

Organometallic nucleoside analogues with ferrocenyl linker groups

Nguyen, Huy V; Sallustrau, Antoine; Balzarini, Jan; Bedford, Matthew R; Eden, John C; Georgousi, Niki; Hodges, Nikolas J; Kedge, Jonathan; Mehellou, Youcef; Tselepis, Chris; Tucker, James H R

DOI:
[10.1021/jm500246h](https://doi.org/10.1021/jm500246h)

License:
None: All rights reserved

Document Version
Peer reviewed version

Citation for published version (Harvard):
Nguyen, HV, Sallustrau, A, Balzarini, J, Bedford, MR, Eden, JC, Georgousi, N, Hodges, NJ, Kedge, J, Mehellou, Y, Tselepis, C & Tucker, JHR 2014, 'Organometallic nucleoside analogues with ferrocenyl linker groups: synthesis and cancer cell line studies', *Journal of Medicinal Chemistry*, vol. 57, no. 13, pp. 5817-22.
<https://doi.org/10.1021/jm500246h>

[Link to publication on Research at Birmingham portal](#)

Publisher Rights Statement:

Final version of record published as: Nguyen, Huy V., et al. "Organometallic nucleoside analogues with ferrocenyl linker groups: synthesis and cancer cell line studies." *Journal of medicinal chemistry* 57.13 (2014): 5817-5822.
Available online at: <http://dx.doi.org/10.1021/jm500246h>

Checked Jan 2016

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Organometallic Nucleoside Analogues with Ferrocenyl Linker Groups: Synthesis and Cancer Cell Line Studies.

Huy V. Nguyen,^a Antoine Sallustrau,^a Jan Balzarini,^b Matthew R. Bedford,^c John C. Eden,^d Niki Georgousi,^d Nikolas J. Hodges,^d Jonathan Kedge,^a Youcef Mehellou,^c Chris Tselepis,^c and James H. R. Tucker,^{*a}

^aSchool of Chemistry, University of Birmingham, Edgbaston, Birmingham, B15 2TT, UK

^bKU Leuven, Rega Institute for Medical Research, Laboratory of Virology and Chemotherapy, Minderbroedersstraat 10, B 3000 Leuven, Belgium

^cSchool of Cancer Sciences, College of Medical and Dental Sciences, University of Birmingham, B15 2TT, UK

^dSchool of Biosciences, College of Life and Environmental Sciences, University of Birmingham, B15 2TT, UK

^eSchool of Clinical and Experimental Medicine, College of Medical and Dental Sciences, University of Birmingham, Edgbaston, Birmingham, B15 2TT, UK

ferrocene, nucleoside analogues, anticancer, bioorganometallic, DNA

ABSTRACT: Examples of organometallic compounds as nucleoside analogues are rare within the field of medicinal bioorganometallic chemistry. We report on the synthesis and properties of two chiral ferrocene derivatives containing both a nucleobase and a hydroxyalkyl group. These so-called ferronucleosides show promising anticancer activity, with cytostatic studies on five different cancer cell lines indicating that both functional groups are required for optimal activity.

Nucleoside analogues have long been established as an effective class of compound that exhibits antiviral or anticancer activity.¹ Two common structural features are a nucleobase moiety and a hydroxymethyl group (Figure 1), which together allow them to act as substrates that adversely affect processes associated with nucleic acid synthesis. These two components are typically connected by an organic linker group that is a modification or a replacement of the sugar ring, which can either be cyclic (e.g. AZT) or acyclic (e.g. acyclovir).² Due to their structural similarities to natural nucleosides, which can lead to resistance and side-effects, there is a continuing need for a diverse range of analogues with different structural features. Ferrocene has attracted active interest in recent years within the field of medicinal and bioorganometallic chemistry,³ with organometallic analogues and derivatives of the antimalarial drug chloroquine (ferroquine) and the breast cancer drug tamoxifen (the ferrocifen family) being the most widely known.⁴ At the same time there has been a number of examples of other ferrocene containing compounds that have shown anticancer,⁵ antibacterial and antifungal properties.⁶ However although there are also some recent examples of ferrocene-conjugated nucleobases⁷ and hydroxylalkyl ferrocenes⁸ that exhibit anticancer activity, as far as we are aware, nucleoside analogues of the type shown in Figure 1 that are bridged solely by an organometallic linker group and show biological activity have not been reported.⁹

As part of our programme to develop novel metal-containing analogues of DNA and its components,¹⁰ we recently reported an organometallic nucleic acid oligomer

designated as Ferrocene Nucleic Acid (FcNA).^{10b} The monomeric components of the reported form of FcNA consist of a tetra-substituted ferrocene unit containing two alkyl hydroxyl groups to allow connectivity via phosphodiester, and two thymine nucleobases. We noticed that these monomeric compounds have the required features for a novel nucleoside analogue, where the 5-membered sugar ring is substituted for a 5-membered cyclopentadienyl ring of a ferrocene unit. Accordingly we now report bisubstituted ferrocenes **1** and **2** (so-called ferronucleosides), that contain both a nucleobase (thymine or adenine) and a hydroxyl group, along with various control compounds (Fig. 2). The cell line studies reported here demonstrate the promise of these ferrocenyl derivatives as a novel class of nucleoside analogue that show anticancer activity.

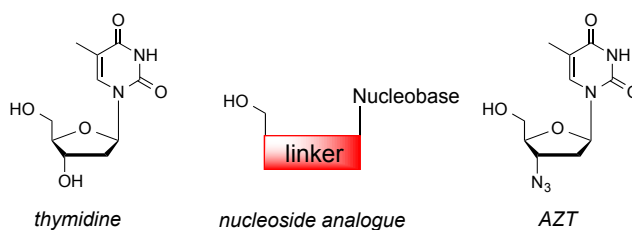
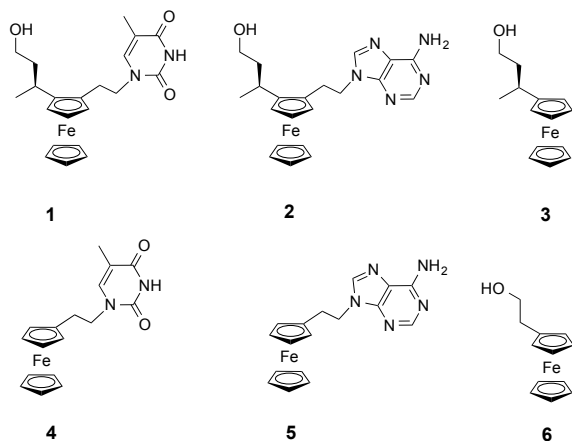


Figure 1. Schematic representation of the structural relationship between thymidine and a nucleoside analogue that contains a variable linker group connecting a hydroxymethyl group with a nucleobase, with AZT (azidothymidine) as a specific example.

Figure 2. Ferronucleosides **1** and **2** and controls **3-6**.



