UNIVERSITYOF **BIRMINGHAM**

University of Birmingham Research at Birmingham

Gums and joints

Lopez-Oliva, Isabel: De Pablo, Paola: Dietrich, Thomas: Chapple, Iain

DOI:

10.1038/s41415-019-0723-7

License:

None: All rights reserved

Document Version Peer reviewed version

Citation for published version (Harvard): Lopez-Oliva, I, De Pablo, P, Dietrich, T & Chapple, I 2019, 'Gums and joints: is there a connection? Part two: the biological link', British Dental Journal, vol. 227, no. 7, pp. 611-617. https://doi.org/10.1038/s41415-019-0723-7

Link to publication on Research at Birmingham portal

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes

- •Users may freely distribute the URL that is used to identify this publication.
- •Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
 •User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- •Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Download date: 11. Apr. 2024

Gums and joints: is there a connection?

PART 2. THE BIOLOGICAL LINK

Isabel Lopez-Oliva, Paola de Pablo, Thomas Dietrich, Iain Chapple

Review for BDJ

3,500 words

ABSTRACT

Rheumatoid arthritis (RA) and periodontitis (PD) are inflammatory diseases characterized by an exacerbated immune-inflammatory reaction that leads to destruction of bone and other connective tissues that share numerous similarities. Although a significant and independent association between has been described, the pathophysiological processes that may explain this relationship remain unknown and multiple theories have been proposed. This review presents the most important theories currently proposed to explain the biological link between RA and PD.

1. INTRODUCTION

The available evidence to date, suggests a significant and independent association between rheumatoid arthritis (RA) and periodontitis (PD). Both are chronic inflammatory diseases characterized by an exacerbated immune-inflammatory reaction that leads to destruction of bone and other connective tissues that share numerous similarities. Hence, the biological mechanisms underpinning such a relationship have been a focus of investigation for many years. ¹ However, the pathophysiological processes that may explain this relationship remain unknown and multiple theories have been proposed.

Given the increasing number of clinical studies suggesting beneficial effects of periodontal therapy on RA outcomes, there is a need for more studies aimed at understanding the mechanisms that connect these two conditions and to help explain why periodontal therapy may be beneficial. This review aims to lay out the most important theories proposed to explain the biological link between RA and PD.

2. Risk factors

RA and PD have multiple risk factors in common (Table 1) and it has therefore been hypothesized that the epidemiological relationship between the two conditions could possibly be explained by the "common risk" hypothesis.

Table 1. Shared risk factors between periodontitis and rheumatoid arthritis (RA). Acronyms: PMNL, polymorphonuclear leukocyte; C.T, connective tissue

PERIODONTITIS	RA
Smoking	\checkmark
Genetics	\checkmark
Host-mediated C.T. destruction	\checkmark
A role for plasma cells in active disease	\checkmark
PMNL infiltration = substantial	\checkmark
Oxidative stress = major feature	\checkmark
Female sex hormones play a role	\checkmark
Symptoms respond to anti-inflammatory drugs	\checkmark

2.1 Genetic risk factors

RA and periodontitis have certain common genetic factors impacting upon the hosts' immune response that translate into, for example, higher levels of proinflammatory cytokine production. The human leukocyte antigen, alleles DR4 (HLA-DR4) epitope is located on the surface of leukocytes and has been found to be associated with both RA and periodontitis. ²

In common with several other immune-mediated diseases, numerous studies report an association between HLA genes and periodontitis. The most recent systematic review reported a protective association with HLA-A2 and B5 and an increased susceptibility for periodontal disease with HLA-A9 and B15 genotypes. ³ Whilst classical twin studies have shown that periodontitis exhibits a genetic component, ⁴ genome wide association studies (GWAS) have been unable to identify significant associations with chronic periodontitis. ⁵ A recent systematic review of 43 studies concluded that there was no evidence for an interaction between any genetic variants (including IL1) and the sub-gingival microflora. ⁶

In RA, robust data are available from genetic association studies, suggesting HLA SE (shared epitope) alleles as the main candidate gene in anticitrullinated protein antibodies (ACPA) positive patients. ⁷ In addition to these alleles, GWAS have revealed another 32 loci associated with RA. ⁸ There is also a clear dissimilarity between genetic risk factors for ACPA-positive and ACPA-negative RA, thus more research is required to understand the genetic differences between these two groups. ⁹

There are however currently few studies investigating a genetic link between RA and periodontitis ¹⁰⁻¹² and no GWAS studies. Therefore, the importance of genetic polymorphisms in the link between these two diseases remains unknown.

2.2 Smoking

Smoking is a shared modifiable risk factor for both periodontitis and RA, ¹³ with recent data demonstrating a 3-fold increased risk of RA in ACPA-positive men who smoke (1.8-fold increase for women smokers), although no association was found in ACPA-negative patients. ¹⁴ This association with smoking is not however evident in patients with early rheumatoid arthritis. This group of patients exhibit a high prevalence of periodontitis but have a low prevalence of smoking (16%) and a young age (mean age 42.2 years old). ¹⁵ In contrast, some scientists have reported a significantly higher risk of periodontitis in non-smokers with RA ¹⁶ whereas others have found an association between RA and periodontitis irrespective of smoking status. ¹⁷ ¹⁸

Therefore, although periodontitis and RA share genetic (HLA-DR4) and lifestyle (e.g. smoking) risk factors, as well as similar inflammatory pathways, these do not appear to be sufficient to explain the connection between the two diseases. ¹⁹ ²⁰

2.3 Poor oral hygiene

Poor oral hygiene has also been proposed as a link between rheumatoid arthritis and periodontitis. The hypothesis that RA patients are unable to maintain good oral hygiene compared to healthy controls due to impaired manual dexterity, is attractive as an explanation for the association between the two. Indeed, it is logical that the hand and joint deformities evident in the most severe forms of RA, alongside the associated functional limitations, predispose to poor levels of plaque removal, and therefore an increased risk of periodontitis (Figure 1). However, studies have consistently failed to show a difference in plaque control that could explain the association with periodontitis. ²¹⁻²³



Figure 1 Patient with rheumatoid arthritis (RA) holding a toothbrush, showing the typical hand deformities of severe RA sufferers.

