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# 1 Measuring biomechanical loads in team sports – from lab to field

2

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20 **ABSTRACT**

21 The benefits of differentiating between the physiological and biomechanical load-response pathways in football  
22 and other (team) sports have become increasingly recognised. In contrast to physiological loads however, the  
23 biomechanical demands of training and competition are still not well understood, primarily due to the difficulty  
24 of quantifying biomechanical loads in a field environment. Although musculoskeletal adaptation and injury are  
25 known to occur at a tissue level, several biomechanical load metrics are available that quantify loads experienced  
26 by the body as a whole, its different structures and the individual tissues that are part of these structures. This  
27 paper discusses the distinct aspects and challenges that are associated with measuring biomechanical loads at  
28 these different levels in laboratory and/or field contexts. Our hope is that through this paper, sport scientists and  
29 practitioners will be able to critically consider the value and limitations of biomechanical load metrics and will  
30 keep pursuing new methods to measure these loads within and outside the lab, as a detailed load quantification is  
31 essential to better understand the biomechanical load-response pathways that occur in the field.

32

33 **INTRODUCTION**

34 Optimal sports performance with minimal injury risk is largely determined by the training an athlete has been  
35 exposed to. Whilst sufficient training loads are required to achieve beneficial physical adaptations for enhanced  
36 performance in the form of improved fitness, excessive loading can introduce fatigue and is known to increase  
37 the risk of injury [1,2]. Training loads are, therefore, widely measured and monitored in football and other (team)  
38 sports, with the aim to better control training prescription and optimise load-response pathways. On the one hand  
39 there is a physiological load-response pathway, where the metabolic challenge to maintain powerful and  
40 prolonged skeletal muscle contractions triggers a broad range of biochemical responses in the body, primarily in  
41 the form of metabolic and cardiorespiratory adaptations [3,4]. On the other hand, there is a biomechanical load-  
42 response pathway, where the mechanical challenges to withstand high forces repetitively applied to the  
43 musculoskeletal system triggers mechanobiological tissue responses of the muscles, tendons, ligaments, bones  
44 and articular cartilage [5–7]. There is a growing belief that monitoring the physiological and biomechanical  
45 loads separately can contribute to the holistic understanding of an athlete’s adaptive mechanisms that ultimately  
46 determine their physical fitness and performance outcomes [8]. However, in contrast to a considerable  
47 understanding of the physiological branch, the extent to which (team) sports imposes loads on the  
48 musculoskeletal system and triggers mechanobiological responses that make the tissues stronger or weaker are  
49 relatively under-investigated and not well understood.

50 A major issue that limits the progress in understanding biomechanical load-response pathways, is that measuring  
51 *in vivo* biomechanical loads to the musculoskeletal system as a whole, to the various structures within it, and to  
52 the tissues making up those structures, remains very difficult or even impossible with the current technologies,  
53 especially in a field-based context. Our aim was therefore 1) to provide an overview of biomechanical load  
54 metrics at different levels, 2) to discuss current methods and challenges for measuring *in vivo* biomechanical  
55 loads, and 3) suggest future considerations and avenues to be explored to enhance field-based biomechanical  
56 load monitoring.

57 **TISSUE LOADS**

58 During training and match-play in football and other (team) sports, the different hard- and soft-tissues of the  
59 body are exposed to an array of forces. These forces cause mechanical tension within the tissues in the form of  
60 stresses and strains that, together with exercise-induced microdamage and metabolic stress, trigger remodelling  
61 and repair responses. Examples of such adaptations include alterations in muscle architecture [9,10], changes in  
62 tendon stiffness and structure [11–14], and increased bone mass and mineral density [15,16], which are generally

63 considered desirable characteristics for enhanced performance (e.g. higher force production, increased storage  
64 and return of elastic energy). Excessive exposure to stresses and strains on the other hand, can outpace repair  
65 mechanisms and cause an accumulation of micro-damage that weakens the tissues over time. This progressive  
66 weakening can ultimately lead to mechanical fatigue and tissue failure, such as muscle tears, tendon rupture or  
67 bone fractures [17,18]. The optimal loading thresholds of individual tissues depend on many factors, including  
68 tissue properties and loading history. In an ideal world one would thus want to quantify and monitor the  
69 accumulation of tissue-specific stresses and strains over time.

70 From a mechanical perspective stress and strain can be defined as the force acting per unit surface area and the  
71 resulting relative tissue deformation, respectively. This direct relationship between force, stress and strain allows  
72 for *in vitro* experiments to be performed to investigate tissue adaptative or failure responses to predefined  
73 biomechanical loads [19,20]. Such experiments can provide a detailed insight into tissue behaviour under  
74 specific loading conditions, but require highly controlled laboratory setups, homogeneous tissue specimens and  
75 strictly constant or repetitive loading patterns. As an alternative, advanced computational modelling approaches  
76 (e.g. finite element analysis) can be used to accurately predict stress and strain distributions throughout tissues *in*  
77 *silico*, and investigate their response mechanisms under different mechanical and biological conditions [21,22].  
78 However, there is extensive physiological, structural and morphological variability within musculoskeletal  
79 structures, and during sports movements tissues are exposed to highly varying non-uniform tensile, compressive  
80 and shear forces. This makes it difficult to translate findings from controlled *in vitro* and/or *in silico* studies to  
81 the field, beyond understanding the expected stress-related deformations and stress tolerances of individual  
82 tissues. Although biomechanical responses to training loads are thus known to take place at a tissue level, the  
83 quantification of tissue-specific loads is primarily restricted to laboratory environments only (Figure 1).

