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Expression of SOAT1 in adrenocortical carcinoma and response to mitotane monotherapy

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1 **Expression of sterol-O-acyl transferase 1 (SOAT1) in adrenocortical carcinoma**
2 **and response to mitotane monotherapy: an ENSAT multicenter study**

3

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51

52 **Precis** (max 200 Zeichen)

53 Mitotane is a cornerstone of adrenal cancer treatment. In this international study, expression
54 of putative mitotane target SOAT1 in tissue did not predict treatment response to mitotane
55 monotherapy.

56

57

58 **Abstract (max 250 words)**

59 **Context** Objective response rate to mitotane in advanced adrenocortical carcinoma (ACC) is
60 approximately 20% and adverse drug effects are frequent. To date there is no marker
61 established that predicts treatment response. Mitotane has been shown to inhibit sterol-O-
62 acyl transferase 1 (SOAT1) which leads to endoplasmic reticulum stress and cell death in
63 ACC cells.

64 **Objective** To investigate SOAT1 protein expression as a marker of treatment response to
65 mitotane.

66 **Patients** 231 ACC patients treated with single agent mitotane as adjuvant (n=158) or
67 advanced disease therapy (n=73) from twelve ENSAT centers were included. SOAT1 protein
68 expression was determined by immunohistochemistry on formalin-fixed paraffin-embedded
69 (FFPE) specimens.

70 **Main outcome measure:** recurrence-free survival (RFS), progression-free survival (PFS),
71 disease-specific survival (DSS)

72 **Results** 61/135 patients (45 %) with adjuvant mitotane treatment had recurrences and 45/68
73 patients (66 %) with mitotane treatment for advanced disease had progressive disease. After
74 multivariate adjustment for sex, age, hormone secretion, tumour stage and Ki67 index, RFS
75 (HR=1.07, 95% CI 0.61-1.85, p=0.82) and DSS (HR=1.30, 95% CI 0.58-2.93, p=0.53) in
76 adjuvantly treated ACC patients did not differ significantly between tumors with high and low
77 SOAT1 expression. Similarly, in the advanced stage setting, PFS (HR=1.34, 95% CI 0.63-
78 2.84, p=0.45) and DSS (HR=0.72, 95% CI 0.31-1.70, p=0.45) were comparable and
79 response rates not significantly different.

80 **Conclusions** SOAT1 expression was not correlated with clinical endpoints RFS, PFS and
81 DSS in ACC patients with mitotane monotherapy. Other factors appear to be relevant for
82 mitotane treatment response and ACC patient survival.

83

84 **Introduction**

85 Adrenocortical carcinoma (ACC) is a rare malignancy with a generally poor prognosis
86 [1] and limited effective treatment options [1, 2]. Mitotane is the only approved drug
87 for metastatic disease [3] but efficacy is very limited and the observed objective
88 response rate is only approximately 20 % [4-6]. Controversy exists regarding
89 adjuvant use which is supported by a large retrospective study [6, 7] and advocated
90 by current guidelines [2] in patients at moderate or high risk of recurrence after
91 complete resection. Adverse drug effects like adrenal insufficiency, diarrhea, nausea
92 and other gastrointestinal symptoms but also central nervous symptoms such as
93 dizziness and speech disturbance may be severe and disabling [8-10] and must be
94 balanced against potential treatment benefits. Mitotane efficacy is correlated with
95 plasma concentrations above 14 mg/l [11]. Therapeutic drug monitoring (TDM) is
96 therefore recommended [2]. Some patients for unknown reasons fail to achieve
97 mitotane plasma concentrations within the therapeutic window which is associated
98 with decreased efficacy [5, 12]. To date, few markers have been suggested for the
99 prediction of response [13, 14], but they have not been validated in a large series.
100 Establishment of such a marker would be a major advancement in ACC treatment
101 and enable tailored treatment of potential responders and avoidance of unnecessary
102 mitotane exposure in non-responders.

103 We have provided evidence that mitotane inhibits sterol-O-acyl transferase 1
104 (SOAT1) also known as ACAT1 [15] (not to be mistaken with acetyl-CoA
105 acetyltransferase known under the same name), an enzyme catalyzing the
106 esterification of cholesterol in the adrenal cortex [16]. This leads to the accumulation
107 of toxic lipids and endoplasmic reticulum (ER) stress which results in apoptosis of
108 adrenocortical cells [15]. Accordingly, a SOAT1 inhibitor has been tested in a phase I
109 clinical trial as a treatment for advanced ACC [17].

110 SOAT1 is strongly expressed in adrenocortical cell lines, normal adrenal glands and
111 different adrenocortical tumor entities, with the highest variation among ACC, while it
112 is only weakly to moderately expressed in non-adrenal tissues [15]. Despite strong
113 evidence of an inhibitory effect on SOAT1, other mechanisms such as impaired
114 mitochondrial respiration and function [18-20] may contribute to the relatively tissue-
115 specific toxicity of mitotane.

116 In a small cohort of patients with advanced ACC, it has been shown that SOAT1
117 expression was correlated with the response to mitotane treatment [15].

118 Here, we aimed to validate in a large multicenter study whether SOAT1 expression is
119 a predictive marker for mitotane efficacy by investigating the association of SOAT1
120 tissue expression with recurrence free survival (RFS) in patients with adjuvant
121 mitotane treatment, progression-free survival (PFS) after mitotane monotherapy
122 administered to patients with advanced disease and disease-specific survival (DSS)
123 for both cases.

