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Flow cytometry and growth-based analysis of the effects of fruit sanitation on the physiology of Escherichia coli in orange juice

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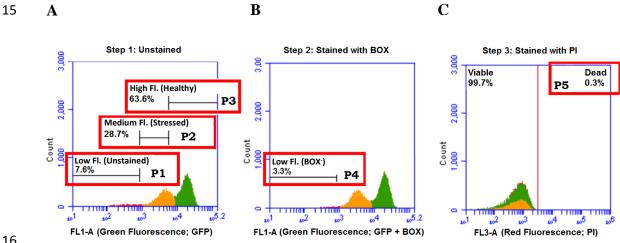
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- 1 Supplemental Information for
- 2
- 3 Flow cytometry and growth-based analysis of the effects of fruit sanitation on the
- 4 physiology of Escherichia coli in orange juice
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14 **Supplemental Figure S1**



Т	ь

Physiological state	Fluorescence characteristics	Population above
Viable healthy	GFP⁺ PI⁻	P3
Viable stressed	Medium GFP	P2
Viable injured	GFP ⁻ BOX ⁺ PI ⁻	(P1 - P4) - P5
Viable low GFP	GFP ⁻ BOX ⁻ PI ⁻	P4
Dead	PI⁺	P5

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Supplemental Figure S1: Simultaneous use of two green fluorophores of GFP and 18 19 BOX along with red fluorescent viability dye of PI in order to study the number of healthy, stressed, injured and dead E. coli K-12 SCC1 cells in OJ. The method of 20 experiment was similar to what was described in Figure 1, with the difference of using BOX 21 22 in addition to PI. (A) Inoculation of cells in OJ resulted in an increase in the number of low 23 GFP and/or GFP⁻ cells. Reduction in GFP fluorescence was presumed to be due to change 24 in internal pH and subsequent denaturation of GFP. Therefore, GFP⁺ were considered 25 healthy whereas low GFP cells were considered to be stressed or injured cells. (B) Samples 26 were then stained with BOX in order to determine the percentage of injured cells without a membrane potential. Addition of BOX caused a reduction in the number of cells with low 27 green fluorescence (compared to histogram A), indicating the staining of the injured GFP-28 cells with BOX. (C) Staining the cells with PI showed the percentage of dead cells. 29 30 Consequently, it was possible to calculate the percentage of injured cells by subtracting the number of dead cells (histogram C) and viable stressed GFP- cells (histogram B) from the 31 32 pre-BOX staining percentage. 33