84 -----

85 *Figure 1 around here*

86 -----

## 87 **STRUCTURAL LOADS**

88 Much research has investigated loads experienced by the musculoskeletal system at a structural level. Individual  
89 organs (e.g. muscles, tendons, ligaments, bones) or a combination thereof (e.g. joints, segments, limbs) form  
90 structures on which forces and moments act. These structural loads thus describe the combination of stresses and  
91 strains working on the individual tissues comprised by the structure. Net moments about the knee joint structure

92 for example, can be used as an indicator of loading magnitude and injury risk of the anterior cruciate ligament  
93 [23,24]. Likewise, measures of joint or leg stiffness, which is the resistance of a structure to withstand the forces  
94 acting on it, have been demonstrated to be sensitive to training status [25], running speed [26] and exercise-  
95 induced fatigue [27,28] (see [29] for an extensive discussion of the use of stiffness measures in sports).  
96 Quantifying structure-specific loading parameters can thus be informative for evaluating the risk of injury or  
97 biomechanical adaptations to training.

98 To indirectly estimate the *in vivo* loads acting on individual structures, including bone and muscle-tendon forces,  
99 and joint moments, reaction forces and stiffness parameters, musculoskeletal modelling techniques can be used  
100 [30,31]. Although such approaches are traditionally laborious and time consuming, recent advancements have  
101 shown the potential for real-time analysis of joint forces and moments, as well as muscle-tendon forces [32–35].  
102 The downside of these methods however, is that they are strongly dependent on kinematic (motion-capture  
103 systems), kinetic (force platforms) and/or neuromuscular (electromyography) input, the combination of which is  
104 yet largely restricted to laboratories. Several studies have, therefore, aimed to directly measure the *in vivo*  
105 structure-specific loads. Surgically implanted force transducers or strain gauges may, for example, be used to  
106 measure muscle-tendon forces [36–38] or bone strains [39] for walking, running and jumping activities, but their  
107 invasive and temporary nature makes the use of implants unsuitable for large-scale human experiments, let alone  
108 day-to-day load monitoring in the field. Very recently, a wearable tensiometer device has shown promising  
109 results for non-invasively assessing mechanical properties and loading of superficial tendons [40], and could be a  
110 first step towards the direct and field-based measurement of structure-specific loads. The difficulty of directly  
111 measuring structural forces has also led to the exploration of various indicators (or surrogate measures) of  
112 structural load. Tibial accelerations measured from shank-mounted accelerometers for example, have been  
113 suggested to provide a valid, reliable and simple field-based indicator of tibial loading [41–43], but it remains  
114 uncertain if tibial accelerations are related to the actual forces, stresses and strains experienced by the bone [44].  
115 In short therefore, despite the availability of several techniques to quantify structural loads directly or indirectly,  
116 their application is still primarily bound to a lab context (Figure 1).

## 117 **WHOLE-BODY LOADS**

118 Besides internal stresses and strains that are experienced by specific tissues and/or structures, the body as a  
119 whole is exposed to external forces. These external loads are primarily caused by interactions with other athletes  
120 (e.g. during tackling), equipment (e.g. kicking or hitting a ball) or the ground. Ground reaction forces (GRFs)  
121 following from foot-ground interactions especially, both drive and are affected by muscular actions, and

122 contribute to impact forces experienced by individual structures. GRFs thus describe the biomechanical loading  
123 experienced by the musculoskeletal system as a whole and have been investigated extensively for their potential  
124 association with running performance features [45–47] or specific overuse related pathologies [48–50]. Such  
125 relationships remain ambiguous though [48,50] and GRF may even be a poor predictor of the loads experienced  
126 at a structural level [20,49].

127 Whilst GRF alone unlikely suffices as a source of information for the prevention or treatment of particular  
128 tissue- or structure-specific pathologies, GRF can still provide a generic indicator of cumulative loading of the  
129 musculoskeletal system as a whole. In contrast to tissue- and structure-specific loads, GRFs can be measured  
130 relatively easily and non-invasively from force platforms. Unfortunately, force platforms are not suitable for  
131 sport-specific training and competition environments, and different approaches have been explored to estimate  
132 GRF from wearable devices in the field. Probably the most intuitive method is by using instrumented insoles,  
133 which are typically worn in or under the shoe and provide a summed measure of the pressure that the foot exerts  
134 on the ground [51]. Although pressure insoles can estimate GRF for running and jumping fairly well [52–56],  
135 their compromised accuracy for high-intensity movements [52,54–56] and practical limitations (e.g. movement  
136 restrictions, added mass in the shoe, discomfort) [51], leaves the feasibility of using insoles for monitoring GRF  
137 on a large-scale in the field currently still questionable.