124 **Patients and Methods**

125 *Setting and data acquisition*

126 Formalin-fixed paraffin-embedded (FFPE) tumor specimens of 231 ACC were included from
127 12 centers belonging to the European Network for the Study of Adrenocortical Tumors
128 (ENSAT; www.ensat.org). Only adult patients with histologically confirmed ACC were
129 included [21]. Patients that have been included in our previous analyses of SOAT1
130 expression [15] have been excluded from this analysis. All patients started mitotane
131 treatment as first medical therapy no later than 3 months after complete resection in the
132 adjuvant setting (n=158) or diagnosis of irresectable or recurrent or metastatic ACC in the
133 advanced stage setting (n=73). The study was conducted as part of the ENSAT registry, has
134 been approved by the ethics committee at each participating institution and was conducted in
135 accordance with the principles of the Declaration of Helsinki. All patients gave informed
136 written consent.

137 Clinical and pathological data, including sex, age at diagnosis, date of diagnosis, tumor stage
138 according to the ENSAT staging system [22], hormone secretion, Weiss score [21], Ki67
139 proliferation index, mitotane plasma concentrations after three and six months and response
140 to treatment during follow-up were either provided by the participant center or collected
141 through the ENSAT registry (<https://registry.ensat.org>).

142 *Chromogenic immunohistochemistry*

143 Full FFPE sections mounted on slides were deparaffinised, rehydrated and antigen retrieval
144 was performed in 10mM citric acid monohydrate buffer (pH 6.5) under pressure for 13 min.
145 Blocking of unspecific binding sites occurred with 20% human AB serum at room
146 temperature (RT) for 1 h and the primary antibody (SOAT1; ab39327 Abcam) was incubated
147 in a 1:1000 dilution for 1h at RT as previously described [15]. The N-Universal negative
148 control anti-rabbit (Dako) was used and signal amplification was achieved by the Advance
149 HRP Link Kit for 40 min and developed for 10 min with the DAB+ Liquid Kit (Dako). Nuclei

150 were counterstained using Mayer's hematoxylin for 3 min and blued for 5 min in running tap
151 water. To ensure specificity of the antibody used [23], we overexpressed human SOAT1 in
152 ACC cells which resulted in an increase of both detected SOAT1 bands and SOAT1 WB of 5
153 normal adrenal glands also resulted only in the two specific bands (Fig. S1).

154 *Semi-quantitative analysis of SOAT1 immunoreactivity*

155 Chromogenic staining intensities were determined by two independent investigators (I.W.
156 and B.A. or L.-S.L.) and graded as 0 (negative), 1 (low), 2 (medium) and 3 (high). The
157 proportion of positive tumor cells was calculated for each slide and scored 0 if 0% were
158 positive, 0.1 if 1-9% were positive, 0.5 if 10-49% were positive and 1 if $\geq 50\%$ were positive
159 [24, 25]. A semi quantitative H-Score was then calculated by multiplying the staining intensity
160 grading score with the proportion score. Where discrepancies were observed, results were
161 jointly assessed by both investigators and the final score was formed by consensus. The
162 Spearman's correlation for inter-observer agreement for each staining was high ($r > 0.85$).

163 *Statistical analysis*

164 RFS and PFS were considered as the time between diagnosis and documented recurrence
165 and progression (based on cross sectional imaging), respectively. DSS was calculated from
166 the time of diagnosis until disease-related death or censored at last follow-up. RFS, PFS and
167 DSS were analysed using the Kaplan–Meier method and groups were compared by using
168 the log-rank test. Assessment of prognostic factors (ENSAT stage, ki67, age, sex, hormone
169 secretion and for the group with advanced disease additionally: preM-TTP (pre mitotane time
170 to progression= time between diagnosis and progress before initiation of mitotane treatment)
171 was performed with the Cox proportional hazard regression model. The Chi-square test was
172 used to investigate dichotomic variables, whereas non-parametric Kruskal-Wallis s test was
173 used for comparison among groups for non-normal distributed variables. Correlations
174 between H-Score and prognostic factors were evaluated by Spearman's correlation. P
175 values < 0.05 were considered statistically significant. Statistical analyses were performed
176 with IBM SPSS Version 23 and GraphPad Prism Version 6.

177 **Results**

178 *Patient characteristics*

179 Clinical characteristics of 231 ACC patients are summarised in Table 1. Median age at
180 diagnosis was 54.2 years (range 17-83) in the adjuvant group and 51 years (range 16-80) in
181 the group with disease. In both groups, approximately 60% of the patients were female and
182 40% were male. At diagnosis, the majority of patients treated with mitotane monotherapy in
183 the adjuvant setting had an ENSAT tumor stage of I-II (62.3%), whereas, in the advanced
184 stage setting, most of the patients had a tumor stage of IV (55.6%). The remaining patients
185 with advanced disease had a localized tumor at diagnosis and started mitotane therapy only

186 after developing local recurrence or metastases. Data regarding Ki67 index were available in
187 91.2% and 83.5% of patients in the adjuvant and advanced stage setting, respectively. 31
188 patients (21%) of the adjuvant group and 18 patients (27.3%) of the advanced stage group
189 had Ki67 index staining below 10% ($p=0.35$, chi-square=0.88). Median Weiss score was 6
190 (range 1-9) in both groups. In both arms, about 70% of the tumors were hormonally active.
191 Median time to start mitotane were one month in the adjuvant group and less than one month
192 in the group with advanced disease. Median mitotane plasma levels at three months of
193 therapy were 9.3 mg/l and 10 mg/l, after six months 13.5 mg/l and 12.8 mg/l in the adjuvant
194 and advanced stage cohort, respectively. In the advanced stage group, preM-TTP was <365
195 days in 51/63 patients (81%) for DSS and <365 days in 52/67 patients (78%) for PFS.

