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Carbamylated LL-37 as a modulator of the immune response

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Running head: Effects of LL-37 carbamylation

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Abstract

Carbamylation of lysine residues and protein N-termini is a ubiquitous, non-enzymatic post-translational modification. Carbamylation at sites of inflammation is due to cyanate formation during the neutrophil oxidative burst and may target lysine residues within the antimicrobial peptide LL-37. The bactericidal and immunomodulatory properties of LL-37 depend on its secondary structure and cationic nature, which are conferred by arginine and lysine residues. Therefore, carbamylation may affect the biological functions of LL-37. The present study examined the kinetics and pattern of LL-37 carbamylation to investigate how this modification affects the bactericidal, cytotoxic, and immunomodulatory function of the peptide. The results indicated that LL-37 undergoes rapid modification in the presence of physiological concentrations of cyanate, yielding a spectrum of diverse carbamylated peptides. Mass spectrometry analyses revealed that the N-terminal amino group of Leu-1 was highly reactive and was modified almost instantly by cyanate to generate the predominant form of the modified peptide, named LL37^{C1}. This was followed by the sequential carbamylation of Lys-8, Lys-12, and Lys-15, to yield LL37^{C8}, and LL37^{C12,15}, respectively. Carbamylation had profound and diverse effects on the structure and biological properties of LL-37. In some cases, anti-inflammatory LL-37 was rapidly converted to pro-inflammatory LL-37.

Introduction

Post-translational modifications (PTMs) are pivotal steps in protein maturation. PTMs increase the functional diversity of the proteome and play a key role in multiple cellular processes, including protein-protein interactions, cell signaling, differentiation, and regulation of gene expression [1]. Carbamylation is a ubiquitous, non-enzymatic PTM, in which cyanate (OCN^-) reacts with primary amino groups (NH_3^+) at the N-termini of proteins, and with lysine residues in the polypeptide chain, to generate a protein:homocitrulline complex [2,3]. Since urea (a by-product of protein metabolism) and cyanate comprise an equilibrium pair, the level of protein carbamylation is markedly increased in diseases associated with chronic uremia (renal dysfunction). A recent study identified a novel pathway linking inflammation, myeloperoxidase (MPO), and carbamylation. MPO, a heme peroxidase released by activated neutrophils, catalyzes the formation of cyanate from hydrogen peroxide and thiocyanate, ultimately leading to homocitrullination of proteins [3]. The chemical conversion of positively charged Lys residues to neutral homocitrulline residues affects the charge distribution within a polypeptide chain in a manner that often results in impairment or even loss of function. For example, loss of function upon carbamylation has been reported for matrix metalloproteinase-2, inhibitor of metalloproteinase-2, and insulin [4,5].

The abundance of MPO at inflammatory foci has sparked significant interest in the role of carbamylation in the context of chronic inflammatory diseases and atherogenesis [3,6,7]. Apart from directly affecting protein function and turnover, homocitrulline residues act as neo-epitopes that can trigger primary immune responses, thereby inducing chemotaxis and proliferation of CD4^+ T cells and the subsequent production of interferon- γ , interleukin (IL)-10, and IL-17. In addition,

antibodies against carbamylated residues have clinical value in that they predict a more erosive progression of rheumatoid arthritis [8,9].

Cationic antimicrobial peptides (CAMPs) are essential components of human innate immunity and are produced by a variety of cells, including epithelial cells [10], keratinocytes [11], and neutrophils [12]. In humans, CAMPs are represented by α - and β -defensins and the cathelicidin-derived LL-37 peptide. The active form of LL-37 is characterized by a high abundance of arginine and lysine residues, which generate a net positive charge of +6 at a neutral pH. Apart from killing a wide spectrum of pathogenic bacteria [13,14], LL-37 neutralizes lipopolysaccharide (LPS), functions as a chemoattractant for immune cells (including T cells, monocytes, neutrophils, and mast cells)[15,16], profoundly affects the course of dendritic cell maturation [17], stimulates production of cytokines, chemokines, and their receptors [18,19], and triggers mast cell degranulation [16]. These effects are mediated, at least in part, by activation of at least four different receptors: formyl peptide receptor-like 1 (FPRL1), epidermal growth factor receptor (EGFR), P2X7, and CXCR2. Finally, recent studies show that LL-37 has a direct effect on the cells and might be hemolytic [20] or cytotoxic to peripheral blood mononuclear cells (PBMCs) at high concentrations [21]. At low concentrations (<5 μ M), LL-37 induces rapid secondary necrosis of apoptotic human neutrophils [22]. To prevent collateral tissue damage due to exacerbation of inflammation, the activity of LL-37 is strictly controlled by serum proteins [23], most likely apolipoprotein-A1 (apoA-1) [24]. Taken together, all available data indicate that the antibacterial activity of LL-37 is secondary to its immunomodulatory functions. In a twist of the paradigm, it is now generally accepted that the major role of CAMPs is not direct killing of invading microbes, but rather acting as signaling molecules for innate and acquired immunity.

