

# The medical therapy of craniopharyngiomas

Alexandraki, Krystallenia I; Kaltsas, Gregory A; Karavitaki, Niki; Grossman, Ashley

DOI:

[10.1210/jc.2019-01299](https://doi.org/10.1210/jc.2019-01299)

License:

Other (please specify with Rights Statement)

Document Version

Peer reviewed version

Citation for published version (Harvard):

Alexandraki, KI, Kaltsas, GA, Karavitaki, N & Grossman, A 2019, 'The medical therapy of craniopharyngiomas: the way ahead', *Journal of Clinical Endocrinology and Metabolism*, vol. 104, no. 12, pp. 5751–5764.  
<https://doi.org/10.1210/jc.2019-01299>

[Link to publication on Research at Birmingham portal](#)

## Publisher Rights Statement:

This is a pre-copyedited, author-produced PDF of an article accepted for publication in *Journal of Clinical Endocrinology and Metabolism* following peer review. The version of record

Krystallenia I Alexandraki, Gregory A Kaltsas, Niki Karavitaki, Ashley B Grossman, The Medical Therapy of Craniopharyngiomas: The Way Ahead, *The Journal of Clinical Endocrinology & Metabolism*, Volume 104, Issue 12, December 2019, Pages 5751–5764,  
<https://doi.org/10.1210/jc.2019-01299>

is available online at: <https://academic.oup.com/jcem/article-abstract/104/12/5751/5540964>

## General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

## Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact [UBIRA@lists.bham.ac.uk](mailto:UBIRA@lists.bham.ac.uk) providing details and we will remove access to the work immediately and investigate.

1 **Title Page**

2 **Full title: THE MEDICAL THERAPY OF CRANIOPHARYNGIOMAS: THE WAY**  
3 **AHEAD**

4

5 Krystallenia I. Alexandraki<sup>1</sup>, Gregory A. Kaltsas<sup>1</sup>, Niki Karavitaki<sup>\*2,3,4</sup> and Ashley B.  
6 Grossman<sup>\*5,6</sup>

7

8 <sup>1</sup>Endocrine Unit, 1st Department of Propaedeutic Medicine, Laiko University Hospital,  
9 Medical School, National and Kapodistrian University of Athens, Athens, Greece

10 <sup>2</sup>Institute of Metabolism and Systems Research, College of Medical and Dental  
11 Sciences, University of Birmingham, Birmingham, UK.

12 <sup>3</sup>Centre for Endocrinology, Diabetes and Metabolism, Birmingham Health Partners,  
13 Birmingham, UK.

14 <sup>4</sup>Department of Endocrinology, Queen Elizabeth Hospital, University Hospitals  
15 Birmingham NHS Foundation Trust, Birmingham, UK.

16 <sup>5</sup>Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford,  
17 Oxford, UK

18 <sup>6</sup>Centre for Endocrinology, William Harvey Institute, Barts and the London School of  
19 Medicine, London, UK

20

21

22 **Short title:** Medical therapy of craniopharyngiomas

23

24 **Key words:** Craniopharyngioma – BRAF –  $\beta$ -catenin – Wnt signaling therapy –  
25 vemurafenib – dabrafenib - trametinib – cobimetinib

26

27 **Correspondence to:**

28 Dr. K. Alexandraki, Endocrine Unit, 1st Department of Propaedeutic Medicine, Laiko  
29 University Hospital, Medical School, National and Kapodistrian University of Athens,  
30 Athens, Greece, Email: [alexandrakik@gmail.com](mailto:alexandrakik@gmail.com)

31

32 **\*NK and ABG are to be considered as joint senior authors**

33 **DISCLOSURE STATEMENT:** The authors have nothing to disclose for this specific  
34 work.

35

## Abstract

**Context:** Craniopharyngiomas which are categorized as adamantinomatous (ACP) or papillary (PCP) have traditionally been treated with surgery and/or radiotherapy, although when the tumors progress or recur further therapeutic possibilities are very limited. albeit with limited therapeutic options available for recurrent or progressive disease. Following recent advances in their molecular pathogenesis, further medical therapeutic options have emerged.

**Evidence Acquisition:** The search strategy that we selected to identify the appropriate evidence involved the following mesh terms: ("Craniopharyngioma"[Mesh] AND "Craniopharyngioma/drug therapy"[Mesh] NOT ("review"[Publication Type] OR "review literature as topic"[MeSH Terms] OR "review"[All Fields]) AND ("2009/05/01"[PDat]: "2019/04/28"[PDat])).

**Evidence Synthesis:** Mutations of  $\beta$ -catenin causing Wnt activation with alterations of the MEK/ERK pathway are encountered in the great majority of patients with ACP; specific alterations also stratify patients to a more aggressive behavior. In most PCP there is primary activation of the Ras/Raf/MEK/ERK pathway secondary to *BRAF*-V600E mutations. BRAF inhibitors, such as dabrafenib or vemurafenib, either alone or in combination with the MEK inhibitors trametinib and cobimetinib, have been administered in patients with PCP producing clinically useful and, in some cases, sustained responses. In contrast to PCP, drugs targeting  $\beta$ -catenin and its downstream MAPK pathway in ACP have so far only been used in *in vitro* studies, but appear to be promising new targets clinically.

**Conclusions:** The identification of specific genetic alterations in patients with craniopharyngiomas has expanded the therapeutic options, providing evidence for a a customized approach using newer molecular agents. more personalized approach with molecular targeted drugs. More studies including a larger number of carefully selected patients are required to evaluate the response to currently available and evolving agents alone and in combination.

65 **Précis**

66 The presence of specific genetic alterations in craniopharyngiomas has important  
67 implications for the diagnosis and potential treatments of these neoplasms with  
68 molecular targeted agents.

69

## Introduction

The title of this review may sound like an oxymoron, as until very recently the mainstays of treatment for craniopharyngiomas (CP) have been surgery and radiotherapy (RT). However, over the last few years there have been major developments in our understanding of the molecular pathology of these tumors that could set the scene for specific targeted therapy.

Craniopharyngiomas develop ~~as malformations of embryonic remnants along the original as embryonic malformations in the sellar and parasellar regions arising along the~~ pathway of the craniopharyngeal duct. They are characterized as mostly benign epithelial tumors according to the World Health Organization (WHO) classification of *Tumors of the Central Nervous System* (CNS), namely WHO grade I tumors (1-3). They are relatively rare, with an incidence of 0.5-2.5/ 10<sup>6</sup> cases/ year (2,4-7), and are divided into two distinct subtypes, adamantinomatous CP (ACP) and papillary CP (PCP), differing in both histological features and genetic alterations (8). ACP is more prevalent and displays a bimodal age distribution, with peaks between the ages 5-15 years and 45-60 years, but can occur at any age, even during the neonatal period (3,9,10); PCP has been classically considered as an adult entity peaking between 40 and 55 years (2,4,7,11), although pediatric cases have occasionally been reported (12).

Harvey Cushing not only suggested the term craniopharyngioma but also recognized the significant challenges in their management (13). Indeed, their anatomic vicinity to vital structures (hypothalamus, optic pathways, brain) and their intrinsic local infiltrative tendency render their growth pattern unpredictable and complete safe

Formatted: Not Highlight

95 surgical removal problematic (14). Furthermore, malignant transformation may also  
96 rarely be seen and, overall, the long-term morbidities and mortality of patients with  
97 craniopharyngioma are disappointingly high, particularly compared to patients with  
98 treated pituitary neuroendocrine tumors (PitNETs) (15,16).

99  
100 Current therapeutic options are relatively limited: surgery, either with the intention to  
101 treat by gross total resection (GTR) or with the intention to reduce their mass by  
102 subtotal/ partial resection (STR) (17,18), with or without post-operative radiotherapy  
103 (RT) (19), is the primary therapeutic modality (3). In a systematic review, STR with  
104 adjuvant RT, and GTR, showed similar rates of long-term tumor control (20).  
105 Currently, STR followed by RT is recommended, unless there is a clear margin  
106 between the tumor and surrounding vital structures allowing complete safe tumor  
107 removal. Whenever feasible, the tumours are approached by the transsphenoidal  
108 route. The trans-sphenoidal approach is used whenever possible (17,18). The intra-  
109 cystic administration of sclerosing substances, mainly bleomycin and interferon-  
110 alpha, has been suggested in the past to attenuate fluid formation and diminish cyst  
111 size (21-23). Bleomycin use is limited by its toxicity (21,22), while interferon-alpha  
112 has shown some benefit with an acceptable safety profile (24); however, no  
113 prospective or randomized study has been performed and there are isolated reports  
114 of relatively poor responses and major adverse effects following interferon-alpha  
115 instillation, limiting its use (21).

116  
117 Recently, progress in molecular biology has unraveled novel potentially druggable  
118 molecular pathways in CPs that offer new promising therapeutic options. The use of  
119 such drugs is especially important to minimize the otherwise adverse effects

Formatted: Not Highlight

associated with surgery and RT, particularly in recurrent tumors where more there  
are few or no alternative options. ~~conventional options have been exhausted.~~ The  
purpose of this mini-review is to present the available literature on clinical studies and  
case reports describing targeted-treatments which are based on the recently-  
identified molecular alterations in both ACP and PCP. The search strategy that we  
selected to identify the appropriate evidence involved the following mesh terms:  
("Craniopharyngioma"[Mesh] AND "Craniopharyngioma/drug therapy"[Mesh] NOT  
("review"[Publication Type] OR "review literature as topic"[MeSH Terms] OR  
"review"[All Fields]) AND ("2009/05/01"[PDat]: "2019/04/28"[PDat]) (**Figure 1**).

