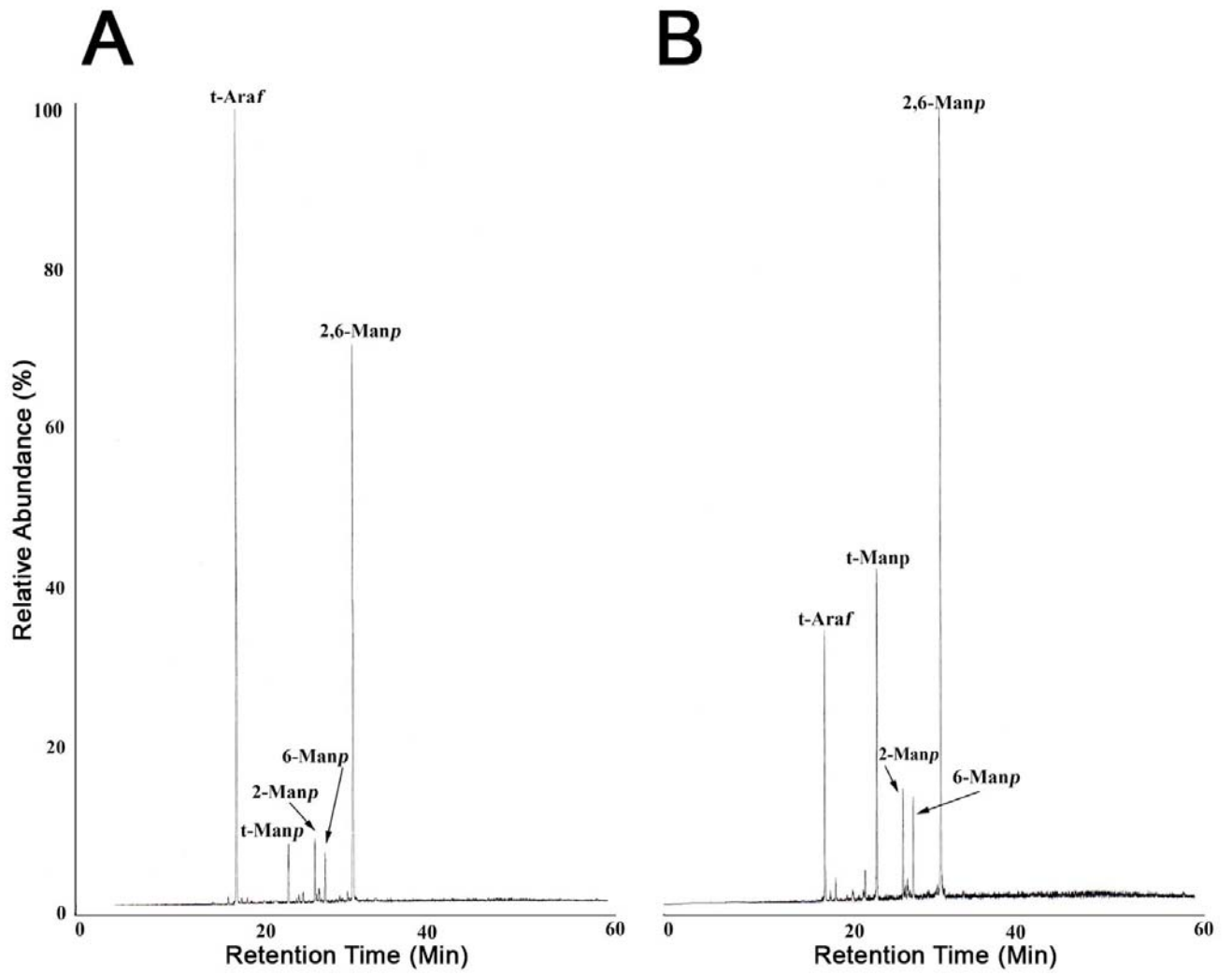


Figure 4

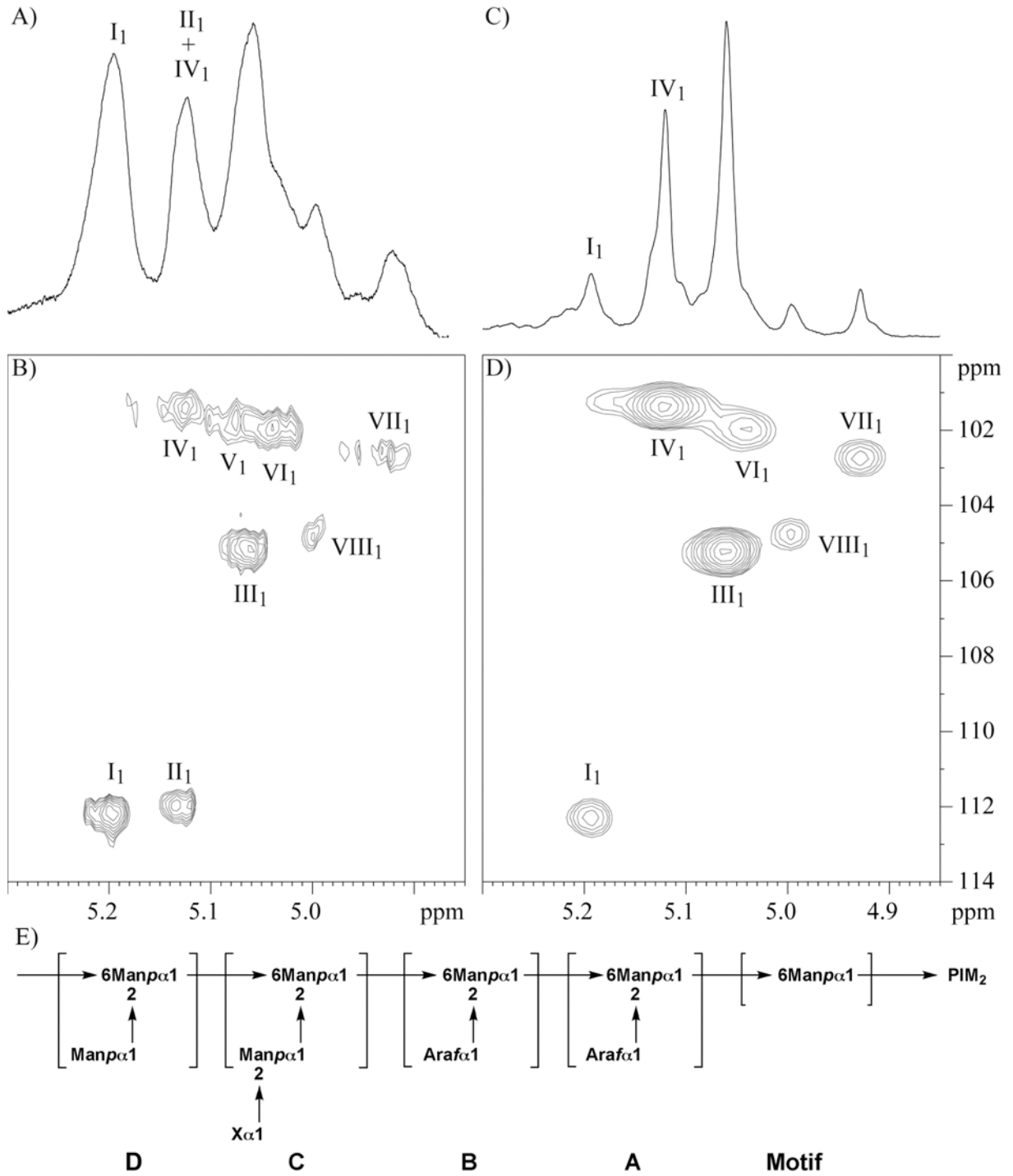