196 No recurrence was observed in 74/135 patients within a median follow-up of 18.5 months
197 (range 1-216 months) in patients treated in adjuvant setting. Best response to advanced
198 stage mitotane was complete (n=1) or partial response in 9, stable disease in 13 and
199 progressive disease in 45 patients. Median follow up of patients still alive (n=18) was 19.5
200 months (range 2-180 months) in this setting.

201 *SOAT1 expression and correlation with known prognostic factors of ACC*

202 Tissue SOAT1 expression differed widely in tumors of both the adjuvant and the group with
203 advanced disease and exhibited different intra-tumoral patterns between homogeneous and
204 heterogeneous staining intensity (Fig. 1). Semiquantitative H-score accounts for this
205 heterogeneity as it takes into account both the staining intensity and percentage of cells
206 being stained and ranged from 0 to 3. Scores from 0 to <2 were designated low expression
207 (Fig. 1J-L) while scores ≥ 2 were indicative of high expression (Fig. 1A-I). No difference in
208 SOAT1 expression was found between hormone producing and endocrine inactive ACC with
209 mean staining intensities of 1.53 ± 0.9 in inactive vs. 1.48 ± 0.9 in hormonally active ACC,
210 $p=0.76$. No correlation of SOAT1 H-score was observed with Ki67, ENSAT stage, Weiss
211 score and age at diagnosis neither in the adjuvant, nor in the advanced stage setting.

212 *SOAT1 expression as factor of survival and response to mitotane treatment in ACC*

213 In the adjuvant setting (Fig. 2A), we did not observe significant differences of RFS between
214 ACC patients with low SOAT1 expression in comparison to those with high SOAT1
215 expression (median 22 months, range 1-153 vs. median 12 months, range 1.5-216 log rank
216 $p=0.12$). When we only included patients with $Ki67 \geq 10\%$ to analyse RFS, we did not observe
217 significant differences between SOAT1 low and high expressing ACC either (log rank
218 $p=0.73$). DSS (Fig. 2B) did not significantly differ between patients whose tumors expressed
219 low levels of SOAT1 compared to those with high SOAT1 expression (median 51 months,
220 range 1-252 vs. 31 months, range 2-216 log rank $p=0.23$). Similarly, in the group with
221 advanced disease, no significant difference in PFS (Fig. 2C) between patients with low

222 SOAT1 expression and those with high SOAT1 expression (median PFS 5 months, range 1-
223 59 vs. median 4 months, range 1-25 log rank $p=0.66$) was observed. Median DSS (Fig. 2D)
224 was likewise not different in tumors with low vs. high SOAT1 (median 22 months, range 4-
225 180 vs. 21 months, 2-83 months, log rank $p=0.47$). When we analysed all patients together
226 (Fig. S2A), low SOAT1 expression was associated with a significantly longer median
227 recurrence-/progression-free survival of 13 months (range 1 -153 months vs 8 months (range
228 1-216 months, log rank $p=0.049$). We did not observe a significant difference in DSS (Fig.
229 S2B) between tumors with low SOAT1 vs high SOAT1 expression (median: 41 months,
230 range 1 -252 vs. median: 28 months, range 2-216, log rank $p=0.41$).

231 The proportion of tumors with low and high SOAT1 expression did not differ between patients
232 in the adjuvant cohort without recurrence (low, $n=44$; high, $n=30$) and with recurrence (low,
233 $n=35$; high, $n=26$) (Fig. 3A). Similarly, in the cohort with advanced disease, there were no
234 differences between tumors with low and high SOAT1 regarding objective response to
235 mitotane (low, $n=6$; high, $n=4$) vs. stable disease (low, $n=6$; high, $n=7$) and progressive
236 disease (low, $n=25$; high, $n=20$), respectively (Fig. 3B).

237 We next aimed at multivariable adjustment for known clinical/histopathological ACC
238 prognostic factors. In the adjuvant arm, univariate analysis revealed only a Ki67-Index $<10\%$
239 as significantly associated with improved DSS and RFS (Table 2). In patients with advanced
240 disease the following factors were significantly associated with improved DSS: male sex,
241 Ki67-Index $<10\%$ and preM-TTP >365 days. After multivariate analysis of all factors,
242 including SOAT1 expression, only preM-TTP >365 days retained statistical significance
243 (Table 3).

244 **SOAT1 expression is not related to mitotane plasma concentrations**

245 We next examined the potential association of SOAT1 expression with mitotane plasma
246 concentrations. Mitotane plasma levels after three months of treatment did not significantly
247 differ between patients whose tumors showed high vs low expression of SOAT1 both in the
248 adjuvant (median mitotane levels: 10.3 mg/l vs 9.1 mg/l) and in the advanced disease setting
249 (median mitotane levels: 11.7 mg/l vs 9.1 mg/l) (Fig. 4A). SOAT1 expression was not
250 associated with mitotane plasma concentrations above 14 mg/l neither in the adjuvant (Fig.
251 4B) nor in the advanced disease arm (Fig. 4C). Similar results were observed after six
252 months of mitotane treatment (median mitotane levels 14.2 mg/l vs 13 mg/l in the adjuvant
253 group and 11.9 mg/l vs 12.8 mg/l in the group with advanced disease). When analyzing only
254 patients reaching the mitotane target level of 14 mg/l after three months, significantly fewer
255 patients with high SOAT1 expression responded to therapy (Fig. 4D) while this difference
256 was no longer observed when considering the six months time point (Fig. 4E).

