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Beverly, David; Lopez-Quiroga, Estefania; Farr, Robert; Melrose, John; Henson, Sian; Bakalis, Serafim; Fryer, Peter

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Modeling Mass and Heat Transfer in Multiphase Coffee Aroma Extraction

David Beverly, Estefanía Lopez-Quiroga, Robert Farr, John Melrose, Sian Henson, Serafim Bakalis, and Peter J. Fryer*



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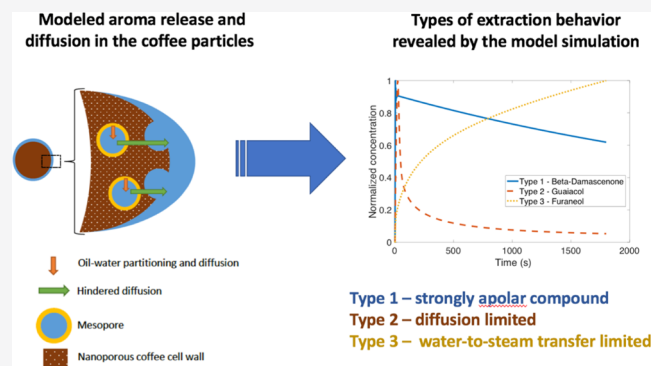


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ABSTRACT: Instant coffee manufacture involves the aqueous extraction of soluble coffee components followed by drying to form a soluble powder. Loss of volatile aroma compounds during concentration through evaporation can lower product quality. One method of retaining aroma is to steam-strip volatiles from the coffee and add them back to a concentrated coffee solution before the final drying stage. A better understanding of the impact of process conditions on the aroma content of the stripped solution will improve product design stages. In this context, we present a multiscale model for aroma extraction describing (i) the release from the matrix, (ii) intraparticle diffusion, (iii) transfer into water and steam, and (iv) advection through the column mechanisms. Results revealed (i) the existence of three different types of compound behavior, (ii) how aroma physiochemistry determines the limiting kinetics of extraction, and (iii) that extraction for some aromas can be inhibited by the interaction with other coffee components.



1. INTRODUCTION

Coffee is the most popularly prepared beverage, with a global production exceeding 9.5 million tonnes of green coffee beans.^{1,2} In the United Kingdom, instant coffee dominates the coffee market with a share of approximately 80%, equivalent to around 50 000 tonnes.³

The instant coffee process begins with the roasting of green beans to develop flavor compounds⁴ (see Table S1 for a list of typical aroma and associated data). Additionally, roasting develops porosity⁵ and vaporizes water, reducing moisture content. Roasted coffee beans are ground to reduce the particle size—increasing the surface area and reducing closed porosity⁵—and undergo multiple stages of high-temperature aqueous extraction of the soluble components. This extract is dried by evaporation and spray- or freeze-drying to form a soluble powder.

To reduce aroma degradation and loss caused by thermal processing, an aroma stream is extracted immediately after grinding, which is then added to the concentrated coffee extract before the final drying stage. Various techniques contacting ground coffee with water and/or steam are described in the patent literatures.^{6–8} These steps largely determine the final product aroma content, so the engineer must understand the mechanisms by which aroma transfers from coffee grounds into aqueous and gaseous media, allowing processes to be optimized to target the desired aromatic content. Steam stripping, analogous to processes^{9,10} in oils, is widely used.

It has been asserted that aroma exists principally within the coffee oil,⁴ but no published mass balance exists to prove this. Schenker¹¹ showed that oil coats the outer walls of the grain cells as discrete micron-scale droplets, which migrate to the bean surface during roasting. The easily accessible aroma on the surface of the grain will dissolve in the water and partition into the headspace, from which the aroma will be carried by the advection of steam. As the surface is depleted and as water fills the pores, aroma will continue to dissolve and diffuse into the surface. Any aroma dissolved within the coffee oil may directly partition into the headspace from the surface oil, but oil extraction from within the grain is poor^{12,13} and such aroma must be extracted by aqueous dissolution first. As the grain is wetted, it swells impacting porosity and diffusion. Particle size analysis showed that swelling of wetted grounds stopped within 10–15 min.^{14,15} A study of the kinetics of aqueous extraction, for domestic coffee brewers, showed how hydrophilic aroma compounds extract significantly faster than lipophilic ones.¹⁶ Sánchez López et al.¹⁷ studied the headspace above espresso coffee and characterized coffee aroma into two clusters: (i) a

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faster extracting group of typically low-molecular-weight compounds (acids, esters, and carbonyls), increasing quickly between 6 and 10 s of extraction; and (ii) a slower group of higher-weight heterocycles and phenols increasing between 6 and 20 s. In another study, when stripping aroma with nitrogen gas from a bed of dry coffee, data could be fitted well to the analytical solution of a Fickian diffusion model and a Weibull model.¹⁸ However, upon wetting the coffee, the behavior of some compounds (including acetic acid, pyridine, and methyl furfural) could not be described using the diffusion model. The addition of the aqueous phase seems to introduce new complexities, such as the interaction with other components in the coffee matrix, potentially involving various functional groups.¹⁹

Several published models of essential oil distillation from plant matter describe extraction purely by fitting mass transfer coefficients.^{10,20} Moroney et al.²¹ modeled coffee extraction in brewing, describing the transfer between inert coffee solid matter, coffee particle (intragranular) pores, and coffee bed (intergranular) pores. They define lengths of diffusion from the solid to intragranular pore and from the intragranular to intergranular pore, and use experiment-derived fitted parameters to describe these processes.

The approach used here is to solve the particle-scale diffusion equation and bed-scale advection simultaneously. It builds upon the particle/bed model used for brew yield by Melrose et al.²² and references therein, adapting for aroma compounds, adding intraparticle interactions and external transfer to a steam flow. The model was prefaced in Beverly et al.,²³ but the aim here is to identify the different rate-limiting extraction mechanisms for different compounds (and, hence, aromatic properties) and predict how features of the process within (or outside) the control of the engineer can impact the chemical and sensory profile of the resulting distillate. The practical result should be a tool to guide process development when optimizing aroma yield, concentration, and desired sensory attributes.

2. EXTRACTION SYSTEM

The focus of this paper is on a process similar to that described by Vitzthum and Koch.²⁴ A packed bed of ground coffee (up to 1.8 mm diameter) is uniformly wetted and steamed for up to 40 min. During this process, heat and mass transfer processes occur simultaneously. On the addition of hot water, there is water absorption into the porous coffee grains, whereupon soluble compounds dissolve and diffuse into the surface. During steaming, the bed is heated by condensation, which provides additional moisture. Volatile compounds will partition into the gas phase and be carried by the steam out of the column.

The system to be modeled is a packed bed of roasted and ground coffee beans, in a cylindrical column (Figure 1a). A defined quantity of water is first added, which is assumed to be perfectly distributed through the column and is sufficient to entirely fill the porous coffee particles. Saturated steam enters via the column base, and a vacuum is applied at the top.

In each column element, the bed consists of loose-packed particles, with free water existing as a surrounding uniform film (Figure 1b). One particle is taken as being representative of the whole population in the element. Each particle is porous, with the porosity consisting of large spherical pores connected by a nanoporous network through the cell walls (Figure 1c), a simplification of the microstructure seen in Figure 2.

The cell wall is built of a matrix of structural plant macromolecules, mainly polysaccharides, containing nano-

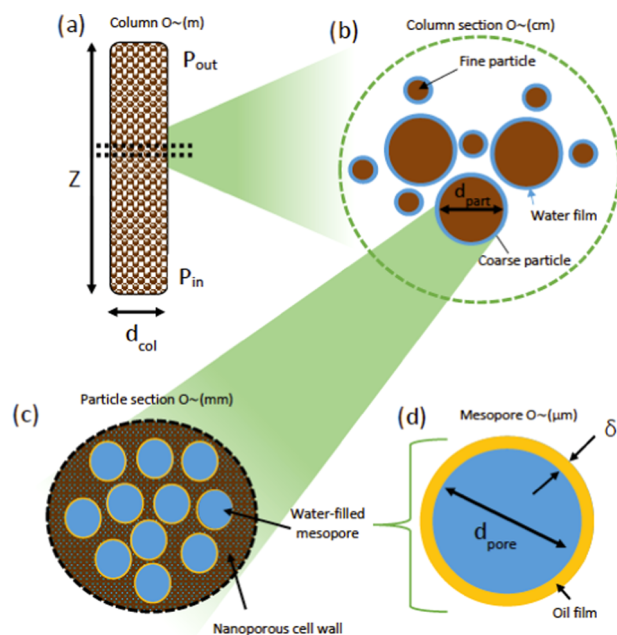


Figure 1. Schematic diagrams of different scales in the system with key geometries: (a) whole column, with the labeled height (Z) inlet and outlet pressures (P_{in} and P_{out}) and column diameter (d_{col}); (b) column section, with the labeled bed pore size ($d_{b,pore}$) and particle diameter (d_{part}); (c) particle section with mesopores distributed in a nanoporous matrix all filled with water; and (d) mesopore with a labeled diameter (d_{pore}) and oil thickness (δ).

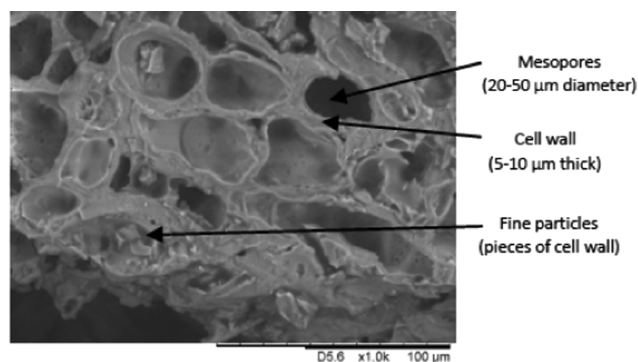


Figure 2. Scanning electron microscope image of a roasted and ground coffee particle (magnification 1000 \times) showing the mesopores and cell walls, taken on a tabletop TM-1000 microscope (Hitachi, Tokyo, Japan).

pores.¹¹ When considering diffusion and thermal conduction, the porous particle is considered homogeneous, with the transport coefficients adjusted for the averaged porosity and tortuosity. The aroma is assumed dissolved in the oil phase, treated here as a uniform thin layer of thickness δ , which coats each large spherical mesopore (Figure 1d), and partitions and diffuses into the free phase according to the octanol–water partition coefficient.