It was decided to first synthesize nucleoside analogues with a 1,2-disubstituted arrangement on one Cp ring, with the other Cp ring unfunctionalized. This would enable us to utilize the synthetic chemistry already developed within the group for FcNA monomer synthesis.^{10b} For the same reason, the compounds would have a three-carbon hydroxyl linker (with a methyl group on the alpha carbon to direct *ortho*-lithiation) and a two-carbon linker to the nucleobase. The synthetic route taken to make the ferronucleoside targets **1** and **2** is outlined in Scheme 1. The chirally pure Ugi amine¹¹ **7** was treated with *n*-BuLi and quenched with iodine to introduce the required planar 1,2-disubstitution pattern. Subsequent functional group inter-conversion gave compound **11**, to provide the chain extension giving a three carbon linker. Treatment of **11** with silyl enol ether, catalyzed by the Lewis acid boron trifluoride, gave compound **12** in good yield. Reduction of the ester, followed by TBDPS protection gave compound **14**. Conversion to aldehyde **15** (via *n*-BuLi halogen exchange and quenching with DMF) enabled a Wittig reaction to be performed to form alkene **16**, with subsequent hydroboration-oxidation giving the mono-protected bis-alcohol **17** in high chiral purity (as checked by chiral HPLC analysis, overall 97% *ee*). The conversion of **17** to the target compounds **1** and **2** proceeded via a Mitsunobu reaction with the appropriate protected nucleobase, followed by deprotection of the protecting groups. The family of compounds was also extended by making the control compounds **3-6** (Fig. 2), to assess the role of the alcohol and nucleobase groups, noting that compound **6** had previously been shown⁸ to display antineoplastic activity against cervix carcinoma (HeLa) tumor cells.

The cytostatic activity of six ferrocene compounds was evaluated in comparison to the established anticancer drugs cisplatin and 5-fluorouracil (5-FU) using a proliferation activity assay carried out on three tumor cell lines: murine leukemia cells (L1210), HeLa, and human T-lymphocyte cells (CEM). The data indicate that the concomitant presence of both the hydroxyl and nucleobase components are of crucial importance to give the highest cytostatic activity, with **1** and **2** exhibiting low to sub micromolar antiproliferative activity comparable with cisplatin (Table 1). In addition,

1 and **2** were almost equally as active as 5-FU in L1210 cell cultures, 2- to 5-fold less active in HeLa cell cultures, but 20- to 50-fold more active in CEM cell cultures. Compounds **1** and **2** proved poorly toxic to non-tumorigenic human embryonic lung (HEL) fibroblast cell cultures (minimal cytotoxic concentration: >50 μ M).

Scheme 1. Synthetic route towards ferrocene nucleoside analogues **1** and **2**.

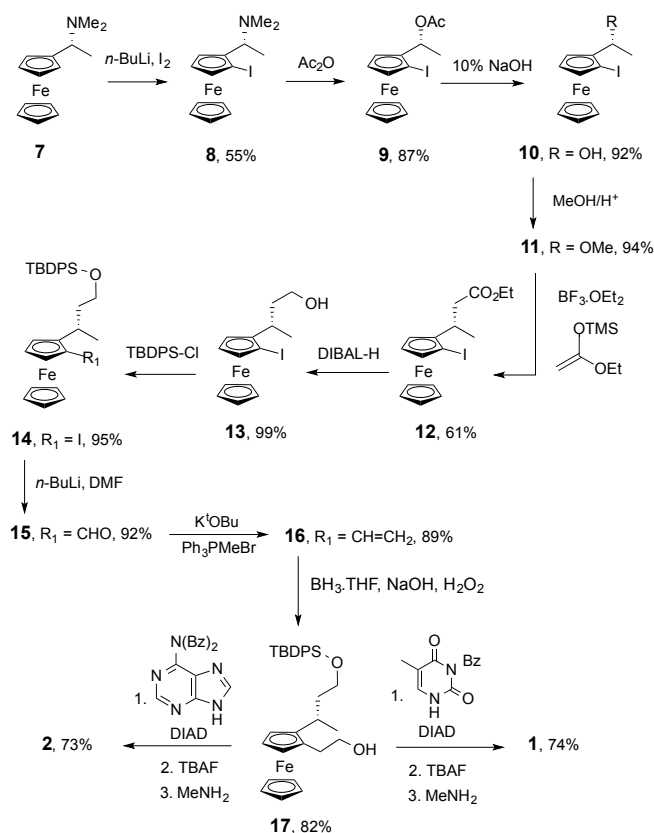


Table 1. Cytostatic activity of compounds **1-4**.

Compound	IC ₅₀ (μ M) ^a		
	L1210	CEM	HeLa
1	0.78	0.9	2.68
2	1.0	0.35	1.1
3	12	39	45
4	417	592	509
5	26	49	94
6	25	43	52
Cisplatin	1	0.9	1.2
5-FU	0.33 \pm 0.17	18 \pm 5	0.54 \pm 0.12

^a50% inhibitory concentration, or compound concentration required to inhibit tumor cell proliferation by 50%. Data are the mean of at least two independent experiments.

Cell growth studies carried out on an oesophageal cancer cell line revealed that compound **1** inhibited growth at

concentrations of 6.25 μM , whereas control compounds **3** and **4** (Figure 3 and supplementary data respectively) had much less or no effect, even up to higher concentrations of 25 μM . The same trend was observed for the adenine compound **2** and its control **5** (see supplementary data). Once again the data indicates that both functional groups (the hydroxyl in addition to the nucleobase) are required for the best cytostatic activities, which is comparable to cisplatin under these conditions.

Assays of cellular viability (MTT assay) and cell proliferation (BrdU assay) were then performed on colorectal cancer cell lines. The results after 48hr exposure (Figure 4) revealed that compounds **1** and **2** had anti-neoplastic activities approaching that 5-FU, whereas the other compounds were less effective. Encouragingly, an AMES assay to investigate the potential mutagenicity of these ferrocenyl derivatives revealed that the compounds were inactive.

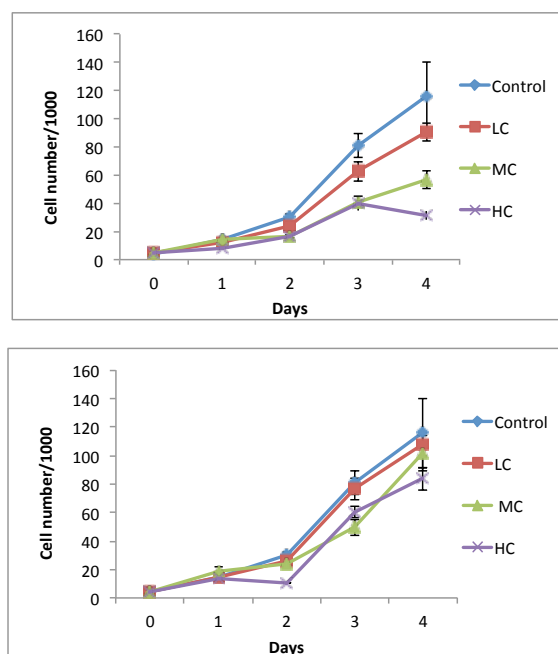


Figure 3. Growth curves for compound **1** (top) and **3** (bottom) at three different concentrations (LC = 6.25 μM , MC = 12.5 μM , HC = 25 μM) over 4 days in cancer cell line OE 19 ($n = 3 \pm \text{SD}$).