257 Median dose of mitotane intake was 4 g/daily (range 1-12 g) in the adjuvant arm and 5
258 g/daily (range 2-12 g/daily) in patients treated for advanced disease and did not significantly
259 differ between the SOAT1 high and low expressing group ($p= 0.6$ (adjuvant) and $p=0.4$
260 (advanced disease)).

261 **Discussion**

262 Mitotane is the only approved drug for the treatment of ACC, however, objective response
263 rates are only approximately 20% [5, 6]. In addition to its limited therapeutic potential,
264 adverse events occur frequently and reliable markers predicting response to therapy are
265 currently not established. Therefore, it is crucial to define a particular subgroup of patients
266 that will take advantage from treatment and to avoid toxicity in patients unlikely to respond.

267 At present, this topic has been addressed only in a limited number of patients [13, 14] and
268 very recently a study demonstrated mitotane sensitivity only in a very specific sub-group of
269 patients [26]. Although mitotane has been used in the clinic for decades, its precise
270 mechanism of action and molecular target remained unknown for decades, despite intense
271 research including several different “omics” approaches [18-20, 27]. We demonstrated that
272 mitotane inhibits SOAT1, leading to ER-stress and cell death of adrenocortical cells [15]. It
273 was also shown that SOAT1 is predominantly expressed in adrenocortical cells, compared to
274 cells of non-adrenal origin [15], possibly explaining the specific adrenolytic toxicity of
275 mitotane. In addition, in glioblastoma, inhibition of SOAT1 has been proposed as a novel
276 treatment [28, 29].

277 In hepatocellular carcinoma high SOAT1 expression was associated with a worse prognosis
278 [30] and has previously been described in prostate cancer as well [31]. An adverse outcome
279 of SOAT1 expression in ACC was recently demonstrated [32]. These results suggest that the
280 elevated expression of SOAT1 could be a prognostic feature of diverse cancers. In a small
281 single center series of patients (n=25) with advanced ACC [15], we had previously shown
282 that SOAT1 expression is associated with improved progression-free survival. This ENSAT
283 multicenter retrospective study aimed at validating the value of SOAT1 as a histologic marker
284 for mitotane response. Our results disprove our initial hypothesis, as no significant
285 differences in response to mitotane treatment could be observed between ACC tissue
286 samples with high and low levels of SOAT1 protein neither in an adjuvant setting nor in
287 patients treated with advanced disease.

288 Our study has the strength of a large collection of tissue samples from specialized ACC
289 centers. SOAT1 expression was histologically determined in a centralized manner. All
290 patients received mitotane monotherapy, no additional therapies were used during mitotane
291 treatment. However, our study has several limitations. First, the clinical data and samples

292 collection were retrospectively retrieved from twelve different ENSAT centers (11 European
293 and one from Brazil) which likely is associated with different treatment strategies. This not
294 only comprises surgery and medical treatment but also documentation and follow-up.
295 Second, mitotane treatment itself is cumbersome and different dosing regimens are in use at
296 different centers [33-35]. In addition, patient-specific factors that are only partially understood
297 lead to a high heterogeneity of mitotane plasma concentrations [36-38]. Accordingly,
298 mitotane plasma concentrations in our cohort after three and six months of treatment were
299 highly variable. When considering only patients who reached mitotane plasma
300 concentrations of >14 mg/L at three or six months, SOAT1 expression was not correlated to
301 clinical response.

302 The lack of an association of SOAT1 expression with survival endpoints and response
303 implicates that additional target molecules different from SOAT1 may be relevant for its toxic
304 effect in adrenal cortical cells. *In vitro*, SOAT1 expression was shown to not be a predictor as
305 demonstrated in few ACC primary cultures [23] which would support the theory that
306 additional targets might be of greater importance. One such potential mechanism includes
307 inhibition of mitochondrial respiratory chain. The novel compound nevanimibe (previously
308 known as ATR101) which has been developed as a new treatment for ACC has been shown
309 to be a potent SOAT1 inhibitor by one group [39] but was also shown to inhibit mitochondrial
310 respiration by a different group [40] similar to mitotane.

311 Importantly, we found pronounced heterogeneity of SOAT1 expression in approximately 20
312 % of tumor samples. It is conceivable that this tissue heterogeneity was not completely
313 accounted for in the monocentric study by Ferreira Lacombe *et al.* [32] in which a tissue
314 microarrays were used to evaluate SOAT1 expression whereas we used full sections.
315 Relationship of SOAT1 with Ki67 index and cortisol secretion was demonstrated in the
316 previous study but not in ours. However, in our study ki67 value was provided by the various
317 participating centers and thus a uniform analysis of this index is not guaranteed.

318 In an adjuvant setting, several other known factors such as resection status or Ki67 index
319 [41], are important to predict tumor recurrence, since even after complete resection,
320 recurrence rates are high [42-44]. In line with previous studies, Ki67-index below 10% (Table
321 2) was significantly associated with a better DSS and TTP in our cohort of patients treated
322 with mitotane in this setting. Similarly, in advanced ACC, Ki67 index, mutational burden [45]
323 but also clinical factors like age or presence of symptoms, have been identified [46, 47] to
324 predict patient outcome independently of mitotane treatment [48]. In our cohort of patients
325 with advanced disease, mitotane monotherapy, Ki67-Index below 10% was also associated
326 with a better DSS (Table 3), which retained significance after multivariate adjustment but was
327 not observed for TTP in a univariate analysis (Table 3). This may be due to the relatively

328 small cohort but is in line with a previous study in which only the DSS, but not the TTP
329 correlated with a Ki67-Index below 10% in advanced ACC [5].