3. MODEL FORMULATION

Aroma is extracted from the oil film (ca. 1 μm thickness) into the water-filled mesopore (ca. 20 μm radius) (Figures 1d and 2). There is stagnant diffusive transport from the oil–water boundary concentration (determined by the octanol–water partition) to the mesopore center (deemed the free aroma

concentration in the particle). An effective diffusion coefficient describes the transport within the particle (ca. 1 mm radius), which combines free diffusion in the mesopores and hindered diffusion in the cell walls (ca. 10 μm thickness) (Figure 1c). A flux boundary condition determines the transfer from the particle surface to free water in the bed, which forms a stagnant film around the particles. Henry's law volatility constants describe the partition into the headspace, and there is convective transport into the steam across the surface area of the particles ($O \sim 10 \text{ m}^2 \text{ kg}^{-1}$) (Figure 1b). Advection by pressure-driven flow through a porous bed describes steam progress through the column [$O \sim (\text{m})$] (Figure 1a). All references to "concentration" hereon relate to molar concentration.

3.1. Assumptions of the Model. At the bed scale:

- The column is perfectly insulated.
- No radial variation in the flow, temperature, or water content.
- The dominant mechanisms of aroma mass transfer are aqueous diffusion within the particle to free water (on the particle external surfaces) to the gas phase and then via advection in the gas phase.
- All material flowing out of the top of the column is condensed.
- The mobile phase is an ideal gas.
- The liquid water surrounding the particles is not mobile (does not flow).

For the particle scale:

- Particles are spherical with evenly distributed porosity,⁵ and only radial gradients will be considered.
- Particle size is bimodal, with the diffusion equation solved for each class of coarse and fine particles using mean particle sizes d_c and d_f .
- Porosity consists of spherical mesopores connected by a nanoporous network, which is entirely saturated with water.
- No particle swelling effects: dissolution of the soluble fraction of the particles is instant upon wetting, i.e., no change in particle size or porosity with time.
- Aroma initially exists dissolved in an oil layer uniformly spread over the inner surface of the mesopores, and dissolved aroma partitions according to the octanol–water partition coefficient, as this is a representative parameter for liquid-phase extraction systems.²⁵
- The octanol–water partition coefficient is independent of temperature (and, thus, time and geometry). The relationship is not studied for many aromas, and, where data exists, it is limited to temperature increases of the order of 20 $^\circ\text{C}$;^{26,27} for some compounds, the difference is insignificant (<10%), but there may be some aromas that change partition behavior strongly over larger ranges.
- Aroma extraction is not limited by solubility.
- Physical properties (density, conductivity, heat capacity, etc.) are weighted averages of solid coffee and water properties.
- Free water in the bed exists as a homogeneous stagnant film coating the particles.²⁸

3.2. Bed Scale. 3.2.1. Water Mass Transfer. The free water in the bed ($\rho_{b,w}$), i.e., the water in excess of that required to fill the porous coffee particles, is expressed as a mass of water per unit volume of bed. The initial value is determined by the added water-to-coffee ratio ($\phi_{wc} = g_{\text{water}} / g_{\text{dry coffee}}^{-1}$), where a homogeneous distribution is assumed

$$\rho_{b,w}(\forall z, t = 0) = (1 - \varepsilon_b)[\phi_{wc}\rho_p - \varepsilon_p\rho_w] \quad (1)$$

where ρ_p and ρ_w are the real densities of the coffee particle and water, respectively; and ε_p is the particle porosity. The amount of free water in the bed is a function of condensation as no flow or loss occurs within the bed

$$\frac{\partial \rho_{b,w}(z, t)}{\partial t} = \frac{\dot{m}_{\text{con}}(z, t)}{\Delta V_{\text{bed}}} \quad (2)$$

The rate of condensation \dot{m}_{con} is defined using the simplified Hertz–Knudsen formula²⁹

$$\dot{m}_{\text{con}}(z, t) = \sqrt{\frac{M_{\text{vap}}}{2\pi R}} \left(\frac{p_{\text{st}}(z)}{\sqrt{T_{\text{st}}(z)}} - \frac{p_w(z, t)}{\sqrt{T_b(z, t)}} \right) a_b \Delta V_{\text{bed}} \quad (3)$$

where M_{vap} is the molar mass of water vapor; R is the gas constant; p_{st} and T_{st} are the saturated steam pressure and temperature, respectively; p_w and T_b are the free water vapor pressure and bed temperature, respectively; a_b is the area per unit volume of bed; and V_{bed} is the bed volume. The surface area per unit volume of the bed a_b is calculated as a function summing the contribution from the coarse and fine particle size classes

$$a_b = \frac{3(1 - \varepsilon_b)\varphi_c}{r_c} + \frac{3(1 - \varepsilon_b)\varphi_f}{r_f} \quad (4)$$

While the column remains below the saturation temperature of the steam, there will be condensation, reducing the flow of steam out of a column element Q_{out}

$$Q_{\text{out}}(z, t) = Q - \frac{\dot{m}_{\text{con}}(z, t)}{\rho_{\text{st}}} \quad (5)$$

where ρ_{st} is the density of the saturated steam. Darcy's Law is used to calculate the volumetric flow of steam (Q) and superficial velocity (v) through the bed

$$v = \frac{Q}{A_{\text{bed}}} = \frac{K}{\mu_{\text{in}}} \frac{(P_{\text{in}} - P_{\text{out}})}{Z} \quad (6)$$

where A_{bed} is the area of the bed and μ_{in} is the gas viscosity at the inlet. The inlet and outlet pressures (P_{in} and P_{out}), bed height (Z), and diameter (d_{bed}) are specified by the process. The bed permeability (K) is calculated using the Kozeny–Carman equation⁵

$$K = \frac{\varepsilon_b^3(d_{3,2})^2}{72\varepsilon_b^{-0.8}(1 - \varepsilon_b)^2} \quad (7)$$

where $d_{3,2}$ is the Sauter mean diameter and ε_b is the bed porosity, which is estimated using the method of Farr and Groot³⁰

$$(1 - \varepsilon_b) = \psi_{\text{max}} = \min \left(\frac{\phi_{\text{RCP}}}{1 - \omega(1 - \phi_{\text{RCP}})}, \frac{\phi_{\text{RCP}}}{\omega} \right) \quad (8)$$

where ϕ_{RCP} is the maximum packing fraction for a random close-packed monodisperse sphere, and ω is defined as

$$\omega = \frac{\varphi_c}{\varphi_c + \varphi_f} \approx 1 \quad (9)$$

In eq 9, φ_c and φ_f are the coarse and fine particle sizes, respectively. The value of $d_{3,2}$ is calculated by modeling the particle size distribution as the sum of two log–normal

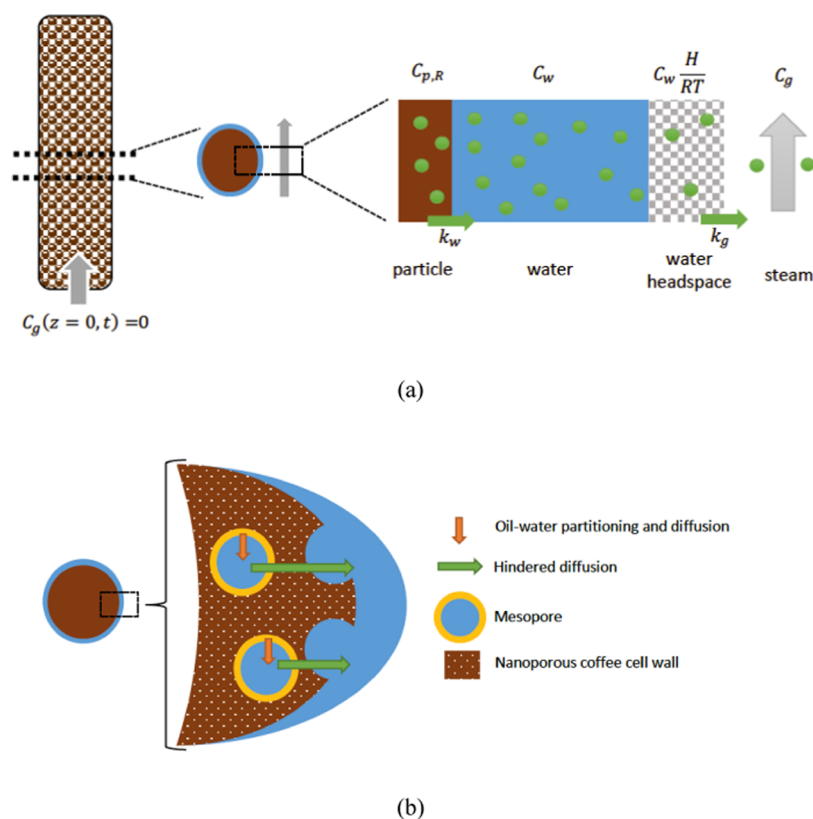


Figure 3. Schematic diagrams of (a) the particle–water–gas transfer stage and (b) aroma release and diffusion into the coffee particles.

distributions, with mean coarse and fine particle sizes (d_c and d_f) similar to the methods used by Melrose et al.²² Water vapor pressure and steam saturation temperature have been estimated by using steam table data³¹ and fitting logarithmic curves for $1.2 < P < 6$ bar and $P < 1.2$ bar ($R^2 > 0.99$).

3.2.2. Aroma Mass Transfer. The concentration (in mol m^{-3}) of each individual aroma is modeled independently. Initially, there is no aroma in the water or gas phase

$$c_w(\forall z, t = 0) = 0; \quad c_g(\forall z, t = 0) = 0 \quad (10)$$

and transfer to the gas phase does not begin until the steam contacts that column element. The concentration in the gas phase c_g (mol m^{-3}) is determined by the sum of advection and source terms—dispersion is negligible

$$\frac{\partial c_g(z, t)}{\partial t} = -\frac{Q(z, t)}{A_{bed}\epsilon_b} \frac{\partial c_g(z, t)}{\partial z} + k_g a_{bp} \left(c_w(z, t) \frac{K_H^{pc}(T_b)}{RT_b(z, t)} - c_g(z, t) \right) \quad (11)$$

where the term a_{bp} is defined as $a_{bp} = a_b/\epsilon_b$ (area that the mobile phase occupies). Henry's law volatility constant $K_H^{pc}(T_b)$ is calculated according to the van't Hoff equation

$$\ln \frac{K_H^{pc}(T_b)}{K_{H,298}^{pc}} = \frac{\Delta H}{R} \left(\frac{1}{T_b} - \frac{1}{298} \right) \quad (12)$$

Values of $K_{H,298}^{pc}$ and ΔH for several aromas can be found in the literatures.^{32,33} For weak acids, the modified value of Henry's law volatility constant can be calculated as a function of the literature values (see Table S1), pH and pK_a , as follows³⁴

$$K_H^{pc} = \frac{K_{H,298}^{pc}}{(1 + 10^{(pH - pK_a)})} \quad (13)$$

by combining van't Hoff and Henderson–Hasselbalch equations. This assumes that there are no bases present to react with the acid. If any such components are present in the aqueous solution, then Henry's constant will be reduced. Required pK_a data have been taken from Harned and Ehlers³⁵ and extrapolated to $T = 100$ °C.