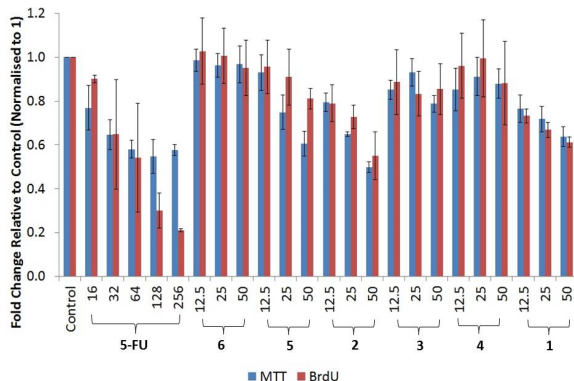


Figure 4. Cellular viability (MTT, blue) and cell proliferation (BrdU, red) assays on colorectal cell lines (48 hours).

In conclusion, novel ferrocenyl nucleoside analogues **1** and **2** appear to exhibit cytostatic activities that are comparable under the conditions used to commercially established anticancer drugs such as cisplatin and 5-FU. Control studies indicate that the presence of both a hydroxyl and a nucleobase group are required for optimal activity. This suggests a mechanistic role for these novel bioorganometallic compounds involving an adverse affect on nucleic acid synthesis, as is the case for nucleobase analogue drugs containing organic linker groups.¹² We are currently planning to carry out further studies to reveal the mode of action of these ferronucleosides and to highlight stereochemical and structure activity relationships for improving biological activity and specificity.

EXPERIMENTAL SECTION

General Information: Unless stated otherwise, all reactions were performed under an Ar atmosphere. Compound **6** was prepared as described previously.⁸ All tested compounds had a purity of $\geq 95\%$, as shown by HPLC (see supplementary information for data and conditions used).

(*R,S_p*)-1-(α -*N,N*-Dimethylaminoethyl)-2-iodo-ferrocene (8**).** The Ugi amine **7**¹¹ (4.00 g, 15.56 mmol) was dissolved in Et₂O (50 ml) at room temperature, *n*-BuLi (12 ml, 30 mmol) was added and the mixture stirred overnight. The reaction mixture was cooled to -78°C and iodine (9.52 g, 37.51 mmol), dissolved in THF (60 ml), was added over 10 min. The reaction mixture was stirred at -78°C for 90 min before being warmed to room temperature, at which point it was stirred for an additional 90 min before being quenched at 0°C with sodium thiosulfate_(aq) (50 ml, 25% w/v). After dilution with Et₂O (30 ml), the layers were separated and the aqueous layer further extracted with Et₂O (3 x 50 ml). The combined organic fractions were dried over MgSO₄, the solvent removed *in vacuo* before purification via flash column chromatography (5% MeOH, 5% TEA in DCM) to yield product (3.18 g, 55%). ¹H NMR (400 MHz, CDCl₃) δ 4.46 (dd, $J = 2.4, 1.4$ Hz, 1H), 4.24 (t, $J = 2.6$ Hz, 1H), 4.15 (dd, $J = 2.7, 1.3$ Hz, 1H), 4.12 (s, 5H), 3.62 (q, $J = 6.8$ Hz, 1H), 2.15 (s, 6H), 1.50 (d, $J = 6.8$ Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 90.21 (ipso Cp), 74.32 (Fc), 71.67 (Fc), 68.19 (Fc), 65.59 (Fc), 57.59 (CH*), 45.49 (ipso Cp), 41.22 (CH₃), 16.01 (CH₃). MS (ES) (m/z) calcd for C₁₄H₁₈N⁵⁶FeI 382.9833, found 382.9820. IR (cm⁻¹): 3078 (=C-H), 2931 (CH₂), 2878 (CH₂), 2809 (CH₂), 1446 (CH₃), 1371 (CH₃), 1243, 1087, 821 (CH=CH), 732 (CH Ar). Mp: 58°C-60°C.

(*R,S_p*)-1-(α -Acetoxyethyl)-2-iodo-ferrocene (9**).** Compound **8** (3.26 g, 8.51 mmol) and acetic anhydride (25.68 ml, 272.17 mmol) were heated at 50°C for 2 hrs. The acetic anhydride was removed under high vacuum (0.1 mmHg) and the residue purified via flash column chromatography (10% EtOAc in hexane) to yield the yellow-brown oily product (2.94 g, 87%). ¹H NMR (400 MHz, CDCl₃) δ 5.89 (q, $J = 6.4$ Hz, 1H), 4.51 (dd, $J = 2.6, 1.4$ Hz, 1H), 4.33 (dd, $J = 2.8, 1.4$ Hz, 1H), 4.28 (t, $J = 2.6$ Hz, 1H), 4.15 (s, 5H), 2.01 (s, 3H), 1.66 (d, $J = 6.5$ Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.30 (C=O), 87.54 (ipso Cp), 75.63 (Fc), 71.76 (Fc), 69.71 (Fc), 68.94 (Fc), 65.80 (CH*), 44.03 (ipso Cp), 21.16 (CH₃), 18.66 (CH₃). IR (cm⁻¹): 3095 (=C-H), 2972

(CH₂), 2928 (CH₂), 2866 (CH₂), 1729 (C=O), 1445 (CH₃), 1371 (CH₃), 1085, 820 (CH=CH), 703 (CH Ar).

(R,S_p)-1-(α -Hydroxyethyl)-2-iodo-ferrocene (10).

Compound **9** (2.937 g, 7.37 mmol) was dissolved in EtOH (35 ml). NaOH_(aq) (30 ml, 10% w/v) was added and the reaction was heated at 95°C for 15 min. After cooling to room temperature, the organic layer was extracted with EtOAc (2 x 40 ml). The organic layers were dried over Na₂SO₄, the solvent removed *in vacuo* and the residue purified via flash column chromatography (25% EtOAc in hexane) to yield the yellow oily product (2.43 g, 92%) ¹H NMR (400 MHz, CDCl₃) δ 4.85 (qd, *J* = 6.5, 2.8 Hz, 1H), 4.46 (dd, *J* = 2.5, 1.4 Hz, 1H), 4.29 (dd, *J* = 2.7, 1.3 Hz, 1H), 4.25 (t, *J* = 2.6 Hz, 1H), 4.14 (s, 5H), 1.88 (d, *J* = 3.6 Hz, 1H), 1.62 (d, *J* = 6.5 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 91.61 (ipso Cp), 75.01 (Fc), 71.59 (Fc), 68.72 (Fc), 66.51 (Fc), 64.98 (CH*), 43.62 (ipso Cp), 21.31 (CH₃). MS (ES) (*m/z*) calcd for C₁₂H₁₃O⁵⁶FeI 355.9361, found 355.9352. IR (cm⁻¹): 3255 (OH), 3093 (=C-H), 2967 (CH₂), 2920 (CH₂), 1445 (CH₃), 1369 (CH₃), 1099 (C-OH), 816 (CH=CH), 684 (CH=CH).

(R,S_p)-1-(α -Methoxyethyl)-2-iodo-ferrocene (11).