## ***Molecular biological advances in ACP and PCP***

It has recently been shown that each CP histological subtype (Figures 2 and 3) is characterized by alterations in oncogenic molecular signaling pathways, harboring distinct mutational, transcriptomic and epigenomic profiles as a result of different gene mutations, gene expression and methylation patterns (25-28). These findings have shed light on the identification of tumor-specific signaling pathway activation that may lead to CP-targeted therapies. An additional role of the different molecular profiles may be to differentiate pathologically indeterminate cases of CP (28-29).

Sekine *et al.* first described activating mutations of the  $\beta$ -catenin encoding gene *CTNNB1* in ACPs as early as 2002 (30). Whole-exome sequencing revealed mutations in *CTNNB1* in the majority of a small group of ACPs (11/12, 92%) (25). In addition, targeted genotyping revealed *CTNNB1* mutations in 96% of ACPs (51/53) (25). Next-generation panel sequencing did not identify any mutations other than those in *CTNNB1* (27). In 76% of the ACPs, a mutation in exon 3 of *CTNNB1* was found, and there was a trend towards a worse event-free survival in cases mutated at Thr41 (27). When primary and recurrent ACP tumor samples from the same patient were tested, the same *CTNNB1* mutation (S45P) was found without additional ones (28). No significant large chromosomal aberrations have been found, although a fraction of ACPs showed recurrent focal gains of chromosomal material, while in other cases ~~there was loss of, showed loss in the~~ chromosomal region Xq28 (27).

These *CTNNB1* mutations are exclusively found in exon 3 which encodes the destruction/degradation complex of  $\beta$ -catenin:  $\beta$ -catenin is usually held in a multi-

Formatted: Not Highlight



157 protein complex and, when phosphorylated at specific residues, is subject to rapid  
158 degradation. Mutation leading to a loss of such phosphorylation sites results in  
159 aberrant  $\beta$ -catenin nucleo-cytoplasmic accumulation (30) (**Figure 3A**). This  
160 represents the most reliable immunohistochemical (IHC) marker for the confirmation  
161 of ACP, aiding in the differential diagnosis from other (para)sellar lesions (31,32);  $\beta$ -  
162 catenin is a component of the adherens junction complex and a central mediator of  
163 the canonical Wnt molecular signaling pathway (8). In the cytoplasm,  $\beta$ -catenin  
164 maintains a low concentration when phosphorylated within the multi-protein  
165 destruction-regulator complex which facilitates its ubiquitination and degradation by  
166 the proteasome. This process corresponds to inactive Wnt signaling (default "off"  
167 state) in the absence of any Wnt ligands (8,33-35) (**Figure 4A**). This regulator  
168 complex includes a number of proteins such as the scaffold proteins Axin and Axin2,  
169 the tumor suppressor protein adenomatosis polyposis coli (APC), and the  
170 phosphokinase glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ), casein kinase 1 $\alpha$  (CK1),  
171 protein phosphatase 2A (PP2A) or alternatively by I kappa B kinase alpha (IKK- $\alpha$ )  
172 (36). APC is phosphorylated by these kinases, resulting in recruitment of  $\beta$ -catenin to  
173 the complex (34,36), which is then tagged by these kinases for degradation through  
174 phosphorylation of the serine (Ser33, Ser37, Ser45) or threonine (Thr41) residues (of  
175 exon 3 on the *CTNNB1*) (34,36-39). This phosphorylation pattern is then recognized  
176 by multiple ubiquitin molecules, such as a component of E3 ubiquitin ligase,  $\beta$ -TrCP,  
177 ubiquitinating  $\beta$ -catenin, resulting in dissociation of  $\alpha$ -catenin from  $\beta$ -catenin with  
178 concomitant loss of cadherin adhesion (34,35,37-41). Exogenous Wnt signaling  
179 ligands 'switch on' i.e., activate Wnt signaling (41,42). The Wnt protein family  
180 includes approximately 20 different proteins that bind to the Frizzled (Fz) family of  
181 receptors; the Wnt pathway is activated (34) resulting in an intracellular signaling

182 cascade that promotes dimerisation of Fz with LRP5/6 (34,43) The Fz/LRP5/6 dimer  
183 can bind to Axin, GSK3 $\beta$ , and CK1, which facilitates their accumulation at the cell  
184 membrane, preventing them from joining the  $\beta$ -catenin destruction complex (34-  
185 36,43,44), i.e. disrupting the regulator complex via Dishevelled (Dvl) by preventing  
186 the phosphorylation of  $\beta$ -catenin and its subsequent degradation (45).Therefore,  $\beta$ -  
187 catenin protein accumulates and ultimately translocates to the nucleus, where it  
188 activates factors allowing transcription of  $\beta$ -catenin target genes, such as CyclinD1,  
189 c-Myc, CD44, Survivin, VEGF or fascin (26,46-49), resulting in stimulation of cellular  
190 proliferation and other Wnt-regulated cellular processes (50). Cellular localisation of  
191  $\beta$ -catenin is not *per se* sufficient to promote target gene activation, as opposed to  
192 *nuclear*  $\beta$ -catenin accumulation which is sufficient to induce Wnt signaling target  
193 gene expression. Target genes (Axin2, bone morphogenetic protein (BMP)4 and  
194 Fascin-1), stem cell markers (CD133 and CD44) and the cell cycle inhibitor  
195 p21<sup>Waf1/CIP1</sup> co-localise in cells with nuclear  $\beta$ -catenin (8). Fascin (48) acts as a  
196 facilitator of tumor cell migration, necessary for invasion and dissociation (36);  $\beta$ -  
197 catenin binds fascin by its armadillo-repeat sequence (essential for the passive  
198 nuclear-pore-complex translocation of  $\beta$ -catenin) (51) which promotes relocation of  $\beta$ -  
199 catenin, changes cell adhesion properties, and reduces  $\beta$ -catenin destruction (36).  
200 Inhibition of fascin or  $\beta$ -catenin expression has been shown to decrease the  
201 migratory capacity of ACP tumor cells in culture (36). Other binding factors for the  
202 armadillo-repeat sequences include the TCP-family transcription factors, Axin2, and  
203 APC (52). In ACP, distinct  $\beta$ -catenin clusters also result by activation of other growth  
204 factor pathways, such as epidermal growth factor receptor (EGFR), or src-driven  
205 phosphorylation at the tyrosine of codon 654 (Y654) and sonic hedgehog (SHH)  
206 signaling pathways (8,26,28,48,52-55), which may cause the dissociation of  $\beta$ -

catenin from the adherens complex by reducing its binding affinity to E-cadherin (40,56), and may facilitate tumor stem cell maintenance (57-59). In addition, Fyn, Fer or c-Met promote phosphorylation of  $\beta$ -catenin tyrosine residue 142, which results in dissociation of  $\alpha$ -catenin from  $\beta$ -catenin (**Figure 4A**). RNA profiling (60) has confirmed that human ACP clusters express high levels of members of the fibroblast growth factor (FGF), BMP and Wnt families (50), representing paracrine growth-related signaling molecules that play a wide range of developmental and physiological roles. FGF-2 is expressed only in recurrent ACPs, while PDGFR- $\alpha$  expression is significantly higher in recurrent ACP compared with non-recurrent tumors, and VEGF and fibronectin are present in both ACP and PCP ~~were expressed in both types of tumors~~ (61). ACP is characterized by high gene expression of genes encoding both BMP2 and BMP4, which are downstream of the Wnt/ $\beta$ -catenin pathway (62,52,53). ~~The~~ EGFR is a member of the human epidermal growth factor receptor (HER) family; the principal ~~while the major~~ downstream signal transduction pathways activated by receptors of the HER family are the Ras-Raf mitogen-activated protein kinase (MAPK) pathway (MAPK/ERK pathway) and the phosphatidylinositol 3-kinase (PI3K)/Akt pathway (8) (**Figure 4B**).

The Ras-Raf-Mek-ERK1/2 is known to be one of the most dysregulated MAPK pathway signaling cascades in human cancer (63,64). Extracellular growth factors (FGFs, EGF, PDGF) bind to and activate receptor tyrosine kinases, causing a downstream phosphorylation cascade which eventually leads to transcription of target genes regulating cellular proliferation, differentiation, apoptosis and senescence (63). Downstream activation of the MAPK/ERK pathway, as evidenced by phosphorylation of ERK1/2, was identified by IHC around the clusters with

Formatted: Not Highlight

particularly prominent staining at the forefront, known as the leading edge, of the  
tumor at the leading edge of tissue invasion (50). Phospholipase C $\gamma$  (PLC $\gamma$ ), the  
signal transducer and activator of transcription (STAT) and src/FAK cascades, are  
also stimulated (8,65,66). SHH signaling pathway gene products, such as Gli2 and  
patched (Ptc)1, are over-expressed in ACP, and their cleaved active forms are  
especially highly expressed (8,26,28,53). Up-regulation of certain matrix  
metalloproteinases (MMPs) has also been reported in ACP transcriptome studies  
(23,50). A variable degree of co-localization between Ki-67 and pERK1/2 expression  
has also been also shown (60). In ACP, whole-exome sequencing showed additional  
mutations in genes listed in the Cancer Gene Census (67), although many of these  
may be 'passenger' mutations (25); there is also evidence for, recurrent copy  
number changes in some tumors, while some ACP showed recurrent focal copy-  
number aberrations at least in subgroups of cases (27). Thus, mutation-produced  
loss of phosphorylation sites on  $\beta$ -catenin leads to a plethora of downstream changes  
initiating and augmenting tumor growth.