138 Based on the relationship between force and acceleration according to Newton's second law ( $F=m \cdot a$ ), segmental  
139 movements may be used to indirectly estimate GRF [57–59]. Currently popular body-worn accelerometers have,  
140 therefore, received special attention for their potential to measure GRF in this manner [41,60–65]. Several  
141 studies have, however, demonstrated that either whole GRF waveforms [60–62], or even specific GRF features  
142 [41,61,63], cannot be estimated well from individual trunk-, pelvis- or shank-mounted accelerometers. In fact,  
143 the majority of segmental accelerations are likely required to accurately estimate GRF [57,58], making the use of  
144 one or even a combination of several accelerometer units to predict GRF probably insufficient.

145 Besides GRF, other accelerometry-based metrics have been suggested to assess whole-body loading, including  
146 vertical stiffness [66–68] and cumulative acceleration metrics [69–74]. Vertical stiffness is assumed to represent  
147 the whole-body response to the dynamic external forces and may be used to assess neuromuscular fatigue and  
148 performance after different types of training [67,68]. Likewise, cumulative acceleration metrics (e.g.  
149 PlayerLoad™, New Body Load, Dynamic Stress Load, Force Load [69–74]) are thought to provide an indication  
150 of the accumulated external impacts the body is exposed to. However, the premise underpinning these metrics

151 that accelerations of individual segments appropriately represent the whole-body acceleration is probably not  
152 valid [60], while evidence for a relationship with loads acting on a structural or tissue level is yet lacking. As  
153 such, if associations between any of these metrics and performance improvements or increased injury risk are  
154 observed, this does not provide an explanation for the underlying mechanisms of such associations. In other  
155 words, although GRF, stiffness or accelerometry-derived metrics offer field-based methods to quantify whole-  
156 body loading (Figure 1), their relevance and intrinsic value for assessing load-response pathways at a structural  
157 or tissue level remains to be determined.

## 158 **FROM LAB TO FIELD**

159 A big hurdle for translating research into the biomechanical load-response pathways from the lab to the field is  
160 the difficulty of quantifying biomechanical loads. This is primarily due to the lack of means to accurately  
161 measure biomechanical information in an athlete's natural training and/or competition environment (e.g. a  
162 football pitch). Recent developments have, however, demonstrated that such information might become more  
163 easily available in applied sport settings in the near future. For example, full-body wireless inertial sensor suits  
164 have been shown to be a reliable and valid method to simultaneously measure kinematic information of all body  
165 segments outside the laboratory (e.g. Xsens MVN [75]), and can already provide GRF and joint moment  
166 estimates during stereotypical activities such as walking [76,77]. To overcome discomfort and movement  
167 restriction issues associated with the use of multiple body-worn devices, markerless motion capture techniques  
168 are a non-invasive method for measuring different biomechanical variables in various sport environments [78–  
169 83]. These techniques may in the future allow for load metrics to be estimated at different levels. If for example,  
170 information from body-worn sensors or markerless motion capture can be used to accurately estimate GRF  
171 [58,84], the combination of kinematics and GRF may eventually be used to estimate structure-specific loading  
172 and thus open the door to field-based measurements and monitoring of internal biomechanical loads.

173 Given the often-limited availability of information in day-to-day football environments (as well as other applied  
174 sports settings), estimating biomechanical loads using conventional mechanical methods that attempt to directly  
175 measure load is not always possible. An imminent area in sports biomechanics that overcomes this issue is the  
176 use of advanced machine learning approaches to identify and/or predict biomechanical variables of interest [85].  
177 For example, neural network methods have been used to predict GRF and moments [86,87] and joint forces [88]  
178 from body-worn inertial sensors for different running tasks. Although these studies show promising results,  
179 interpreting the underlying biomechanical mechanisms of the predicted variable can be difficult [85,89], which  
180 could limit their application for e.g. explaining adaptation criteria or injury mechanisms. If similar techniques



181 can be used to accurately predict tissue- or structure-specific forces however, this may enable large-scale and  
182 non-invasive internal load monitoring in the field.

183 To effectively investigate and describe biomechanical load-response pathways in the field, the relevance of  
184 metrics used to quantify loads acting on the musculoskeletal system, as well as the outcome measures against  
185 which these loads are validated, should be considered. Popular body-worn sensor technologies especially, have  
186 opened the door for relatively easy measurements of several indicators of whole-body loading, but the applied  
187 researcher or practitioner should be reminded that their relationship with established tissue or structural load  
188 metrics, or their relevance in the context of the adaptive or injury mechanisms, has not been validated. For  
189 example, changes observed at a whole-body level (e.g. technique changes in a fatigued state) can be insightful  
190 when assessing generic whole-body adaptations to training but as yet, cannot be used to directly infer on load-  
191 response pathways experienced by individual tissues or structures. Therefore, careful validation is required for  
192 such field-based metrics against measures of tissue and/or structural responses (e.g. from tissue biopsies or  
193 ultrasound scanning) to establish the relationships between available biomechanical load metrics and the  
194 adaptive or injury mechanisms occurring at internal levels.