2.4 Gender

One of the biggest disparities we find between the incidence of RA and periodontitis is the gender frequencies. While 75% of RA patients are female, men are believed to have worse periodontal health. ²⁴

This gender difference has not been fully investigated although it may well be explained by a stronger effect of smoking and obesity in the development of periodontitis. However, it is known that in both diseases female hormones

play a role, worsening periodontal status in female puberty, pregnancy and at the postmenopausal stage. ²⁵

3. Protein citrullination

Protein citrullination or "deimination" has been widely studied due to its role in autoimmune diseases especially in RA. This post-translational modification of proteins occurs when a family of enzymes called peptidyl-arginine deiminase (PAD) transforms the amino acid arginine into citrulline. In the presence of calcium, PAD replaces the ketamine group (=NH) with a ketone group (=O), converting the positively charged arginine into a neutrally charged citrulline amino acid. ²⁶ This renders the parent protein hydrophobic, resulting in a change in protein folding and thus function (Figure 2).

While citrullination is a physiological phenomenon in healthy individuals, in RA citrullinated proteins are recognized as an antigen and activate the immune system. ⁴² Therefore, why the immune system produces antibodies against citrullinated proteins in RA patients remains unknown and this unique production of auto-antibodies in RA suggests the necessity for an external factor to trigger this auto-immune reaction. It has also been shown that PAD (in particular PAD-2 and PAD-4) are expressed in inflamed periodontal tissues and this may be one important source of protein citrullination, arising many years before the RA develops. ²⁷ This event could break the immune tolerance to citrullinated proteins and predispose to RA following a second insult in the joints themselves.

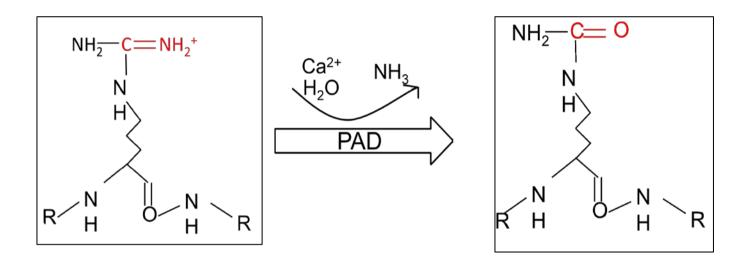


Figure 2. Protein citrullination by Peptidyl-Arginine Deiminase (PAD). In the presence of calcium, PAD catalyses the conversion of Arginine in to Citrulline, changing the Ketamine group (=NH) for a Ketone group (=O).

3.1. Porphyromonas gingivalis citrullination

Porphyromonas gingivalis (*P. gingivalis*) is Gram-negative anaerobic bacteria strongly associated with periodontitis which is considered key to disruption of the host-microbial homeostasis that characterizes the disease. ²⁸ ²⁹ ³⁰ Interestingly, *P. gingivalis* uniquely expresses peptidyl arginine deiminase (PPAD) which is capable of citrullinating both host and bacterial peptides ³¹ for its survival within the periodontal pocket. ³² It has been observed in some *in vitro* studies that PPAD can only citrullinate C-terminal arginine and not internal arginine, in contrast to its homologues human PAD2 and PAD4, creating citrullinated peptides that would not normally occur in the absence of *P. gingivalis*. By presenting these neoepitopes to the immune system, *P. gingivalis* could break immune tolerance to citrullinated proteins and lead to the subsequent generation of ACPAs characteristic of RA patients (Figure 3).

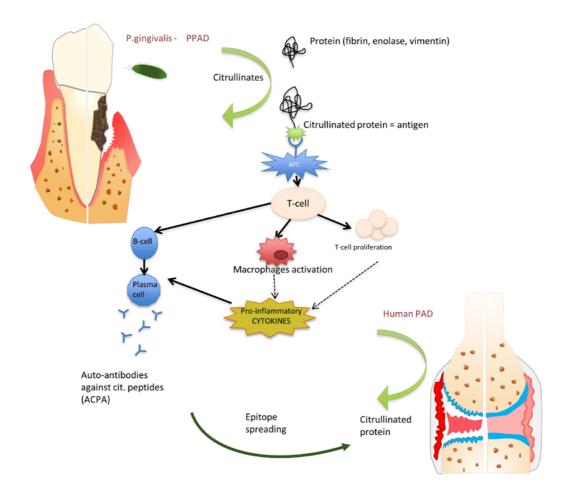


Figure 3. Model of the biological link between rheumatoid arthritis and periodontitis. *P. gingivalis* breaking immune tolerance to citrullinated proteins in rheumatoid arthritis (RA) by bacterial citrullination of proteins through PPAD, initiating an immune response leading to loss of tolerance to citrullinated proteins in the joints. Abbreviations: *P.gingivalis*, *Porphyromonas gingivalis*; PPAD, peptidyl arginine deiminase from P.*gingivalis*; APC, antigen presenting cell; TNF, tumour necrosis factor; IL, interleukin; GMCSF, granulocyte-macrophage colony-stimulating factor.