Considering PCPs, a *BRAF*-V600E mutation was documented (25) by targeted  
genotyping in 95% of PCPs (36/39), while whole-exome sequencing revealed  
recurrent mutations in *BRAF*-V600E in all PCPs (3/3) (25). In a parallel study, an  
analysis of known oncogenes revealed that *BRAF*-V600E mutations were seen in  
81% (17/21) of PCPs by targeted Sanger sequencing and in 86% (18/21) of PCPs by  
IHC (68). Agreement between methods was seen in 95% (20/21) of cases; however,  
interpretation of anti-*BRAF*-V600E staining was challenging due to occasional non-  
specific reactivity (68). It is well established that the gain-of-function mutation *BRAF*-  
V600E, a critical serine/threonine kinase in the Ras-Raf-Mek-ERK1/2 pathway,

Formatted: Not Highlight

renders it a potent oncogene leading to increased cell proliferation and survival, resulting in cell transformation and tumorigenesis (69,70) (**Figure 4B**). Such mutations render the BRAF kinase constitutively-active. Mutations in BRAF transform the BRAF kinase into a constitutively-active kinase. The MAPK pathway also plays a role in controlling stem-cell specification during development and perhaps during stem-cell homeostasis in the post-natal period (70), modulating the balance between the proliferation and differentiation of Sox2+ cells, implying that persistent proliferative capacity of Sox2+ cells may underlie the pathogenesis of PCP (70). Sox2+ progenitor cells appear to show tumor-inducing potential, driving tumor formation in a paracrine manner by inducing tumorigenic events in adjacent cells but without gains or losses of genetic material (53,54,71). No gains or losses of genetic material were detected in the PCPs (27). In PCPs (25), whole-exome sequencing with a recently described novel technique - *MuTect* – also showed isolated mutations of cancer-related genes encoding chromatin remodeling factors (CHD5, CHD6) and cell adhesion molecules (CDH26, PTPRT), and one (KIAA1549) that is fused to BRAF in most cases of pilocytic astrocytoma (72,73). However, it is presently unclear whether these mutations are pathogenetic or merely ‘passenger’ mutations (25).

~~Brastianos et al first postulated that~~ *CTNNB1* and *BRAF* mutations were mutually exclusive and clonal in ACP and PCP, according to an original observation by Brastianos et al, and there were no each CP subtype, since they did not detect any other recurrent mutations or genomic aberrations in either subtype (25) (**Figures 3A and 3B**). Thus, the specific oncogenic change ~~The crucial pathogenetic event~~ appears to be Wnt activation in ACP, and activation of the Ras/Raf/MEK/ERK pathway by *BRAF*-V600E mutations in PCP (27). It was then suggested that one

Formatted: Not Highlight

Formatted: Not Highlight

282 should classify CPs as ACP *CTNNB1*-mutated, ACP *CTNNB1*-wild-type, PCP *BRAF*-  
283 V600E-mutated and PCP *BRAF*-wild-type (74). Comparison microarray analysis of  
284 18 ACP and 10 PCP samples revealed significant up-regulation of several direct  
285 targets of the Wnt/ $\beta$ -catenin signaling pathway in ACPs compared to PCP, including  
286 LEF1 and AXIN, while components of the SHH such as GLI2, PTCH1 and SHH were  
287 also over-expressed specifically in ACP in comparison to PCP (8,36,48,52,71,75,76).  
288 Unsupervised consensus clustering of the gene expression values of the 5000 most  
289 variable genes resulted in two distinct and stable clusters that perfectly separated  
290 ACP and PCP samples. Gene expression data showed increased expression of  
291 microtubule-associated protein 2 (MAP2), tenascin C (TNC) and the stem cell marker  
292 CD133 (PROM1) in ACPs. CD44 and claudin 1 (CLDN1) were significantly down-  
293 regulated in ACPs, but showed a recently described distinguishing marker for the  
294 two variants, were significantly down-regulated in ACPs but exhibited significantly  
295 higher expression in PCPs (28,52,75). Moreover, reduction of claudin-1 protein levels  
296 point towards an invasive growth pattern in ACP (75). Down-regulation of microRNA  
297 (miRNA) miR-132 was identified as a marker of aggressiveness, playing a role in  
298 epithelial–mesenchymal transition (EMT), in a study of 754 miRNAs from childhood  
299 CP (77). However, in contrast to the view of a complete molecular separation  
300 between these two tumor subtypes, Larkin *et al* in a small cohort of ACPs reported a  
301 dual aberration of *CTNNB1* mutated at Thr41 together with *BRAF*-V600E mutation in  
302 two tumors (68). This finding was validated by review of morphology and comparison  
303 with IHC findings. Further validation was obtained by Sequencing in forward and  
304 reverse directions sequencing in forward and reverse directions from two DNA  
305 samples extracted on different occasions confirmed these findings. Whether this  
306 represents the rare occurrence of a collision tumor needs to be established in future

Formatted: Not Highlight

Formatted: Not Highlight

307 larger studies (68). The expression of additional stem cell markers SOX9, KLF4 and  
308 Okt4 was also identified by IHC in both ACPs and PCPs (8,78).

309

310 Other transcriptional molecular analyses of ACP have revealed the expression of  
311 immune-system gene expression, including Interleukin (IL)-1 $\beta$ , IL-6, IL-8 (CXCL-8),  
312 IL-10, IL-18 and TNF (TNF- $\alpha$ ). In particular, there is a characteristic inflammatory  
313 cytokine/chemokine reaction in both ACP cyst fluid and solid tumor components.  
314 ACP cyst fluid and solid tumor components are characterized by an inflammatory  
315 cytokine and chemokine expression pattern. Cytokines and chemokines with  
316 elevated concentrations in ACPs included IL-6, CXCL-1 (GRO), CXCL-8 and the  
317 immunosuppressive cytokine IL-10, and their relevant receptors are also expressed  
318 in ACPs., as were IL-10, plus . ~~Most receptors for these cytokines and chemokines~~  
319 ~~are also over-expressed in ACPs. In addition to IL-10, the established~~  
320 immunosuppressive factor IDO-1 ~~was over-expressed by ACPs~~ at both mRNA and  
321 protein levels (79). This interaction between tumor cells and chemokines is not  
322 dissimilar to that seen in some PitNETs (80).

323

324 *Summing up, in the era of molecular targeted treatments, the molecular pathways*  
325 *that characterize either ACP or PCP are under vigorous investigation. Currently, ACP*  
326 *is characterized and identified by activating mutations of the  $\beta$ -catenin-encoding gene*  
327 *CTNNB1, while PCP are characterized by the activating BRAF-V600E mutation. In*  
328 *both cases, there is up-regulation of the MAP-kinase pathway which is critically*  
329 *involved in cell proliferation.*

330

331 **Targeting molecular aberrations in craniopharyngiomas**

Formatted: Not Highlight

332

333 Currently, there are no approved targeted or cytotoxic therapies available for the  
334 systemic treatment of CPs. The development of molecular targeted treatments in  
335 oncology has opened new horizons in the pharmacological treatment of tumors  
336 caused by genetic alterations. Recently, in cancers harboring *BRAF*-V600E  
337 alterations, V600E mutation-specific BRAF inhibitors, such as dabrafenib or  
338 vemurafenib (81,82), have been shown to be effective anti-tumor agents. In addition,  
339 the MEK inhibitors trametinib and cobimetinib have been administered in combination  
340 with BRAF inhibitors, since these compounds override any resistance to BRAF  
341 inhibition (81,83). These data have led to the use of BRAF inhibitors in patients with  
342 aggressive PCPs (see Table 1 for summary). Aylwin *et al.* (83) described the  
343 beneficial effect of the off-label administration of vemurafenib (960 mg twice daily) in  
344 a 41 year-old female with a 16-year history of a recurrent PCP (carrying a *BRAF*-  
345 V600E mutation) with progressive visual failure following three trans-sphenoidal  
346 operations. Magnetic resonance imaging (MRI) two weeks after starting treatment  
347 showed marked reduction in the size of the tumor with resolution of the surrounding  
348 edema, while three months later there was evidence of radiological improvement.  
349 after starting vemurafenib radiological remission was seen. The craniopharyngioma  
350 recurred six weeks later and vemurafenib was re-started with reduction of tumor size  
351 followed by stabilization for seven months but this was followed by further growth. In  
352 the same year, Brastianos *et al.* (84), reported on the use of dabrafenib (150mg twice  
353 daily) in a 39 year-old male with a PCP harboring a *BRAF*-V600E mutation. This  
354 patient had undergone four craniotomies along with cyst decompression over a  
355 period of 11 months: as early as four days after treatment, the tumor showed a a  
356 23% size reduction while the cyst volume decreased by 32%. in tumor size and a

Formatted: Not Highlight



32% decrease in cyst volume were identified. The beneficial effect was even more obvious 17 days after treatment, achieving a 52% reduction in tumor size and 70% decrease in the cyst volume. The MEK inhibitor trametinib (2mg, orally, twice daily) was added to the therapeutic scheme on day 21 aiming to enhance the effect. Fourteen days after this combined therapy, there was a 85% reduction in tumor size and 81% decrease in the cyst volume. On day 38 of treatment, the patient underwent endoscopic trans-sphenoidal surgery: two weeks later the combined treatment was discontinued, and a week later RT was offered (50.4 Gy in 28 fractions). Seven months later, the patient has remained free of symptomatic recurrence. In the following year, Roque & Odia (29) described a 47 year-old female with acute visual loss and a 4-month history of amenorrhea, cold intolerance and headache. A 2.7cm cystic lesion was identified in the suprasellar area harboring a *BRAF-V600E* mutation (29). Subsequently, she underwent right frontal craniotomy but the tumor regrew causing bilateral visual impairment necessitating an Ommaya catheter placement into the cyst and then RT (54 Gy in 30 fractions). Seven months later, the tumor regrew and dabrafenib (150mg twice daily) and trametinib (2-mg daily) were administered. After two months, a 52% reduction in tumor volume was documented, and after five months, the tumor size was reduced by 75%. The patient was continuing to improve on this treatment both radiologically and clinically 7 months post-treatment. Subsequently, Rostami *et al.* (85) reported a 65 year-old male with visual deficits who underwent a partial trans-sphenoidal resection of a PCP harbouring a *BRAF-V600E* mutation. After rapid tumor growth, dabrafenib (150mg twice daily) was initiated, followed by trametinib (2mg daily) three weeks later for a total of 7 weeks. An 11% tumor reduction was seen at four weeks and 91% by the 8<sup>th</sup> week.