## 195 **CONCLUSION**

196 Biomechanical load-response pathways can be explained at different levels of the musculoskeletal system. Due  
197 to the currently limited availability of field-based biomechanical load metrics, enhancing our understanding of  
198 what biomechanical load metrics can and cannot be used for is essential. Our hope is that through this paper,  
199 sport scientists and practitioners alike will revisit their views on the value and limitations of biomechanical load  
200 metrics at different levels. Moreover, we would like to encourage sport scientists and biomechanics researchers  
201 to keep pursuing ways to overcome the challenges of measuring these loads within and outside the lab, as a  
202 detailed quantification of biomechanical loads experienced during football and other (team) sports is essential to  
203 further understand the *in vivo* biomechanical load-response pathways and ultimately monitor them in the field.

204

205 **REFERENCES**

- 206 1 Eckard TG, Padua DA, Hearn DW, *et al.* *The Relationship Between Training Load and Injury in*  
207 *Athletes: A Systematic Review.* Springer International Publishing 2018. doi:10.1007/s40279-018-0951-z
- 208 2 Drew MK, Finch CF. The Relationship Between Training Load and Injury, Illness and Soreness: A  
209 Systematic and Literature Review. *Sport Med* Published Online First: 2016. doi:10.1007/s40279-015-  
210 0459-8
- 211 3 Impellizzeri FM, Rampinini E, Marcora SM. Physiological assessment of aerobic training in soccer. *J*  
212 *Sports Sci* 2005;**23**:583–92. doi:10.1080/02640410400021278
- 213 4 MacInnis MJ, Gibala MJ. Physiological adaptations to interval training and the role of exercise intensity.  
214 *J Physiol* 2017;**595**:2915–30. doi:10.1113/JP273196
- 215 5 Rosa N, Simoes R, Magalhães FD, *et al.* From mechanical stimulus to bone formation: A review. *Med*  
216 *Eng Phys* 2015;**37**:719–28. doi:10.1016/j.medengphy.2015.05.015
- 217 6 Bohm S, Mersmann F, Arampatzis A. Human tendon adaptation in response to mechanical loading: a  
218 systematic review and meta-analysis of exercise intervention studies on healthy adults. *Sport Med - Open*  
219 2015;**1**:7. doi:10.1186/s40798-015-0009-9
- 220 7 Wisdom KM, Delp SL, Kuhl E. Use it or lose it: multiscale skeletal muscle adaptation to mechanical  
221 stimuli. *Biomech Model Mechanobiol* 2015;**14**:195–215. doi:10.1007/s10237-014-0607-3
- 222 8 Vanrenterghem J, Nedergaard NJ, Robinson MA, *et al.* Training Load Monitoring in Team Sports: A  
223 Novel Framework Separating Physiological and Biomechanical Load-Adaptation Pathways. *Sport Med*  
224 2017;**47**:2135–42. doi:10.1007/s40279-017-0714-2
- 225 9 Nimphius S, McGuigan MR, Newton RU. Changes in Muscle Architecture and Performance During a  
226 Competitive Season in Female Softball Players. *J Strength Cond Res* 2012;**26**:2655–66.
- 227 10 Secomb JL, Farley OR, Nimphius S, *et al.* The training-specific adaptations resulting from resistance  
228 training, gymnastics and plyometric training, and non-training in adolescent athletes. *Sport Sci Coach*  
229 2017;**12**:762–73. doi:10.1177/1747954117727810
- 230 11 Coupe C, Kongsgaard M, Aagaard P, *et al.* Habitual loading results in tendon hypertrophy and  
231 increased stiffness of the human patellar tendon. *J Appl Physiol* 2008;**105**:805–10.