Compound **10** (2.43 g, 6.83 mmol) was dissolved in a MeOH/AcOH (20 ml, 9:1) mixture and the solution was stirred at room temperature for 48 hrs. The reaction was quenched with water (10 ml) and extracted with DCM (2x20 ml). The combined organic fractions were dried over MgSO₄, the solvent removed *in vacuo* and the residue purified via flash column chromatography (25% EtOAc in hexane) to yield the yellow oily product (2.37 g, 94%). ¹H NMR (400 MHz, CDCl₃) δ 4.49 (dd, *J* = 2.4, 1.4 Hz, 1H), 4.34 (q, *J* = 6.5 Hz, 1H), 4.29 – 4.25 (m, 2H), 4.13 (s, 5H), 3.26 (s, 3H), 1.64 (d, *J* = 6.5 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 89.78 (ipso Cp), 74.78 (Fc), 74.22 (Fc), 71.66 (Fc), 68.86 (Fc), 65.35 (CH*), 56.00 (CH₃), 39.48 (ipso Cp), 19.63 (CH₃). MS (ES) (*m/z*) calcd for C₁₃H₁₅O⁵⁶FeI 369.9517, found 369.9513. IR (cm⁻¹): 3094 (=C-H), 2974 (CH₂), 2926 (CH₂), 2871 (CH₂), 2815 (CH₂), 1448 (CH₃), 1371 (CH₃), 1085 (C-O-C), 820 (CH=CH).

(S,S_p)-1-[α -Methyl(2-ethylpropanoate)]-2-iodo-ferrocene (12). Compound **11** (2.37 g, 6.42 mmol) and 1-ethoxyvinyltrimethylsilane (8.234 g, 51.37 mmol) were dissolved in DCM (30 ml). The mixture was cooled to -78°C and BF₃·OEt₂ (1.77 ml, 14.12 mmol) was then added dropwise. The reaction mixture was stirred for 15 min at -78°C before being warmed to room temperature and quenched with saturated NaHCO₃ (40 ml). The organic layer was separated and the aqueous layer was further extracted with DCM (40 ml). The combined organic fractions were dried over MgSO₄, the solvent removed *in vacuo* and the residue purified via flash column chromatography (10% EtOAc in hexane) to yield the yellow oily product (1.676 g, 61%). ¹H NMR (400 MHz, CDCl₃) δ 4.42 (dd, *J* = 2.4, 1.4 Hz, 1H), 4.18 – 4.08 (m, 7H+2H), 3.14 – 3.05 (m, 1H), 2.53 (dd, *J* = 15.0, 3.7 Hz, 1H), 2.11 (dd, *J* = 15.0, 10.3 Hz, 1H), 1.43 (d, *J* = 6.9 Hz, 3H), 1.25 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 172.00 (C=O), 94.07 (ipso Cp), 74.12 (Fc), 71.52 (Fc), 67.84 (Fc), 64.58 (Fc), 60.26 (CH₂), 44.08 (ipso Cp), 43.19 (CH₂), 30.72 (CH*), 18.90 (CH₃),

14.27 (CH₃). MS (ES) (*m/z*) calcd for C₁₆H₁₉O₂⁵⁶FeI 425.9779, found 425.9782.

(S,S_p)-1-[α -Methyl-(3-(hydroxy)propyl)]-2-iodo-ferrocene (13).

Compound **12** (1.592 g, 3.73 mmol) was dissolved in Et₂O (50 ml) and the solution was cooled to 0°C. After standing for 5 min, diisobutylaluminum hydride (11.2 ml, 11.2 mmol) was added slowly at that temperature. The mixture was stirred at 0°C for 1 hr before the reaction was quenched with saturated sodium potassium tartrate in water (30 ml). The layers were separated and the aqueous layer was further extracted with Et₂O (30 ml). The combined organic fractions were dried over Na₂SO₄, the solvent removed *in vacuo* and the residue purified via flash column chromatography (50% EtOAc in hexane) to yield the product (1.413 g, 99%). ¹H NMR (400 MHz, CDCl₃) δ 4.42 (dd, *J* = 2.4, 1.4 Hz, 1H), 4.17 (td, *J* = 2.6, 0.6 Hz, 1H), 4.13 (s, 5H), 4.06 (dd, *J* = 2.7, 1.3 Hz, 1H), 3.59 (t, *J* = 6.6 Hz, 2H), 2.78 – 2.69 (m, 1H), 1.72 – 1.52 (m, 2H), 1.41 (d, *J* = 7.0 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 95.87 (ipso Cp), 73.76 (Fc), 71.47 (Fc), 67.87 (Fc), 64.18 (Fc), 60.84 (CH₂), 44.73 (ipso Cp), 42.09 (CH₂), 29.81 (CH*), 19.69 (CH₃). MS (ES) (*m/z*) calcd for C₁₄H₁₇O⁵⁶FeI 383.9674, found 383.9678. IR (cm⁻¹): 3282 br (OH), 3088 (=CH Fc), 2971 (CH₂), 2932 (CH₂), 2854 (CH₂), 1556, 1452 (CH₂), 1376 (CH₃), 680 (C=C). Mp: 96°C-98°C.

(S,S_p)-1-[α -Methyl-(3-(tert-butyl)phenylsilyloxy)propyl]-2-iodo-ferrocene (14).

Compound **13** (1.413 g, 3.67 mmol) was dissolved in DCM (20 mL) at room temperature. TEA (0.77 ml, 5.52 mmol), *tert*-butyldiphenylsilyl chloride (1.44 ml, 5.51 mmol) and DMAP (catalytic amount) were added to the mixture. The solution was then stirred overnight at room temperature before quenching with water (10 ml). The phases were separated and the aqueous layer was extracted with Et₂O (2 x 20 ml). The combined organic fractions were dried over Na₂SO₄, the solvent removed *in vacuo* and the residue purified via flash column chromatography (10% EtOAc in hexane) to yield a yellow oily product (2 g, 95%). ¹H NMR (400 MHz, CDCl₃) δ 7.70 – 7.65 (m, 4H), 7.43 – 7.32 (m, 6H), 4.38 (dd, *J* = 2.4, 1.3 Hz, 1H), 4.10 (s, 5H+1H), 4.00 (dd, *J* = 2.7, 1.3 Hz, 1H), 3.70 – 3.65 (m, 2H), 2.77 – 2.68 (m, 1H), 1.88 – 1.80 (m, 1H), 1.43 – 1.34 (m, 1H), 1.31 (d, *J* = 6.9 Hz, 3H), 1.05 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 135.63 (Ph), 134.10 (ipso Ph), 134.05 (ipso Ph), 129.47 (Ph), 127.58 (Ph), 96.25 (ipso Cp), 73.81 (Fc), 71.38 (Fc), 67.61 (Fc), 64.27(Fc), 62.10 (CH₂), 44.45 (ipso Cp), 41.60 (CH₂), 30.03 (CH*), 26.94 (tBu), 19.24 (ipso tBu), 18.91 (CH₃). MS (ES) (*m/z*) calcd for C₃₀H₃₅O⁵⁶FeISiNa 622.0851, found 622.0846. IR (cm⁻¹): 3071 (=CH Fc), 2958 (CH₂), 2929 (CH₂), 2856 (CH₂), 1472 (CH₂), 1387 (CH₃), 1361, 1106, 1085, 821 (CH Ar TBDPS), 700 (C=C).