382

383 Very recently, Himes *et al.* (86) presented the longer-term effects of monotherapy  
384 with dabrafenib in a patient with a history of non-Hodgkin's lymphoma and colon  
385 cancer who had a PCP with the *BRAF*-V600E mutation treated by carinal surgery  
386 and RT. who had undergone craniotomy and RT for a PCP with the *BRAF*-V600E  
387 mutation. Three years after the RT he developed tumor recurrence, and dabrafenib  
388 was initiated (150mg twice daily, reduced to 150mg daily due to joint pains but then  
389 increased to 225mg daily). Two months following treatment, an enlargement of the  
390 cystic component was documented in parallel with a reduction of the solid  
391 component. However, six months post-treatment significant reduction in the size of  
392 both components was identified, while by 9 months only minimal residual tumor was  
393 evident and this remained stable until 12 months; treatment was discontinued, and  
394 the tumor remained stable for further 12 months. A sixth case was recently reported  
395 at the *European Congress of Endocrinology 2019* (87) by Juratli and colleagues, who  
396 administered dabrafenib in a neo-adjuvant setting to a 21 year-old male prior to  
397 surgery and found a more than 80% reduction of tumor size. All these cases support  
398 the potential benefits of BRAF inhibitors, at least in achieving transient tumor  
399 responses, either as neo-adjuvant treatment or following recurrence (**Table 1**). What  
400 is unclear from these single reports relates to the efficacy of different inhibitors, the  
401 mechanisms of tumor resistance, and the means to overcome such resistance. An  
402 interesting observation from one of these case reports was the documentation of a  
403 circulating *BRAF*-V600E mutation in the peripheral blood of the patient during  
404 treatment with the BRAF-inhibitor (84). The authors stated that their finding of  
405 detectable *BRAF*-V600E in peripheral blood was unexpected, since there was not any  
406 previously reported case of circulating tumor cells or cell free DNA in patients with

Formatted: Font: Italic

407 intracranial benign tumors. However, if this is not a result of the surgical procedures  
408 or drug treatment, and the presence of *BRAF*V600E can be detected in the blood  
409 before any treatment, then this assessment may be of value as a selection criterion  
410 of the patients that may benefit from BRAF-inhibitor neoadjuvant therapy (84).  
411 Following these observations, an ongoing cooperative group trial (*Alliance A071601*)  
412 is testing the combination of vemurafenib and cobimetinib in an open-label, phase II  
413 study in patients with *BRAF*-V600E-mutant CP (88) to determine the frequency,  
414 durability, and extent of BRAF-treatment responses in patients with PCP. In this  
415 study, vemurafenib is administered twice daily on days 1-28, and cobimetinib four-  
416 times daily on days 1-21; treatment is repeated every 28 days to a maximum of 5  
417 cycles as long as the disease shows a failure to progress and there is no  
418 unacceptable toxicity. for up to 5 cycles in the absence of disease progression or  
419 unacceptable toxicity. Patients may then be treated with RT, surgery, or to continue  
420 the combination of medical treatment. In another published series, a patient in whom  
421 GTR was achieved had a recurrence of a cystic tumor component after two years,  
422 and has been referred to a phase-II trial of combined BRAF/MEK inhibitors  
423 (NCT03224767) (88,89).

#### 424 425 *Adverse effects of compounds currently used in the management of* 426 *craniopharyngiomas*

427 Data on the adverse effects profile of these treatments in patient with CP are limited.  
428 The phenomenon of “pseudo-progression” with dabrafenib, defined as early initial  
429 cystic enlargement together with solid component reduction, followed by lesion  
430 shrinkage, has been observed, similar to that seen in gliomas after RT or  
431 chemotherapy (86). In addition, transient fever has been described with BRAF

432 inhibitors (29,84,85). Finally, the addition of the MEK inhibitor to BRAF inhibition may  
433 play a role in reducing the incidence of secondary squamous cell skin carcinomas, as  
434 previously shown in patients with melanoma (90).

435

#### 436 *Other potential treatments targeting craniopharyngiomas*

437 Despite the identification of the deranged molecular pathways in ACP pathogenesis,  
438 drugs targeting  $\beta$ -catenin and its downstream MAPK pathway have been used only in  
439 *in vitro* studies. A transcriptome study used mRNA microarray gene expression  
440 analysis and unraveled several 'druggable' pharmaceutical targets that were  
441 consistently over-expressed in a panel of 15 ACP, compared to other brain tumors  
442 and normal brain tissue (26). LCK, EPHA2, SRC were identified as targets of the  
443 tyrosine kinase inhibitor (TKI) dasatinib. AREG, EGFR, ERBB-3 in the EGFR  
444 pathway may respond to inhibitors such as cetuximab, erlotinib, lapatinib; and MMP9  
445 and MMP12 could potentially respond to the oral MMP9/12 inhibitor, AZD1236  
446 (26,50). Recurrence in ACP has been associated with increased VEGF expression  
447 suggesting a possible therapeutic role for the VEGF inhibitor bevacizumab (49,61).

448 Indeed, some of these targets can be shown to be expressed in the primary ACP  
449 tumor using Western blot analysis. ~~Western blot analysis confirmed that a subset of~~  
450 ~~these targets is highly expressed in ACP primary tumor samples~~ (26). On the other  
451 hand, although the SHH pathway may be implicated in the pathogenesis of ACP,  
452 inhibition of this pathway may be tumorigenic (91). This relies on data showing that  
453 vismodegib, a well-established inhibitor of SHH, results in premature tumor formation  
454 and increased tumor cell proliferation in both genetically-engineered and patient-  
455 derived xenograft mouse models (91). Inhibition of EGFR signaling by the TKI,  
456 gefitinib, reduced ACP cell migration *in vitro* by decreasing Fascin expression; this

Formatted: Not Highlight

457 was attributed to the finding of nuclear co-localization of activated EGFR,  $\beta$ -catenin  
458 and Fascin in ACP cell migration *in vitro* (48), implying a crosstalk of these pathways  
459 in ACP. Since Annexin A2 expression, a  $\text{Ca}^{2+}$ -regulated binding protein, correlates  
460 with gefitinib sensitivity *in vitro*, this may serve as a biomarker of a response to  
461 treatment (92). Small samples of human ACP tumors were cultured with and without  
462 trametinib, a specific MEK inhibitor; the inhibition of the MAPK/ERK pathway resulted  
463 in reduced proliferation along with a dose-dependent significant decrease in the Ki-67  
464 proliferation index in the trametinib treated tumors, suggesting a possible role in the  
465 management of not only PCP but also ACP (91). Notably, the well-established  
466 synergistic effects of combined targeted therapies in other neuroendocrine tumor  
467 models provide support for the potential use of combinatorial targeted therapy in CPs  
468 (93,94).

469  
470 Given the potential role of inflammation in the pathogenesis of craniopharyngioma  
471 and previous studies on intra-tumor administration of interferon-alpha in patients with  
472 CP (22), an open label, phase II study is underway aiming to estimate the one-year  
473 disease stabilization rate associated with the use of Peg-interferon alfa-2b in patients  
474 with progressive unresectable or recurrent CP after surgery alone (without adjuvant  
475 RT). The study will also estimate the sustained objective response rate (partial and  
476 complete response) to Peg-interferon alfa-2b in patients with CP which progress or  
477 recur following RT (95). Finally, noting the immune microenvironment in ACP, IL1R  
478 inhibitors, such as anakinra, may have a place in our therapeutic armamentarium  
479 (60).

480  
481

## Conclusions

The discovery of distinct oncogenic molecular signaling pathways in CPs has opened new avenues for the personalized treatment of patients with these challenging tumors. The key pathogenetic event in ACP appears to be Wnt activation along with alterations of the MEK/ERK pathway, whereas in PCP there is primary activation of the Ras/Raf/MEK/ERK pathway by *BRAF*-V600E mutations. Current literature on PCPs includes single case reports, with both BRAF and MAPK inhibitors, in patients with V600E mutations. Their results are promising but well-designed studies are needed aiming to provide robust data on treatment regimens and protocols, as well as on efficacy and safety. It also remains unclear whether single or combination therapies will be necessary, although *in vitro* data from other endocrine tumors have highlighted the use of multiple simultaneous combination treatments. The use of MAPK inhibitors for the more common ACPs also requires assessment in clinical trials, particularly in view of the risk of acquired drug resistance. The value of combination therapies aiming to target multiple molecular mediators implicated in the pathogenesis of ACP and possibly minimizing drug toxicities also requires further investigation. ~~needs to be evaluated.~~

Nevertheless, the recently expanding data on the pathogenesis of CPs offer substantial ground for the translation of molecular insights into practical therapies for primary, recurrent or very aggressive tumors which will, hopefully, improve the prognosis of the patients. The eventual use of neo-adjuvant medical treatment or even an entirely non-operative treatment strategy for CP should improve the quality of life of these patients, minimizing the neurological and endocrinological sequelae

caused either by the tumor extension or by the neurosurgical or radiotherapeutic complications. Combination therapy strategies could be also be effective in cases of ACP in order to target the multiple molecular mediators that are implicated in ACP pathogenesis and to minimize any adverse treatment effects. ~~and minimizing the side effects of treatment.~~

## References

1. Karavitaki N, Cudlip S, Adams CB, Wass JA. Craniopharyngiomas. *Endocr Rev.* 2006;27(4):371-397.
2. Louis D, Ohgaki H, Wiestler O, Cavenee W, Ellison D, Figarella-Branger D, et al (2016) WHO classification of tumours of the central nervous system, Revised 4th Edition. IARC, Lyon, pp 324–328. ISBN 978-92-832-4492-9
3. Lithgow K, Pohl U, Karavitaki N. Craniopharyngiomas. In: Feingold KR, Anawalt B, Boyce A, Chrousos G, Dungan K, Grossman A, Hershman JM, Kaltsas G, Koch C, Kopp P, Korbonits M, McLachlan R, Morley JE, New M, Perreault L, Purnell J, Rebar R, Singer F, Trencle DL, Vinik A, Wilson DP, editors. *Endotext* [Internet]. South Dartmouth (MA): MDText.com, Inc.; 2000-2019 Mar 14.
4. Bunin GR, Surawicz TS, Witman PA, Preston-Martin S, Davis F, Bruner JM. The descriptive epidemiology of craniopharyngioma. *J Neurosurg.* 1998;89(4):547-551.
5. Bulow B, Attewell R, Hagmar L, Malmstrom P, Nordstrom CH, Erfurth EM. Postoperative prognosis in craniopharyngioma with respect to cardiovascular

mortality, survival, and tumor recurrence. *J Clin Endocrinol Metab.* 1998;83(11):3897-3904.