- 232 doi:10.1152/jappphysiol.90361.2008
- 233 12 Mersmann F, Bohm S, Schroll A, *et al.* Muscle and tendon adaptation in adolescent athletes: A  
234 longitudinal study. *Scand J Med Sci Sports* 2017;**27**:75–82. doi:10.1111/sms.12631
- 235 13 Esmaeili A, Stewart AM, Hopkins WG, *et al.* Effects of Training Load and Leg Dominance on Achilles  
236 and Patellar Tendon Structure. *Int J Sports Physiol Perform* 2017;**12**:S2-122-S2-126.
- 237 14 Rabello LM, Zwerver J, Stewart RE, *et al.* Patellar tendon structure responds to load over a 7 - week  
238 preseason in elite male volleyball players. *Scand J Med Sci Sports* 2019;:1–8. doi:10.1111/sms.13428
- 239 15 Fredericson M, Chew K, Ngo J, *et al.* Regional bone mineral density in male athletes: a comparison of  
240 soccer players, runners and controls. *Br J Sports Med* 2007;**41**:664–8. doi:10.1136/bjism.2006.030783
- 241 16 Helge EW, Andersen T., Schmidt JF, *et al.* Recreational football improves bone mineral density and  
242 bone turnover marker profile in elderly men. *Scand J Med Sci Sports* 2014;**24**:98–104.  
243 doi:10.1111/sms.12239
- 244 17 Edwards WB. Modeling Overuse Injuries in Sport as a Mechanical Fatigue Phenomenon. *Exerc Sport*  
245 *Sci Rev* 2018;**46**:224–31. doi:10.1249/JES.0000000000000163
- 246 18 Bertelsen ML, Hulme A, Petersen J, *et al.* A framework for the etiology of running-related injuries.  
247 *Scand J Med Sci Sport* 2017;**27**:1170–80. doi:10.1111/sms.12883
- 248 19 Wang T, Lin Z, Day RE, *et al.* Programmable mechanical stimulation influences tendon homeostasis in a  
249 bioreactor system. *Biotechnol Bioeng* 2013;**110**:1495–507. doi:10.1002/bit.24809
- 250 20 Loundagin LL, Schmidt TA, Edwards WB. Mechanical Fatigue of Bovine Cortical Bone Using Ground  
251 Reaction Force Waveforms in Running. *J Biomech Eng* 2018;**140**. doi:10.1115/1.4038288
- 252 21 Amirouche F, Bobko A. Bone Remodeling and Biomechanical Processes- A Multiphysics Approach.  
253 *Austin J Biotechnol Bioeng* 2015;**2**:id1041.
- 254 22 Smith DW, Rubenson J, Lloyd D, *et al.* A conceptual framework for computational models of Achilles  
255 tendon homeostasis. *WIREs Syst Biol Med* 2013;**5**:523–38. doi:10.1002/wsbm.1229
- 256 23 Hewett TE, Myer GD, Ford KR, *et al.* Biomechanical Measures of Neuromuscular Control and Valgus  
257 Loading of the Knee Predict Anterior Cruciate Ligament Injury Risk in Female Athletes: A Prospective

258 Study. *Am J Sports Med* 2005;**33**:492–501.

259 24 Lin C, Liu H, Gros MT, *et al.* Biomechanical risk factors of non-contact ACL injuries: A stochastic  
260 biomechanical modeling study. *J Sport Heal Sci* 2012;**1**:36–42. doi:10.1016/j.jshs.2012.01.001

261 25 Verheul J, Clansey AC, Lake MJ. Adjustments with running speed reveal neuromuscular adaptations  
262 during landing associated with high mileage running training. *J Appl Physiol* 2017;**122**:653–665.  
263 doi:10.1152/jappphysiol.00801.2016

264 26 Arampatzis A, Brüggemann G-P, Metzler V. The effect of speed on leg stiffness and joint kinetics in  
265 human running. *J Biomech* 1999;**32**:1349–53.

266 27 Morin JB, Samozino P, Millet GY. Changes in running kinematics, kinetics, and spring-mass behavior  
267 over a 24-h run. *Med Sci Sports Exerc* 2011;**43**:829–36. doi:10.1249/MSS.0b013e3181fec518

268 28 Oliver JL, De Ste MBA, Lloyd RS, *et al.* Altered neuromuscular control of leg stiffness following  
269 soccer-specific exercise. *Eur J Appl Physiol* 2014;**114**:2241–9. doi:10.1007/s00421-014-2949-z

270 29 Maloney SJ, Fletcher IM. Lower limb stiffness testing in athletic performance: a critical review. *Sport*  
271 *Biomech* 2018;**3141**:1–22. doi:10.1080/14763141.2018.1460395

272 30 Seth A, Hicks JL, Uchida TK, *et al.* OpenSim: Simulating musculoskeletal dynamics and neuromuscular  
273 control to study human and animal movement. *PLoS Comput Biol* 2018;**14**:e1006223.  
274 doi:10.1371/journal.pcbi.1006223

275 31 Scott SH, Winter DA. Internal forces at chronic running injury sites. *Med Sci Sports Exerc* 1990;**22**:357–  
276 69.

277 32 Pizzolato C, Saxby DJ, Ceseracciu E, *et al.* Biofeedback for gait retraining based on real-time estimation  
278 of tibiofemoral joint contact forces. *Trans Neural Syst Rehabil Eng* 2017;**25**:1612–21.

279 33 Pizzolato C, Reggiani M, Modenese L, *et al.* Real-time inverse kinematics and inverse dynamics for  
280 lower limb applications using OpenSim. *Comput Methods Biomech Biomed Engin* 2017;**20**:436–45.  
281 doi:10.1080/10255842.2016.1240789

282 34 van den Bogert AJ, Geijtenbeek T, Even-Zohar O, *et al.* A real-time system for biomechanical analysis  
283 of human movement and muscle function. *Med Biol Eng Comput* 2013;**51**:1069–77.  
284 doi:10.1007/s11517-013-1076-z