(S,S_p)-1-[α -Methyl-(3-(tert-butyl)phenylsilyloxy)propyl]-2-formyl-ferrocene (15).

Compound **14** (2.182 g, 3.51 mmol) was dissolved in Et₂O (30 ml), the mixture was cooled to -78°C and *n*-BuLi (2.32 ml, 7.01 mmol) was added. After 30 min, DMF (0.68 ml, 8.76 mmol) was added and the mixture was stirred at -78°C for another 30 min before being allowed to warm to room temperature before quenching with water (20 ml). The phases were separated and the aqueous layer was extracted

with Et₂O (2 x 20 ml). The combined ethereal fractions were dried over Na₂SO₄, the solvent removed *in vacuo* and the residue purified via flash column chromatography (10% EtOAc in hexane) to yield the red oily product (1.686 g, 92%). ¹H NMR (400 MHz, CDCl₃) δ 10.11 (s, 1H), 7.68 – 7.59 (m, 4H), 7.42 – 7.33 (m, 6H), 4.75 (dd, *J* = 2.7, 1.4 Hz, 1H), 4.48 (t, *J* = 2.6 Hz, 1H), 4.43 (dd, *J* = 2.6, 1.4 Hz, 1H), 4.21 (s, 5H), 3.61 (t, *J* = 7.1, 2H), 3.21 – 3.10 (m, 1H), 1.73 – 1.50 (m, 2H), 1.34 (d, *J* = 6.9 Hz, 3H), 1.04 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 193.25 (C=O), 135.55 (Ph), 133.94 (ipso Ph), 133.80 (ipso Ph), 129.55 (Ph), 127.61 (Ph), 99.14 (ipso Cp), 76.31 (ipso Cp), 71.04 (Fc), 70.80 (Fc), 70.03 (Fc), 68.89 (Fc), 61.75 (CH₂), 43.28 (CH₂), 27.90 (CH*), 26.89 (tBu), 22.66 (ipso tBu), 19.17 (CH₃). MS (ES) (*m/z*) calcd for C₃₁H₃₆O₂⁵⁶FeSiNa 547.1732, found 547.1727. IR (cm⁻¹): 3071 (=CH Fc), 2958 (CH₂), 2929 (CH₂), 2856 (CH₂), 1673 (C=O), 1589 (C=N), 1427 (CH₂), 1376 (tBu), 1106 (Si-OR), 1086 (Si-OR), 821 (CH Ar Ph), 700 (C=C).

(S,R_p)-1-[α-Methyl-(3-*tert*-butyldiphenylsilyloxy)propyl]-2-vinyl-ferrocene (16). Trimethylmethylphosphonium bromide (1.722 g, 4.82 mmol), potassium *tert*-butoxide (0.541 g, 4.82 mmol) and a catalytic amount of dibenzo-18-crown-6-ether were dissolved in THF (20 ml). The mixture was stirred for 30 min and then **15** (1.686 g, 3.21 mmol), dissolved in THF (30 ml), was added to the mixture. The reaction mixture was stirred overnight at room temperature, before quenching with water (10 ml) and extracting with Et₂O (2 x 20 ml). The combined ethereal fractions were dried over Na₂SO₄, solvent removed *in vacuo* and purified via flash column chromatography (5% EtOAc in hexane) to yield the product as a yellow oil (1.497 g, 89%). ¹H NMR (400 MHz, CDCl₃) δ 7.69 – 7.63 (m, 4H), 7.44 – 7.33 (m, 6H), 6.62 (dd, *J* = 17.4, 10.8 Hz, 1H), 5.34 (dd, *J* = 17.5, 1.8 Hz, 1H), 5.01 (dd, *J* = 10.8, 1.7 Hz, 1H), 4.43 (dd, *J* = 2.5, 1.4 Hz, 1H), 4.12 (t, *J* = 2.6 Hz, 1H), 4.06 (dd, *J* = 2.5, 1.4 Hz, 1H), 4.03 (s, 5H), 3.62 (dd, *J* = 7.2, 5.4 Hz, 2H), 2.94 – 2.86 (m, 1H), 1.72 – 1.61 (m, 1H), 1.45 – 1.37 (m, 1H), 1.30 (d, *J* = 6.8 Hz, 3H), 1.06 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 135.59 (Ph), 134.08 (ipso Ph), 134.02 (ipso Ph), 133.50 (CH vinyl), 129.49 (Ph), 127.56 (Ph), 110.96 (CH₂ vinyl), 94.84 (ipso Cp), 81.37 (ipso Cp), 69.66 (Fc), 66.55 (Fc), 66.27 (Fc), 64.08 (Fc), 61.89 (CH₂), 42.80 (CH₂), 27.65 (CH*), 26.89 (tBu), 19.23 (ipso tBu), 18.92 (CH₃). MS (ES) (*m/z*) calcd for C₃₂H₃₈O⁵⁶FeSi 522.2041, found 522.2055. IR (cm⁻¹): 3072 (=CH Fc), 2958 (CH₂), 2930 (CH₂), 2857 (CH₂), 1625 (Ar Ph), 1589, 1427 (CH₂), 1388 (CH₃), 1105 (Si-OR), 1086 (Si-OR), 821 (CH Ar), 699 (vinyl/C=C).

(S,R_p)-1-[α-Methyl-(3-*tert*-butyldiphenylsilyloxy)propyl]-[2-(hydroxy)ethyl]-ferrocene (17). Compound **16** (1.497 g, 2.87 mmol) was dissolved in THF (30 ml). BH₃.THF (1M, 8.2 ml, 8.2 mmol) was then added dropwise at room temperature and the mixture stirred for 2 hrs. EtOH (9.76 ml), NaOH (3M, 9.76 ml, 29.28 moles) and H₂O₂ (30 wt. % in water, 7.17 ml, 63.24 moles) were then successively added and the mixture stirred for 1 hr at room temperature. The reaction mixture was extracted with DCM (30 ml), washed with

brine (20 ml) and then dried over Na₂SO₄. The solvent was removed *in vacuo* and the residue purified via flash column chromatography (10% EtOAc in hexane) to yield the yellow oily product (1.434 g, 82%). ¹H NMR (400 MHz, CDCl₃) δ 7.68 – 7.64 (m, 4H), 7.43 – 7.36 (m, 6H), 4.11 – 4.09 (m, 1H), 4.05 (s, 5H), 4.00 (t, *J* = 2.5 Hz, 1H), 3.97 (dd, *J* = 2.5, 1.3 Hz, 1H), 3.75 (tq, *J* = 6.8, 2.6 Hz, 2H), 3.67 – 3.63 (m, 2H), 2.77 – 2.68 (m, 1H), 2.66 – 2.49 (m, 2H), 1.74 – 1.66 (m, 1H), 1.43 – 1.32 (m, 1H), 1.26 (d, *J* = 6.8 Hz, 3H), 1.06 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 135.56 (Ph), 133.96 (ipso Ph), 129.59 (Ph), 127.63 (Ph), 95.01 (ipso Cp), 82.31 (ipso Cp), 69.03 (Fc), 67.36 (Fc), 65.39 (Fc), 65.15 (Fc), 63.00 (CH₂), 61.98 (CH₂), 42.43 (CH₂), 30.93 (CH₂), 27.51 (CH*), 26.91 (tBu), 19.37 (CH₃), 19.22 (ipso tBu). MS (ES) (*m/z*) calcd for C₃₂H₄₀O₂⁵⁶FeSiNa 563.2045, found 563.2039. IR (cm⁻¹): 3378 br (OH), 3072 (=CH Fc), 2930 (CH₂), 2857 (CH₂), 1589, 1472 (CH₃), 1427 (CH₂), 1388 (tBu), 1361, 1105 (Si-OR), 1086 (Si-OR), 819 (CH Ar Ph), 705 (C=C). HPLC: retention time 16.85 min; chiral AD column, 1% IPA in hexane, isocratic over 40 min (1 mL/min), *ee* 97%.