330 In conclusion, in this multicenter study, we could not confirm SOAT1 expression to be a
331 clinically useful marker to predict treatment response to mitotane.

332

333 **Contributors:**

334 The following scientists contributed tissue samples and clinical data in addition to those listed
335 as co-authors:

336 Marcus Quinkler (Berlin), Masanori Murakami (Munich), Felix Beuschlein (Munich, Zurich),
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343 **References**

- 344 1. Fassnacht, M., M. Kroiss, and B. Allolio, *Update in adrenocortical carcinoma*. J Clin Endocrinol
345 Metab, 2013. **98**(12): p. 4551-64.
- 346 2. Fassnacht, M., et al., *European Society of Endocrinology Clinical Practice Guidelines on the*
347 *Management of Adrenocortical Carcinoma in Adults, in collaboration with the European*
348 *Network for the Study of Adrenal Tumors*. Eur J Endocrinol, 2018.
- 349 3. Schteingart, D.E., et al., *Management of patients with adrenal cancer: recommendations of*
350 *an international consensus conference*. Endocr Relat Cancer, 2005. **12**(3): p. 667-80.
- 351 4. Reidy-Lagunes, D.L., et al., *Complete Responses to Mitotane in Metastatic Adrenocortical*
352 *Carcinoma-A New Look at an Old Drug*. Oncologist, 2017. **22**(9): p. 1102-1106.
- 353 5. Megerle, F., et al., *Mitotane Monotherapy in Patients With Advanced Adrenocortical*
354 *Carcinoma*. J Clin Endocrinol Metab, 2018. **103**(4): p. 1686-1695.
- 355 6. Terzolo, M., et al., *Adjuvant mitotane treatment for adrenocortical carcinoma*. N Engl J Med,
356 2007. **356**(23): p. 2372-80.
- 357 7. Berruti, A., et al., *Long-Term Outcomes of Adjuvant Mitotane Therapy in Patients With*
358 *Radically Resected Adrenocortical Carcinoma*. J Clin Endocrinol Metab, 2017. **102**(4): p. 1358-
359 1365.
- 360 8. Fassnacht, M. and B. Allolio, *Clinical management of adrenocortical carcinoma*. Best Pract
361 Res Clin Endocrinol Metab, 2009. **23**(2): p. 273-89.
- 362 9. Else, T., et al., *Adrenocortical carcinoma*. Endocr Rev, 2014. **35**(2): p. 282-326.
- 363 10. Daffara, F., et al., *Prospective evaluation of mitotane toxicity in adrenocortical cancer*
364 *patients treated adjuvantly*. Endocr Relat Cancer, 2008. **15**(4): p. 1043-53.
- 365 11. Hermsen, I.G., et al., *Plasma concentrations of o,p'DDD, o,p'DDA, and o,p'DDE as predictors*
366 *of tumor response to mitotane in adrenocortical carcinoma: results of a retrospective ENS@T*
367 *multicenter study*. J Clin Endocrinol Metab, 2011. **96**(6): p. 1844-51.
- 368 12. Terzolo, M., et al., *Mitotane levels predict the outcome of patients with adrenocortical*
369 *carcinoma treated adjuvantly following radical resection*. Eur J Endocrinol, 2013. **169**(3): p.
370 263-70.
- 371 13. Ronchi, C.L., et al., *CYP2W1 is highly expressed in adrenal glands and is positively associated*
372 *with the response to mitotane in adrenocortical carcinoma*. PLoS One, 2014. **9**(8): p. e105855.
- 373 14. Volante, M., et al., *Ribonucleotide reductase large subunit (RRM1) gene expression may*
374 *predict efficacy of adjuvant mitotane in adrenocortical cancer*. Clin Cancer Res, 2012. **18**(12):
375 p. 3452-61.
- 376 15. Sbiera, S., et al., *Mitotane Inhibits Sterol-O-Acyl Transferase 1 Triggering Lipid-Mediated*
377 *Endoplasmic Reticulum Stress and Apoptosis in Adrenocortical Carcinoma Cells*.
378 Endocrinology, 2015. **156**(11): p. 3895-908.
- 379 16. Lee, H.T., et al., *Inhibitors of acyl-CoA:cholesterol O-acyltransferase (ACAT) as*
380 *hypcholesterolemic agents: synthesis and structure-activity relationships of novel series of*
381 *sulfonamides, acylphosphonamides and acylphosphoramidates*. Bioorg Med Chem Lett, 1998.
382 **8**(3): p. 289-94.
- 383 17. Smith, D.C., et al., *A phase 1 study of nevanimibe HCl, a novel adrenal-specific sterol O-*
384 *acyltransferase 1 (SOAT1) inhibitor, in adrenocortical carcinoma*. Invest New Drugs, 2020.
- 385 18. Hescot, S., et al., *Mitotane alters mitochondrial respiratory chain activity by inducing*
386 *cytochrome c oxidase defect in human adrenocortical cells*. Endocr Relat Cancer, 2013. **20**(3):
387 p. 371-81.
- 388 19. Poli, G., et al., *Morphofunctional effects of mitotane on mitochondria in human*
389 *adrenocortical cancer cells*. Endocrine-Related Cancer, 2013. **20**(4): p. 537-550.
- 390 20. Hescot, S., et al., *Identifying mitotane-induced mitochondria-associated membranes*
391 *dysfunctions: metabolomic and lipidomic approaches*. Oncotarget, 2017. **8**(66): p. 109924-
392 109940.
- 393 21. Weiss, L.M., L.J. Medeiros, and A.L. Vickery, Jr., *Pathologic features of prognostic significance*
394 *in adrenocortical carcinoma*. Am J Surg Pathol, 1989. **13**(3): p. 202-6.