At the column inlet, there is no aroma in the gas phase. As the upwind scheme is used to define the concentration gradient in eq 11, each node's properties are calculated as a function of the preceding nodes. This means the outlet is an open boundary

$$c_g(z = 0, \forall t) = 0; \quad c_g(z > Z, t) = c_g(z, t) \quad (14)$$

The concentration in the stationary free water in the bed (c_w in mol m^{-3}) is described by source and loss terms from/to the particle surface ($c_{p,R}$) and gas (c_g), respectively (see Figure 3).

There is also a dilution effect caused by condensing steam that increases the volume of water in which the aroma is dissolved

$$\frac{\partial c_w(z, t)}{\partial t} = \left(k_w a_w(z, t) [c_{p,R}(z, t) - c_w(z, t)] - k_g a_w(z, t) \left[c_w(z, t) \frac{K_H^{pc}(T_b)}{RT_b(z, t)} - c_g(z, t) \right] - \frac{c_w(z, t)}{\rho_{b,w}(z, t)} \frac{\partial \rho_{b,w}(z, t)}{\partial t} \right) \quad (15)$$

where k_w and k_g are the water- and gas-phase mass transfer coefficients, respectively; and a_w is the water surface area per unit

volume of water, which is equivalent to the particle surface area, since the water film is thin

$$a_w(z, t) = a_b \frac{\rho_w}{\rho_{b,w}(z, t)} \quad (16)$$

The mass transfer coefficient k_g has been obtained as³⁶

$$k_g = 1.15 \frac{\nu}{\varepsilon_b} Re^{-0.5} Sc^{-2/3} \quad (17)$$

for $Re < 1000$. For the stagnant water layer, with $Re \cong 0$ and $Sh \cong 2$. The mass transfer coefficient k_w was calculated as³⁷

$$k_w = \frac{2D_0(z, t)}{d_{3,2}} \quad (18)$$

where D_0 is the free diffusivity in water.

3.2.3. Heat Transfer. The rate of heat transfer from condensing steam to the coffee particles is significantly faster (ca. 100 times faster) than mass transfer, so the wetted coffee bed can be considered a homogeneous material with material-averaged thermal properties. We may regard axial conduction effects through the bed to be insignificant compared to latent heat contributions, so the corresponding energy balance on the bed leads to

$$\frac{\partial(\bar{\rho}_b(z, t)\bar{C}_p T_b(z, t))}{\partial t} = \dot{m}_{con}(z, t)\lambda + \dot{m}_{con}(z, t)\bar{C}_p(T_{sat}(z) - T_b(z, t)) \quad (19)$$

where $\bar{\rho}_b$ and \bar{C}_p are, respectively, the mass-averaged density ($\text{kg}_{\text{material}} \text{m}_{\text{bed}}^{-3}$) and specific heat capacity; T_b and T_{sat} are the bed and saturation temperatures, respectively; and λ is the latent heat of steam. The mass-averaged temperature of the column contents (T_b) is calculated as

$$T_b(\forall z, t = 0) = \frac{T_c C_{p,c} + T_w C_{p,w} \phi_{wc}}{C_{p,c} + C_{p,w} \phi_{wc}} \quad (20)$$

where $C_{p,w}$ and $C_{p,c}$ are the specific heat capacities of the water and dry coffee, respectively. The ground coffee is initially at an ambient temperature ($T_c = 298 \text{ K}$), and the added water temperature (T_w) can be varied.

3.3. Particle Scale. 3.3.1. Aroma Mass Transfer. Two concentrations in the particle are modeled: (i) dissolved aroma in the oil layer of the mesopores (c_{do} in mol m^{-3}) and (ii) free aroma that diffuses through the particle in the water-filled mesopores (c_p in mol m^{-3}), as depicted in Figure 3b. The release of dissolved aroma is described by

$$\frac{\partial c_{do}(r, t)}{\partial t} = -k_{por} a_{por,do} \left(\frac{c_{do}(r, t)}{K_{o/w}} - c_p(r, t) \right) \quad (21)$$

where $a_{por,do}$ is the oil–mesopore interfacial area per unit volume of oil, assuming spherical mesopores; and $K_{o/w}$ is the octanol–water partition coefficient, considered constant throughout the column for the duration of extraction. The mass transfer coefficient k_{por} is defined as

$$k_{por} = \frac{D_0(z, t)}{r_{por}} \quad (22)$$

where r_{por} is the pore radius. Equilibration between oil and water in the mesopore is rapid (of the order of 1 s). Diffusion of free

aroma is described using Fick's second law with a source term from the dissolved aroma being released

$$\frac{\partial c_p(r, t)}{\partial t} = D_{eff}(r, t) \left(\frac{2}{r} \frac{\partial c_p(r, t)}{\partial r} + \frac{\partial^2 c_p(r, t)}{\partial r^2} \right) + k_{por} a_{por,p} \left(\frac{c_{do}(r, t)}{K_{o/w}} - c_p(r, t) \right) \quad (23)$$

where $a_{por,p} = 3/r_{por}$ is the oil–mesopore interfacial area per unit volume of the mesopore and D_{eff} is the effective diffusivity. As there are two phases through which the mobile aroma diffuses—an unhindered mesopore region and a tortuous nanoporous cell wall— D_{eff} is calculated by using the Maxwell homogenization model³⁸

$$D_{eff} = D_h \left(\frac{2D_h + D_0 - 2\varepsilon_p(D_h - D_0)}{2D_h + D_0 - \varepsilon_p(D_h - D_0)} \right) \quad (24)$$

where the spherical mesopores are the inclusions with free diffusivity in water D_0 and the continuum is the cell wall material with hindered diffusivity D_h . Free diffusion coefficient values were obtained from the literature for several compounds (see Table 1), and a correlation with molecular weight was made to

Table 1. Diffusivity in Water and Molecular Weights of Some Aromas⁴⁰

compound	diffusivity at 298 K ($\text{m}^2 \text{s}^{-1}$)	molecular weight (g mol^{-1})
phenol	9.7×10^{-10}	94.1
4-methylphenol	8.5×10^{-10}	108
4-ethylphenol	7.7×10^{-10}	122
guaiacol	8.2×10^{-10}	124
vanillin	7.2×10^{-10}	152
indole	8.4×10^{-10}	117
acetic acid	11.9×10^{-10}	60.1
3-methylbutanoic acid	8.3×10^{-10}	102
2,3-pentanedione	8.8×10^{-10}	100

estimate others. The diffusion coefficient for loosely packed beds (D_h) depends on the coffee grain microstructure and the fluid properties (i.e., viscosity). Various correlations exist, relating diffusivity to both porosity and tortuosity, and tortuosity has itself been related to the porosity through various power law and logarithmic functions.³⁹ Corrochano et al.⁵ combined these and used the following relation for loosely packed beds

$$D_h = \varepsilon_p^{1.4} D_0 \quad (25)$$

The Stokes–Einstein equation also gives rise to the relation of diffusivity with temperature and fluid viscosity

$$D(T_b) = D(T^*) \frac{T_b}{T^*} \frac{\mu(T^*)}{\mu(T_b)} \quad (26)$$

where $T^* = 298 \text{ K}$.

The viscosity of coffee solution filling the mesopores is estimated using⁴¹

$$\mu = \exp\left(-12.96 - 9.43x_{s,p} + 8.12x_{s,p}^2 + \frac{1789 + 4382x_{s,p}}{T_b}\right) \quad (27)$$

where the mass fraction of coffee solids [$x_{s,p}$ ($\text{kg}_{\text{dry coffee}}/\text{kg}_{\text{solution}}^{-1}$)] is obtained as

$$x_{s,p} = \frac{0.3}{\left(\frac{\varepsilon_p \rho_w}{\rho_c(1-\varepsilon_p)} + 0.3\right)} \quad (28)$$

assuming that the maximum extractable solid content (0.3 w/w²⁸) is entirely dissolved into the water in the coffee pores.

Initially, there is uniform aroma distribution in the oil layer (thickness = δ) within the mesopores (radius = r_{por}) and no free aroma

$$c_{\text{do}}(\forall r, t = 0) = \frac{m_0 \rho_p}{\varepsilon_p \left(1 - \left(1 - \frac{\delta}{r_{\text{por}}}\right)^3\right)}; \quad c_p(\forall r, t = 0) = 0 \quad (29)$$

where m_0 is the aroma content per unit mass of coffee, ρ_p is the particle density, and ε_p is the particle porosity. Symmetry is assumed in the center of the particles, and a flux boundary condition is imposed on the boundary between the particle mesopores (c_p in mol m^{-3}) and surface water (c_w in mol m^{-3})

$$\frac{\partial c_p(r = 0, \forall t)}{\partial r} = 0; \quad \varepsilon_p D_{\text{eff}}(R, t) \frac{\partial c_p(r = R, \forall t)}{\partial r} = -k_w(c_p(R, t) - c_w(z, t)) \quad (30)$$

3.3.2. Matrix Interactions. An irreversible reaction mechanism that is first order in each species, and with a 1:1 molar ratio of the reactants, is used to model interactions between aroma and the phenolic component of the unextractable coffee matrix⁴²



where c_{do} is the concentration of the aroma (mol m^{-3}), $c_{\text{do,PP}}$ is the phenolic (binding species) concentration (mol m^{-3}), and $c_{\text{do,comp}}$ is the bound complex concentration (mol m^{-3}), all within the oil phase. For compounds undergoing this binding, an additional reaction term is added to eq 21, leading to

$$\frac{\partial c_{\text{do}}}{\partial t} = -k_{\text{por}} a_{\text{por,p}} \left(\frac{c_{\text{do}}(r, t)}{K_{\text{o/w}}} - c_p(r, t) \right) - k_{\text{on}} c_{\text{do}} c_{\text{do,PP}} \quad (32)$$