(S,R_p)-1-[α-Methyl-(3-(hydroxy)propyl)]-2-[(thyminy)ethyl]-ferrocene (1). Triphenylphosphine (137 mg, 0.516 mmol), 3-benzoyl thymine¹³ (95 mg, 0.447 mmol) and compound **17** (0.186 g, 0.344 mmol) were dissolved in THF (10 ml) and stirred for 10 min at room temperature. The flask was then covered with foil and DIAD (0.11 ml, 0.516 mmol) was added at room temperature before the mixture was heated at 65°C for 2 hrs. The reaction mixture was evaporated, extracted with EtOAc (30 ml), washed with brine (20 ml) followed by water (20 ml) and dried over Na₂SO₄. The solvent was removed *in vacuo* and the residue purified via flash column chromatography (30% EtOAc in hexane) to give the fully protected product (219 mg, 85%). Deprotection was achieved firstly by stirring the compound in 5 ml of TBAF for 2 hr before the solvent was removed. The mixture was then redissolved in methylamine (33 wt. % in ethanol, 2 ml) and stirred at room temperature for an addition 30 min. The methylamine was evaporated and the crude was purified via flash column chromatography (5% MeOH in DCM) to give the product as a yellow oil (105 mg, 74%). ¹H NMR (400 MHz, CDCl₃) δ 9.44 (s, 1H), 7.07 (d, *J* = 1.2 Hz, 1H), 4.16 – 4.02 (m, 8H), 3.99 – 3.88 (m, 1H), 3.74 – 3.55 (m, 3H), 2.96 – 2.47 (m, 4H), 1.95 (d, *J* = 1.1 Hz, 3H), 1.75 – 1.64 (m, 1H), 1.56 – 1.46 (m, 1H), 1.39 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 164.22 (C=O), 151.11 (C=O), 140.41 (CH-thymine), 111.00 (ipso thymine), 95.36 (ipso Fc), 80.77 (ipso Fc), 69.30 (CH Cp), 67.63 (CH Cp), 65.97 (CH Cp), 65.37 (CH Cp), 60.27 (CH₂), 49.84 (CH₂), 43.24 (CH₂), 27.99 (CH₂), 27.07 (CH), 19.25 (CH₃ thymine), 12.30 (CH₃). MS (ES) (*m/z*) calcd for C₂₁H₂₆N₂O₃Na⁵⁶Fe 433.1191, found 433.1182. IR (cm⁻¹): 3520-3291 br (OH), 2957 (CH), 1677 (C=O).

(S,R_p)-1-[α-Methyl-(3-(hydroxy)propyl)]-2-[2-(adenin-9-yl)ethyl]-ferrocene (2). Triphenylphosphine (0.291 g, 1.109 mmol), *N,N*-6-dibenzoyl-adenine¹⁴ (0.381 g, 1.109 mmol) and compound **17** (0.300 g, 0.555 mmol) were dissolved in THF (10 ml) and stirred for 10 min at room temperature. The flask was then covered with foil and

DIAD (0.24 ml, 1.109 mmol) was added at room temperature before the mixture was warmed up to 65°C for 2 hrs. The reaction mixture was evaporated, extracted with EtOAc (30 ml), washed with brine (20 ml) followed by water (20 ml) and dried over Na₂SO₄. The solvent was removed *in vacuo* and the residue purified via flash column chromatography (30% EtOAc in hexane) to give the protected product (0.343 g, 72%). Deprotection was achieved firstly by stirring the compound in TBAF (5 ml, 1M) for 2 hr. The solvent was then removed and the residue redissolved in methylamine (33 wt. % in ethanol, 2 ml) and stirred at room temperature for an additional 30 min. The methylamine was then evaporated and the crude mixture purified via flash column chromatography to give the product as a yellow solid (170 mg, 73%). ¹H NMR (400 MHz, DMSO-d₆) δ 8.19 (s, 1H), 8.17 (s, 1H), 7.21 (s, 2H), 4.41 – 4.33 (m, 2H+1H (OH)), 4.11 (s, 5H), 4.02 (d, *J* = 2.4 Hz, 2H), 3.99 (t, *J* = 2.4 Hz, 1H), 3.42 – 3.28 (m, 2H), 2.97 – 2.75 (m, 2H), 2.75 – 2.67 (m, 1H), 1.53 – 1.45 (m, 1H), 1.32 (d, *J* = 6.8 Hz, 3H+1H). ¹³C NMR (101 MHz, DMSO-d₆) δ 155.95 (ipso adenine), 152.43 (CH adenine), 149.39 (ipso adenine), 140.71 (CH adenine), 118.75 (ipso adenine), 94.43 (ipso Cp), 81.89 (ipso Cp), 68.80 (Cp), 66.46 (Cp), 64.92 (Cp), 64.74 (Cp), 58.66 (CH₂), 43.10 (CH₂), 42.43 (CH₂), 27.98 (CH₂), 27.10 (CH*), 19.38 (CH₃). MS (ES) (*m/z*) calcd for C₂₁H₂₆N₅O⁵⁶Fe 420.1487, found 420.1484. IR (cm⁻¹): 3348 br (OH), 3270 (NH₂), 3240 (NH₂), 3098 (=CH Fc), 2955 (CH₂), 2926 (CH₂), 2871 (CH₂), 1674 (C=N), 1604 (NH₂), 1574 (NH₂), 1305 (OH), 1076 (C-O), 814 (CH Ar). Mp: 90°C-92°C.

1-[α -Methyl-(3-(hydroxy)propyl)]ferrocene (3). (*S*)-3-Ethoxy-1-methyl-3-oxopropylferrocene¹⁵ (220 mg, 0.733 mmol) was dissolved in diethyl ether (10 ml). LiAlH₄ (56 mg, 1.466 mmol) was added carefully and the resulting suspension was left to stir for 1 hr. The reaction was quenched with saturated sodium potassium tartrate (10 ml), extracted with diethyl ether (2 x 20ml), dried over MgSO₄ and the solvent removed *in vacuo*. The residue was purified via flash column chromatography to give the product as a yellow oil (100 mg, 58%). ¹H NMR (300 MHz, CDCl₃) δ 4.13 (s, 5H), 4.09 – 4.04 (m, 4H), 3.67 (q, *J* = 6.2 Hz, 2H), 2.74 – 2.53 (m, 1H), 1.85 – 1.61 (m, 2H), 1.27 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 95.41 (ipso Cp), 68.51 (CH Cp), 67.22 (CH Cp), 67.12 (CH Cp), 67.07 (CH Cp), 65.70 (CH Cp), 61.12 (CH₂), 41.47 (CH₂), 29.62 (CH), 20.64 (CH₃). MS (ES) (*m/z*) calcd for C₁₄H₁₈O₂⁵⁶Fe 258.0707, found 258.0708. IR (cm⁻¹): 3512-3146 br (OH), 2933 (CH), 1052 (C-O).