The potential role of *P. gingivalis* in the etiology of RA has been investigated over the last decade. ³³ Martinez-martinez et al ³⁴ found DNA from *P.gingivalis* in the synovial fluid of patients with RA and established that periodontal pathogens could be a trigger for the autoimmune response in RA. However, PPAD itself to date has not been found within synovial fluid. Hitchon et al. ³⁵ reported an association between immune responses to *P. gingivalis* and the presence of ACPA in a population with a high background prevalence of the RA predisposing HLA alleles. A study by Mikuls et al., showed that high levels of anti-P.*gingivalis* antibodies in RA subjects correlated with levels of ACPAs, suggesting that this organism plays a role as a risk factor in RA. ³⁶

Interestingly, it has been observed that the periodontium of periodontitis patients with no signs of RA express citrullinated proteins ³⁷ and that serum from these patients contain higher levels of antibodies to citrullinated and non-citrullinated human peptides compared to healthy controls. ³⁸ De Pablo et al. suggested that the greater citrullination arising in periodontitis leads to a loss of tolerance to citrullinated and un-citrullinated peptides, which may evolve in a cross-reaction against the citrullinated proteins in the joint that may in turn lead to RA.

Based on these findings, researchers propose that bacterial and human PADs could be a therapeutic target in RA. ³⁹ Moreover, it has been hypothesized that periodontal therapy could decrease the load of *P.gingivalis* and PPAD in the periodontal pocket and that by reducing gingival inflammation, the expression of human PADs could be reduced weakening the autoimmune response in RA.

However, it remains unclear whether there is a causal role for antibodies to citrullinated proteins in the evolution of RA, and there is no strong evidence that periodontal therapy reduces citrullination systemically yet.

3.2. Other periodontal bacterial candidates to play a role in RA

Recently, other periodontal pathogens have been identified as possible triggers for RA. In a study conducted by Konig and collaborators, authors found that *Aggregatibacter actinomycetemcomitans* was the only bacteria able to induce hypercitrullination within human neutrophils when investigating a number of periodontal pathogens and oral commensals. Through secretion of leukotoxin A (LtxA), *A. actinomycetemcomitans* induces calcium influx and hyperactivation of PAD in the neutrophil, leading to neutrophil hypercitrullination. ⁴⁰ Interestingly, the effect of human lymphocyte antigen—DRB1 shared epitope alleles on auto-antibody positivity was limited to RA patients who were exposed to *A. actinomycetemcomitans*.

Last year, a study investigating the oral microbiome in RA, identified *Cryptobacterium curtum* (previously misclassified as *Eubacterium saburreum*) as a predominant member of the RA-influenced periodontal microbiome with a 100-fold greater abundance in RA and with a 39-fold greater odds of detection compared to non-RA controls. ⁴¹ Interestingly, this gram-positive anaerobic rod/periodontal pathogen ⁴² degrades arginine through the arginine deiminase pathway and produces substantial amounts of citrulline, ornithine and ammonia. ⁴³ Others have previously found *C. curtum* to be enriched in the oral and gut microbiomes of early RA cases ¹⁵ ⁴⁴ thus the authors suggest that *C.curtum* is a candidate for further studies.

3.3. Citrullinated human proteins targeted in RA

Anti-citrullinated protein antibody (ACPA) testing kits are regularly used by rheumatologists to measure antibodies against citrullinated proteins and to diagnose patients as ACPA+ and ACPA- which can be used to anticipate disease severity and treatment responses. ⁴⁵ This test uses a synthetic citrullinated cyclic peptide as an antigen to capture antibodies against any citrullinated proteins present in patient sera. Moreover, investigating the proteins that are citrullinated in RA can help us understand the development of the autoimmune response and development of the disease. To date, numerous studies have reported that proteins including fibrinogen, vimentin,

enolase and tenascin are targeted in RA and antibodies against these citrullinated proteins are associated with the severity of the disease and also elevated in periodontitis patients. ^{45 46, 47,48 49 47 50}

The current data demonstrate that both RA and periodontitis induce citrullination, which may be the link between these two diseases. Recently, it has been found that the production of citrullinated proteins in the periodontal tissues is associated with gingival inflammation, ²⁷ occurring in 80% of periodontitis-affected stroma. ³⁷ This is supported by the discovery of higher serum antibodies against citrullinated proteins in periodontitis patients compared to controls. ³⁸ For this reason, investigators believe that the gingiva of periodontitis patients might be an extra-articular source of citrullinated proteins, which can lead to ACPA production contributing to RA progression.

3.4. Carbamylation and Malondialdehyde-acetaldehyde adducts

As detailed above, post-translational modification of proteins can lead to the production of autoantibodies and loss of immune tolerance. Apart from citrullination, several other post-translational modifications of proteins have been associated with RA. Of these, malondialdehyde-acetaldehyde adducts and carbamylated proteins are receiving particular attention. ⁵¹ A recent study evidence of identified immunohistochemical the presence malondialdehyde-acetaldehyde adducts, citrullinated and carbamylated proteins in inflamed human gingival tissue biopsies but not in biopsies of healthy, non-inflamed gingivae. 52 Identification of such modified proteins in inflamed gingiva suggests that inflammation of the periodontal tissues may influence the development of RA.

3.5. Porphyromonas gingivalis antigens targeted in RA

Proteins from *P.gingivalis* may be recognized as an antigen by the host and initiate an immune response that could break tolerance to human antigens such as citrullinated proteins. Based on this hypothesis, researchers have

investigated antibodies against some *P.gingivalis* -antigens and their role in RA.

Antibodies against citrullinated PPAD (CPP3 and CPP5) are increased in ACPA+ RA patients as well as in periodontitis patients (CPP5). ⁵³ In RA, and in patients in the pre-clinical period of the disease (pre-RA patients), anti-CPP3 antibodies are elevated, ⁵⁴ although other researchers have been unable to find an association between anti-PPAD antibodies and early RA. ⁵⁵ Another limitation of this hypothesis is the lack of studies investigating the presence of PPAD in inflamed periodontal tissues from RA patients.