- 395 22. Fassnacht, M., et al., *Limited prognostic value of the 2004 International Union Against Cancer*
396 *staging classification for adrenocortical carcinoma: proposal for a Revised TNM Classification.*
397 *Cancer*, 2009. **115**(2): p. 243-50.
- 398 23. van Koetsveld, P.M., et al., *The Efficacy of Mitotane in Human Primary Adrenocortical*
399 *Carcinoma Cultures.* *J Clin Endocrinol Metab*, 2019.
- 400 24. Olaussen, K.A., et al., *DNA repair by ERCC1 in non-small-cell lung cancer and cisplatin-based*
401 *adjuvant chemotherapy.* *N Engl J Med*, 2006. **355**(10): p. 983-91.
- 402 25. Weigand, I., et al., *Differential expression of the protein kinase A subunits in normal adrenal*
403 *glands and adrenocortical adenomas.* *Sci Rep*, 2017. **7**(1): p. 49.
- 404 26. Altieri, B., et al., *Effects of Germline CYP2W1*6 and CYP2B6*6 Single Nucleotide*
405 *Polymorphisms on Mitotane Treatment in Adrenocortical Carcinoma: A Multicenter ENSAT*
406 *Study.* *Cancers (Basel)*, 2020. **12**(2).
- 407 27. Zsippai, A., et al., *Effects of mitotane on gene expression in the adrenocortical cell line NCI-*
408 *H295R: a microarray study.* *Pharmacogenomics*, 2012. **13**(12): p. 1351-61.
- 409 28. Geng, F., et al., *Inhibition of SOAT1 Suppresses Glioblastoma Growth via Blocking SREBP-1-*
410 *Mediated Lipogenesis.* *Clin Cancer Res*, 2016. **22**(21): p. 5337-5348.
- 411 29. Geng, F. and D. Guo, *Lipid droplets, potential biomarker and metabolic target in*
412 *glioblastoma.* *Intern Med Rev (Wash D C)*, 2017. **3**(5).
- 413 30. Jiang, Y., et al., *Proteomics identifies new therapeutic targets of early-stage hepatocellular*
414 *carcinoma.* *Nature*, 2019. **567**(7747): p. 257-261.
- 415 31. Stopsack, K.H., et al., *Cholesterol uptake and regulation in high-grade and lethal prostate*
416 *cancers.* *Carcinogenesis*, 2017. **38**(8): p. 806-811.
- 417 32. Lacombe, A.M.F., et al., *Sterol O-Acyl Transferase 1 as a Prognostic Marker of Adrenocortical*
418 *Carcinoma.* *Cancers (Basel)*, 2020. **12**(1).
- 419 33. Kerkhofs, T.M., et al., *Comparison of two mitotane starting dose regimens in patients with*
420 *advanced adrenocortical carcinoma.* *J Clin Endocrinol Metab*, 2013. **98**(12): p. 4759-67.
- 421 34. Mauclere-Denost, S., et al., *High-dose mitotane strategy in adrenocortical carcinoma:*
422 *prospective analysis of plasma mitotane measurement during the first 3 months of follow-up.*
423 *Eur J Endocrinol*, 2012. **166**(2): p. 261-8.
- 424 35. Baudin, E., et al., *Impact of monitoring plasma 1,1-dichlorodiphenildichloroethane (o,p'DDD)*
425 *levels on the treatment of patients with adrenocortical carcinoma.* *Cancer*, 2001. **92**(6): p.
426 1385-92.
- 427 36. Kerkhofs, T.M., et al., *Development of a pharmacokinetic model of mitotane: toward*
428 *personalized dosing in adrenocortical carcinoma.* *Ther Drug Monit*, 2015. **37**(1): p. 58-65.
- 429 37. Arshad, U., et al., *Enzyme autoinduction by mitotane supported by population*
430 *pharmacokinetic modelling in a large cohort of adrenocortical carcinoma patients.* *Eur J*
431 *Endocrinol*, 2018. **179**(5): p. 287-297.
- 432 38. Puglisi, S., et al., *Mitotane Concentrations Influence the Risk of Recurrence in Adrenocortical*
433 *Carcinoma Patients on Adjuvant Treatment.* *J Clin Med*, 2019. **8**(11).
- 434 39. LaPensee, C.R., et al., *ATR-101, a Selective and Potent Inhibitor of Acyl-CoA Acyltransferase 1,*
435 *Induces Apoptosis in H295R Adrenocortical Cells and in the Adrenal Cortex of Dogs.*
436 *Endocrinology*, 2016. **157**(5): p. 1775-88.
- 437 40. Cheng, Y., R.E. Kerppola, and T.K. Kerppola, *ATR-101 disrupts mitochondrial functions in*
438 *adrenocortical carcinoma cells and in vivo.* *Endocr Relat Cancer*, 2016. **23**(4): p. 1-19.
- 439 41. Beuschlein, F., et al., *Major prognostic role of Ki67 in localized adrenocortical carcinoma after*
440 *complete resection.* *J Clin Endocrinol Metab*, 2015. **100**(3): p. 841-9.
- 441 42. Luton, J.P., et al., *Clinical features of adrenocortical carcinoma, prognostic factors, and the*
442 *effect of mitotane therapy.* *N Engl J Med*, 1990. **322**(17): p. 1195-201.
- 443 43. Fassnacht, M., et al., *Adrenocortical carcinoma: a clinician's update.* *Nat Rev Endocrinol*,
444 2011. **7**(6): p. 323-35.
- 445 44. Scollo, C., et al., *Prognostic Factors for Adrenocortical Carcinoma Outcomes.* *Front Endocrinol*
446 *(Lausanne)*, 2016. **7**: p. 99.