6. Nielsen EH, Feldt-Rasmussen U, Poulsen L, Kristensen LO, Astrup J, Jørgensen JO, Bjerre P, Andersen M, Andersen C, Jørgensen J, Lindholm J, Laurberg P. Incidence of craniopharyngioma in Denmark (n = 189) and estimated world incidence of craniopharyngioma in children and adults. *J Neurooncol.* 2011;104(3):755-763

7. Olsson DS, Andersson E, Bryngelsson IL, Nilsson AG, Johannsson G. Excess mortality and morbidity in patients with craniopharyngioma, especially in patients with childhood onset: a population-based study in Sweden. *J Clin Endocrinol Metab.* 2015;100(2):467-474

8. Hölsken A. Pathogenesis of Human ACP, pp 1-26. In: J.P. Martinez-Barbera, C.L. Andoniadou (eds.), *Basic Research and Clinical Aspects of Adamantinomatous Craniopharyngioma* Springer International Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_1

9. Bailey W, Freidenberg GR, James HE, Hesselink JR, Jones KL (1990) Prenatal diagnosis of a craniopharyngioma using ultrasonography and magnetic resonance imaging. *Prenat Diagn.* 1990;10(10):623-629.

10. Müller-Scholden J, Lehrnbecher T, Müller HL, Bensch J, Hengen RH, Sørensen N, Stockhausen HB. Radical surgery in a neonate with craniopharyngioma—report of a case. *Pediatr Neurosurg.* 2000;33(5):265-269.

11. Nielsen EH, Lindholm J, Laurberg P. Excess mortality in women with pituitary disease: a meta-analysis. *Clin Endocrinol (Oxf).* 2007;67(5):693-697.

12. Borrill R, Cheesman E, Stivaros S, Kamaly-Asl ID, Gnanalingham K, Kilday JP. Papillary craniopharyngioma in a 4-year-old girl with BRAF V600E



- 556 mutation: a case report and review of the literature. *Childs Nerv Syst.*  
557 2019;35:169-173
- 558 13. Cushing H. *Intracranial Tumors. Notes upon series of two thousand verified*  
559 *cases with surgical mortality percentages pertaining thereto.* Thomas,  
560 Springfield,, 1932
- 561 14. Vardon AJ and Karavitaki N. Clinical Diagnosis of Human ACP, pp 57-66. In:  
562 J.P. Martinez-Barbera, C.L. Andoniadou (eds.), *Basic Research and Clinical*  
563 *Aspects of Adamantinomatous Craniopharyngioma* Springer International  
564 Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_4
- 565 15. Erfurth EM. Obesity and Metabolic Disturbances in Adamantinomatous  
566 Craniopharyngioma Patients, pp 85-98. In: J.P. Martinez-Barbera, C.L.  
567 Andoniadou (eds.), *Basic Research and Clinical Aspects of*  
568 *Adamantinomatous Craniopharyngioma* Springer International Publishing AG  
569 2017; DOI 10.1007/978-3-319-51890-9\_6
- 570 16. Asa SL, Casar-Borota O, Chanson P, Delgrange E, Earls P, Ezzat S,  
571 Grossman A, Ikeda H, Inoshita N, Karavitaki N, Korbonits M, Laws ER Jr,  
572 Lopes MB, Maartens N, McCutcheon IE, Mete O, Nishioka H, Raverot G,  
573 Roncaroli F, Saeger W, Syro LV, Vasiljevic A, Villa C, Wierinckx A, Trouillas J;  
574 attendees of 14th Meeting of the International Pituitary Pathology Club,  
575 Annecy, France, November 2016. From pituitary adenoma to pituitary  
576 neuroendocrine tumor (PitNET): an International Pituitary Pathology Club  
577 proposal. *Endocr Relat Cancer.* 2017;24(4):C5-C8.
- 578 17. Aquilina K and Buchfelder M. Surgical Treatment of Human ACP, pp 137-158.  
579 In: J.P. Martinez-Barbera, C.L. Andoniadou (eds.), *Basic Research and*

- Clinical Aspects of Adamantinomatous Craniopharyngioma Springer International Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_8
18. Müller HL. Long-Term Management and Clinical Trials in Adamantinomatous Craniopharyngioma, pp 179-214 In: J.P. Martinez-Barbera, C.L. Andoniadou (eds.), Basic Research and Clinical Aspects of Adamantinomatous Craniopharyngioma Springer International Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_10
19. Jyoti B, Indelicato DJ, Bradley JA, Rotondo RL. Radiology and Radiotherapy of Craniopharyngioma, pp 101-135. In: J.P. Martinez-Barbera, C.L. Andoniadou (eds.), Basic Research and Clinical Aspects of Adamantinomatous Craniopharyngioma Springer International Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_7
20. Yang I, Sughrue ME, Rutkowski MJ, Kaur R, Ivan ME, Aranda D, Barani IJ, Parsa AT. Craniopharyngioma: a comparison of tumor control with various treatment strategies. *Neurosurg Focus*. 2010;28(4):E5.
21. Kilday JP, Bartels U. Intracystic Administration of Interferon-Alpha for Reduction of Cystic Tumour Burden, pp 159-177. In: J.P. Martinez-Barbera, C.L. Andoniadou (eds.), Basic Research and Clinical Aspects of Adamantinomatous Craniopharyngioma Springer International Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_9
22. Mrowczynski OD, Langan ST, Rizk EB. Craniopharyngiomas: A systematic review and evaluation of the current intratumoral treatment landscape. *Clin Neurol Neurosurg*. 2018;166:124-130.
23. Pettorini BL, Inzitari R, Massimi L, Tamburrini G, Caldarelli M, Fanali C, Cabras T, Messana I, Castagnola M, Di Rocco C. The role of inflammation in

the genesis of the cystic component of craniopharyngiomas. *Childs Nerv Syst.* 2010;26(12):1779-178424. Kilday JP, Caldarelli M, Massimi L, Chen RH, Lee YY, Liang ML, Parkes J, Naiker T, van Veelen ML, Michiels E, Mallucci C, Pettorini B, Meijer L, Dorfer C, Czech T, Diezi M, Schouten-van Meeteren AYN, Holm S, Gustavsson B, Benesch M, Müller HL, Hoffmann A, Rutkowski S, Flitsch J, Escherich G, Grotzer M, Spoudeas HA, Azquikina K, Capra M, Jiménez-Guerra R, MacDonald P, Johnston DL, Dvir R, Constantini S, Kuo MF, Yang SH, Bartels U. Intracystic interferon-alpha in pediatric craniopharyngioma patients: an international multicenter assessment on behalf of SIOPE and ISPN. *Neuro Oncol.* 2017 Oct 1;19(10):1398-1407

24. Kilday JP, Caldarelli M, Massimi L, Chen RH, Lee YY, Liang ML, Parkes J, Naiker T, van Veelen ML, Michiels E, Mallucci C, Pettorini B, Meijer L, Dorfer C, Czech T, Diezi M, Schouten-van Meeteren AYN, Holm S, Gustavsson B, Benesch M, Müller HL, Hoffmann A, Rutkowski S, Flitsch J, Escherich G, Grotzer M, Spoudeas HA, Azquikina K, Capra M, Jiménez-Guerra R, MacDonald P, Johnston DL, Dvir R, Constantini S, Kuo MF, Yang SH, Bartels U. Intracystic interferon-alpha in pediatric craniopharyngioma patients: an international multicenter assessment on behalf of SIOPE and ISPN. *Neuro Oncol.* 2017 Oct 1;19(10):1398-1407

25. Brastianos PK, Taylor-Weiner A, Manley PE, Jones RT, Dias-Santagata D, Thorner AR, Lawrence MS, Rodriguez FJ, Bernardo LA, Schubert L, Sunkavalli A, Shillingford N, Calicchio ML, Lidov HG, Taha H, Martinez-Lage M, Santi M, Storm PB, Lee JY, Palmer JN, Adappa ND, Scott RM, Dunn IF, Laws ER Jr, Stewart C, Ligon KL, Hoang MP, Van Hummelen P, Hahn WC, Louis DN, Resnick AC, Kieran MW, Getz G, Santagata S. Exome sequencing