1-(Thyminyloethyl)ferrocene (4). Triphenylphosphine (348 mg, 1.30 mmol), *N*-3-benzoylthymine¹³ (223 mg, 1.04 mmol), and 2-ferrocenylethanol¹⁶ (200 mg, 0.87 mmol) were dissolved in THF (10 ml) and stirred for 10 min at room temperature. The flask was then covered with foil and DIAD (0.28 ml, 1.30 mmol) was added at room temperature before the mixture was heated at 65°C for 2 hrs. The solvent was then evaporated and the residue extracted with EtOAc (30 ml), washed with brine (20 ml) and water (20 ml) and dried over Na₂SO₄, before the solvent was removed *in vacuo*. Deprotection was achieved by treating the crude mixture with methylamine solution (33 wt. % in ethanol, 5

ml) for 30 min. The solvent was then evaporated *in vacuo* and the residue purified via flash column chromatography (40% EtOAc in hexane) to give the product (176 mg, 60%). ¹H NMR (300 MHz, CDCl₃) δ 8.29 (s, 1H), 6.70 (d, *J* = 1.2 Hz, 1H), 4.15 (s, 5H), 4.13 – 4.09 (m, 2H), 4.04 (t, *J* = 1.8 Hz, 2H), 3.79 (t, *J* = 7.1 Hz, 2H), 2.73 (t, *J* = 7.1 Hz, 2H), 1.85 (d, *J* = 1.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃, trace of MeOD) δ 216.94 (ipso thymine), 193.73 (C=O), 181.71 (C=O), 141.17 (CH-thymine), 83.57 (ipso-Cp), 68.63(CH-Cp), 68.37(CH-Cp), 67.94 (CH-Cp), 50.20 (CH₂), 29.17 (CH₂), 11.98 (CH₃). MS (ES) (*m/z*) calcd for C₁₇H₁₈N₂O₂⁵⁶Fe 338.0718, found 338.0720. Mp: degraded at 235°C. IR (cm⁻¹): 3146 (NH), 2999 (CH), 1683 (C=O), 1644 (NH bending).

1-[2-(Adenin-9-yl)ethyl]-ferrocene (5). Triphenylphosphine (0.383 g, 1.46 mmol), *N,N*-6,6-dibenzoyl-adenine¹⁴ (0.500 g, 1.46 mmol) and 2-ferrocenylethanol¹⁶ (0.201 g, 0.73 mmol) were dissolved in THF (10 ml) and stirred for 10 min at room temperature. The flask was then covered with foil and DIAD (0.24 ml, 1.18 mmol) was added at room temperature before the mixture was warmed up to 65°C for 2 hrs. The reaction mixture was evaporated, extracted with EtOAc (30 ml), washed with brine (20 ml) followed by water (20 ml) and then dried over Na₂SO₄. The solvent was removed *in vacuo* and purified via flash column chromatography (40% EtOAc in hexane) to give the bis-protected product (0.134 g, 33%). Deprotection was achieved by dissolving the compound (0.055 g, 0.1 mmol) in methylamine (33 wt. % in ethanol, 3 ml) and stirring at room temperature for 30 min. The methylamine was then evaporated and the residue purified via flash column chromatography (95/5 DCM/MeOH) to give the product as a yellow solid (0.019 g, 58%). ¹H NMR (400 MHz, DMSO-d₆) δ 8.17 (s, 1H), 8.05 (s, 1H), 7.17 (s, 2H), 4.31 (dd, *J* = 8.2, 6.8 Hz, 2H), 4.15 (s, 5H), 4.08 – 4.02 (m, 4H), 2.85 (t, *J* = 7.5 Hz, 2H). ¹³C NMR (101 MHz, DMSO-d₆) δ 155.89 (ipso adenine), 152.35 (CH adenine), 149.40 (ipso adenine), 140.76 (CH adenine), 118.73 (ipso adenine), 84.39 (ipso Cp), 69.00 (Fc), 68.55 (Fc), 43.75 (CH₂), 29.34 (CH₂). MS (ES) (*m/z*) calcd for C₁₇H₁₈N₅⁵⁶Fe 348.0912, found 348.0920 (M⁺ + H⁺). IR (cm⁻¹): 3399 (NH₂), 3316 (NH₂), 3084 (=CH Fc), 2980 (CH₂), 2931 (CH₂), 2907 (CH₂), 1653 (C=C), 1596 (NH₂), 1435 (CH₂), 1245, 797 (CH Ar). Mp: 142°C (decomp).

ASSOCIATED CONTENT

Supporting Information. ¹H NMR spectra and ¹³C NMR spectra for compounds **1-5** and **8-17**, HPLC data for compounds **1-6** and **17**. Cell study procedures, cell growth data for **2**, **4** and **5** on an oesophageal cancer cell line. This material is available free of charge via the Internet at <http://pubs.acs.org>.

AUTHOR INFORMATION

Corresponding Author

*Email: j.tucker@bham.ac.uk. Phone: +44 (0)121 414 4422.

ACKNOWLEDGMENT

JHRT acknowledges financial support from the EPSRC (Leadership Fellowship, EP/G007578/1), an ISSF Grant from the Wellcome Trust and a grant from KU Leuven (GOA 10/014).

ABBREVIATIONS

DCM, dichloromethane; DIAD, diisopropyl azodicarboxylate; DMSO, dimethyl sulfoxide; TEA, triethylamine; DMAP, 4-dimethylaminopyridine; *n*-BuLi, *n*-butyllithium; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; BrdU, bromodeoxyuridine; THF, tetrahydrofuran