While only a few studies have investigated these antibodies and more research is needed, considering these results, the hypothesis that *P.gingivalis* -antigens may break immune tolerance in RA, requires further investigation and if proven, these epitopes could be targeted in novel therapies for RA patients. ⁵⁶

4. The role of the oral microbiome in RA

The human microbiome is defined as "the genes carried by the particular community of microorganisms that live in and on the human body". Using DNA-based technologies to investigate bacteria has opened a new chapter in our understanding of the microbes that live with us, ⁵⁷ as traditional cultivation techniques limited understanding to the minority of organisms that could be grown outside the body.

Currently, three microbiome studies have investigated the oral microbiome in RA patients. ⁵⁸ ¹⁵ Zhang and collaborators reported the sub-gingival, oral and gut microbiome to be altered in RA, and certain species such as *Lactobacillus salivarius* and *Cryptobacterium curtum* were enriched, therefore offering potential diagnostic tools. ⁵⁸ In a study of early RA patients, Scher and colleagues noted that some oral bacterial taxa were significantly different in

abundance, however overall measures of diversity were no different to systemically healthy controls. In a more recent study investigating the periodontally healthy oral microbiome in RA found that compared to controls, RA patients had a distinct microbiome with a significantly higher percentage of anaerobes, including *Cryptobacterium curtum.* ⁴¹ However, the effect of periodontal therapy on the oral microbiome remains to be fully investigated using next generation sequencing techniques, and the effect in RA patients remains unknown.

5. The cytokine imbalance in RA and Periodontitis

In RA, there is an imbalance between the pro/anti-inflammatory host systems ⁵⁹, similar to that reported in periodontitis patients ⁶⁰. Understanding the cytokine networks involved in the pathobiology of RA has led to a revolution in the therapeutic arsenal available for this disease in the last decade ⁶¹ and may also represent a key biological link between RA and periodontitis.

To date, several studies have evaluated the effect of TNF inhibitors in periodontitis and RA ^{62 63 64 65 66}. Although the authors concluded that anti-TNF therapy could benefit periodontal status, they suggest that it is difficult to predict the collateral harm that may result from targeting cytokines therapeutically. ⁶⁷

Interestingly, periodontal therapy has been shown to reduce serum TNF levels in some studies 68 69 , whilst other studies failed to find this reduction 70 . For this reason, it is important to investigate periodontal therapy as a potentially safe and non-pharmacological treatment that could reduce inflammation in systemic inflammatory conditions such as RA. 71 72 73

6. Neutrophils and neutrophil extracellular traps

Neutrophils are the first leukocyte to arrive at the site of infection and their principal functions include immunological surveillance, recognising and

phagocytosing microorganisms, and activation of and collaboration with humoral immunity. When activated by multiple inflammatory signals, neutrophils may release their enzymatic content to the exterior of the cell, together with their DNA, forming a web-like structure called a neutrophil extracellular trap (NET), whose purpose is to immobilise microbes and prevent their spread. Although effective, the release of these contents, along with reactive oxygen species (ROS) provokes collateral tissue damage to the surrounding tissues, exacerbates the inflammatory response and exposes possible autoantigens. ^{74 75}

Elevated NET formation occurs in both RA and periodontitis patients, ⁷⁶ ⁷⁷ and there is a clear correlation between levels of DNA in the joints, serum ACPA levels and neutrophil counts, as well as a clear mapping of citrullinated proteins to neutrophils within cell pellets prepared from inflamed joints. NET release was also reported to be correlated with high ACPA and rheumatoid factor (RF) levels. ⁷⁸

Taken together, these results suggest that, in periodontitis, a chronic exposure of gingival tissues to PAD4 and citrullination may result in the breakdown of immune tolerance to citrullinated peptides in susceptible individuals which could lead or contribute to RA pathogenesis. NET generation may therefore not only contribute to periodontitis and RA pathogenesis, but also provides a potential causal link between these two conditions (Figure 4). ⁷

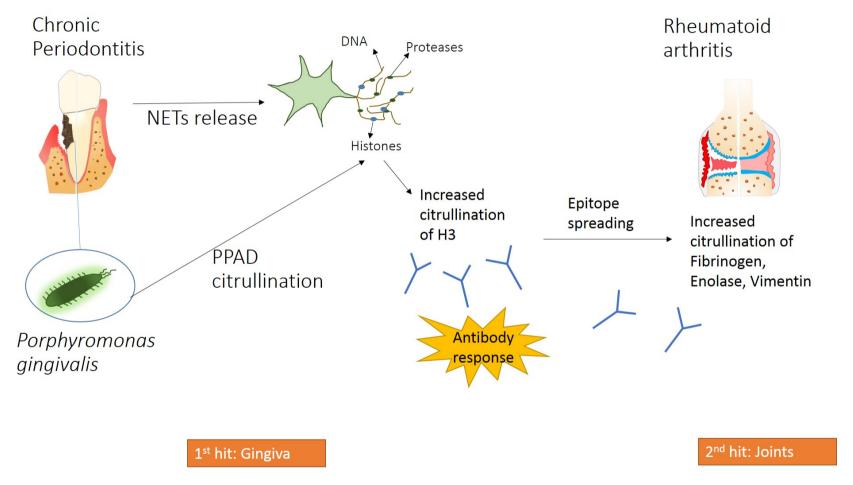


Figure 4. Hypothesis of the role of NETs in the connection between RA and periodontitis. Abbreviations: NETs, neutrophil extracellular traps; PPAD, peptidyl-arginine deiminase from *Porphyromonas gingivalis*; H3, histone 3.

7. The "Two-hit" model of RA pathogenesis

Golub et al. were the first to describe a theory in which the association between periodontitis and systemic diseases could be explained by a "two hit" model ⁸⁰. In this theory, a first "hit" arises within the periodontum, initiated by an infection that activates a destructive inflammatory cascade in the periodontal tissues. In susceptible patients, a second systemic "hit" then occurs, characterized by increased serum levels of pro-inflammatory cytokines that amplifies the inflammatory cascade, with production of local and systemic pro-inflammatory mediators (cytokines and prostaglandins).