Within the inflammatory milieu, the concomitant release of LL-37 and MPO by activated neutrophils may create conditions required for LL-37 carbamylation. Additionally, lysine residues within CAMPs are crucial for peptide structure and activity, and conversion of these positively charged residues to neutral homocitrulline residues would be expected to abrogate the biological activity of LL-37.

Here, we performed mass spectrometry based sequence analysis and found that LL-37 undergoes rapid carbamylation in the presence of cyanate in a time- and concentration-dependent manner. Unexpectedly, we found that the free amino group of the N-terminal leucine was most susceptible to carbamylation under conditions that reflected the cyanate concentration in the inflammatory milieu. Prolonged incubation resulted in the generation of a mixture of variably carbamylated LL-37 molecules with impaired antimicrobial activity against both Gram-negative and Gram-positive species.

Taken together, these results suggest that carbamylation of LL-37 within an inflammatory environment might actually exacerbate inflammation and be detrimental to the host.

Materials and methods

In vitro carbamylation of LL-37 and mass spectrometry analysis. LL-37 (ProImmune, Oxford, U.K) was carbamylated by incubation with 10, 50 and 100 mM KCNO (Sigma-Aldrich, Oslo, Norway) in 100 mM HEPES (pH 7.8) for 3 hours at 37°C. The reaction was quenched by addition of formic acid to a final concentration of 5% and samples were immediately purified using StageTips (ThermoScientific). The samples were lyophilized and either analyzed directly by LC-MS/MS or subjected to proteolytic digestion with *S. aureus* protease V8 (1:25 w/w) at 37°C for 16 hours prior to LC-MS/MS analysis. NanoESI-MS/MS analyses were performed on an EASY-nLC II system (ThermoScientific) connected to a TripleTOF 5600 mass spectrometer (AB Sciex) equipped with a NanoSpray III source (AB Sciex) operated under Analyst TF 1.5.1 control. The samples were suspended in 0.1% formic acid, injected, trapped and desalted on a Biosphere C18 column (5 µm, 2 cm x 100 µm I.D; Nano Separations) after which the peptides were eluted from the trap column and separated on a 15-cm analytical column (75 µm i.d.) packed in-house in a pulled emitter with RP ReproSil-Pur C18-AQ 3 µm resin (Dr. Marisch GmbH, Ammerbuch-Entringen, Germany) and connected in-line to the mass spectrometer. The peptides were eluted using a 20 min gradient from either 5%–35% phase B or 5–90% phase B (0.1% formic acid and 90% acetonitrile). The collected MS files were converted to Mascot generic format (MGF) using the AB SCIEX MS Data Converter beta 1.1 (AB SCIEX). The peptide sequence was identified using in-house Mascot search engine (matrix science). Search parameters were allowing two missed cleaving sites and carbamylation as a variable modification. Peptide tolerance and MS/MS tolerance were set to 10 ppm and 0.1 Da respectively.

Peptide synthesis. Native and carbamylated LL-37 were synthesized by ProImmune by using F_{moc} solid-phase peptide synthesis, diluted in 0.01% v/v acetic acid and stored at -70°C until use.

Circular dichroism (CD) spectroscopy. The secondary structure of the LL-37 analogues was investigated by Circular dichroism (CD) spectroscopy. The experiments were performed using a Jasco J-810 spectropolarimeter. Far UV-spectra were acquired at 37°C in the 195 – 260 nm range at a scan rate of 50 nm/min and a band width of 1 nm. Three scans were accumulated for each sample and appropriate blanks were subtracted from each spectrum by using the software provided by the instrument manufacturers. The peptides (10 µM) were analyzed upon dilution in a 10 mM sodium phosphate buffer containing 50% v/v trifluorethanol (TFE) or a physiological salt solution resembling plasma (113 mM NaCl, 24 mM NaHCO₃, 0.6 mM MgCl₂, 1.3 mM CaCl₂, 3.9mM KCl) in 1.0-mm quartz cuvettes (Hellma-Analytics, Oslo, Norway). The mean ellipticity was calculated using the formula $[\theta] = \theta / (10 \cdot c \cdot l)$, where θ is the ellipticity (mdeg), 10 is a scaling factor, c is the protein concentration (M) and l is the path length of the cuvette (cm). The helical content (percentage of helix) was estimated by using the CDNN program from the molar ellipticity θ [deg.cm² dmol⁻¹].