Figure 5

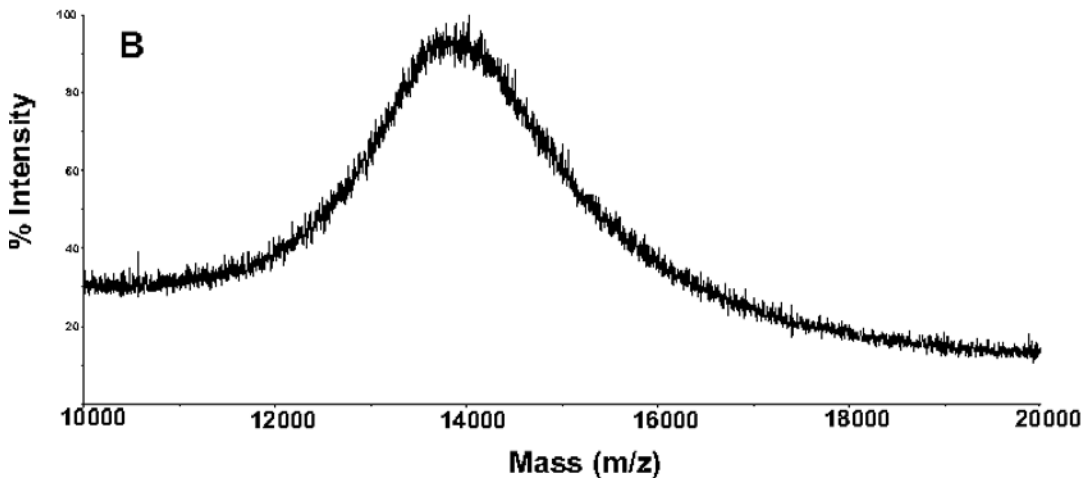
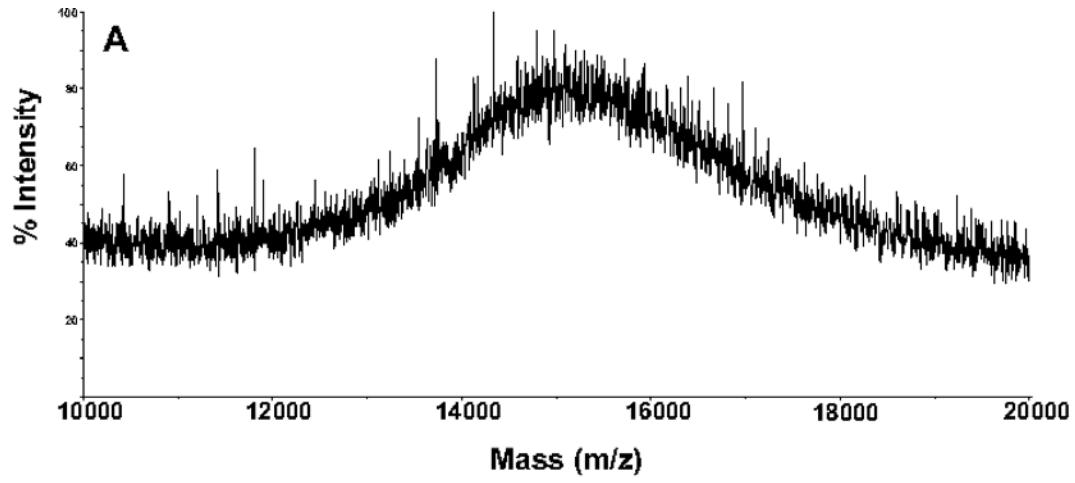


Figure 6