- 285 35 Manal K, Gravare-Silbernagel K, Buchanan TS. A real-time EMG-driven musculoskeletal model of the  
286 ankle. *Multibody Syst Dyn* 2012;**28**:169–80. doi:10.1007/s11044-011-9285-4
- 287 36 Fukashiro S, Komi P V., Järvinen M, *et al.* In vivo achilles tendon loading' during jumping in humans.  
288 *Eur J Appl Physiol Occup Physiol* 1995;**71**:453–458.
- 289 37 Komi P V., Salonen M, Järvinen M, *et al.* In Vivo Registration of Achilles Tendon Forces in Man. I.  
290 Methodological Development. *Int J Sport Med* 1987;**8**:3–8.
- 291 38 Komi P V. Relevance of in vivo force measurements to human biomechanics. *J Biomech* 1990;**23**:23–34.  
292 doi:10.1016/0021-9290(90)90038-5
- 293 39 Burr DB, Milgrom C, Fyhrie D, *et al.* In Vivo Measurement of Human Tibial Strains During Vigorous  
294 Activity. *Bone* 1996;**18**:405–10.
- 295 40 Martin JA, Brandon SCE, Keuler EM, *et al.* Gauging force by tapping tendons. *Nat Commun* 2018;**9**:1–  
296 9. doi:10.1038/s41467-018-03797-6
- 297 41 Raper DP, Witchalls J, Philips EJ, *et al.* Use of a tibial accelerometer to measure ground reaction force in  
298 running: A reliability and validity comparison with force plates. *J Sci Med Sport* 2018;**21**:84–8.  
299 doi:10.1016/j.jsams.2017.06.010
- 300 42 Milner CE, Ferber R, Pollard CD, *et al.* Biomechanical factors associated with tibial stress fracture in  
301 female runners. *Med Sci Sports Exerc* 2006;**38**:323–8. doi:10.1249/01.mss.0000183477.75808.92
- 302 43 Hennig EM, Lafortune MA. Relationships between ground reaction force and tibial bone acceleration  
303 parameters. *Int J Sport Biomech* 1991;**7**:303–9.
- 304 44 Sheerin KR, Reid D, Besier TF. The measurement of tibial acceleration in runners. A review of the  
305 factors that can affect tibial acceleration during running and evidence based guidelines for its use. *Gait*  
306 *Posture* 2019;**67**:12–24. doi:10.1016/j.gaitpost.2018.09.017
- 307 45 Nagahara R, Mizutani M, Matsuo A, *et al.* Association of sprint performance with ground reaction forces  
308 during acceleration and maximal speed phases in a single sprint. *J Appl Biomech* 2017;**;**1–20.  
309 doi:10.1123/jab.2016-0356
- 310 46 Bezodis NE, North JS, Razavet JL. Alterations to the orientation of the ground reaction force vector  
311 affect sprint acceleration performance in team sports athletes. *J Sports Sci* 2017;**35**:1817–24.

312 doi:10.1080/02640414.2016.1239024

313 47 Rabita G, Dorel S, Slawinski J, *et al.* Sprint mechanics in world-class athletes: A new insight into the  
314 limits of human locomotion. *Scand J Med Sci Sports* 2015;**25**:583–94. doi:10.1111/sms.12389

315 48 Zadpoor AA, Nikooyan AA. The relationship between lower-extremity stress fractures and the ground  
316 reaction force: A systematic review. *Clin Biomech* 2011;**26**:23–8.  
317 doi:10.1016/j.clinbiomech.2010.08.005

318 49 Matijevich ES, Branscombe LM, Scott LR, *et al.* Ground reaction force metrics are not strongly  
319 correlated with tibial bone load when running across speeds and slopes: Implications for science, sport  
320 and wearable tech. *PLoS One* 2019;**14**:e0210000. doi:10.1371/journal.pone.0210000

321 50 van der Worp H, Vrielink JW, Bredeweg SW. Do runners who suffer injuries have higher vertical  
322 ground reaction forces than those who remain injury-free? A systematic review and meta-analysis. *Br J*  
323 *Sports Med* 2016;**50**:450–7. doi:10.1136/bjsports-2015-094924

324 51 Ramirez-Bautista JA, Huerta-Ruelas JA, Chaparro-Cárdenas SL, *et al.* A Review in Detection and  
325 Monitoring Gait Disorders Using In-Shoe Plantar Measurement Systems. *IEEE Rev Biomed Eng*  
326 2017;**10**:299–309. doi:10.1109/RBME.2017.2747402

327 52 Renner KE, Williams DSB, Queen RM. The Reliability and Validity of the Loadsol® under Various  
328 Walking and Running Conditions. *Sensors* 2019;**19**:1–14. doi:10.3390/s19020265

329 53 Burns GT, Zandler JD, Zernicke RF. Validation of a wireless shoe insole for ground reaction force  
330 measurement. *J Sports Sci* 2018;:1–10. doi:10.1080/02640414.2018.1545515

331 54 Seiberl W, Jensen E, Merker J, *et al.* Accuracy and precision of loadsol® insole force- sensors for the  
332 quantification of ground reaction force-based biomechanical running parameters parameters. *Eur J Sport*  
333 *Sci* 2018;**18**:1100–9. doi:10.1080/17461391.2018.1477993