Broth microdilution assay. Frozen samples of *S. aureus* LS-1, *E. coli* ATCC 25922 and *B. subtilis* ATCC 3366 were cultured on horse blood-agar plates at 37°C over night. Few colonies were selected and pre-cultured in 50 ml LB-broth in a shaking incubator (220 rpm, 37°C) over night. The bacteria were diluted 1:100 times in fresh LB-broth and cultured to its mid-log-phase. The bacteria were washed four times at 4000 x g for 5 min (*E. coli* and *S. aureus*) or at 6000 x g for 8 min (*B. subtilis*) at 4°C and thereafter suspended to 1 x 10⁶ CFU/ml in PBS without calcium and magnesium.

Native and carbamylated LL-37 described above were diluted in 0.01% v/v acetic acid containing 0.2% w/v bovine serum albumin (Sigma-Aldrich). Thereafter, one part of the peptide solutions were mixed with nine parts bacteria solution to get a final peptide concentration of 1 µg/ml (i.e., 0.2 µM). In addition, one positive control sample containing bacterial solution without additives and one negative control without bacteria were prepared. All samples were incubated for 2 hours at 37°C. Samples containing *B. subtilis* were incubated on a shaking plate at 220 rpm. 100 µl aliquots of the bacterial mixture were spread on blood agar plates in duplicate or triplicate after being 10-fold diluted in 4 steps (undiluted, 1:10, 1:100, 1:1000, 1:10000). The plates were incubated at 37°C over night. Colony-forming units were counted and the total number was determined from the dilution factor. The experiments were performed under sterile conditions.

Culture of human monocyte-derived macrophages (hMDMs). Human blood samples were collected from healthy volunteers by using heparin-coated tubes (BD) and diluted 1:1 in PBS without calcium and magnesium. PBMCs were isolated by density gradient separation on Lymphoprep (Axis-Shield Poc AS) and diluted in macrophage medium (RPMI 1640 containing UltraGlutamin supplemented with 10% v/v autologous human serum and penicillin streptavidin; Sigma-Aldrich). Subsequently, 3×10^6 cells/well were plated in 24-well plates (Sarstedt, Nümbrecht, Germany) and incubated for 24 hours at 37°C in 5% CO₂. Non-adherent PBMCs were removed by washing with PBS. Adherent cells were cultured in regularly changed macrophage medium for a minimum of 10 days, at which time approximately 10% of the cells had differentiated into human monocyte-derived macrophages (hMDMs).

Cytokine production of hMDMs. Following washing (3x) with PBS mature hMDMs were incubated with RPMI medium containing 0.5 µM native or

carbamylated LL-37 together with 100 ng/ml **E-coli LPS** (Sigma-Aldrich) in triplicated wells for 20 hours at 37°C in 5% CO₂. RPMI without any additives was used as a negative control and RPMI with LPS was used as a positive control. The supernatants were removed from the wells, centrifuged at 300 x g for 5 min and frozen at -70°C until cytokine analysis. The concentrations of GM-CSF, IFN- γ , IL1 β , IL-2, IL-4, IL-5, IL-6, IL-8, IL-10 and TNF- α were measured using a cytokine human 10-plex panel (Life technologies, Oslo, Norway).

Hemolysis of erythrocytes. Peripheral blood was collected in heparin coated tubes (BD) from healthy volunteers and centrifuged at 800 x g for 10 min at 10°C. The pellet was gently suspended in PBS, twice the original volume, and washed (2x) by centrifugation. The pellet was once again suspended in PBS to the initial blood volume. Thereafter, 4% v/v erythrocyte suspension was mixed with 1-20 μ M native and carbamylated LL-37 diluted in PBS, added to a V-bottomed 96-well plate and incubated for 1-6 hours at 37°C. The plate was mixed every 15 min by shaking at 600 rpm. After incubation, the plates were centrifuged at 500 x g for 5 min. The supernatant was collected and the released of hemoglobin was measured at 405 nm.

Chemotaxis of neutrophils. The Insall chamber was used to visualise chemotaxis [25]. For each sample, **discontinuous Percoll gradient** isolated neutrophils (400 μ l in RPMI, final density 1×10^6 /ml) were added to acid washed (0.2 M HCl), dried and blocked (7.5%, BSA 400 μ l, Sigma) coverslips (22 mm, VWR International), which were then incubated at room temperature (approximately 23°C) for 30 min. to allow the cells to adhere. The coverslip was then inverted and placed at the top of the chemotaxis chamber ensuring that the chemoattractant loading bays were exposed. The desired chemoattractant (80 μ l), fMLP (10 nM) or LL-37 and its modified forms (used at 20 μ M after assessing a range of concentrations) or control (RPMI media)

was injected into the chemoattractant channels. The cell movement was analyzed using a Zeiss Primovert microscope (Carl Zeiss Imaging, Thornwood, NY, USA) and Images captured every 30 s for up to 40 frames per condition using a Q Imaging Retiga 2000R camera (Qimaging, Surry, Canada).