- 447 45. Lippert, J., et al., *Targeted Molecular Analysis in Adrenocortical Carcinomas: A Strategy*
448 *Toward Improved Personalized Prognostication*. J Clin Endocrinol Metab, 2018. **103**(12): p.
449 4511-4523.
- 450 46. Zhang, F., et al., *Prognostic Role of Ki-67 in Adrenocortical Carcinoma After Primary*
451 *Resection: A Retrospective Mono-Institutional Study*. Adv Ther, 2019. **36**(10): p. 2756-2768.
- 452 47. Libe, R., *Clinical and molecular prognostic factors in adrenocortical carcinoma*. Minerva
453 Endocrinol, 2019. **44**(1): p. 58-69.
- 454 48. Libe, R., et al., *Prognostic factors in stage III-IV adrenocortical carcinomas (ACC): an European*
455 *Network for the Study of Adrenal Tumor (ENSAT) study*. Ann Oncol, 2015. **26**(10): p. 2119-25.

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458

459 **Table 1: Patient characteristics.**

	adjuvant
Patients n	158
Age in years: median (range)	54.2 (17-83)
Sex - n (%)	
female	97 (61)
male	61 (39)
Tumor stage - n (%)	
I	10 (6.5)
II	88 (56)
III	52 (33)
IV	7 (4.5)
R status (n=156)	
0	121 (77.6)
X	30 (19.2)
1	5 (3.2)
Ki67-Index (n=144) - n (%)	
<10%	30 (21)
≥10%	114 (79)
Weiss score (n=142): median (range)	6 (1-9)
Endocrine activity (n=143) - n (%)	
hormone secretion	96 (67)
no over-secretion	47 (33)
Months to mitotane start (median (IQR))	1 (2)
Mitotane plasma concentration at 3 months (median ±SD) (n=132)	9.05 ±5.9
Mitotane plasma concentration at 6 months (median ±SD) (n=125)	13 ±5.7

	advanced disease
Patients n	72
Age in years: median (range)	51 (16-80)
Sex n (%)	
female	42 (58)
male	30 (42)
Tumor stage- n (%)	
I	2 (2.8)
II	19 (26.4)
III	11 (15.3)
IV	40 (55.6)
R status (n=68)	
0	28 (41)
X	15 (22)
1	13 (19)
2	12 (18)
Ki67-Index (n=66) - n (%)	
<10%	18 (27.3)
≥10%	48 (66.4)
Weiss score (n=55): median (range)	6 (1-9)
Endocrine activity (n=62) - n (%)	
hormone secretion	45 (72.6)
no over-secretion	17 (27.4)
Months to mitotane start (median (IQR))	0 (2)
Mitotane level at 3 months (median ±SD) (n=61)	10±6.3
Mitotane level at 6 months (median ±SD) (n=51)	12.5±7

460

461 **Table 2: Impact of SOAT1 expression and known prognostic parameters on RFS and**
462 **DSS in the adjuvant (R0 or RX) cohort.**

RFS variables	univariate analysis			multivariate analysis		
	HR	95% CI	p	HR	95% CI	p
Sex						
female (n=91)						
male (n=56)	1.13	0.72-1.76	0.60	1.30	0.77-2.19	0.33
Age						
<50 (n=90)						
≥50 (n=58)	0.78	0.49-1.23	0.29	0.72	0.40-1.29	0.27

Hormone over-secretion							
Yes (n=90)							
No (n=43)	1.39	0.84-2.32	0.20	1.55	0.86-2.77	0.14	
Tumor stage							
I+ II (n=90)							
III + IV (n=56)	1.51	0.97-2.34	0.07	1.54	0.90-2.62	0.11	
Ki67							
<10 (n=28)							
≥10 (n=107)	3.810	1.64-8.84	0.002*	2.86	1.18-6.96	0.02	
Mitotane levels 3 months (median:9.1 mg/l) n=122							
<9.1		0.92-					
≥9.1	1.50	2.44	0.11	-	-	-	
Mitotane levels 6 months (median:13 mg/l) n=116							
<13		0.62 -					
≥13	1.02	1.67	0.95	-	-	-	
SOAT1							
H-Score low: <2 (n=89)							
H-Score high: ≥2 (n=59)	1.42	0.91-2.21	0.12	1.07	0.61-1.85	0.82	