A similar "two hit model" was described 6-years later to explain the breakdown of immune tolerance to citrullinated proteins in RA patients triggered by smoking. ⁸¹ Similar to smoking, periodontitis could represent a first hit of citrullinated peptide production. In a first extra-articular hit, chronic periodontitis could break immune tolerance to citrullinated proteins, due to abnormal levels of protein citrullination within periodontal tissues (the periodontal citrullinome). Through epitope spreading (a process in which the immune response does not remain fixed towards a specific epitope, but extends to include other epitopes on the same protein or other proteins in the same tissue), this local autoimmune response to citrullinated proteins could lead to the production of ACPAs systemically. These ACPAs react against citrullinated proteins in the joint since, in joint diseases there is an increase of protein citrullination in the synovium. This model describes a primary "hit" of ACPA production due to chronic periodontitis followed by a secondary "hit" in the joint, that could induce RA. ^{19 82} (Figure 5)

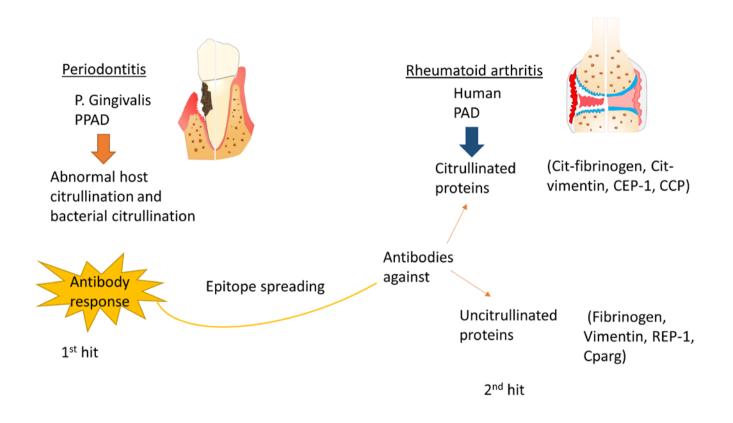


Figure 5. Two hit model proposed to explain the link between periodontitis and RA. Abbreviations: PAD, peptidylarginine deiminase; PPAD, peptidylarginine deiminase from *Phorphyromonas gingivalis*; Cit-, citrullinated; CEP enolase 1; REP non-citrullinated enolase 1; Cparg negative control for anti-CCP; CCP, anti-cyclic citrullinated protein.

8. SUMMARY AND CONCLUSIONS

Although promising, the theories described in this paper to link RA and periodontitis remain to be proven and studies supporting them have limitations. None of the theories described above explain fully why this inflammatory response, initiated or exacerbated by periodontitis, would be specific to the joint structures in RA, and it remains unproven whether this link is coincidental or due to common risk factors.

A causal relationship between RA and periodontitis cannot be established with the current evidence, and if proven, the relationship may only exist in a subset of RA patients.

Therefore, future research investigating the oral micobiome, inflammatory and autoimmune response of pre-RA patients, RA patients and periodontitis patients, is required to clarify gaps in knowledge. Depending upon the outcomes of such studies, therapies targeting the microbiome/specific microorganisms or components of the immune response could help to prevent both diseases.

BIBLIOGRAPHY

- 1. Bingham III CO, Moni M. Periodontal disease and rheumatoid arthritis: the evidence accumulates for complex pathobiologic interactions. *Current opinion in rheumatology* 2013;25(3):345.
- 2. Gregersen PK, Silver J, Winchester RJ. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis & Rheumatology* 1987;30(11):1205-13.
- 3. Stein JM, Machulla HK, Smeets R, et al. Human leukocyte antigen polymorphism in chronic and aggressive periodontitis among Caucasians: a meta-analysis. *J Clin Periodontol* 2008;35(3):183-92. doi: 10.1111/j.1600-051X.2007.01189.x [published Online First: 2008/01/15]
- 4. Michalowicz BS, Diehl SR, Gunsolley JC, et al. Evidence of a substantial genetic basis for risk of adult periodontitis. *Journal of periodontology* 2000;71(11):1699-707.
- 5. Vaithilingam R, Safii S, Baharuddin N, et al. Moving into a new era of periodontal genetic studies: relevance of large case—control samples using severe phenotypes for genome- wide association studies. *Journal of periodontal research* 2014;49(6):683-95.
- 6. Nibali L, Di Iorio A, Onabolu O, et al. Periodontal infectogenomics: systematic review of associations between host genetic variants and subgingival microbial detection. *Journal of clinical periodontology* 2016;43(11):889-900.
- 7. van der Helm- van Mil AH, Verpoort KN, Breedveld FC, et al. The HLA–DRB1 shared epitope alleles are primarily a risk factor for anti–cyclic citrullinated peptide antibodies and are not an independent risk factor for development of rheumatoid arthritis. *Arthritis & Rheumatism* 2006;54(4):1117-21.
- 8. Genetics in rheumatoid arthritis beyond HLA genes: what meta-analyses have shown? Seminars in arthritis and rheumatism; 2013. Elsevier.
- 9. Padyukov L, Seielstad M, Ong RT, et al. A genome-wide association study suggests contrasting associations in ACPA-positive versus ACPA-negative