Binding of LL-37 and apo-A1 by surface plasmon resonance. The SPR experiments were performed using a BIACORE 3000 instrument (GE Healthcare). The N-HisTag containing ApoA1 (NH-ApoA1) was immobilized on the surface of a NTA sensor chip (GE Healthcare) according to manufacturer's protocol. Shortly, for direct capture of NH-ApoA1, the surface of NTA chip was prepared by 1 min injection of 0.5 mM NiCl₂, then NH-ApoA1 at concentration of 2,5 µg/ml diluted in running buffer (10 mM HEPES, 150 mM NaCl, and 0.005% surfactant (v/v), pH 7.4) was injected with a flow rate of 5 µl/min to achieve capture level between 800-1000 resonance units (RU). The surface stabilized ApoA1 was primed with running buffer before subsequent assay steps. The binding of all LL-37 analogs were each tested in triplicate in concentration range up to 1000 nM in running buffer. All samples were injected at a flow rate of 5 µl/min. Between experiments, the surfaces were strictly regenerated with two pulses (30 s) of 1 M NaCl at a flow rate of 20 µl/min, followed by an extensive wash procedure with running buffer. All measurements were performed at a constant temperature of 25 °C. Sensorgrams corrected for the reference were aligned, and a blank run was subtracted.

Results

LL-37 undergoes rapid carbamylation, which affects the peptide structure.

Multiple studies based on circular dichroism [23], Fourier Transform Infrared [26] and NMR spectroscopy [27] reveal that LL-37 exists as a linear cationic, amphipathic α -helical structure within vesicles, both within the lipid bilayer and in solutions with an ionic composition resembling that of intracellular fluid or plasma. We used mass spectrometry to show that LL-37 undergoes rapid modification in the presence of cyanate, with the number of carbamylated residues increasing in a concentration- and time-dependent manner (Fig. 1A–D). In the presence of 10 mM KCNO (equivalent to cyanate levels found in the inflammatory milieu), multiple forms of carbamylated LL-37 were detected in the reaction mixture after only 10 minutes. Closer analysis of the elution pattern of modified peptides from a RP-HPLC column revealed two distinct forms of LL-37 bearing a single homocitrulline, which were the most commonly detected modifications. Although almost 70% of the modifications were represented by a single site substitution, a variety of peptides carrying multiple (two, three or four) carbamylated residues (Fig. 1B) were also detected. Increasing either the cyanate concentration (to 50 mM and 100 mM) or incubation time shifted the modification pattern. In 50 mM KCNO, previously predominant peptides bearing a single modification were replaced by peptides bearing multiple carbamylations on different Lys residues, resulting in a highly heterogeneous mixture. Peptides of the same molecular mass bearing one or more carbamylated amino groups resolved into several peaks (same color in Fig. 1C), which depended on the particular combination of the amino groups modified. This suggests that all six lysine side chains in LL-37 were equally susceptible to carbamylation. This was confirmed by analysis of LL-37 in the presence of 100 mM cyanate, which showed the presence of a heterogeneous

population of LL-37 with both 5 and 6 carbamylations (Several peaks with same colour Fig. 1C).

To address the functional significance of LL-37 carbamylation, we focused our attention on modifications identified after incubating the peptide in an environment mimicking that of the inflammatory milieu. To this end, we first identified the most common carbamylated variants of LL-37 under these conditions. KCNO-treated samples were digested with *S. aureus* protease V8 and analyzed by LC-MS/MS. By calculating the number of spectra matching the identified peptides (spectral count), we showed that LL-37 bearing a carbamylated N-terminus (α -carbamyl-Leu-1 LL-37; LL37^{C1}) was the predominant peptide form present after 1 h of exposure to 10 mM KCNO. The second most common single-modified peptide was LL-37 bearing homocitrulline residue at position 8 (LL37^{C8}) (Fig. 1D).

Prolonged incubation with KCNO generated a mixture of LL-37 peptide derivatives containing homocitrulline at random positions. Among them, LL-37 bearing a double modification (on Lys-12 and Lys-15; LL-37^{C12,15}) was frequently detected (data not shown). Based on these results, LL37^{C1}, LL-37^{C8}, and LL-37^{C12,15} were synthesized for further analysis (Table1).

In aqueous solution, LL-37 forms a random coil structure, but adopts an α -helical conformation under physiological conditions; the α -helical structure is also adopted in TFE or lipid bilayer vesicles [26]. In the latter case, NMR studies show that LL-37 comprises three basic parts: an N-terminal α -helix, a C-terminal α -helix, and a C-terminal tail [27]. The hydrophobic surface of LL-37 is bordered by positively charged residues, which enable interaction with negatively charged molecules or structures such as LPS [28], genetic material, and bacterial cell wall components [13,14]. We hypothesized that carbamylation-induced changes in charge

and hydrophobicity would have a significant impact on the secondary structure of the peptide and, consequently, its biological activity (which is strictly related to the physicochemical properties of the peptide). We used CD spectroscopy to examine the impact of carbamylation on the capacity of LL-37 to form a helical structure. The far UV CD spectrum of LL-37 in a physiological salt solution resembling blood plasma showed two minima (at 208 and 222 nm) (Fig. 2A). These are characteristic for an α -helical secondary structure. Interestingly, neither single carbamylation of Lys-8 (LL-37^{C8}) nor double carbamylation of Lys-12 and Lys-15 (LL-37^{C12,15}) had any impact on the α -helical structure of LL-37. Conversely, carbamylation of the N-terminal amino group (LL37^{C1}) led to a significant increase in the propensity of the peptide to adopt an α -helical structure (Fig. 2A).