334 55 Park J, Na Y, Gu G, *et al.* Flexible insole ground reaction force measurement shoes for jumping and  
335 running. *Proc IEEE RAS EMBS Int Conf Biomed Robot Biomechatronics* 2016;**2016-July**:1062–7.  
336 doi:10.1109/BIOROB.2016.7523772

337 56 Peebles AT, Maguire LA, Renner KE, *et al.* Validity and Repeatability of Single-Sensor Loadsol Insoles  
338 during Landing. *Sensors* 2018;**18**:1–10. doi:10.3390/s18124082

- 339 57 Pavei G, Seminati E, Cazzola D, *et al.* On the estimation accuracy of the 3D body center of mass  
340 trajectory during human locomotion: Inverse vs. forward dynamics. *Front Physiol* 2017;**8**:1–13.  
341 doi:10.3389/fphys.2017.00129
- 342 58 Verheul J, Gregson W, Lisboa PJ, *et al.* Whole-body biomechanical load in running-based sports: The  
343 validity of estimating ground reaction forces from segmental accelerations. *J Sci Med Sport*  
344 2019;**22**:716–22. doi:10.1016/j.jsams.2018.12.007
- 345 59 Bobbert MF, Schamhardt HC, Nigg BM. Calculation of vertical ground reaction force estimates during  
346 running from positional data. *J Biomech* 1991;**24**:1095–105. doi:10.1016/0021-9290(91)90002-5
- 347 60 Nedergaard NJ, Robinson MA, Eusterwiemann E, *et al.* The Relationship Between Whole-Body  
348 External Loading and Body-Worn Accelerometry During Team-Sport Movements. *Int J Sports Physiol*  
349 *Perform* 2017;**12**:18–  
350 26. <https://search.ebscohost.com/login.aspx?direct=true&db=sph&AN=121191349&site=ehost-live>
- 351 61 Edwards S, White S, Humphreys S, *et al.* Caution using data from triaxial accelerometers housed in  
352 player tracking units during running. *J Sports Sci* 2019;**37**:810–8. doi:10.1080/02640414.2018.1527675
- 353 62 Nedergaard NJ, Verheul J, Drust B, *et al.* The feasibility of predicting ground reaction forces during  
354 running from a trunk accelerometry driven mass-spring-damper model. *PeerJ* 2018;**6**:e6105.  
355 doi:10.7717/peerj.6105
- 356 63 Wundersitz DWT, Netto KJ, Aisbett B, *et al.* Validity of an upper-body-mounted accelerometer to  
357 measure peak vertical and resultant force during running and change-of-direction tasks. *Sport Biomech*  
358 2013;**12**:403–12. doi:10.1080/14763141.2013.811284
- 359 64 Gurchiek RD, McGinnis RS, Needle AR, *et al.* The use of a single inertial sensor to estimate 3-  
360 dimensional ground reaction force during accelerative running tasks. *J Biomech* 2017;**61**:263–8.  
361 doi:10.1016/j.jbiomech.2017.07.035
- 362 65 Neugebauer JM, Collins KH, Hawkins DA. Ground Reaction Force Estimates from ActiGraph GT3X+  
363 Hip Accelerations. *PLoS One* 2014;**9**:e99023. doi:10.1371/journal.pone.0099023
- 364 66 Gaudino P, Gaudino C, Alberti G, *et al.* Biomechanics and predicted energetics of sprinting on sand:  
365 Hints for soccer training. *J Sci Med Sport* 2013;**16**:271–5. doi:10.1016/j.jsams.2012.07.003

- 366 67 Buchheit M, Gray A, Morin J-B. Assessing stride variables and vertical stiffness with GPS-embedded  
367 accelerometers: preliminary insights for the monitoring of neuromuscular fatigue on the field. *J Sport Sci*  
368 *Med* 2015;:698–701.
- 369 68 Buchheit M, Lacombe M, Cholley Y, *et al.* Neuromuscular Responses to Conditioned Soccer Sessions  
370 Assessed via GPS-Embedded Accelerometers: Insights Into Tactical Periodization. *Int J Sports Physiol*  
371 *Perform* 2018;**13**:577–83.
- 372 69 Page RM, Marrin K, Brogden CM, *et al.* Biomechanical and physiological response to a contemporary  
373 soccer match-play simulation. *J Strength Cond Res* 2015;**29**:2860–6.
- 374 70 Colby MJ, Dawson B, Heasman J, *et al.* Accelerometer and GPS-Derived Running Loads and Injury  
375 Risk in Elite Australian Footballers. *J Strength Cond Res* 2014;**28**:2244–52.
- 376 71 Barrett S, Midgley A, Lovell R. PlayerLoad™: Reliability, convergent validity, and influence of unit  
377 position during treadmill running. *Int J Sports Physiol Perform* 2014;**9**:945–52. doi:10.1123/ijsp.2013-  
378 0418
- 379 72 Gaudino P, Alberti G, Iaia FM. Estimated metabolic and mechanical demands during different small-  
380 sided games in elite soccer players. *Hum Mov Sci* 2014;**36**:123–33. doi:10.1016/j.humov.2014.05.006
- 381 73 Ehrmann FE, Duncan CS, Sindhusake D, *et al.* GPS and Injury Prevention in Professional Soccer. *J*  
382 *Strength Cond Res* 2016;**30**:360–7.
- 383 74 Boyd LJ, Ball K, Aughey RJ. The reliability of minimaxX accelerometers for measuring physical  
384 activity in australian football. *Int J Sports Physiol Perform* 2011;**6**:311–21.
- 385 75 Roetenberg D, Luinge H, Slycke P. Xsens MVN: Full 6DOF Human Motion Tracking Using Miniature  
386 Inertial Sensors. 2013. doi:10.1.1.569.9604
- 387 76 Karatsidis A, Bellusci G, Schepers MH, *et al.* Estimation of Ground Reaction Forces and Moments  
388 During Gait Using Only Inertial Motion Capture. *Sensors* 2017;**17**:1–22. doi:10.3390/s17010075
- 389 77 Konrath J, Karatsidis A, Schepers MH, *et al.* Estimation of the Knee Adduction Moment and Joint  
390 Contact Force during Daily Living Activities Using Inertial Motion Capture. *Sensors* 2019;**19**:1–12.  
391 doi:10.3390/s19071681
- 392 78 Corazza S, Mündermann L, Chaudhari AM, *et al.* A markerless motion capture system to study