Equation 32 can also be adapted for the phenolic and complexed species

$$\frac{\partial c_{\text{do,PP}}}{\partial t} = -k_{\text{on}} c_{\text{do}} c_{\text{do,PP}} \quad (33)$$

$$\frac{\partial c_{\text{p,comp}}}{\partial t} = k_{\text{on}} c_{\text{do}} c_{\text{do,PP}} \quad (34)$$

The initial phenolic content has been taken or estimated from the literatures.^{43,44}

4. RESULTS AND DISCUSSION

The extraction problem formed by eqs 1–34 was solved using a self-developed one-dimensional forward time centered space (FTCS) finite difference (FD) scheme, which was implemented in MATLAB. Two sets of numerical simulations were performed:

- Small-scale extraction simulations, which were used to calibrate the model for the range of operating conditions and coffee beds studied here. Values for the cell wall porosity (ε_{cw}), the ratio $\frac{\Delta H}{R}$, and the binding rate constant (k_{on}) were estimated by fitting the proposed model to the published data¹⁸ for similar extraction systems (i.e., water-saturated-nitrogen stripping of a 5 g bed of coffee wetted in a 1:1 ratio of water to coffee¹⁸).
- Industrial-scale simulations (i.e., large scale), which were used to evaluate how different process conditions (i.e., a variation on the process parameters) might affect the aroma profile characteristic of the resulting distillate.

4.1. Small-Scale Simulations. To calibrate the model, published experimental data¹⁸ corresponding to the extraction kinetics of three key aroma compounds, i.e., acetaldehyde, acetic acid, and pyridine, was used. These three aromas display different extraction kinetics, so they were used to estimate (i) cell wall porosity, (ii) ratio $\frac{\Delta H}{R}$, and (iii) binding rate constant, respectively. Estimated values were then used for all other aromas.

Numerical simulations used 50 mesh nodes for the coarse particles, 5 mesh nodes for the fine ones, and 25 nodes for the bed domain. Mesh convergence was ensured by a sensitivity analysis on mesh sizes, as well as by comparing simulated results with analytical solutions³⁸ for Fickian diffusion systems. Table 2 lists the model parameter values used to simulate the small-scale extraction system.

Table 2. Parameters Used for Model Calibration^{5,18,28}

parameter	validation settings	parameter	validation settings
d_c	750 μm	Z	0.024 m
d_{part}	40 μm	d_{bed}	0.024 m
ϕ_f	0.15	ΔP	8.3 kPa
$d_{3,2}$	198 μm	K	$1.8 \times 10^{-12} \text{ m}^2$
Q	$1.24 \times 10^{-5} \text{ m}^3 \text{ s}^{-1}$	Re_{N_2}	1.2
ε_p	0.42	S_{CN_2}	1.2
ε_b	0.24	ϕ_{wc}	1
ρ_c	1337 kg m^{-3}	T_w	353 K
σ_c	0.28	max extractable solids	0.3 w/w
σ_f	0.22	ϕ_{RCP}	0.6435
r_{por}	20 μm	polyphenolic content	0.63 $\text{g kg}_{\text{coffee}}^{-1}$
δ	1 μm	D_{pp}	$1 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$
K_{pp}	10		

Estimates for the unknown parameters (i.e., ε_{cw} , $\frac{\Delta H}{R}$ and k_{on}) that minimized the error (in a least-squares sense) between published extraction data¹⁸ and model outcomes for each aroma extraction curve were obtained using regression analysis. Corresponding values for root-mean-square error (RMSE), χ^2 , and the coefficient of determination (R^2) are presented in Table 3.

Table 3. Error Analysis Using Root-Mean-Square Error (RMSE), χ^2 , and Coefficient of Determination (R^2) Corresponding to the Model Calibration

simulation	RMSE	χ^2	R^2
acetaldehyde, monomodal distribution	0.0681	0.00474	0.990
acetaldehyde, bimodal distribution	0.0561	0.00321	0.994
acetic acid, monomodal distribution	0.165	0.0278	0.976
pyridine, monomodal distribution	0.0545	0.00300	0.983

4.1.1. Acetaldehyde. A cell wall porosity (ϵ_{cw}) value was estimated by fitting the proposed extraction model to experimental data for acetaldehyde.¹⁸ No reaction with the soluble coffee solids ($k_{on} = 0$) was assumed in this case. To assess the effect of multiple particle sizes on the extraction process, both monodisperse (coarse particles) and bidisperse (fine and coarse particles) beds were simulated, resulting in values of $\epsilon_{cw} = 3.30$ and 2.76%, respectively. Figure 4 presents a comparison between reference data¹⁸ and the fitted extraction curves for each particle size, showing the goodness of the fit; i.e., experimental trends observed in Mateus et al.¹⁸ can be accurately reproduced.

There is limited literature on the porosity of plant cell walls. Schenker et al.⁴⁵ published mercury porosimetry measurements, from which the contribution to total porosity made by the cell wall (typically <10 nm) can be estimated

$$\epsilon_p = \epsilon_{mp} + \epsilon_{cw}(1 - \epsilon_{mp}) = V_{por}\rho_p \quad (35)$$

The coffee particle density (ρ_p) was 622 kg m⁻³, $V_{por} = 850$ mm³ g⁻¹, and the cell wall porosity contribution was estimated⁴⁵ to be 130 mm³ g⁻¹, generating a total porosity (ϵ_p) of 0.53, a mesopore contribution (ϵ_{mp}) of 0.45, and a cell wall porosity (ϵ_{cw}) of 0.17.

A cell wall porosity of 17% is clearly larger than that estimated by the model; however, there are several reasons for why the “effective porosity” would be much lower. First, porosity below a certain size may not be appropriate for aroma transport or the restricted pore channel may not permit free diffusion. The hydrodynamic radius of arabinose was calculated to be around 0.4 nm⁴⁶—aroma molecules of this size are approaching the size of the smallest nanopores. Larger molecules, including soluble coffee solids, would thus be trapped in these nanopores, blocking diffusion of other substances. Second, organic molecules interact with the polysaccharide matrix, further inhibiting diffusion. Any association of larger coffee solutes to the cell wall material will further inhibit diffusion through those

nanopores via additional steric hindrance. Finally, the addition of water will cause absorption, which could compress the nanopores as the polysaccharide matrix swells.

Using the estimated cell wall porosity values, the effective diffusivity values were then obtained through eq 25, leading to values of $D_{eff} = 6.0 \times 10^{-12}$ and 4.7×10^{-12} m² s⁻¹ for the monodisperse and bidisperse particle sizes, respectively. This results in hindrance factors (H_f) of 204 and 262, respectively, as given by

$$H_f = \frac{D_0}{D_{eff}} \quad (36)$$

Hindrance values (H_f) obtained here were larger than those previously reported for caffeine and mineral ions,^{47,48} which range between $H_f = 9$ and 48. In addition to geometric factors considered in those other works,^{47,48} there are several possible explanations for these differences. First, the cited studies performed an extraction in very dilute conditions, whereas here, the wetted grain retains its soluble solid content (up to ca. 30% of the total mass), which will provide an additional steric hindrance. Second, interactions between solids and aroma molecules will further hinder diffusion, as well as influence the effective Henry's constant. Third, the viscosity of the fluid within the grain is higher here (a viscosity ratio of 2.7), and diffusion coefficients in concentrated carbohydrate solutions can present very high hindrance factors due to viscosity effects.⁴⁹ Finally, the bed may not experience uniform gas flow and total particle surface area contact with steam, further limiting mass transfer, in contrast to well-mixed systems with lower hindrance factors.

4.1.2. Acetic Acid. Using the estimated porosity values in Section 4.1.1, and assuming no reaction with the soluble coffee solids ($k_{on} = 0$), the acetic acid extraction curve was simulated and compared to that of Mateus et al.¹⁸ data, giving a significant underprediction of extraction (see Figure 5a).

The almost linear extraction behavior at the beginning of the process (Figure 5a, solid line) may suggest that the extraction of acetic acid is limited principally by its ability to partition into the headspace. The sensitivity of the model to Henry's law constant was thus studied

The ratio $\frac{\Delta H}{R}$ (see eq 12), was fitted to the experimental data¹⁸ available, resulting in an estimate that improved the model performance (i.e., simulated kinetics for acetic acid followed experimental trends, as shown in Figure 5b). This estimated

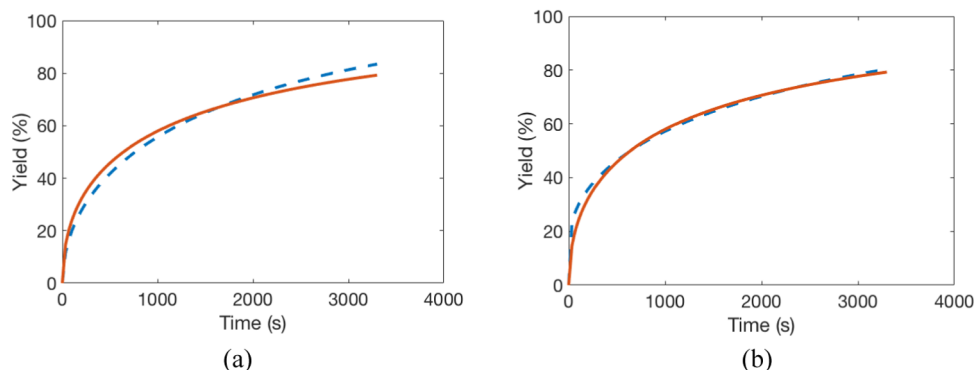


Figure 4. Comparison graphs between experimental data from Mateus et al.¹⁸ (solid line) and fitted model outputs (dashed line); acetaldehyde yield curves using (a) monodisperse coarse particles with $\epsilon_{cw} = 3.30\%$ and (b) bidisperse coarse and fine particles using $\epsilon_{cw} = 2.76\%$.