As expected, incubation with TFE increased the ellipticity and the helical content of the peptides beyond that observed in plasma buffer (Fig. 2B). Under these conditions, CD analysis of the far UV spectra predicted an α -helical content of approximately 80% for LL37^{C1}, but approximately 50% for the native peptide (Fig. 2B). However, the difference between the peptides in TFE was similar to that observed in plasma buffer, i.e., the α -helical composition of LL37^{C1} was about 30% greater than that of the other peptides (Fig. 2C).

Carbamylation abrogate the bactericidal capacity of LL-37.

LL-37 interacts with bacteria via electrostatic and hydrophobic interactions resulting in membrane permeabilisation and disruption. A broth micro-dilution assay revealed that the bactericidal activity of LL-37 was profoundly affected by carbamylation. At a peptide concentration of 1 $\mu\text{g/ml}$ (i.e., 0.2 μM), LL37^{C8} and LL37^{C12,15} demonstrated impaired ability to inhibit bacterial growth of *B. subtilis* when compared with the

native peptide ($p < 0.05$ and $p < 0.001$, respectively; Figure 3A). Additionally, the potential of LL37^{C12,15} to inhibit the growth of *E. coli* ($p < 0.001$; Fig. 3B) and *S. aureus* ($p < 0.05$; Fig. 3C) was significantly decreased as compared to native LL-37. Confirming previous observations, that reducing the number of residues on the N terminus of LL-37 had only minor impact on its bactericidal properties, carbamylation of the N-terminal amino group did not affect the antimicrobial capacity of LL37^{C1} as compared to that of the native peptide (Fig. 3A – C). In addition to its direct microbicidal role, LL-37 is a potent regulator of innate immunity, controlling the response to pathogen-associated molecular patterns including endotoxin (LPS). Electrostatic interaction of the cationic LL-37 with the strongly anionic Lipid A domain of LPS prevents it from binding to TLR receptors expressed by monocytes and macrophages, a keystone event in inflammatory response. Therefore, we investigated whether carbamylation, and thus conversion of cationic Lys residues into neutral homocitrulline **in LL-37 will affect the capacity of LPS to stimulate cells.** hMDMs were exposed to LPS (100 ng/ml) in the presence of the either native LL-37 or carbamylated peptides. Thereafter, the cytokine profile of the supernatants was examined by multiplex analysis. LL-37^{C1} and LL-37^{C8} attenuated the pro-inflammatory activity of LPS and blocked secretion of TNF- α and IL-6 from the cells as effectively as the native peptide. Interestingly, we observed statistically significant decrease of TNF- α and IL-6 in supernatants from LPS stimulated macrophages in the presence of LL-37^{C12,15} as compared to the native peptide (Fig.4A, B). **This may indicate that,** as compared to the native peptide, LL-37^{C12,15} exhibits higher affinity for endotoxin thereof limiting its binding to the TLR4 receptors.

Carbamylation of LL-37 affects its affinity for Apo-A1.

In vivo, LL-37 circulates in a complex with Apo-A1 [24]. This carrier protein might function as a scavenger to inhibit the cytotoxic effects of the peptide secondary to its release at inflammatory sites. Therefore, we used surface plasmon resonance to examine whether carbamylation alters the affinity of LL-37 for Apo-A1, which would effectively modulate the concentration of the bioavailable peptide within the inflammatory milieu. We found that Lys-12 and Lys-15 are critical for the interaction between the peptide and Apo-A1. LL-37^{C12,15} showed a 3-fold lower affinity for Apo-A1 than the native peptide. By contrast, both peptides carrying a single modification, LL-37^{C8} and LL-37^{C1}, showed significantly stronger binding (1.5 and 3 times higher, respectively) than native LL-37. Taken together, these data indicate that any interference with the charged side chains of the amino acids within the LL-37 polypeptide chain has a significant impact on the peptide's ability to associate with its carrier protein. This may either reduce or amplify the observed *in vivo* effects by modulating the amount of accessible LL-37 in the environment (Fig. 5).

Carbamylation affects the chemotactic capacity of LL-37.