- identifies BRAF mutations in papillary craniopharyngiomas. *Nat Genet.* 2014;46(2):161-165.
26. Gump JM, Donson AM, Birks DK, Amani VM, Rao KK, Griesinger AM, Kleinschmidt-DeMasters BK, Johnston JM, Anderson RC, Rosenfeld A, Handler M, Gore L, Foreman N, Hankinson TC. Identification of targets for rational pharmacological therapy in childhood craniopharyngioma. *Acta Neuropathol Commun.* 2015;3:30.
27. Goschzik T, Gessi M, Dreschmann V, Gebhardt U, Wang L, Yamaguchi S, Wheeler DA, Lauriola L, Lau CC, Müller HL, Pietsch T. Genomic Alterations of Adamantinomatous and Papillary Craniopharyngioma. *J Neuropathol Exp Neurol.* 2017;76(2):126-134.
28. Hölsken A, Sill M, Merkle J, Schweizer L, Buchfelder M, Flitsch J, Fahlbusch R, Metzler M, Kool M, Pfister SM, von Deimling A, Capper D, Jones DT, Buslei R. Adamantinomatous and papillary craniopharyngiomas are characterized by distinct epigenomic as well as mutational and transcriptomic profiles. *Acta Neuropathol Commun.* 2016;4:20
29. Roque A, Odia Y. BRAF-V600E mutant papillary craniopharyngioma dramatically responds to combination BRAF and MEK inhibitors. *CNS Oncol.* 2017;6(2):95-99.
30. Sekine S, Shibata T, Kokubu A, Morishita Y, Noguchi M, Nakanishi Y, Sakamoto M, Hirohashi S. Craniopharyngiomas of adamantinomatous type harbor beta-catenin gene mutations. *Am J Pathol.* 2002;161(6):1997-2001 .
31. Hofmann BM, Kreutzer J, Saeger W, Buchfelder M, Blumcke I, Fahlbusch R, Buslei R. Nuclear beta-catenin accumulation as reliable marker for the

- differentiation between cystic craniopharyngiomas and rathke cleft cysts: a clinico-pathologic approach. *Am J Surg Pathol*. 2006;30(12):1595-1603.
32. Martinez-Barbera JP. Molecular and cellular pathogenesis of adamantinomatous craniopharyngioma. *Neuropathol Appl Neurobiol*. 2015;41(6):721-732.
33. Kemler R. From cadherins to catenins: cytoplasmic protein interactions and regulation of cell adhesion. *Trends Genet*. 1993;9(9):317-21.
34. Larkin SJ, Ansorge O. Pathology and pathogenesis of craniopharyngiomas. *Pituitary*. 2013;16(1):9-17
35. Cani CM, Matushita H, Carvalho LR, Soares IC, Brito LP, Almeida MQ, Mendonça BB. PROP1 and CTNNB1 expression in adamantinomatous craniopharyngiomas with or without b-catenin mutations. *Clinics (Sao Paulo)*. 2011;66(11):1849-1854.
36. Hölsken A, Buchfelder M, Fahlbusch R, Blümcke I, Buslei R. Tumour cell migration in adamantinomatous craniopharyngiomas is promoted by activated Wnt-signalling. *Acta Neuropathol*. 2010;119(5):631-639.
37. Behrens J, Jerchow BA, Würtele M, Grimm J, Asbrand C, Wirtz R, Kühl M, Wedlich D, Birchmeier W. Functional interaction of an axin homolog, conductin, with beta-catenin, APC, and GSK3beta. *Science*. 1998;280(5363):596-599.
38. Rubinfeld B, Albert I, Porfiri E, Fiol C, Munemitsu S, Polakis P. Binding of GSK3beta to the APC-beta-catenin complex and regulation of complex assembly. *Science*. 1996;272(5264):1023-1026.

39. Rubinfeld B, Souza B, Albert I, Müller O, Chamberlain SH, Masiarz FR, Munemitsu S, Polakis P. Association of the APC gene product with beta-catenin. *Science*. 1993;262(5140):1731-1734.
40. Lilien J, Balsamo J. The regulation of cadherin-mediated adhesion by tyrosine phosphorylation/dephosphorylation of beta-catenin. *Curr Opin Cell Biol*. 2005;17(5):459-465
41. Giles RH, van Es JH, Clevers H. Caught up in a Wnt storm: Wnt signaling in cancer. *Biochim Biophys Acta*. 2003;1653(1):1-24
42. Hölsken A, Stache C, Schlaffer SM, Flitsch J, Fahlbusch R, Buchfelder M, Buslei R. Adamantinomatous craniopharyngiomas express tumor stem cell markers in cells with activated Wnt signaling: further evidence for the existence of a tumor stem cell niche? *Pituitary*. 2014;17(6):546-556
43. Esheba GE, Hassan AA. Comparative immunohistochemical expression of b-catenin, EGFR, ErbB2, and p63 in adamantinomatous and papillary craniopharyngiomas. *J Egypt Natl Canc Inst*. 2015;27(3):139-145
44. Buslei R, Nolde M, Hofmann B, Meissner S, Eyupoglu IY, Siebzehnriibl F, Hahnen E, Kreutzer J, Fahlbusch R. Common mutations of b-catenin in adamantinomatous craniopharyngiomas but not in other tumors originating from the sellar region. *Acta Neuropathol*. 2005;109(6):589-597.
45. Andoniadou CL, Martinez-Barbera JP. Genetically Modified Mouse Models of Adamantinomatous Craniopharyngioma. In: J.P. Martinez-Barbera, C.L. Andoniadou (eds.), *Basic Research and Clinical Aspects of Adamantinomatous Craniopharyngioma* Springer International Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_3

46. Chen Z, He X, Jia M, Liu Y, Qu D, Wu D, Wu P, Ni C, Zhang Z, Ye J, Xu J, Huang J.  $\beta$ -catenin Overexpression in the Nucleus Predicts Progress Disease and Unfavourable Survival in Colorectal Cancer: A Meta-Analysis. *PLoS One*. 2013;8(5):e63854.
47. Weiss V, Dueber J, Wright JP, Cates J, Revetta F, Parikh AA, Merchant NB, Shi C. Immunohistochemical analysis of the Wnt/ $\beta$ -catenin signaling pathway in pancreatic neuroendocrine neoplasms. *World J Gastrointest Oncol*. 2016;8(8):615-622.
48. Hölsken A, Gebhardt M, Buchfelder M, Fahlbusch R, Blümcke I, Buslei R. EGFR signaling regulates tumor cell migration in craniopharyngiomas. *Clin Cancer Res*. 2011;17(13):4367-4377.
49. Gupta S, Bi WL, Giantini Larsen A, Al-Abdulmohsen S, Abedalthagafi M, Dunn IF. Craniopharyngioma: a roadmap for scientific translation. *Neurosurg Focus*. 2018;44(6):E12.
50. Robinson L, Santagata S, Hankinson TC. Transcriptomic and Genomic Analyses of Human Craniopharyngioma, pp 27-39. In: J.P. Martinez-Barbera, C.L. Andoniadou (eds.), *Basic Research and Clinical Aspects of Adamantinomatous Craniopharyngioma* Springer International Publishing AG 2017; DOI 10.1007/978-3-319-51890-9\_2
51. Koike M1, Kose S, Furuta M, Taniguchi N, Yokoya F, Yoneda Y, Imamoto N. beta-Catenin shows an overlapping sequence requirement but distinct molecular interactions for its bidirectional passage through nuclear pores. *J Biol Chem*. 2004;279(32):34038-34047
52. Hölsken A, Kreutzer J, Hofmann BM, Hans V, Oppel F, Buchfelder M, Fahlbusch R, Blümcke I, Buslei R. Target gene activation of the Wnt signaling

pathway in nuclear beta-catenin accumulating cells of adamantinomatous craniopharyngiomas. *Brain Pathol.* 2009;19(3):357-364.

53. Andoniadou CL, Gaston-Massuet C, Reddy R, Schneider RP, Blasco MA, Le Tissier P, Jacques TS, Pevny LH, Dattani MT, Martinez-Barbera JP. Identification of novel pathways involved in the pathogenesis of human adamantinomatous craniopharyngioma. *Acta Neuropathol.* 2012;124(2):259-271

54. Gaston-Massuet C1, Andoniadou CL, Signore M, Jayakody SA, Charolidi N, Kyeyune R, Vernay B, Jacques TS, Taketo MM, Le Tissier P, Dattani MT, Martinez-Barbera JP. Increased Wingless (Wnt) signaling in pituitary progenitor/stem cells gives rise to pituitary tumors in mice and humans. *Proc Natl Acad Sci U S A.* 2011;108(28):11482-11487.

55. Sekine S, Takata T, Shibata T, Mori M, Morishita Y, Noguchi M, Uchida T, Kanai Y, Hirohashi S. Expression of enamel proteins and LEF1 in adamantinomatous craniopharyngioma: evidence for its odontogenic epithelial differentiation. *Histopathology.* 2004;45(6):573-579.

56. Roura S, Miravet S, Piedra J, García de Herreros A, Duñach M. Regulation of E-cadherin/Catenin association by tyrosine phosphorylation. *J Biol Chem.* 1999;274(51):36734-36740.