- rheumatoid arthritis. *Annals of the rheumatic diseases* 2010:annrheumdis126821.
- 10. Havemose- Poulsen A, Sørensen LK, Bendtzen K, et al. Polymorphisms within the IL- 1 gene cluster: Effects on cytokine profiles in peripheral blood and whole blood cell cultures of patients with aggressive periodontitis, juvenile idiopathic arthritis, and rheumatoid arthritis. *Journal of periodontology* 2007;78(3):475-92.
- 11. Kobayashi T, Yokoyama T, Ishida K, et al. Serum cytokine and periodontal profiles in relation to disease activity of rheumatoid arthritis in Japanese adults. *Journal of periodontology* 2010;81(5):650-57.
- 12. Domínguez-Pérez RA, Loyola-Rodriguez JP, Abud-Mendoza C, et al. Association of cytokines polymorphisms with chronic peridontitis and rheumatoid arthritis in a Mexican population. *Acta Odontologica Scandinavica* 2017;75(4):243-48.
- 13. Silman AJ, Newman J, Macgregor AJ. Cigarette smoking increases the risk of rheumatoid arthritis: results from a nationwide study of disease-discordant twins. *Arthritis & rheumatism* 1996;39(5):732-35.
- 14. Eriksson K, Nise L, Alfredsson L, et al. Seropositivity combined with smoking is associated with increased prevalence of periodontitis in patients with rheumatoid arthritis. *Annals of the rheumatic diseases* 2017;annrheumdis-2017-212091.
- 15. Scher JU, Ubeda C, Equinda M, et al. Periodontal disease and the oral microbiota in new-onset rheumatoid arthritis. *Arthritis & Rheumatology* 2012;64(10):3083-94.
- 16. Potikuri D, Dannana KC, Kanchinadam S, et al. Periodontal disease is significantly higher in non-smoking treatment-naive rheumatoid arthritis patients: results from a case-control study. *Annals of the rheumatic diseases* 2012;71(9):1541-44.
- 17. Dissick A, Redman RS, Jones M, et al. Association of Periodontitis With Rheumatoid Arthritis: A Pilot Study. *Journal of Periodontology* 2009;81(2):223-30. doi: 10.1902/jop.2009.090309

- 18. Mikuls TR, Payne JB, Yu F, et al. Periodontitis and Porphyromonas gingivalis in patients with rheumatoid arthritis. *Arthritis & Rheumatology* 2014;66(5):1090-100.
- 19. Kaur S, White S, Bartold P. Periodontal disease and rheumatoid arthritis a systematic review. *Journal of dental research* 2013:0022034513483142.
- 20. Marotte H, Farge P, Gaudin P, et al. The association between periodontal disease and joint destruction in rheumatoid arthritis extends the link between the HLA-DR shared epitope and severity of bone destruction. *Annals of the rheumatic diseases* 2006;65(7):905-09. doi: 10.1136/ard.2005.036913
- 21. Torkzaban P, Hjiabadi T, Basiri Z, et al. Effect of rheumatoid arthritis on periodontitis: a historical cohort study. *J Periodontal Implant Sci* 2012;42(3):67-72.
- 22. Mercado FB, Marshall RI, Klestov AC, et al. Relationship Between Rheumatoid Arthritis and Periodontitis. *Journal of Periodontology* 2001;72(6):779-87. doi: 10.1902/jop.2001.72.6.779
- 23. Pischon N, Pischon T, Kröger J, et al. Association Among Rheumatoid Arthritis, Oral Hygiene, and Periodontitis. *Journal of Periodontology* 2008;79(6):979-86. doi: 10.1902/jop.2008.070501
- 24. Brown L, Brunelle J, Kingman A. Periodontal status in the United States, 1988–91: prevalence, extent, and demographic variation. *Journal of dental research* 1996;75(2_suppl):672-83.
- 25. Jafri Z, Bhardwaj A, Sawai M, et al. Influence of female sex hormones on periodontium: A case series. *Journal of Natural Science, Biology, and Medicine* 2015;6(Suppl 1):S146-S49. doi: 10.4103/0976-9668.166124
- 26. Baka Z, György B, Géher P, et al. Citrullination under physiological and pathological conditions. *Joint Bone Spine* 2012;79(5):431-36.
- 27. Harvey G, Fitzsimmons T, Dhamarpatni A, et al. Expression of peptidylarginine deiminase- 2 and- 4, citrullinated proteins and anti-citrullinated protein antibodies in human gingiva. *Journal of Periodontal Research* 2013;48(2):252-61.
- 28. Hajishengallis G, Darveau RP, Curtis MA. The keystone-pathogen hypothesis. *Nature Reviews Microbiology* 2012;10(10):717-25.

- 29. Darveau R, Hajishengallis G, Curtis M. Porphyromonas gingivalis as a potential community activist for disease. *Journal of dental research* 2012;91(9):816-20.
- 30. Darveau RP. Periodontitis: a polymicrobial disruption of host homeostasis. *Nature Reviews Microbiology* 2010;8(7):481-90.
- 31. Wegner N, Wait R, Sroka A, et al. Peptidylarginine deiminase from Porphyromonas gingivalis citrullinates human fibrinogen and α enolase: Implications for autoimmunity in rheumatoid arthritis. *Arthritis & Rheumatism* 2010;62(9):2662-72.
- 32. Nomura K. Specificity and mode of action of the muscle-type proteinarginine deiminase. *Archives of biochemistry and biophysics* 1992;293(2):362-69.
- 33. Rosenstein ED, Greenwald RA, Kushner LJ, et al. Hypothesis: the humoral immune response to oral bacteria provides a stimulus for the development of rheumatoid arthritis. *Inflammation* 2004;28(6):311-18.
- 34. Martinez-Martinez RE, Abud-Mendoza C, Patiño-Marin N, et al. Detection of periodontal bacterial DNA in serum and synovial fluid in refractory rheumatoid arthritis patients. *Journal of clinical periodontology* 2009;36(12):1004-10.
- 35. Hitchon CA, Chandad F, Ferucci ED, et al. Antibodies to Porphyromonas gingivalis are associated with anticitrullinated protein antibodies in patients with rheumatoid arthritis and their relatives. *The Journal of rheumatology* 2010;37(6):1105-12.
- 36. Mikuls TR, Thiele GM, Deane KD, et al. Porphyromonas gingivalis and disease- related autoantibodies in individuals at increased risk of rheumatoid arthritis. *Arthritis & Rheumatism* 2012;64(11):3522-30.
- 37. Nesse W, Westra J, van der Wal JE, et al. The periodontium of periodontitis patients contains citrullinated proteins which may play a role in ACPA (anti-citrullinated protein antibody) formation. *J Clin Periodontol* 2012;39(7):599-607. doi: 10.1111/j.1600-051X.2012.01885.x [published Online First: 2012/04/26]
- 38. de Pablo P, Dietrich T, Chapple IL, et al. The autoantibody repertoire in periodontitis: a role in the induction of autoimmunity to citrullinated proteins in