The ability of neutrophils to efficiently reach sites of inflammation is pivotal for efficient elimination of pathogens and restriction of potential tissue damage. LL-37 is a strong chemoattractant for neutrophils, monocytes, and T cells via the FPRL1 receptor [15]. To investigate how carbamylation impacts the chemotactic activity of LL-37, we used a “state of the art” approach that allowed us to observe cell migration in real time. The Insall chambers used in the study provide a gradient for the cells to migrate against, rather than simply exposing cells to the chemoattractant alone. This in turn allows observation of cell migration in more detail as information about speed (average speed of the cell over time in any direction), velocity (average speed of the

cell in the direction of the gradient over time), and the directional accuracy of chemotaxis (expressed as the chemotactic index) can be obtained. We found that a peptide concentration of 20 μM was optimal in our assay (tested concentrations, 10 μM –40 μM ; data not shown). Both native LL-37 and the carbamylated forms induced neutrophil migration without any apparent toxicity (Fig. 6A, B, C). However, neutrophils were significantly less responsive to the carbamylated versions of LL-37. Both the speed and velocity of migrating neutrophils were significantly lower when LL37^{C1}, LL37^{C8}, and LL37^{C12,15} were used as chemoattractants. Interestingly, LL37^{C1} was not only a significantly weaker chemoattractant than the other carbamylated peptides, but it also showed lower directional accuracy. Taken together, these observations suggest that, although all of the homocitrullinated peptides were able to trigger cell migration, carbamylation of the N-terminal amino group had a strong negative effect on directional accuracy.

Carbamylation affects the cytotoxicity of LL-37.

LL-37 can be cytotoxic to host cells. Therefore, we used a highly sensitive hemolytic assay to examine the impact of carbamylation on the cell-permeabilizing effects of LL-37. Human red blood cells (hRBC) were incubated with the peptide for 2 h at 37°C. The results showed that LL37^{C1} exerted a very strong and concentration-dependent lytic effect. The peptide caused a significant increase in hemoglobin release when used at a concentration of 2 μM (# $p < 0.05$; Fig. 7) as compared to PBS. The half-maximal effective concentration for LL37^{C1} was 17.9 μM . Conversely, LL37^{C8} and LL37^{C12,15} had significantly less capacity to induce membrane permeabilization (Fig. 7); indeed, hRBC were resistant to permeabilization by LL37^{C12,15} at concentrations of up to 20 μM . Given that the concentration of LL-37

can easily reach 20 μM at sites of inflammation (with levels up to 250 μM reported in psoriatic lesions) [29], permeabilization of membranes might facilitate the extracellular release of various potentially deleterious substances from lysed cells, which may then activate neutrophils and eventually lead to further increases in carbamylation.

Discussion

Host antimicrobial peptides, in particular LL-37, are potential novel therapeutics, mainly due to their wound healing, antiseptic, and antimicrobial properties[14,30,31]. In the era of rapidly increasing drug resistance among pathogens, novel treatments based on LL-37-derived peptides are very much needed. In addition, the immunostimulatory and cytotoxic properties of LL-37 may be useful tools for cancer treatment [32], or even as a vaginal contraceptive [33]. It is, however, important to keep in mind that the effector functions of LL-37 are most often executed at local sites of inflammation, and mainly in the context of neutrophil activation and subsequent release of MPO and H_2O_2 [34]. Thus, LL-37 is mainly present in microenvironments that foster carbamylation. Therefore, it is important to examine the impact of carbamylation on the immunomodulatory and cytotoxic functions of LL-37 prior to the clinical administration of cathelicidin-derived peptides. Here, we explored the effects of carbamylation on the biological functions of LL-37.

We found that in the presence of cyanate, LL-37 undergoes rapid modification to generate a pool of peptides, each with a diverse carbamylation pattern. Mass spectrometry analyses revealed that the N-terminal amino group of leucine residues is highly reactive and is modified almost instantaneously in the presence of 10 mM KCNO. Thus, LL37^{C1} is most likely the predominant form of carbamylated LL-37 *in*

in vivo. Even though carbamylation at this site did not affect the bactericidal properties of the peptide, LL-37^{C1} lost the ability to function as a chemoattractant for neutrophils. Concurrently, LL-37^{C1} gained significant RBC-lysing capacity; indeed, it was almost 3-fold more cytotoxic than native LL-37 in a hemolytic assay. Because tissue injury/necrosis results in increased receptor-dependent immune cell migration in response to released intracellular components[35,36], we cannot rule out the possibility that LL37^{C1} indirectly triggers cell migration *in vivo* in response to damage to surrounding tissue. By stark contrast, although the chemoattractive capacity of LL-37^{C8} and LL-37^{C12,15} was diminished, neutrophils retained directional accuracy in response to the stimuli. This suggests that, despite the reduced affinity of Lys-8 and Lys-12/Lys-15 carbamylated peptides for FPRL-1, they were still able to bind the receptor. At the same time, LL-37^{C8} and LL-37^{C12,15} were significantly less toxic to hBRCs, with the latter exhibiting no hemolytic effects at a concentration of 20 μ M. This observation confirms those of previous studies showing that truncation or blocking [26] of the N-terminus reduces the cytotoxicity of LL-37 while at the same time leaving bactericidal activity unchanged [37]. Indeed, only LL-37^{C12,15} showed lower bactericidal activity than the native peptide against *B. subtilis*, *E. coli*, and *S. aureus*. In addition, Xhindoli and colleagues showed that oligomerization of LL-37 results in increased formation of α -helices, leading to an increased capacity to permeabilize erythrocytes and monocytes; this suggests that a parallel arrangement of the peptides favors aggregation *via* interaction between the N-termini [26]. In agreement with these results, we showed that LL37^{C1} was about 50% more α -helical than the native peptide, both in a secondary structure-inducing environment (50% TFE buffer) and in a plasma-mimicking buffer. Since loss of the N-terminal charge due to carbamylation increases the hydrophobicity of this region [38], it is possible