DSS variables	univariate analysis			multivariate analysis		
	HR	95% CI	p	HR	95% CI	p
Sex						
female (n=81)		0.462-				
male (n=53)	1.19	2.28	0.61	1.65	0.74-3.67	0.22
Age						
<50 (n=80)						
≥50 (n=55)	0.64	0.32-1.29	0.21	0.60	0.24-1.55	0.29
Hormone over-secretion						
Yes (n=85)						
No (n=36)	1.52	0.68-3.40	0.31	1.48	0.58-3.79	0.42
Tumor stage						
I + II (n=85)						
III + IV (n=48)	1.43	0.74-2.76	0.28	1.23	0.54-2.78	0.63
Ki67						
<10 (n=24)						
≥10 (n=99)	4.91	1.17-20.67	0.03*	3.60	0.80-16.24	0.10
Mitotane levels 3						

months (median:9.1 mg/l) n=112							
<9.1		0.76 –					
≥9.1	1.52	3.06	0.24	-	-	-	
Mitotane levels 6 months (median:13 mg/l) n=103							
< 13							
≥13	0.74	0.34-1.60	0.44	-	-	-	
SOAT1							
H-Score low: <2 (n=81)							
H-Score high: ≥2 (n=54)	1.49	0.77-2.86	0.24	1.30	0.58-2.93	0.53	

463 **Table 3: Impact of SOAT1 expression and known prognostic parameters on PFS and**
464 **DSS in the cohort with advanced disease.** preM-TTP: pre mitotane time to progression

PFS variables	univariate analysis			multivariate analysis		
	HR	95% CI	<i>p</i>	HR	95% CI	<i>p</i>
Sex						
female (n=40)						
male (n=27)	0.75	0.44-1.27	0.28	0.81	0.43-1.53	0.51
Age						
<50 (n=29)						
≥50 (n=38)	0.84	0.50-1.42	0.52	0.73	0.36-1.50	0.40
Hormone over- secretion						
Yes (n=43)						
No (n=14)	1.46	0.77-2.78	0.25	1.98	0.97-4.03	0.06-
preM-TTP						
<365 days						
≥365 days	0.37	0.18-0.72	0.004*	0.49	0.21-1.11	0.09
Ki67						
<10 (n=17)						
≥10 (n=45)	1.19	0.66-2.14	0.55	0.92	0.46-1.83	0.81
Mitotane levels 3 months (median:10 mg/l) n=58						
<10						
≥10	0.70	0.40-1.22	0.21	-	-	-
Mitotane levels 6 months (median:12.5 mg/l) n=48						

<12.5							
≥12.5	0.61	0.33-1.14	0.12	-	-	-	
SOAT1							
H-Score low: <2 (n=37)							
H-Score high: ≥2 (n=30)	1.11	0.68-1.86	0.68	1.34	0.63-2.84	0.45	

DSS variables	univariate analysis			multivariate analysis		
	HR	95% CI	p	HR	95% CI	p
Sex						
female (n=36)						
male (n=27)	0.48	0.26-0.92	0.026*	0.92	0.40-2.11	0.83
Age						
<50 (n=27)						
≥50 (n=36)	0.82	0.46-1.48	0.52	1.39	0.63-3.04	0.42
Hormone over- secretion						
Yes (n=40)						
No (n=13)	1.04	0.52-2.07	0.92	1.20	0.52-2.80	0.67
Ki67						
<10 (n=14)						
≥10 (n=45)	2.47	1.14-5.32	0.021*	1.83	0.73-4.60	0.20
preM-TTP						
<365 days		0.014-				
>365 days	0.60	0.257	<0.001*	0.10	0.02-0.49	0.004*
Mitotane levels 3 months (median:10 mg/l) n=54						
<10		0.56-				
≥10	1.05	1.98	0.88	-	-	-
Mitotane levels 6 months (median:12.5 mg/l) n=45						
<12.5						
≥12.5	0.62	0.30-1.27	0.19	-	-	-
SOAT1						
H-Score low: <2 (n=35)						
H-Score high: ≥2 (n=28)	0.81	0.44-1.46	0.48	0.72	0.31-1.70	0.45

465 **Figure legends**

466 **Figure 1: SOAT1 immunohistochemistry staining of full ACC FFPE sections.** First

467 column shows an overview of SOAT1 staining intensities within the same tumors (scale bars:

468 3mm). Second column shows 3x magnification of the representative slide in first column
469 (scale bars: 700µm) and third column shows 20x magnification of the slide shown in column
470 A (scale bars: 200µm) (A-C: SOAT1 H-score 3, inhomogeneous staining; D-F: SOAT1 H-
471 Score 3, inhomogeneous staining; G-I: SOAT1 H-score 2, homogeneous staining, J-L:
472 SOAT1 H-score 0, homogenous staining).

473 **Figure 2: Kaplan-Meier plots of SOAT1 low and high expressing ACC. (A)** Recurrence-
474 /progression-free survival and **(B)** disease-specific survival of all ACC patients. **(C)**
475 Recurrence-free survival **(D)** and disease-specific survival of ACC patients in the adjuvant
476 group. **(E)** progression-free survival **(F)** and disease-specific survival of ACC patients with
477 advanced disease.

478 **Figure 3: SOAT1 expression and treatment response.** No significant differences
479 regarding mitotane response between SOAT1 high and SOAT1 low expressing tumors were
480 observed in the adjuvant arm **(A)**, nor in advanced stages **(B)**.

481 **Figure 4: Correlation of SOAT1 expression and mitotane plasma concentrations. (A)** In
482 both arms, high SOAT1 expression was not correlated with higher mitotane plasma levels.
483 Patients with high SOAT1 expression are not more likely to reach mitotane plasma levels
484 above 14 mg/l not in the adjuvant setting **(B)**, nor in patients with advanced disease **(C)**.
485 When only patients reaching the mitotane target level of 14 mg/l were analysed, high SOAT1
486 expression was significantly correlated with higher rates of recurrences after three months
487 **(D)** which did not retain significance after six months **(E)**.