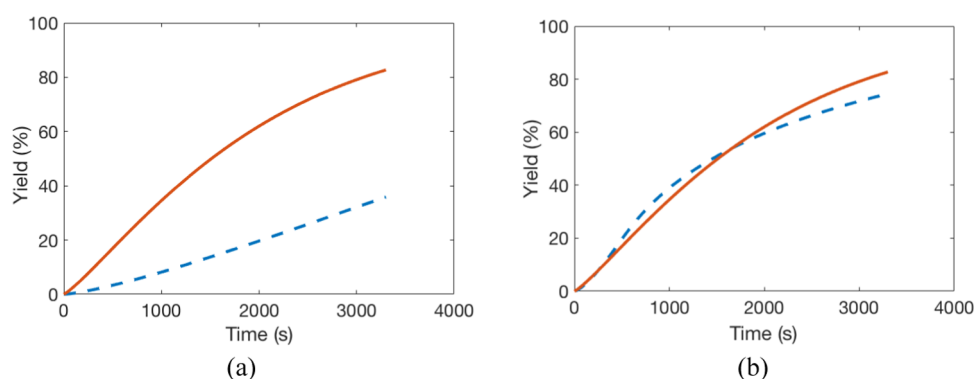


Figure 5. Comparison of experimental data from Mateus et al.¹⁸ (solid line) and model outputs (dashed line) for acetic acid yield curves using Henry's volatility constant values of $H_{298} = 0.025 \text{ Pa mol}^{-1} \text{ m}^3$ with (a) $\frac{\Delta H}{R} = -6400 \text{ K}^{-1}$ and (b) $\frac{\Delta H}{R} = -9408 \text{ K}^{-1}$ (estimated).

value (i.e., $\frac{\Delta H}{R} = -9408 \text{ K}^{-1}$) was 13% larger than the largest value given by Sander.³²

While partitioning acetic acid in water is well studied, its relationship with temperature is less consistently reported.³² There may also be interaction with minerals and other ions. Navarini and Rivetti⁵⁰ showed that many equilibria between carbonates and minerals exist in a coffee solution. It is feasible that species may neutralize acids but could act as a sink for H^+ that is accessed at later times. Alternatively, the "salting-out" effect of other soluble coffee species could have an effect.

When the model incorporates the fine particle fraction, little change in extraction behavior or yield was observed. Total extraction will, therefore, be limited by the flow rate of gas (replenishing the concentration gradient).

4.1.3. Pyridine. The extraction profile of pyridine displays a sharp plateau after about 10 min, after which no significant extraction occurs (see Figure 6).

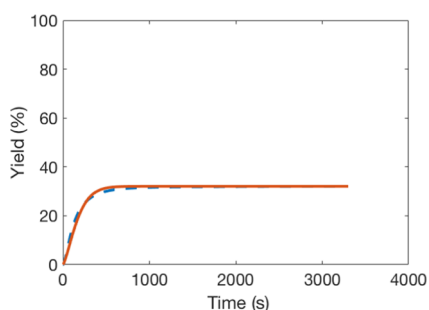


Figure 6. Comparison between experimental data from Mateus et al.¹⁸ (solid line) and fitted (dashed line) pyridine extraction curve with the estimated binding rate constant of $k_{\text{on}} = 1.9 \times 10^{-5} \text{ m}^3 \text{ s}^{-1} \text{ mol}^{-1}$.

Mateus et al.¹⁸ attributed this behavior to the protonation of pyridine, rendering it involatile, but at the pH of coffee solutions (ca. 5.3), 40% of pyridine should still be unprotonated and free to diffuse and vaporize. It is, instead, hypothesized that pyridine binds irreversibly to some phenolic components.

The binding rate constant k_{on} (see eq 31) was estimated by fitting the model for pyridine extraction.¹⁸ Simulations used the monodisperse particle distribution, the estimate for the cell wall porosity, and Henry's law volatility constant taken from the literature. The resulting estimate for the irreversible first-order binding constant was $k_{\text{on}} = 1.9 \times 10^{-5} \text{ m}^3 \text{ s}^{-1} \text{ mol}^{-1}$ (Figure 6).

Literature values of rate constants for aroma-binding reactions are rare, but Harrison and Hill⁴² gave model rates for aroma

release of the order of 10^0 – 10^{-3} m s^{-1} . However, these are different systems, and irreversible binding to phenolics will differ from reversible binding on proteins.

4.2. Large-Scale Column Simulations. The patent of Vitzthum and Koch²⁴ describes ranges of process parameters that may be employed to produce an aromatized distillate. The model can be used to explore how variation in the process parameters might affect the aroma profile of the distillate. A range of aromas with different physical and sensory properties were taken to study the effect of the stripping process (data in Table S1):

- Acetaldehyde—polar and volatile; high Henry's constant.
- Furanol—polar and nonvolatile; very low Henry's constant.
- 2-Furfurylthiol—apolar; high Henry's constant.
- Guaiacol—moderately apolar; moderate value of Henry's constant (a compound with no extreme properties, useful for comparison with other aromas).
- β -Damascenone—a strongly apolar compound with very high Henry's constant.
- Pyridine—representative of an aroma undergoing fast, irreversible binding.

Table 4 lists the process parameters of the reference system used for numerical simulation of the large-scale column.

The yield (y) is defined as the product of distillate concentration (c_{dist}) and volume (V_{dist})

Table 4. Parameters Used in the Simulation of Plant-Scale Operation^{5,24}

parameter	validation settings	parameter	validation settings
d_c	1800 μm	Z	2 m
d_{part}	100 μm	d_{bed}	0.625 m
φ_f	0.02	ΔP	50 kPa
$d_{3,2}$	1306 μm	K	$3.4 \times 10^{-10} \text{ m}^2$
Q	$0.263 \text{ m}^3 \text{ s}^{-1}$	Re_{N_2}	192
ϵ_p	0.42	Sc_{N_2}	0.807
ϵ_b	0.34	ϕ_{wc}	0.7
ρ_c	1337 kg m^{-3}	T_w	353 K
k_c	$0.13 \text{ W m}^{-1} \text{ K}^{-1}$	ϕ_{RCP}	0.6435
$C_{p,c}$	$1430 \text{ J kg}^{-1} \text{ K}^{-1}$	max solids extractable	0.3 w/w
σ_c	0.28	r_{por}	20 μm
σ_f	0.22	δ	1 μm
D_{pp}	$1 \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$	polyphenolic content	$0.63 \text{ g kg}_{\text{coffee}}^{-1}$
K_{pp}	10		

$$y = c_{\text{dist}} V_{\text{dist}} \quad (37)$$

where

$$V_{\text{dist}}(t) = \int Q_{\text{out}}(Z, t) \rho_{\text{st}}(Z) dt \quad (38)$$

$$c_{\text{dist}} = \frac{\int c_{\text{g}}(Z, t) Q_{\text{out}}(Z, t) dt}{V_{\text{dist}}(t)} \quad (39)$$

The steam density (ρ_{st}) is calculated assuming it is an ideal gas. If yield decreases with time, it shows that the extraction rate is slowing, perhaps as the aromas become stripped from the particle surface and the process becomes internally diffusion-limited.

4.2.1. Steaming Time. A simple variable to control is the time taken to complete the steam strip. The time available is limited by effects on downstream processes and plant capacity—if longer is spent steaming every batch of coffee, then plant productivity falls. However, yield (both aroma and soluble solids) must be balanced with throughput, and aroma yield is linked to product quality (by potentially being perceived as more aromatic). The concentration and yield results of simulating a 20 and 40 min steam strip are shown in Figure 7.

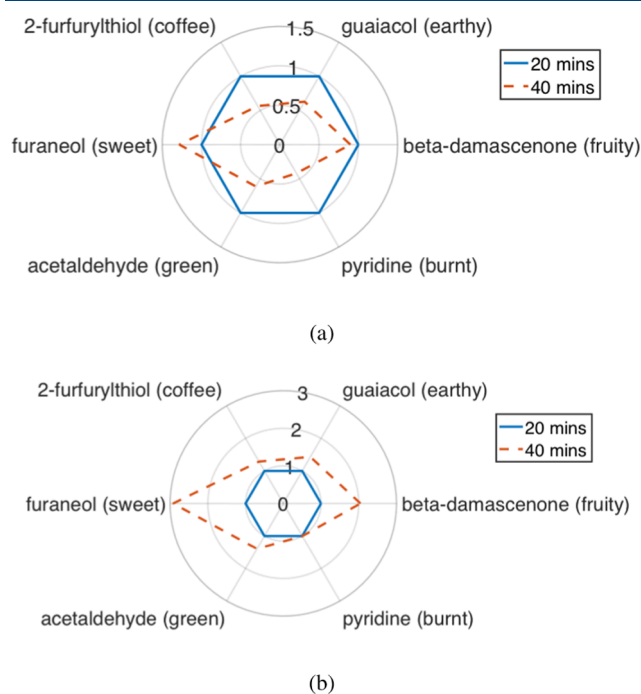


Figure 7. (a) Normalized concentration and (b) normalized yields (both normalized to values after 20 min of steam stripping) of some key aromas in the distillate when simulating 20 and 40 min steam stripping.

When the steaming time is doubled, the model predicts a dilution in many aromas to between 50 and 70% of the concentration obtained after 20 min. Figure 7a also shows that, for pyridine that is consumed by binding reactions with the coffee matrix, the concentration is <40% of the value after 20 min. In contrast, the very apolar compound (β -damascenone) is only slightly more dilute after 40 min, and a compound with a very low Henry's volatility partition coefficient (furaneol) is over 20% more concentrated at the longer time.

Yields shown in Figure 7b for 2-furfurylthiol, guaiacol, and acetaldehyde increase by between 20 and 40% after doubling the

steaming time. β -Damascenone and furaneol yields increase to beyond 200% and nearly 300%, respectively, showing that their rate of extraction is increasing over the 20–40 min time frame. Pyridine yield is almost unchanged, showing that by this point most have been either extracted or bound irreversibly to soluble coffee solids.