57. Mimeault M, Batra SK. Frequent deregulations in the hedgehog signaling network and cross-talks with the epidermal growth factor receptor pathway involved in cancer progression and targeted therapies. *Pharmacol Rev.* 2010;62(3):497-524

58. Reya T, Clevers H. Wnt signalling in stem cells and cancer. *Nature.* 2005;434(7035):843-850



59. Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature*. 2001;414(6859):105-111.
60. Apps JR, Carreno G, Gonzalez-Meljem JM, Haston S, Guiho R, Cooper JE, Manshaei S, Jani N, Hölsken A, Pettorini B, Beynon RJ, Simpson DM, Fraser HC, Hong Y, Hallang S, Stone TJ, Virasami A, Donson AM, Jones D, Aquilina K, Spoudeas H, Joshi AR, Grundy R, Storer LCD, Korbonits M, Hilton DA, Tossell K, Thavaraj S, Ungless MA, Gil J, Buslei R, Hankinson T, Hargrave D, Goding C, Andoniadou CL, Brogan P, Jacques TS, Williams HJ, Martinez-Barbera JP. Tumour compartment transcriptomics demonstrates the activation of inflammatory and odontogenic programmes in human adamantinomatous craniopharyngioma and identifies the MAPK/ERK pathway as a novel therapeutic target. *Acta Neuropathol*. 2018;135(5):757-777
61. Sun H, Akgun E, Bicer A, Ozkan A, Bozkurt SU, Kurtkaya O, Koc DY, Pamir MN, Kilic T. Expression of angiogenic factors in craniopharyngiomas: implications for tumor recurrence. *Neurosurgery*. 2010;66(4):744-750
62. Baker JC, Beddington RS, Harland RM. Wnt signaling in *Xenopus* embryos inhibits *bmp4* expression and activates neural development. *Genes Dev*. 1999;13(23):3149-3159.
63. Dhillon AS, Hagan S, Rath O, Kolch W. MAP kinase signaling pathways in cancer. *Oncogene*. 2007;26(22):3279-3290
64. Zhang W, Liu HT. MAPK signal pathways in the regulation of cell proliferation in mammalian cells. *Cell Res*. 2002;12(1):9-18.
65. Herbst RS. Review of epidermal growth factor receptor biology. *Int J Radiat Oncol Biol Phys*. 2004;59(2 Suppl):21-6

66. Laurent-Puig P, Lievre A, Blons H. Mutations and response to epidermal growth factor receptor inhibitors. *Clin Cancer Res.* 2009;15(4):1133-1139
67. Futreal PA, Coin L, Marshall M, Down T, Hubbard T, Wooster R, Rahman N, Stratton MR. A census of human cancer genes. *Nat Rev Cancer.* 2004;4(3):177-183.
68. Larkin SJ, Preda V, Karavitaki N, Grossman A, Ansorge O. BRAF V600E mutations are characteristic for papillary craniopharyngioma and may coexist with CTNNB1-mutated adamantinomatous craniopharyngioma. *Acta Neuropathol.* 2014;127(6):927-929
69. Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, Teague J, Woffendin H, Garnett MJ, Bottomley W, Davis N, Dicks E, Ewing R, Floyd Y, Gray K, Hall S, Hawes R, Hughes J, Kosmidou V, Menzies A, Mould C, Parker A, Stevens C, Watt S, Hooper S, Wilson R, Jayatilake H, Gusterson BA, Cooper C, Shipley J, Hargrave D, Pritchard-Jones K, Maitland N, Chenevix-Trench G, Riggins GJ, Bigner DD, Palmieri G, Cossu A, Flanagan A, Nicholson A, Ho JW, Leung SY, Yuen ST, Weber BL, Seigler HF, Darrow TL, Paterson H, Marais R, Marshall CJ, Wooster R, Stratton MR, Futreal PA. Mutations of the BRAF gene in human cancer. *Nature.* 2002 Jun 27;417(6892):949-954
70. Haston S, Pozzi S, Carreno G, Manshaei S, Panousopoulos L, Gonzalez-Meljem JM, Apps JR, Virasami A, Thavaraj S, Gutteridge A, Forshaw T, Marais R, Brandner S, Jacques TS, Andoniadou CL, Martinez-Barbera JP. MAPK pathway control of stem cell proliferation and differentiation in the embryonic pituitary provides insights into the pathogenesis of papillary craniopharyngioma. *Development.* 2017;144(12):2141-2152.

71. Larkin S, Karavitaki N. Recent advances in molecular pathology of craniopharyngioma. *F1000Res*. 2017;6:1202
72. Jones DT, Kocialkowski S, Liu L, Pearson DM, Ichimura K, Collins VP. Oncogenic RAF1 rearrangement and a novel BRAF mutation as alternatives to KIAA1549:BRAF fusion in activating the MAPK pathway in pilocytic astrocytoma. *Oncogene*. 2009;28(20):2119-2123
73. Tian Y, Rich BE, Vena N, Craig JM, Macconail LE, Rajaram V, Goldman S, Taha H, Mahmoud M, Ozek M, Sav A, Longtine JA, Lindeman NI, Garraway LA, Ligon AH, Stiles CD, Santagata S, Chan JA, Kieran MW, Ligon KL. Detection of KIAA1549-BRAF fusion transcripts in formalin-fixed paraffin-embedded pediatric low-grade gliomas. *J Mol Diagn*. 2011;13(6):669-677.
74. Brastianos PK, Santagata S. ENDOCRINE TUMORS: **BRAF** V600E mutations in papillary craniopharyngioma. *Eur J Endocrinol*. 2016;174(4):R139-144
75. Stache C, Hölsken A, Fahlbusch R, Flitsch J, Schlaffer SM, Buchfelder M, Buslei R. Tight junction protein claudin-1 is differentially expressed in craniopharyngioma subtypes and indicates invasive tumor growth. *Neuro Oncol*. 2014;16(2):256-264.
76. Stache C, Hölsken A, Schlaffer SM, Hess A, Metzler M, Frey B, Fahlbusch R, Flitsch J, Buchfelder M, Buslei R. Insights into the infiltrative behavior of adamantinomatous craniopharyngioma in a new xenotransplant mouse model. *Brain Pathol*. 2015;25(1):1-10.
77. Samis J, Vanin EF, Sredni ST, de Bonaldo Mde F, Costa FF, Tomita T, Habiby R, Zimmerman D, Soares MB. Extensive miRNA expression analysis in craniopharyngiomas. *Childs Nerv Syst*. 2016;32(9):1617-1624

78. Garcia-Lavandeira M, Saez C, Diaz-Rodriguez E, Perez-Romero S, Senra A, Dieguez C, Japon MA, Alvarez CV. Craniopharyngiomas express embryonic stem cell markers(SOX2, OCT4, KLF4, and SOX9) as pituitary stem cells but do not coexpress RET/GFRA3 receptors. *J Clin Endocrinol Metab.* 2012;97(1):E80-87.
79. Donson AM, Apps J, Griesinger AM, Amani V, Witt DA, Anderson RCE, Niazi TN, Grant G, Souweidane M, Johnston JM, Jackson EM, Kleinschmidt-DeMasters BK, Handler MH, Tan AC, Gore L, Virasami A, Gonzalez-Meljem JM, Jacques TS, Martinez-Barbera JP, Foreman NK, Hankinson TC; Advancing Treatment for Pediatric Craniopharyngioma Consortium. Molecular Analyses Reveal Inflammatory Mediators in the Solid Component and Cyst Fluid of Human Adamantinomatous Craniopharyngioma. *J Neuropathol Exp Neurol.* 2017;76(9):779-788
80. Barry S, Carlsen E, Marques P, Stiles CE, Gadaleta E, Berney DM, Roncaroli F, Chelala C, Solomou A, Herincs M, Caimari F, Grossman AB, Crnogorac-Jurcevic T, Haworth O, Gaston-Massuet C, Korbonits M. Tumor microenvironment defines the invasive phenotype of AIP-mutation-positive pituitary tumors. *Oncogene.* 2019 Mar 12.
81. Ascierto PA, Minor D, Ribas A, Lebbe C, O'Hagan A, Arya N, Guckert M, Schadendorf D, Kefford RF, Grob JJ, Hamid O, Amaravadi R, Simeone E, Wilhelm T, Kim KB, Long GV, Martin AM, Mazumdar J, Goodman VL, Trefzer U. Phase II Trial (BREAK-2) of the BRAF Inhibitor Dabrafenib (GSK2118436) in Patients With Metastatic Melanoma. *J Clin Oncol.* 2013;31(26):3205-3211
82. Tsoukalas N, Tsapakidis K, Alexandraki KI. The role of palbociclib in thyroid carcinoma with BRAF mutation. *Gland Surg.* 2018;7(Suppl 1):S82-S85

83. Aylwin SJ, Bodi I, Beaney R. Pronounced response of papillary craniopharyngioma to treatment with vemurafenib, a BRAF inhibitor. *Pituitary*. 2016;19(5):544-546.
84. Brastianos PK, Shankar GM, Gill CM, Taylor-Weiner A, Nayyar N, Panka DJ, Sullivan RJ, Frederick DT, Abedalthagafi M, Jones PS, Dunn IF, Nahed BV, Romero JM, Louis DN, Getz G, Cahill DP, Santagata S, Curry WT Jr, Barker FG 2nd. Dramatic Response of BRAF V600E Mutant Papillary Craniopharyngioma to Targeted Therapy. *J Natl Cancer Inst*. 2015;108(2)
85. Rostami E, Witt Nyström P, Libard S, Wikström J, Casar-Borota O, Gudjonsson O. Recurrent papillary craniopharyngioma with BRAFV600E mutation treated with neoadjuvant-targeted therapy. *Acta Neurochir (Wien)*. 2017;159(11):2217-2221.
86. Himes BT, Ruff MW, Van Gompel JJ, Park SS, Galanis E, Kaufmann TJ, Uhm JH. Recurrent papillary craniopharyngioma with BRAF V600E mutation treated with dabrafenib: case report. *J Neurosurg*. 2018:1-5.
87. Juratli T. New potential treatment alternatives in patients with papillary craniopharyngioma. *Endocrine Abstracts* 2019 63 S12.2
88. Vemurafenib and Cobimetinib in Treating Patients With BRAF V600E Mutation Positive Craniopharyngioma. *ClinicalTrials.gov Identifier: NCT03224767*
89. La Corte E, Younus I, Pivari F, Selimi A, Ottenhausen M, Forbes JA, Pisapia DJ, Dobri GA, Anand VK, Schwartz TH. BRAF V600E mutant papillary craniopharyngiomas: a single-institutional case series. *Pituitary*. 2018;21(6):571-583
90. Flaherty KT, Infante JR, Daud A, Gonzalez R, Kefford RF, Sosman J, Hamid O, Schuchter L, Cebon J, Ibrahim N, Kudchadkar R, Burris HA 3rd, Falchook