that LL37^{C1} is more prone to aggregation due to reduced electrostatic repulsion between the molecules. This would increase its propensity to form α -helices and explain the increased cytotoxicity. Intriguingly, the ability of LL-37^{C8} and LL-37^{C12,15} to lyse neutrophils and erythrocytes was different. Neutrophil membranes are zwitterionic and interact with LL-37 independently of the overall charge of the peptide [22,39]. By contrast, the erythrocyte membrane contains sialic acid, which results in a negatively charged cell surface; therefore, it is more susceptible to lysis by cationic peptides. Thus, it is not surprising that the reduced cationicity conferred by carbamylation results in impaired hemolysis but does not affect the lysis of neutrophils. Interestingly, this difference was not evident in the case of LL37^{C1}. However, the uncompromising capacity of LL37^{C1} to lyse both erythrocytes and neutrophils (data not shown) emphasizes the involvement of a hydrophobic N-terminus in this process.

The antimicrobial and cytotoxic activities of LL-37 are effectively inhibited in human plasma. This latency is due to the interaction between LL-37 and its carrier protein, apoA-1 [24]; this interaction is dependent on the hydrophobicity of the N-terminus of LL-37 [40] as well as on its α -helical content [41]. In line with these results, we found that LL37^{C1} showed significantly higher affinity for apo-A1 (300%) than the native peptide, indicating that the cytotoxic effects of LL37^{C1} may be limited by an Apo-A1-dependent protective mechanism. However, LL37^{C1} may be detrimental to the host if generated at inflammatory foci that are poorly infiltrated by blood plasma. By contrast, LL37^{C12,15} had lower affinity for apo-A1. Since Apo-A1 has a net negative charge [42], it is likely that the significant loss of electrostatic interactions upon carbamylation of two or more Lys residues impairs L-37 binding to apo-A1.

Upon carbamylation, the bactericidal activity of LL-37 is compromised. Again, this is most likely related to the loss of two positive charges when Lys residues are converted to homocitrulline. It is likely that the reduced electrostatic attraction between LL-37^{C12,15} and bacterial membranes results in impaired antimicrobial activity against *B. subtilis*, *E. coli*, and *S. aureus*. Interestingly, even though the bactericidal domain of LL-37 has been mapped to the C-terminal region (amino acids 17–29) [43], LL-37^{C8} showed impaired bactericidal activity against *B. subtilis*. Thus, the lysine at position 8 is important for efficient killing of this Gram-positive bacterium.

Interestingly, LL-37^{C12,15} was the most effective LPS-detoxifying agent among all tested forms of LL-37, i.e., this carbamylated peptide efficiently attenuated the LPS-induced production of TNF- α , an important mediator of endotoxic shock [44]. In addition to directly binding LPS via electrostatic and hydrophobic interactions, LL-37 also interacts with the LPS receptor, CD14, to block LPS-induced macrophage activation [45]. We hypothesize that carbamylation of Lys-12 and Lys-15 increases the binding of LL-37 to this receptor. From a clinical point of view, it is tempting to speculate that LL37^{C12,15} provides therapeutic protection against septic shock without being cytotoxic. Nevertheless, it must be kept in mind that further carbamylation may alter the biological activity of the peptide.

In summary, carbamylation has a profound impact on the bactericidal, cytotoxic, and pro-inflammatory activity of LL-37, which may have detrimental consequences for the host. The pattern of carbamylation is dependent on the OCN⁻ concentration and time of exposure, suggesting that the subsequent effects of this modification will be difficult to foresee and, therefore, control *in vivo*. Thus, we suggest that caution should be exercised when administering cathelicidin-derived

peptides to patients with diseases manifested by inflammation, such as severe infections or sepsis.

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Tables and Figures

Table 1. LL-37 peptides synthesized and examined in this study.