4.2.2. Types of Extraction Behavior. After simulating the behavior of aromas from all of the major aroma groups (see Table S1), we propose that the nonbinding compounds can be classified into three groups. Each group's extraction is limited by either (i) partition into the aqueous phase, (ii) internal diffusion, or (iii) partition into the steam phase. An example of each characteristic extraction kinetic is shown in Figure 8:

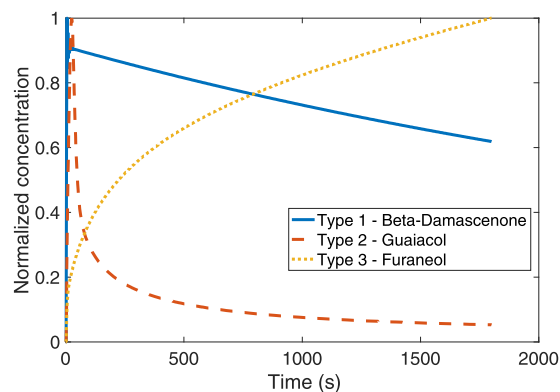


Figure 8. Normalized concentration of aromas in the exiting steam, demonstrating the limiting factors in extraction (time shown is after the column reaches saturation temperature).

- **Type 1**—low solubility in water. For very apolar compounds, such as β -damascenone, the aromas remain partitioned strongly in the mesopore oil layer; extraction is limited by the low solubility/partitioning in the aqueous phase. To enhance mass transfer, large internal concentration gradients between pore water and oil are needed. Over time scales of minutes, where the characteristic time for diffusion (R^2/D_h) is of the order of hours, only mesopore water within the surface of the particles is depleted and sets up these large concentration gradients. As a result, extraction is strictly limited by surface area, and distillate concentration slowly falls with time.
- **Type 2**—internal diffusion-controlled. Most compounds fall into the second category, as seen for acetaldehyde in the small-scale study (Figure 4a,b). After an initial peak in concentration in both water and steam phases as extraction from near the particle surface takes place, the aroma is quickly stripped out and concentration in the gas falls. Extraction is subsequently determined by internal diffusion.
- **Type 3**—low volatility. Some very polar compounds, such as furaneol, have very low Henry's law constants and remain strongly partitioned in the water phase. They quickly reach saturation concentration in the steam and their extraction is limited by how much steam can be contacted with the water in the time available. As a result, their concentration increases slowly as material accumulates in the water (increasing the saturation concentration in the steam). Over the time scale of practical steam stripping, this is the only effect to be seen; however,

eventually, diffusion limitations would lower the concentration.

Figure 7a shows that there is some difference between the type 2 aromas when comparing 20 and 40 min stripping times, based on diffusion coefficients. The most polar and fastest diffusing compound (acetaldehyde—see Section 4.2 and Table S1) has the smallest (25%) reduction in concentration at the longer time compared to the greater diffusional limitations of 2-furfurylthiol (48%) and guaiacol (44%), which have diffusivities in water 60–70% that of acetaldehyde. There is a smaller differentiation between these two slower-diffusing type 2 compounds. This is probably due to the difference in octanol–water partition coefficients, which contribute to the total internal resistance to extraction, even though the release from the oil does not dominate.

4.2.3. Water Addition. To achieve saturation of particles in the bed, increasing amounts of water can be added to the column. In real systems, water may not be evenly distributed in the column, so “excess” water can be added to ensure saturation in the wetted regions of the bed. This both adds thermal mass, which must be heated by the steam (extending the column heating time) but will also dilute the material that diffuses into the free water. Figure 9 shows the effect of doubling the water–

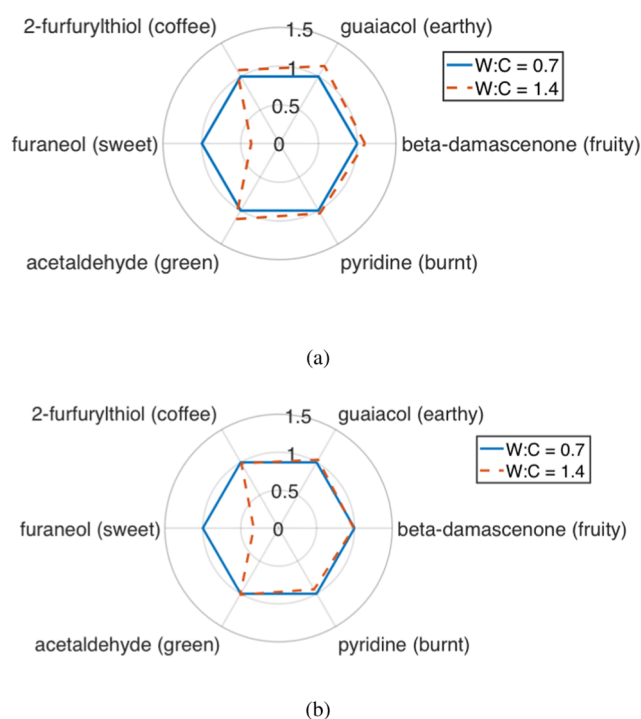


Figure 9. (a) Normalized concentrations and (b) yields (normalized to values obtained at a water-to-coffee ratio of 0.7) of some key aromas in the distillate when simulating water-to-coffee ratios of 0.7 and 1.4 prior to steam stripping.

coffee ratio from 0.7 to 1.4 before steam stripping. The effects of larger amounts of water addition can be seen in these simulation results. The addition of water increases the column heating time from 339 to 428 s, although the heating phase is still short in comparison to the process time.

Figure 9 shows that concentrations were largely unaffected except for the type 3 compound, furaneol (see Figure 9). The diluting effect of adding extra water into the column reduces the equilibrium gas-phase concentration and slows the mass transfer

from water to steam by reducing the concentration gradient. Pyridine concentration changes only slightly at higher water additions, suggesting that the binding reaction rate is not strongly dependent on dilution.

Yields of all compounds, however, fall with increasing water addition because of the dilution effect. Furaneol is the case, where the yield decreases significantly, reflecting the fall in concentration. This is probably due to the longer heating times associated with larger water additions. The longer the column takes to heat, the more time there is for binding reactions to occur prior to stripping from the column. Column heating time is therefore an important parameter for binding susceptible compounds.

4.2.4. Column Aspect Ratio. It has been assumed that there are no radial gradients in the bed. Changing the height of the coffee bed (either through different column fill settings or when developing new process equipment) will, however, affect the steam-stripping process.

By changing the height over which the pressure gradient is exerted, fluid flow will change. In addition, the ratio of aromas enriched through longer advection lengths to those that quickly reach saturation in the headspace due to poor headspace partitioning will change. The extremes of the height-to-diameter ratios (0.9:1 and 3.2:1), as described in the patent of Vitzthum and Koch,²⁴ are simulated, and the results are shown in Figure 10.

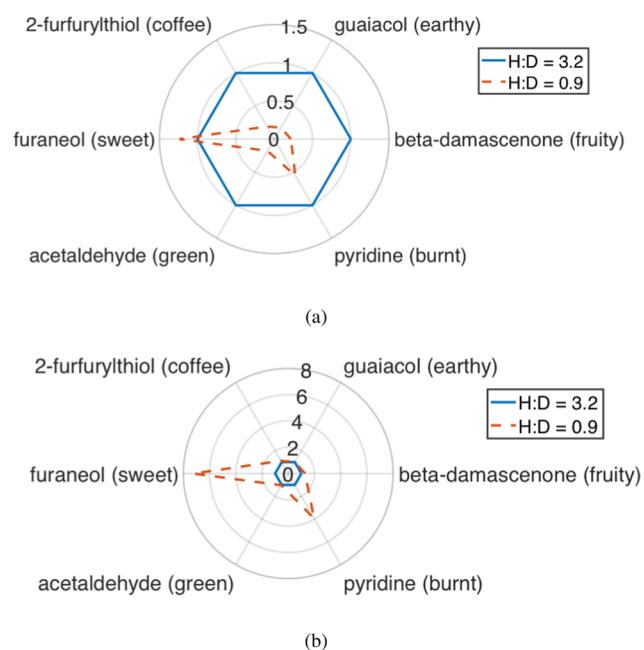


Figure 10. (a) Normalized concentration and (b) yields (normalized to the values at a height-to-diameter ratio of 3.2) of some key aromas in the distillate when simulating bed height-to-diameter ratios of 3.2 and 0.9.

Figure 10a shows that furaneol and pyridine are the compounds most affected by the faster-flowing steam. Although all compound yields increase with the faster flow (due to larger water-to-steam concentration gradients), the effect is particularly strong for type 3 compounds. For these, the water-to-gas limitation means that its yield (at these time scales) is largely determined by the amount of steam that can be contacted with the water in the given time period. For pyridine, reducing the

column heating time and faster extraction relative to the binding reaction speed increases the overall yield.

The yield of furaneol increases significantly. Figure 11 shows a plot of the concentration of furaneol within the two columns.

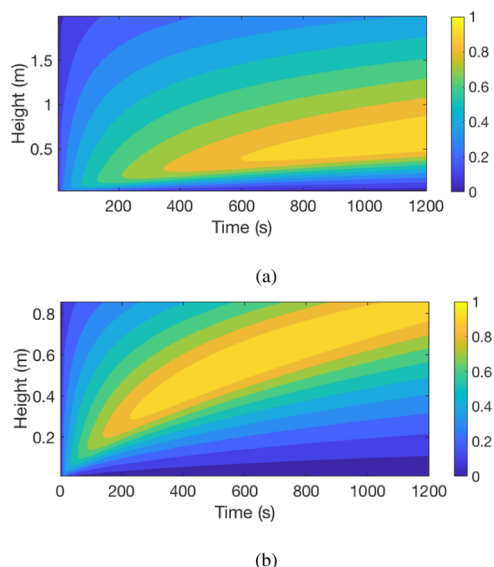


Figure 11. Contour plots of furaneol concentration in steam for columns of two height-to-diameter ratios (a) 3.2 and (b) 0.9. Data is normalized to the maximum concentration.

Steam quickly becomes saturated with furaneol toward the base of the column, but as the steam rises into the lower pressure (and cooler) part of the column, the material transfers back into the free water phase, lowering the concentration. By altering the aspect ratio, the shorter column, with its faster flow, does not allow as much time for this back-transfer to occur, and, as such, more material remains in the flowing steam and the distillate concentration increases. Figure 11b thus shows how the plume of high concentration reaches the top of the short column, while in the taller column, the stripping time is not long enough to extract much furaneol.

5. CONCLUSIONS

A model for the steam stripping of aromas has been built and validated against the published data. A study of a selection of key aroma compounds has been presented. The saturated coffee bed model can describe the gas-stripping extraction of some aromas at the small-scale well, having been validated against the published data. The extraction of acetic acid is least well described, and this is probably related to the as-yet-undescribed weak and reversible interactions with other coffee components.