- G, Algazi A, Lewis K, Long GV, Puzanov I, Lebowitz P, Singh A, Little S, Sun P, Allred A, Ouellet D, Kim KB, Patel K, Weber J. Combined BRAF and MEK inhibition in melanoma with BRAF V600 mutations. *N Engl J Med*. 2012;367(18):1694-1670
91. Carreno G, Boulton JKR, Apps JR, Gonzalez-Meljem JM, Haston S, Guiho R, Stache C, Danielson LS, Koers A, Smith LS, Virasami A, Panousopoulos L, Buchfelder M, Jacques TS, Chesler L, Robinson S, Martinez-Barbera JP. SHH pathway inhibition is protumorigenic in adamantinomatous craniopharyngioma. *Endocr Relat Cancer*. 2019 Jan 1
92. Wang Y, Deng J, Guo G, Tong A, Peng X, Chen H, Xu J, Liu Y, You C, Zhou L. Clinical and prognostic role of annexin A2 in adamantinomatous craniopharyngioma. *J Neurooncol*. 2017;131(1):21-29.
93. Nölting S, Garcia E, Alusi G, Giubellino A, Pacak K, Korbonits M, Grossman AB. Combined blockade of signalling pathways shows marked anti-tumour potential in pheochromocytoma cell lines. *J Mol Endocrinol*. 2012;49(2):79-96.
94. Giubellino A, Bullova P, Nölting S, Turkova H, Powers JF, Liu Q, Guichard S, Tischler AS, Grossman AB, Pacak K. Combined inhibition of mTORC1 and mTORC2 signaling pathways is a promising therapeutic option in inhibiting pheochromocytoma tumor growth: in vitro and in vivo studies in female athymic nude mice. *Endocrinology*. 2013;154(2):646-655.
95. Peginterferon Alfa-2b in Treating Younger Patients With Craniopharyngioma That is Recurrent or Cannot Be Removed By Surgery. *ClinicalTrials.gov* Identifier: NCT01964300

**Table 1.** Case reports and current clinical trials of patients with papillary craniopharyngioma (PCP) and BRAF-V600E alterations treated with V600E mutation-specific BRAF inhibitors (dabrafenib or vemurafenib) with or without combined treatment with MEK inhibitors (trametinib or cobimetinib).

Case series/ reports	CP	Previous treatment	Histopathological and molecular genetic analysis	Novel treatment	Duration of treatment	Outcome (mean of measurement)	Comments
Aylwin et al, 2015	PCP	TSSx2, RT.	Pyrosequencing analysis indicated the presence of the BRAF mutation c.1799T[A (p.Val600Glu).	Vemurafenib 960 mg bd.	3 months.	Near complete resolution (MRI.)	Recurrence 6 weeks off treatment; restart of treatment and stabilization for 7 months with a regrowth.
Brastianos et al, 2015	PCP	Craniotomy x3.	Uniform staining of the BRAFV600E protein (VE1 antibody) by IHC; V600E mutation confirmed by allele-specific genetic testing. The mutation was detectable in peripheral blood during treatment. Absence of nuclear staining for $\beta$ -catenin by IHC.	Dabrafenib 150 mg bd + after 3 weeks trametinib 2 mg od.	35 days (followed by other treatments).	85% reduction tumor volume and 81% reduction tumor-associated Cyst (MRI).	
Roque et al, 2016	PCP	Craniotomy, Ommaya catheter with percutaneous aspiration of cyst fluid, RT 54 Gy in 30 fractions.	(+)ve staining for BRAFV600E by IHC; next-generation sequencing confirmed BRAFV600E mutation and FGFR3 splice site (437_445+3del12) with two variants of unknown significance (BRCA2-G1771D and FGFR4-S551F).	Dabrafenib 150 mg orally twice daily and trametinib 2-mg orally od.	7 months	Almost disappearance of 2.6 $\times$ 2.3 $\times$ 3.2-cm tumour (MRI); reduced volume and intensity of FDG uptake on PET.	
Rostami et al, 2017	PCP	TSS	Weak staining of mutated BRAFV600E (VE1 antibody) by IHC; BRAFV600E genotype confirmed by pyrosequencing mutational analysis.	Dabrafenib 150 mg bd + after 3 weeks trametinib 2 mg od	7 weeks	91% reduction tumour size (MRI).	Combined BRAF and MEK-targeted therapy.
Himes et al, 2019	PCP	Craniotomy, RT 36 Gy in 12 fractions.	BRAF V600E mutation was confirmed in specimens from his original resection.	Dabrafenib 150 mg bd (shortly) $\rightarrow$ od (several weeks) $\rightarrow$ 225 mg od	1 year under treatment and follow-up for 1 year off treatment.	Minimal residual tumor.	
Jurati et al, 2019	PCP			Dabrafenib 150 mg bd + trametinib 2 mg od.		>80% reduction of tumour size (MRI).	
NCT03224767	PCP	Cohort A: $\pm$ surgery Cohort B: RT $\pm$ other treatment (except for BRAF or MEK inhibitors)	IHC for BRAF V600E mutation (VE1) and $\beta$ -catenin IHC (membranous, non-nuclear pattern).	Vemurafenib and Cobimetinib.	Vemurafenib bd day 1-28 + cobimetinib QD days 1-21.	Every 28 days for up to 5 courses in the absence of disease progression or unacceptable toxicity (MRI).	Patients may then receive RT, surgery, or continued treatment.

bd: bis die (twice a day); CP: craniopharyngioma; IHC: immunohistochemistry; MRI: magnetic resonance imaging; od: once a day; PCP: papillary craniopharyngioma; RT: radiation therapy; TSS: trans-sphenoidal surgery;  $^{18}$ F-FDG PET/CT:  $^{18}$ fluoro-D-glucose Positron emission tomography/ computed tomography; RT: radiotherapy; (+)ve: positive

912 **FIGURE 1.** The mesh terms used to the search strategy : ("Craniopharyngioma"[Mesh] AND  
913 "Craniopharyngioma/drug therapy"[Mesh] NOT ("review"[Publication Type] OR "review literature as  
914 topic"[MeSH Terms] OR "review"[All Fields]) AND ("2009/05/01"[PDat]: "2019/04/28"[PDat])  
915  
916



917 **FIGURE 2.** Photomicrograph of a representative case of a Papillary Craniopharyngioma. Hematoxylin  
918 and eosin staining, x100 magnification.

919

920 (Dr A. Ramanathan and Professor A.B.Grossman, unpublished data)

921

922

923 **FIGURE 3A.** Histology of adamantinomatous craniopharyngioma. Basal epithelium demonstrating the  
924 aberrant nuclear accumulation of beta-catenin in nodular whorls (arrowheads) due to beta-catenin  
925 mutation; Anti-beta-catenin, x400 magnification.

926 **FIGURE 3B.** Histology of papillary craniopharyngioma. Squamous epithelium showing membranous  
927 immunoreactivity of beta-catenin, lacking clusters with aberrant nuclear accumulation, x400  
928 magnification.

929  
930  
931 Reproduced from Lithgow K, Pohl U, Karavitaki N. Craniopharyngiomas. In: Endotext [Internet].  
932 South Dartmouth (MA): MDText.com, Inc.; 2000-2019 Mar 14.

933  
934

**FIGURE 4A** . Schematic presentation of adherens junction complex (including  $\beta$ - and  $\alpha$ -catenin and E-cadherin) and the molecular signaling pathways that may crosstalk in both, ACP or PCP. In ACP,  $\beta$ -catenin clusters cells show an activation of the Wnt, SHH and EGFR pathway, which are reported to crosstalk with each other. **2A.** When Wnt signaling is inactive, axin recruits and binds to GSK3 $\beta$ , CK1 $\alpha$ , and PP2A. APC is phosphorylated by these kinases, recruit  $\beta$ -catenin to the complex and tag it for degradation through phosphorylation of the serine (S33, S37, S45) or threonine (T41) residues of exon 3 on the CTNNB1 gene. This phosphorylation pattern is recognised by  $\beta$ -TrCP, which ubiquitinates  $\beta$ -catenin. When Wnt signaling is active, by the activation of a Wnt ligand or when extracellular SHH binds to the transmembrane receptor PTCH1, the protein degradation complex is inhibited and  $\beta$ -catenin is accumulated firstly in cytoplasm and then in the nucleus. There,  $\beta$ -catenin represents a transcription co-factor by interacting with transcription factors of the TCF family TCF/LEF1, initiating the target genes expression.

**FIGURE 4B.** In ACP, paracrine growth-related signaling molecules such as SOX2, FGFs, EGF, PDGF bind to their receptors activating the Ras/Raf/MAPK/ERK1/2 pathway and the PI3K/Akt pathway. Similarly, in PCP the BRAF-V600E serine/threonine kinase is a critical component of the Ras-Raf-Mek-ERK1/2 pathway that has been mutated. Red lines show the inhibitors that may be used as molecular-targeted therapy for CP.

*ACP: adamantinomatous craniopharyngioma; APC: adenomatosis polyposis coli; BMP: bone morphogenetic protein; CK1: casein kinase 1 $\alpha$ ; CP: craniopharyngiomas; Dvl: Dishevelled; EGFR: epidermal growth factor receptor; FGF: fibroblast growth factor; Fz: Frizzled; GSK-3 $\beta$ : glycogen synthase kinase-3 $\beta$ ; IKK- $\alpha$ : I kappa B kinase alpha; MAPK: mitogen-activated protein kinase; PCP: papillary craniopharyngioma; PI3K: phosphatidylinositol 3-kinase; PP2A: protein phosphatase 2A; PTCH1: patched 1; SHH: sonic hedgehog; TCF: T-cell factor; LEF1: lymphocyte enhancer factor 1. VEGF-R: vascular endothelial growth factor receptor*