REFERENCES

- (1) (a) Galmarini, C. M.; Mackey, J. R.; Dumontet, C. Nucleoside analogues and nucleobases in cancer treatment. *Lancet Oncol.* **2002**, *3*, 415. (b) Cihlar T.; Ray A. S. Nucleoside and nucleotide HIV reverse transcriptase inhibitors: 25 years after zidovudine. *Antiviral Res.* **2010**, *85*, 39.
- (2) (a) Romeo, G.; Chiacchio, U.; Corsaro, A.; Merino, P. *Chem. Rev.* **2010**, *110*, 3337-3370. (b), Mathé, C.; Périgaud, C. Recent approaches in the synthesis of conformationally restricted nucleoside analogues. *Eur. J. Org. Chem.* **2008**, 1489.
- (3) (a) van Staveren, D. R.; Metzler-Nolte, N. Bioorganometallic chemistry of ferrocene. *Chem. Rev.* **2004**, *104*, 5931-5985. (b) Fouda, M. F. R.; Abd-Elzaher, M. M.; Abdelsamaia, R. A.; Labib, A. A. On the medicinal chemistry of ferrocene. *Appl. Organomet. Chem.* **2007**, *21*, 613-625. (c) Gasser, G.; Ott, I.; Metzler-Nolte, N. Organometallic anticancer compounds. *J. Med. Chem.* **2011**, *54*, 3-25. (d) Hillard, E. A.; Jaouen, G. Bioorganometallics: Future trends in drug discovery, analytical chemistry, and catalysis. *Organometallics.* **2011**, *30*, 20-27.
- (4) (a) Dive, D.; Biot, C. Ferrocene conjugates of chloroquine and other antimalarials: the development of ferroquine, a new antimalarial. *ChemMedChem.* **2008**, *3*, 383-391. (b) Cázares-Marinero, J. J.; Top, S.; Vessières A., Jaouen, G. Synthesis and antiproliferative activity of hydroxyferrocifen hybrids against triple-negative breast cancer cells. *Dalton Trans.*, **2014**, *43*, 817 and references therein.
- (5) (a) For a recent review, see: Ornelas, C.; Application of ferrocene and its derivatives in cancer research. *New J. Chem.*, **2011**, *35*, 1973-1985. (b) Mooney, A.; Corry, A. J.; Ruairc, N.; Mahgoub, T.; O'Sullivan, D.; O'Donovan, N.; Crown, J.; Varughese, S.; Draper, S. M.; Rai, D. K.; Kenny, P. T. M. Synthesis, characterization and biological evaluation of N-(ferrocenyl)naphthoyle amino acid esters as anticancer agents. *Dalton Trans.* **2010**, *39*, 8228-8239. (c) Schobert, R.; Seibt, S.; Mahal, K.; Ahmad, A.; Biersack, B.; Effenberger-Neidnicht, K.; Padhye, S.; Sarkar, F. H.; Mueller, T. Cancer selective metallocenedicarboxylates of the fungal cytotoxin illudin M. *J. Med. Chem.* **2011**, *54*, 6177-6182. (d) Librizzi, M.; Longo, A.; Chiarelli, R.; Amin, J.; Spencer, J.; Luparello, C. Cytotoxic effects of jay amin hydroxamic acid (JAHA), a ferrocene-based class I histone deacetylase inhibitor, on triple-negative MDA-MB231 breast cancer cells. *Chem. Res. Toxicol.* **2012**, *25*, 2608-2616.
- (6) (a) Edwards, E. I.; Epton, R.; Marr, G. The synthesis and reactions of homonuclear ferrocene acid anhydrides and their Use in the preparation of ferrocenylpenicillins and cephalosporins. *J. Organomet. Chem.* **1979**, *168*, 259-272. (b) Cohan, Z. H. Antibacterial and antifungal ferrocene incorporated dithiothione and dithioketone compounds. *Appl. Organomet. Chem.* **2006**, *20*, 112-116. (c) Patra, M.; Gasser, G.; Wenzel, M.; Merz, K.; Bandow, J. E.; Metzler-Nolte, N. Synthesis and biological evaluation of ferrocene-containing bioorganometallics inspired by the antibiotic platensimycin lead structure. *Organometallics.* **2010**, *29*, 4312-4219. (d) Zaheer, M.; Shah, A.; Akhter, Z.; Qureshi, R.; Mirza, B.; Tauseef, M.; Bolte, M. Synthesis, characterization, electrochemistry and evaluation of biological activities of some ferrocenyl schiff bases. *Appl. Organomet. Chem.* **2011**, *25*, 61-69.
- (7) (a) Kowalski, K.; Koceva-Chyla, A.; Pieniazek, A.; Bernasinska, J.; Skiba, J.; Rybarczyk-Pirek, J. A.; Józwiak, Z. The synthesis, structure, electrochemistry and in vitro anticancer activity studies of ferrocenyl-thymine conjugates. *J. Organomet. Chem.* **2012**, *700*, 58-68. (b) Simenela, A. A.; Morozovaa, A. E.; Snegura, V. L.; Zykovaa, I. S.; Kachalab, V. V.; Ostrovskayac, A. L.; Bluchterovac, V. N.; Fomina, M. M. Simple route to ferrocenylalkyl nucleobases. Antitumor activity in vivo. *Appl. Organomet. Chem.* **2009**, *23*, 219-224. (c) Kowalski, K.; Skiba, J.; Oehninger, L.; Ott, I.; Solecka, J.; Rajnisz, A.; Therrien, B. Metallocene-modified uracils: synthesis, structure, and biological activity. *Organometallics.* **2013**, *32*, 5766-5733. (d) James, P.; Neudörfl, J.; Eissmann, M.; Jesse, P.; Prokop, A.; Schmalz, H. -G. Enantioselective synthesis of ferrocenyl nucleoside analogues with apoptosis-inducing activity. *Org. Lett.*, **2006**, *8*, 2763-2766.
- (8) Shago, F. R.; Swarts, C. J.; Kreft, E.; Van Rensburg, E. J. C. Antineoplastic activity of a series of ferrocene-containing alcohols. *Anticancer Res.* **2007**, *27*, 3431-3434.
- (9) A ferrocene derivative containing a nucleobase and a hydroxyl group (but not bridged solely by a ferrocene group) was reported previously, which did not show apoptosis-inducing activity against tumor cells (see Reference 7d).
- (10) (a) Nguyen, H. V.; Sallustrau, A.; Male, L.; Thornton, P. J.; Tucker, J. H. R. 1,1'-Homodisubstituted ferrocenes containing adenine and thymine nucleobases: Synthesis, electrochemistry, and formation of H-bonded arrays. *Organometallics* **2011**, *30*, 5284-5290. (b) Nguyen, H. V.; Zhao, Z. Y.; Sallustrau, A.; Horswell, S. L. Male, L.; Mulas, A.; Tucker, J. H. R. A ferrocene nucleic acid oligomer as an organometallic structural mimic of DNA. *Chem. Comm.* **2012**, *48*, 12165.
- (11) Marquarding, D.; Klusacek, H.; Gokel, G.; Hoffmann, P.; Ugi, I. Correlation of central and planar chirality in ferrocene derivatives. *J. Am. Chem. Soc.* **1970**, *92*, 5389-5393.
- (12) Initial cell line studies indicate that compound **1** also displays antiviral activity (Patent application - GB1322752.5). Further studies are underway to establish mechanistic pathways (e.g. phosphorylation).
- (13) Cruickshank, K. A.; Jiricny, J.; Reese, C. B. The benzylation of uracil and thymine. *Tetrahedron. Lett.* **1984**, *25*, 681-684.
- (14) Sallustrau, A. Synthesis of ferrocenyl derivatives as novel nucleic acid monomers. PhD Thesis, **2013**, University of Birmingham.
- (15) Locke, A. J.; Richards, C. J. Asymmetric synthesis of [3](1,1')- and [3](1,1')[3](3,3')-ferrocenophanes. *Organometallics.* **1999**, *18*, 3750-3759.
- (16) Barry, K. P.; Nataro, C. A new synthesis and electrochemistry of 1,1'-bis(β-hydroxyethyl)ferrocene. *Inorg. Chim. Acta.* **2009**, *362*, 2068-2070.

(TOC)