- 393 musculoskeletal biomechanics: Visual hull and simulated annealing approach. *Ann Biomed Eng*  
394 2006;**34**:1019–29. doi:10.1007/s10439-006-9122-8
- 395 79 Abrams GD, Harris AHS, Andriacchi TP, *et al.* Biomechanical analysis of three tennis serve types using  
396 a markerless system. *Br J Sports Med* 2014;**48**:339–42. doi:10.1136/bjsports-2012-091371
- 397 80 Fung SK, Sundaraj K, Ahamed NU, *et al.* Hybrid markerless tracking of complex articulated motion in  
398 golf swings. *J Bodyw Mov Ther* 2014;**18**:220–7. doi:10.1016/j.jbmt.2013.05.011
- 399 81 Grigg J, Haakonssen E, Rathbone E, *et al.* The validity and intra-tester reliability of markerless motion  
400 capture to analyse kinematics of the BMX Supercross gate start. *Sport Biomech* 2018;**17**:383–401.  
401 doi:10.1080/14763141.2017.1353129
- 402 82 Saylor K, Nicolella D, Chambers D, *et al.* Markerless biomechanics analysis for optimization of soldier  
403 physical performance. *J Sci Med Sport* 2017;**20S**:S119. doi:10.1016/j.jsams.2017.09.431
- 404 83 Perrott MA, Pizzari T, Cook J, *et al.* Comparison of lower limb and trunk kinematics between markerless  
405 and marker-based motion capture systems. *Gait Posture* 2017;**52**:57–61.  
406 doi:10.1016/j.gaitpost.2016.10.020
- 407 84 Skals S, Jung MK, Damsgaard M, *et al.* Prediction of ground reaction forces and moments during sports-  
408 related movements. *Multibody Syst Dyn* 2017;**39**:175–95. doi:10.1007/s11044-016-9537-4
- 409 85 Halilaj E, Rajagopal A, Fiterau M, *et al.* Machine Learning in Human Movement Biomechanics: Best  
410 Practices, Common Pitfalls, and New Opportunities. *J Biomech* Published Online First: 2018.  
411 doi:10.1016/j.jbiomech.2018.09.009
- 412 86 Wouda FJ, Giuberti M, Bellusci G, *et al.* Estimation of Vertical Ground Reaction Forces and Sagittal  
413 Knee Kinematics During Running Using Three Inertial Sensors. *Front Physiol* 2018;**9**:1–14.  
414 doi:10.3389/fphys.2018.00218
- 415 87 Johnson WR, Mian A, Robinson MA, *et al.* Multidimensional ground reaction forces and moments from  
416 wearable sensor accelerations via deep learning. *arXiv Prepr* 2019.
- 417 88 Stetter BJ, Ringhof S, Krafft FC, *et al.* Estimation of Knee Joint Forces in Sport Movements Using  
418 Wearable Sensors and Machine Learning. *Sensors* 2019;**19**:3690. doi:10.3390/s19173690
- 419 89 Doshi-Velez F, Kim B. Towards A Rigorous Science of Interpretable Machine Learning. *ArXiv Prepr*

420 2017;**1702.08608**.<http://arxiv.org/abs/1702.08608>

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423 **FIGURE CAPTIONS**

424

425 **Figure 1** Schematic overview of currently available biomechanical load metrics. The feasibility of measuring  
426 these metrics, ranging from strictly limited to the laboratory to viable in field environments, is indicated along  
427 the y-axis. The level at which loads act on the musculoskeletal system is indicated along the x-axis. The different  
428 hard- and soft-tissues affected by each load metric are shown in red (muscles), green (tendons and ligaments)  
429 and/or blue (bones and cartilage). Metrics to assess tissue- or structure-specific loads that are viable to be  
430 measured in the field are still lacking.