The model has been used to study a range of representative aroma compounds. A classification of compounds can be made in terms of their partitioning and binding behavior. Type 1 aromas (strongly apolar compounds) fall in concentration slowly with increased steaming. At practical steam-stripping time scales, compounds with high Henry's law constants quickly become diffusion-limited (type 2), while those that partition poorly into the headspace are water-to-steam transfer-limited (type 3). Irreversibly binding compounds are best extracted quickly before being consumed by the reaction.

The model has been used to assess the significance of steaming time, and column geometry varies across the spectrum of aroma compounds. Further work will incorporate new

process variables and explore the capacity for process optimization. It should be noted that many compounds share physical and chemical properties to those simulated but have different sensory properties. Sensory perception is also not directly proportional to aroma concentration, so it should not be assumed that an increase in the concentration of a "sweet" compound will make for a sweeter instant coffee. There is some cross-over between the sensory attributes of the categories, so relating sensory attributes to process variables will be complex.

Further model development should include the steam stripping of dry and partially wet coffee. This must incorporate a wetting model, where condensing steam is absorbed and then enhances the extraction of certain compounds. Experimental results should validate the suitability of the aforementioned wetting model, the Carman–Kozeny and Darcy equations for steam flow, and the trends predicted in aroma extraction. This will lead to a more versatile and reliable predictive model for use in tailoring process conditions to the desired chemical and sensory outcomes.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.iecr.0c01153>.

Table S1 contains a comprehensive list of aroma compounds and corresponding chemical structures, octanol–water partition coefficients, boiling points, sensory notes, and Henry's law constant values at 298 K (PDF)

■ AUTHOR INFORMATION

Corresponding Author

Peter J. Fryer – School of Chemical Engineering, University of Birmingham, Birmingham B15 2TT, U.K.; orcid.org/0000-0003-4767-7839; Email: p.j.fryer@bham.ac.uk

Authors

David Beverly – Jacobs Douwe Egberts R&D, Banbury OX16 2QU, U.K.; School of Chemical Engineering, University of Birmingham, Birmingham B15 2TT, U.K.

Estefanía Lopez-Quiroga – School of Chemical Engineering, University of Birmingham, Birmingham B15 2TT, U.K.; orcid.org/0000-0003-2111-0121

Robert Farr – Jacobs Douwe Egberts R&D, Banbury OX16 2QU, U.K.

John Melrose – Jacobs Douwe Egberts R&D, Banbury OX16 2QU, U.K.; orcid.org/0000-0003-4234-9000

Sian Henson – Jacobs Douwe Egberts R&D, Banbury OX16 2QU, U.K.

Serafim Bakalis – Faculty of Engineering, University Park, Nottingham NG7 2RD, U.K.

Complete contact information is available at <https://pubs.acs.org/10.1021/acs.iecr.0c01153>

Author Contributions

D.B., methodology, software, formal analysis, writing (original and editing), and visualization; E.L.-Q., methodology, writing (review and editing), and supervision; R.F., methodology, writing (review and editing), and supervision; J.M., conceptualization, methodology, writing (review and editing), and supervision; S.H., conceptualization, supervision, writing (review and editing), and project administration; S.B., conceptu-

alization, supervision, writing (review and editing), project administration, and funding acquisition; and P.J.F., supervision, writing (review and editing), project administration, and funding acquisition.

Notes

The authors declare no competing financial interest.

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NOMENCLATURE

Symbol Parameter

a_{bp}	surface area per unit volume of bed pore occupied by the gas phase ($m^2 m_{bed\ pore}^{-3}$)
a_w	surface area per unit volume of water ($m^2 m^{-3}$)
$a_{por,p}$	oil–mesopore interfacial area per unit volume of mesopore ($m^2 m_{bed\ pore}^{-3}$)
$a_{por,do}$	oil–mesopore interfacial area per unit volume of oil ($m^2 m_{oil}^{-3}$)
a_b	surface area per unit volume of bed ($m^2 m^{-3}$)
A_{bed}	bed area (m^2)
c_w	aroma concentration in the free water ($mol m_{water}^{-3}$)
c_g	aroma concentration in the gas phase ($mol m_{bed\ pore}^{-3}$)
c_p	aroma concentration in the particle pore ($mol m_{particle\ pore}^{-3}$)
c_{do}	aroma concentration in the particle oil layer ($mol m_{particle\ pore}^{-3}$)
$c_{do,pp}$	phenolic concentration in the oil layer ($mol m_{particle\ pore}^{-3}$)
$c_{do,comp}$	bound complex concentration in the oil layer ($mol m_{particle\ pore}^{-3}$)
c_{dist}	distillate concentration ($mol m_{distillate}^{-3}$)
$C_{p,c}$	specific heat capacity of coffee ($J kg^{-1} K^{-1}$)
$C_{p,w}$	specific heat capacity of water ($J kg^{-1} K^{-1}$)
C_p	average specific heat capacity of the bed ($J kg^{-1} K^{-1}$)
d_c	mean coarse particle size (μm)
d_f	mean fine particle size (μm)
$d_{b,pore}$	bed pore diameter (m)
d_{part}	particle diameter (m)
d_{pore}	mesopore diameter (m)
d_{bed}	coffee bed diameter (m)
$d_{3,2}$	Sauter mean diameter (μm)
D_0	aroma free diffusivity in water at 298 K ($m^2 s^{-1}$)
D_{eff}	effective diffusivity ($m^2 s^{-1}$)
D_h	hindered diffusivity in cell wall region ($m^2 s^{-1}$)
D_{pp}	polyphenol free diffusivity in water at 298 K ($m^2 s^{-1}$)
$\Delta H/R$	van't Hoff slope (K)
H_f	hindrance factor
k_w	water-phase mass transfer coefficient ($m s^{-1}$)
k_g	gas-phase mass transfer coefficient ($m s^{-1}$)
k_c	thermal conductivity of wetted coffee ($W m^{-1} K^{-1}$)
k_{por}	oil-phase mass transfer coefficient in the pore ($m s^{-1}$)
k_{on}	bonding rate constant ($m^3 s^{-1} mol^{-1}$)
K	bed permeability (m^2)
$K_{o/w}$	octanol–water partition coefficient
K_{pp}	polyphenol octanol–water partition coefficient
K_H^c	Henry's law volatility constant ($Pa mol^{-1} m^3$)

$x_{s,p}$	mass fraction of coffee solids ($kg_{soluble\ solids} kg_{solution}^{-1}$)
m_0	mass fraction of aroma ($kg_{aroma} kg_{coffee}^{-1}$)
\dot{m}_{con}	rate of condensation ($kg s^{-1}$)
M_{vap}	molar mass of water vapor ($kg mol^{-1}$)
P_{in}	inlet pressure (Pa)
P_{out}	outlet pressure (Pa)
p_{st}	saturated steam pressure (Pa)
p_{vap}	free water vapor pressure (Pa)
ΔP	pressure drop (Pa)
Q	gas flow rate ($m^3 s^{-1}$)
Q_{out}	gas flow rate out of a column element ($m^3 s^{-1}$)
r_{por}	mesopore radius (μm)
Re	Reynolds number
r_c	coarse particle radius (m)
r_f	fine particle radius (m)
R	gas constant ($J K^{-1} mol^{-1}$)
Sc	Schmidt number
Sh	Sherwood number
T_w	added water temperature (K)
T_{st}	saturated steam temperature (K)
T_b	bed temperature (K)
T_c	coffee particle temperature (K)
v	steam superficial velocity ($m s^{-1}$)
V_{bed}	bed volume (m^3)
V_{por}	specific mesopore volume of pore per unit mass of coffee ($mm^3 g^{-1}$)
V_{dist}	volume of distillate (m^3)
y_{dist}	yield of aroma (mol)
Z	coffee bed height (m)
δ	oil layer thickness (μm)
ϵ_p	particle porosity
ϵ_b	bed porosity
ϵ_{mp}	mesoporosity
ϵ_{cw}	cell wall porosity
μ_{in}	gas viscosity at the inlet (Pa s)
μ	liquid viscosity (Pa s)
$\rho_{b,w}$	free water in the bed ($kg m_{bed}^{-3}$)
ρ_p	coffee particle density ($kg m^{-3}$)
ρ_w	water density ($kg m^{-3}$)
ρ_{st}	saturated steam density ($kg m^{-3}$)
$\bar{\rho}_b$	averaged bed density ($kg m_{bed}^{-3}$)
ρ_c	intrinsic coffee density ($kg m^{-3}$)
σ_c	coarse particle distribution variance
σ_f	fine particle distribution variance
ϕ_{wc}	added water-to-coffee ratio ($kg_w kg_{coffee}^{-1}$)
ϕ_{RCP}	maximum packing fraction of monodisperse spheres
ψ_{max}	maximum packing fraction
φ_c	coarse particle fraction
φ_f	fine particle fraction
ω	mass fraction of large spheres on the total particle volume

REFERENCES

- (1) Farah, A. Coffee as a Speciality and Functional Beverage. In *Functional and Speciality Beverage Technology*; Paquin, P., Ed.; Woodhead Publishing Series in Food Science, Technology and Nutrition; Woodhead Publishing: Cambridge, 2009; pp 370–395.
- (2) International Coffee Organization, ICO. *Coffee Market Continues Downward Trend*, Coffee Market Report; ICO, 2018, <http://www.ico.org/documents/cy2017-18/cmr-0318-e.pdf> (accessed April 12, 2018).
- (3) International Coffee Organization, ICO. *Trends in Coffee Consumption in Selected Importing Countries*, International Coffee Council, 109th Session, 24–28 Sept 2012, London, United Kingdom,