- rheumatoid arthritis? *Annals of the rheumatic diseases* 2014;73(3):580-6. doi: 10.1136/annrheumdis-2012-202701 [published Online First: 2013/02/26]
- 39. Mangat P, Wegner N, Venables PJ, et al. Bacterial and human peptidylarginine deiminases: targets for inhibiting the autoimmune response in rheumatoid arthritis? *Arthritis Research & Therapy* 2010;12(3):209-09. doi: 10.1186/ar3000
- 40. Konig MF, Abusleme L, Reinholdt J, et al. Aggregatibacter actinomycetemcomitans—induced hypercitrullination links periodontal infection to autoimmunity in rheumatoid arthritis. *Science translational medicine* 2016;8(369):369ra176-369ra176.
- 41. Lopez- Oliva I, Paropkari AD, Saraswat S, et al. Dysbiotic subgingival microbial communities in periodontally healthy patients with rheumatoid arthritis. *Arthritis & Rheumatology* 2018
- 42. Kumar P, S, Griffen A, L, Barton J, A, et al. New bacterial species associated with chronic periodontitis. *J Dent Res* 2003;5(82):338-44.
- 43. Uematsu H, Sato N, Djais A, et al. Degradation of arginine by Slackia exigua ATCC 700122 and Cryptobacterium curtum ATCC 700683. *Oral Microbiology and Immunology* 2006;21(6):381-84. doi: 10.1111/j.1399-302X.2006.00307.x
- 44. Vaahtovuo J, Munukka E, Korkeamaki M, et al. Fecal microbiota in early rheumatoid arthritis. *J Rheumatol* 2008;35(8):1500-5. [published Online First: 2008/06/06]
- 45. Nielen MM, van Schaardenburg D, Reesink HW, et al. Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. *Arthritis and rheumatism* 2004;50(2):380-6. doi: 10.1002/art.20018 [published Online First: 2004/02/12]
- 46. Montgomery AB, Venables PJ, Fisher BA. The case for measuring antibodies to specific citrullinated antigens. *Expert review of clinical immunology* 2013;9(12):1185-92.
- 47. Raza K, Schwenzer A, Juarez M, et al. Detection of antibodies to citrullinated tenascin-C in patients with early synovitis is associated with the development of rheumatoid arthritis. *RMD open* 2016;2(2):e000318. doi: 10.1136/rmdopen-2016-000318 [published Online First: 2016/12/10]

- 48. Zhao X, Okeke NL, Sharpe O, et al. Circulating immune complexes contain citrullinated fibrinogen in rheumatoid arthritis. *Arthritis research & therapy* 2008;10(4):R94.
- 49. Gilliam BE, Reed MR, Chauhan AK, et al. Evidence of fibrinogen as a target of citrullination in IgM rheumatoid factor-positive polyarticular juvenile idiopathic arthritis. *Pediatric Rheumatology* 2011;9(1):8.
- 50. Schwenzer A, Jiang X, Mikuls TR, et al. Identification of an immunodominant peptide from citrullinated tenascin-C as a major target for autoantibodies in rheumatoid arthritis. *Annals of the rheumatic diseases* 2016;75(10):1876-83. doi: 10.1136/annrheumdis-2015-208495 [published Online First: 2015/12/15]
- 51. Thiele GM, Duryee MJ, Anderson DR, et al. Malondialdehyde-Acetaldehyde Adducts and Anti–Malondialdehyde-Acetaldehyde Antibodies in Rheumatoid Arthritis. *Arthritis & rheumatology* 2015;67(3):645-55.
- 52. Bright R, Thiele G, Manavis J, et al. Gingival tissue, an extrasynovial source of malondialdehyde- acetaldehyde adducts, citrullinated and carbamylated proteins. *Journal of periodontal research* 2018;53(1):139-43.
- 53. Quirke A-M, Lugli EB, Wegner N, et al. Heightened immune response to autocitrullinated Porphyromonas gingivalis peptidylarginine deiminase: a potential mechanism for breaching immunologic tolerance in rheumatoid arthritis. *Annals of the rheumatic diseases* 2013:annrheumdis-2012-202726.
- 54. Johansson L, Sherina N, Kharlamova N, et al. Concentration of antibodies against Porphyromonas gingivalis is increased before the onset of symptoms of rheumatoid arthritis. *Arthritis research & therapy* 2016;18(1):201.
- 55. Fisher BA, Cartwright AJ, Quirke A-M, et al. Smoking, Porphyromonas gingivalis and the immune response to citrullinated autoantigens before the clinical onset of rheumatoid arthritis in a Southern European nested case—control study. *BMC musculoskeletal disorders* 2015;16(1):331.
- 56. Quirke A-M, Lugli EB, Wegner N, et al. Heightened immune response to autocitrullinated Porphyromonas gingivalis peptidylarginine deiminase: a potential mechanism for breaching immunologic tolerance in rheumatoid arthritis. *Annals of the rheumatic diseases* 2014;73(1):263-69. doi: 10.1136/annrheumdis-2012-202726