Peptide	Amino acid sequence
LL-37	LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNLVPRTES
LL-37 ^{C1}	(α -carb-L)LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNLVPRTES
LL-37 ^{C8}	LLGDFFR(ϵ -carb-K)SKEKIGKEFKRIVQRIKDFLRNLVPRTES
LL-37 ^{C12,15}	LLGDFFRKSKE(ϵ -carb-K)IG(ϵ -carb-K)EFKRIVQRIKDFLRNLVPRTES

α -carb-L, leucine carbamylated on the α -carbon; ϵ -carb-K, lysine carbamylated on the ϵ -carbon.

Figure 1. The number of carbamylated amino acid residues increases with KCNO concentration. LL-37 was incubated for 3 h with increasing amounts of KCNO. (A) 0 mM, (B) 10 mM, (C) 50 mM. The samples were then analyzed by liquid chromatography (LC)-MS/MS and the m/z values corresponding to LL-37 bearing 0–7 carbamylated amino acid residues were extracted (see legend for color code). KCNO-mediated carbamylation results in a heterogeneous population of carbamylated LL-37 peptides, as evidenced by the appearance of multiple peaks. (D) The N-terminal leucine (L1) and the lysine at position 8 within LL-37 are highly accessible for carbamylation. LL-37 was incubated with 10 mM or 50 mM KCNO for 1 h at 37°C. The position of the modifications was determined using NanoESI-

MS/MS after V8 digestion. Spectral counts were calculated by summing the number of spectra matching the identified peptides.

Figure 2. CD spectra of native and carbamylated LL-37. The peptide concentration was 10 μ M. The TFE buffer comprised 50% TFE in 10 mM sodium phosphate buffer. The plasma buffer contained 113 mM NaCl, 24 mM NaHCO₃, 0.6 mM MgCl₂, 1.3 mM CaCl₂, and 3.9 mM KCl. (A) Spectra represent the mean of two (50% TFE buffer) or three (plasma buffer) independent experiments. (B) Differences in the predicted α -helical content of the peptide analogues in 50% TFE buffer at 190–260 nm. (C) Difference in the predicted α -helical content of the peptide analogues in plasma buffer at 190–260 nm. (B–C) Data are expressed as the mean \pm SD. Statistical significance was evaluated by one-way ANOVA, followed by Tukey's multiple comparison test. $p < 0.05$, $**p < 0.01$, and $***p < 0.001$. TFE, trifluoroethanol.

Figure 3. Carbamylation abrogates the antimicrobial activity of LL-37. (A–C) The number of colony-forming units formed after incubation with (A) *B. subtilis*, (B) *E. coli*, and (C) *S. aureus*. All bacterial species were incubated with carbamylated or native LL-37. Bacterial growth in the presence of native LL-37 was set to 100%. Data are expressed as the mean \pm SD. Statistical significance was evaluated by one-way ANOVA, followed by Tukey's multiple comparisons post-test. $*p < 0.05$, $**p < 0.01$, and $***p < 0.001$; and $\#p < 0.05$, $\##p < 0.01$, and $\###p < 0.001$ compared with negative control (broth).

Figure 4. LPS induced cell activation remain suppressed in the presence of carbamylated LL-37. Human monocyte-derived macrophages were incubated with 100 ng/ml LPS in the absence or presence of modified peptides. The levels of (A) TNF- α and (B) IL-6 in the supernatants were then measured. LL-37+LPS was set at

100%. Data are expressed as the mean \pm SD. Statistical significance was evaluated by one-way ANOVA, followed by Tukey's multiple comparisons post-test. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$; and # $p < 0.05$, ## $p < 0.01$, and ### $p < 0.001$, compared with LPS.

Figure 5. Carbamylation of LL-37 reduces its affinity for apolipoprotein A1.

Surface plasmon resonance curves showing the interaction between Apo-A1 and carbamylated and native LL-37 at a concentration of 1 μ M. RU, resonance units.

Figure 6. Carbamylation reduces the chemotactic potential of LL-37.

Analysis of chemotaxis at a peptide concentration of 20 μ M. Neutrophils from four healthy volunteers were treated with either native or modified peptide. Values for speed, velocity, chemotactic index, and the resultant vector length were analyzed. The midline of each box represents the median value. Statistical significance was evaluated by one-way ANOVA, followed by Tukey's multiple comparisons post-test: * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$; and # $p < 0.05$, ## $p < 0.01$, and ### $p < 0.001$, compared with the native peptide.

Figure 7. Carbamylation significantly affects the hemolytic capacity of LL-37.

Erythrocytes were incubated for 2 h with each LL-37 analogue at concentrations ranging from 1–20 μ M. The hemolytic activity of the peptides was evaluated by recording the release of hemoglobin from human erythrocytes at 405 nm. Data are representative of four individual experiments and are expressed as the mean \pm SD. Statistical significance was evaluated by one-way ANOVA, followed by Dunnett's multiple comparisons post-test: * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$, compared with LL-37. # $p < 0.05$ in case of LL-37^{C1} as compared with PBS.