2012. <http://www.ico.org/documents/icc-109-8e-trends-consumption.pdf> (accessed June 21, 2018).
- (4) Buffo, R. A.; Cardelli-Freire, C. Coffee Flavour: An Overview. *Flavour Fragrance J.* **2004**, *19*, 99–104.
- (5) Corrochano, B. R.; Melrose, J.; Bentley, A. C.; Fryer, P. J.; Bakalis, S. A New Methodology to Estimate the Steady-state Permeability of Roast and Ground Coffee in Packed Beds. *J. Food Eng.* **2015**, *150*, 106–116.
- (6) Mahlmann, J. P. Method of Producing Aromatized Coffee Extract. US3,132,947, May 12, 1964.
- (7) Grubbs, D. R. Process for Steaming Coffee. European Patent EP0086526A3, Oct 3, 1984.
- (8) Furrer, M.; Gretschi, C. Coffee Aroma Recovery Process. European Patent EP1211948B1, Sept 26, 2012.
- (9) Usseglio, M.; Salvadori, V.; Siri, G. Advanced Modeling of Vegetable Oils Steam Stripping with Structured Packing Columns. *Comput. Chem. Eng.* **2019**, *121*, 654–669.
- (10) Cerpa, M. G.; Mato, R. B.; Cocero, M. J. Modeling Steam Distillation of Essential Oils: Application to Lavandin Super Oil. *AIChE J.* **2008**, *54*, 909–917.
- (11) Schenker, S. Investigations on the Hot Air Roasting of Coffee Beans. Ph.D. Thesis, ETH Zurich, 2000.
- (12) Zhang, C.; Linforth, R.; Fisk, I. D. Cafestol Extraction Yield from Different Coffee Brew Mechanisms. *Food Res. Int.* **2012**, *49*, 27–31.
- (13) Couto, R. M.; Fernandes, J.; Gomes da Silva, M. D. R.; Simões, P. C. Supercritical Fluid Extraction of Lipids from Spent Coffee Grounds. *J. Supercrit. Fluids* **2009**, *51*, 159–166.
- (14) Mateus, M. L.; Rouvet, M.; Gumy, J. C.; Liardon, R. Interactions of Water with Roasted and Ground Coffee in the Wetting Process Investigated by a Combination of Physical Determinations. *J. Agric. Food Chem.* **2007**, *55*, 2979–2984.
- (15) Mateus, M. L.; Champion, D.; Liardon, R.; Voilley, A. Characterization of Water Mobility in Dry and Wetted Roasted Coffee Using Low-field Proton Nuclear Magnetic Resonance. *J. Food Eng.* **2007**, *81*, 572–579.
- (16) Mestdagh, F.; Davidek, T.; Chaumonteuil, M.; Folmer, B.; Blank, I. The Kinetics of Coffee Aroma Extraction. *Food Res. Int.* **2014**, *63*, 271–274.
- (17) Sánchez López, J. A.; Wellinger, M.; Gloess, A. N.; Zimmermann, R.; Yeretian, C. Extraction Kinetics of Coffee Aroma Compounds Using a Semi-automatic Machine: On-line Analysis by PTR-ToF-MS. *Int. J. Mass Spectrom.* **2016**, *401*, 22–30.
- (18) Mateus, M. L.; Lindinger, C.; Gumy, J. C.; Liardon, R. Release Kinetics of Volatile Organic Compounds from Roasted and Ground Coffee: Online Measurements by PTR-MS and Mathematical Modelling. *J. Agric. Food Chem.* **2007**, *55*, 10117–10128.
- (19) Guichard, E. Interaction of Aroma Compounds with Food Matrices. In *Flavour Development, Analysis and Perception in Food and Beverages*; Parker, J. K.; Elmore, S.; Methven, L.; José, M., Eds.; Woodhead Publishing Series in Food Science, Technology and Nutrition; Woodhead Publishing: Cambridge, 2015; pp 273–295.
- (20) Xavier, V. B.; Vargas, R. M. F.; Cassel, E.; Lucas, A. M.; Santos, M. A.; Mondin, C. A.; Santarem, E. R.; Astarita, L. V.; Sartor, T. Mathematical Modelling for Extraction of Essential Oil from *Baccharis* spp. by Steam Distillation. *Ind. Crop. Prod.* **2011**, *33*, 599–604.
- (21) Moroney, K. M.; Lee, W. T.; O'Brien, S. B. G.; Suijver, F.; Marra, J. Modelling of Coffee Extraction During Brewing Using Multiscale Methods: An Experimentally Validated Model. *Chem. Eng. Sci.* **2015**, *137*, 216–234.
- (22) Melrose, J. R.; Corrochano, B.; Montoya-Guerra, M.; Bakalis, S. Towards a New Brewing Control Chart for the 21st Century. *J. Agric. Food Chem.* **2018**, *66*, 5301–5309.
- (23) Beverly, D.; Fryer, P. J.; Bakalis, S.; Lopez-Quiroga, E.; Farr, R. Mathematical Modelling of the Steam Stripping of Aroma from Roast and Ground Coffee. *Energy Procedia* **2019**, *161*, 157–164.
- (24) Vitzthum, O.; Koch, K. D. Process for the Preparation of Soluble Coffee. European Patent EP0489401B1, June 10, 1992.
- (25) Amézqueta, S.; Subirats, X.; Fuguet, E.; Rosés, M.; Ràfols, C. Octanol–Water Partition Constant. In *Liquid-Phase Extraction*; Poole, C. F., Ed.; Handbooks in Separation Science; Elsevier, 2020; pp 183–208.
- (26) Sangster, J. Octanol-Water Partition Coefficients of Simple Organic Compounds. *J. Phys. Chem. Ref. Data* **1989**, *18*, 1111–1227.
- (27) Noubigh, A.; Mgaidi, A.; Abderrabba, M. Temperature Effect on the Distribution of Some Phenolic Compounds: An Experimental Measurement of 1 Octanol/Water Partition Coefficients. *J. Chem. Eng. Data* **2010**, *55*, 488–491.
- (28) Corrochano, B. R. Advancing the Engineering Understanding of Coffee Extraction. Eng.D. Thesis, University of Birmingham, 2017.
- (29) Barrett, J.; Clement, C. Kinetic Evaporation and Condensation Rates and Their Coefficients. *J. Colloid Interface Sci.* **1992**, *150*, 352–364.
- (30) Farr, R.; Groot, R. Close Packing Density of Polydisperse Hard Spheres. *J. Chem. Phys.* **2009**, *131*, No. 244104.
- (31) International Association for the Properties of Water and Steam, IAPWS. *Knovel Steam Tables*; Knovel Corporation, 2018. <https://app.knovel.com/web/toc.v/cid:kpKST00001/viewerType:toc/> (accessed June 28, 2018).
- (32) Sander, R. Compilation of Henry's Law Constants (Version 4.0) for Water as a Solvent. *Atmos. Chem. Phys.* **2015**, *15*, 4399–4981.
- (33) Wieland, F.; Neff, A.; Gloess, A. N.; Poisson, L.; Atlan, S.; Larrain, D.; Prêtre, D.; Blank, I.; Yeretian, C. Temperature Dependence of Henry's Law Constants: An Automated, High-throughput Gas Stripping Cell Design Coupled to PTR-ToF-MS. *Int. J. Mass Spectrom.* **2015**, *387*, 69–77.
- (34) Rigaki, A.; Webb, C.; Theodoropoulos, C. Double substrate limitation model for the bio-based production of succinic acid from glycerol. *Biochem. Eng. J.* **2020**, *153*, No. 107391.
- (35) Harned, H. S.; Ehlers, R. W. The Dissociation Constant of Acetic Acid from 0 to 60° Centigrade. *J. Am. Chem. Soc.* **1933**, *55*, 652–656.
- (36) Carberry, J. A Boundary-Layer Model of Fluid-Particle Mass Transfer in Fixed Beds. *AIChE J.* **1960**, *6*, 460–462.
- (37) Wakao, N.; Funazkri, T. Effect of Fluid Dispersion Coefficients on Particle-to-fluid Mass Transfer Coefficients in Packed Beds. *Chem. Eng. Sci.* **1978**, *33*, 1375–1384.
- (38) Crank, J. *The Mathematics of Diffusion*, 2nd ed.; Clarendon Press: Oxford, 1975.
- (39) Shen, L.; Chen, Z. Critical Review of the Impact of Tortuosity on Diffusion. *Chem. Eng. Sci.* **2007**, *62*, 3748–3755.
- (40) Yaws, C. L. *Yaws' Handbook of Thermodynamic and Physical Properties of Chemical Compounds: Physical, Thermodynamic and Transport Properties for 5,000 Organic Chemical Compounds*; Knovel Corporation, 2003. <https://app.knovel.com/hotlink/toc/id:kpYHTPPCC4/yaws-handbook-thermodynamic/yaws-handbook-thermodynamic>.
- (41) Sobolík, V.; Žitný, R.; Tovcigrečko, V.; Delgado, M.; Allaf, K. Viscosity and Electrical Conductivity of Concentrated Solutions of Soluble Coffee. *J. Food Eng.* **2002**, *51*, 93–98.
- (42) Harrison, M.; Hills, B. Mathematical Model of Flavor Release from Liquids Containing Aroma-Binding Macromolecules. *J. Agric. Food Chem.* **1997**, *45*, 1883–1890.
- (43) Wang, X.; Lim, L.-T. Physicochemical Characteristics of Roasted Coffee. In *Coffee in Health and Disease Prevention*; Preedy, V. R., Ed.; Academic Press, 2015; pp 247–254.
- (44) Pedras, B. M.; Nascimento, M.; Sá-Nogueira, I.; Simões, P.; Paiva, A.; Barreiros, S. Semi-continuous Extraction/Hydrolysis of Spent Coffee Grounds with Subcritical Water. *J. Ind. Eng. Chem.* **2019**, *72*, 453–456.
- (45) Schenker, S.; Handschin, S.; Frey, B.; Perren, R.; Escher, F. Pore Structure of Coffee Beans Affected by Roasting Conditions. *J. Food Sci.* **2000**, *65*, 452–457.
- (46) Schultz, S. G.; Solomon, A. K. Determination of the Effective Hydrodynamic Radii of Small Molecules by Viscometry. *J. Gen. Physiol.* **1961**, *44*, 1189–1199.
- (47) Spiro, M.; Chong, Y. Y. The Kinetics and Mechanism of Caffeine Infusion from Coffee: the Temperature Variation of the Hindrance Factor. *J. Sci. Food Agric.* **1997**, *74*, 416–420.

- (48) Jaganyi, D.; Madlala, S. P. Kinetic of Coffee Infusion: a Comparative Study on the Extraction Kinetics of Mineral Ions and Caffeine from Several Types of Medium Roasted Coffees. *J. Sci. Food Agric.* **2000**, *80*, 85–90.
- (49) Oosting, E. M.; Gray, J. I.; Grulke, E. A. Correlating Diffusions Coefficients in Concentrated Carbohydrate Solutions. *AIChE J.* **1985**, *31*, 773–780.
- (50) Navarini, L.; Rivetti, D. Water quality for Espresso coffee. *Food Chem.* **2010**, *122*, 424–428.