- 57. Turnbaugh PJ, Ley RE, Hamady M, et al. The human microbiome project: exploring the microbial part of ourselves in a changing world. *Nature* 2007;449(7164):804.
- 58. Zhang X, Zhang D, Jia H, et al. The oral and gut microbiomes are perturbed in rheumatoid arthritis and partly normalized after treatment. *Nature medicine* 2015;21(8):895-905. doi: 10.1038/nm.3914 [published Online First: 2015/07/28]
- 59. Feldmann M, Brennan FM, Maini RN. Role of cytokines in rheumatoid arthritis. *Annual review of immunology* 1996;14(1):397-440.
- 60. Gemmell E, Carter CL, Seymour GJ. Chemokines in human periodontal disease tissues. *Clinical and experimental immunology* 2001;125(1):134-41. [published Online First: 2001/07/27]
- 61. Azizi G, Jadidi- Niaragh F, Mirshafiey A. Th17 Cells in Immunopathogenesis and treatment of rheumatoid arthritis. *International journal of rheumatic diseases* 2013;16(3):243-53.
- 62. Ortiz P, Bissada N, Palomo L, et al. Periodontal therapy reduces the severity of active rheumatoid arthritis in patients treated with or without tumor necrosis factor inhibitors. *Journal of periodontology* 2009;80(4):535-40.
- 63. Pers J-O, Saraux A, Pierre R, et al. Anti–TNF-α Immunotherapy Is Associated With Increased Gingival Inflammation Without Clinical Attachment Loss in Subjects With Rheumatoid Arthritis. *Journal of periodontology* 2008;79(9):1645-51.
- 64. Mayer Y, Balbir-Gurman A, Machtei EE. Anti-tumor necrosis factor-alpha therapy and periodontal parameters in patients with rheumatoid arthritis. *Journal of periodontology* 2009;80(9):1414-20.
- 65. Üstün K, Erciyas K, Kısacık B, et al. Host modulation in rheumatoid arthritis patients with TNF blockers significantly decreases biochemical parameters in periodontitis. *Inflammation* 2013;36(5):1171-77.
- 66. Kobayashi T, Yokoyama T, Ito S, et al. Periodontal and serum protein profiles in patients with rheumatoid arthritis treated with tumor necrosis factor inhibitor adalimumab. *Journal of periodontology* 2014;85(11):1480-88.
- 67. Antoni C, Braun J. Side effects of anti-TNF therapy: current knowledge. *Clinical and experimental rheumatology* 2002;20(6; SUPP/28):S-152.

- 68. O'connell PA, Taba Jr M, Nomizo A, et al. Effects of periodontal therapy on glycemic control and inflammatory markers. *Journal of periodontology* 2008;79(5):774-83.
- 69. Nishimura F, Iwamoto Y, Mineshiba J, et al. Periodontal disease and diabetes mellitus: the role of tumor necrosis factor-α in a 2-way relationship. *Journal of Periodontology* 2003;74(1):97-102.
- 70. Yamazaki K, Honda T, Oda T, et al. Effect of periodontal treatment on the C- reactive protein and proinflammatory cytokine levels in Japanese periodontitis patients. *Journal of periodontal research* 2005;40(1):53-58.
- 71. Shimada Y, Komatsu Y, Ikezawa-Suzuki I, et al. The effect of periodontal treatment on serum leptin, interleukin-6, and C-reactive protein. *Journal of periodontology* 2010;81(8):1118-23.
- 72. D'aiuto F, Nibali L, Parkar M, et al. Short-term effects of intensive periodontal therapy on serum inflammatory markers and cholesterol. *Journal of dental research* 2005;84(3):269-73.
- 73. Kurgan Ş, Fentoğlu Ö, Önder C, et al. The effects of periodontal therapy on gingival crevicular fluid matrix metalloproteinase- 8, interleukin- 6 and prostaglandin E2 levels in patients with rheumatoid arthritis. *Journal of periodontal research* 2016;51(5):586-95.
- 74. Liou TG, Campbell EJ. Quantum proteolysis resulting from release of single granules by human neutrophils: a novel, nonoxidative mechanism of extracellular proteolytic activity. *The Journal of Immunology* 1996;157(6):2624-31.
- 75. Kolaczkowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nature Reviews Immunology* 2013;13(3):159.
- 76. Cooper PR, Palmer LJ, Chapple IL. Neutrophil extracellular traps as a new paradigm in innate immunity: friend or foe? *Periodontology 2000* 2013;63(1):165-97.
- 77. Spengler J, Lugonja B, Jimmy Ytterberg A, et al. Release of active peptidyl arginine deiminases by neutrophils can explain production of extracellular citrullinated autoantigens in rheumatoid arthritis synovial fluid. *Arthritis & rheumatology* 2015;67(12):3135-45.

- 78. Khandpur R, Carmona-Rivera C, Vivekanandan-Giri A, et al. NETs are a source of citrullinated autoantigens and stimulate inflammatory responses in rheumatoid arthritis. *Science translational medicine* 2013;5(178):178ra40-78ra40.
- 79. White P, Sakellari D, Roberts H, et al. Peripheral blood neutrophil extracellular trap production and degradation in chronic periodontitis. *Journal of clinical periodontology* 2016;43(12):1041-49.
- 80. Golub L, Payne JB, Reinhardt RA, et al. Can systemic diseases co-induce (not just exacerbate) periodontitis? A hypothetical "two-hit" model. *Journal of Dental Research* 2006;85(2):102-05.
- 81. Farquharson D, Butcher J, Culshaw S. Periodontitis, Porphyromonas, and the pathogenesis of rheumatoid arthritis. *Mucosal immunology* 2012;5(2):112.
- 82. Lundberg K, Wegner N, Yucel-Lindberg T, et al. Periodontitis in RA—the citrullinated enolase connection. *Nature Reviews Rheumatology* 2010;6(12):727-30.