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## Accepted Manuscript

Groundwater flooding: Ecosystem structure following an extreme recharge event

Julia Reiss, Daniel M. Perkins, Katarina E. Fussmann, Stefan Krause, Cristina Canhoto, Paul Romeijn, Anne L. Robertson



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**GROUNDWATER FLOODING: ECOSYSTEM STRUCTURE FOLLOWING AN  
EXTREME RECHARGE EVENT**

Julia Reiss<sup>1\*</sup>, Daniel M. Perkins<sup>1</sup>, Katarina E. Fussmann<sup>1</sup>, Stefan Krause<sup>2</sup>, Cristina Canhoto<sup>3</sup>,  
Paul Romeijn<sup>2</sup> and Anne L. Robertson<sup>1</sup>

<sup>1</sup>*Department of Life Sciences, Whitelands College, Roehampton University, London SW15  
4JD, United Kingdom.*

<sup>2</sup>*School of Geography, Earth and Environmental Sciences, University of Birmingham,  
Birmingham, B15 2TT, United Kingdom*

<sup>3</sup>*Departamento de Ciências da Vida, Faculdade de Ciências e Tecnologia, Universidade de  
Coimbra, Calçada Martim de Freitas, 3000-456 Coimbra, Portugal*

\* corresponding author Email: [julia.reiss@roehampton.ac.uk](mailto:julia.reiss@roehampton.ac.uk) (JR)

**Author Contributions**

Conceived and designed the study: AR, JR, SK. Performed the study: KF, AR, JR, SK, CC,  
PR. Analysed the data: DP, JR, SK. R-codes and Figures: DP. Wrote the paper: JR, KF, DP,  
SK, CC, AR.

**ABSTRACT**

1) Aquifers are recharged by surface water percolating through soil and rock and by connections with surface streams and rivers. Extreme rainfall can cause extensive flooding of surface waters and, eventually, of groundwaters. However, how the resultant changes in nutrients impact groundwater organisms and the structure of groundwater food webs is largely unknown.

2) We monitored abiotic (nutrients, temperature and more) and biotic (all organismal groups except viruses) conditions in eight groundwater boreholes in two locations in a chalk aquifer over the course of 25 weeks (ten sampling occasions), following an extreme rainfall- and groundwater-flooding event in the UK.

3) We show that groundwater flooding can cause substantial nutrient fertilisation of aquifers – nutrient concentrations (especially dissolved organic carbon) in the groundwater were highest when we started the sampling campaign, directly following the flood event, and then decreased over time while groundwater levels also declined back to their baseline.

4) Bacteria in the open water (i.e. bacteria not associated with sediment) became more abundant as the water table and DOC concentrations decreased. Importantly their functional richness tracked the DOC patterns, illustrating that bacteria were responsible for respiring DOC. Microbial metabolic activity and bacterial respiration, measured using smart tracers, supported this finding; DOC and microbial respiration showed a positive correlation.

5) The other biota (protists, micro- and macro-metazoans) showed different abundance patterns over time, but importantly, the entire sediment community, ranging from bacteria to macrofaunal species, showed a strong community size structure (mean size spectra slope: -1.12). Size spectra changed gradually through time towards steeper slopes, except in the very deep aquifer.

6) Our approach allowed us to demonstrate that groundwater communities track extreme changes in their usually stable environment, highlighting that they potentially buffer environmental change, although we still do not know what the limits of this ‘service’ might be.

Keywords: DOC, protozoan, bacteria, recharge, stygobite, metabolism

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## 1. Introduction

Aquifers hold more than 97% of our world's unfrozen fresh water and more than two billion people world-wide rely on groundwater for their daily supply of drinking water (see review on groundwater services by Griebler and Avramov, 2014). Aquifers are constantly recharged with water from the land's surface, as groundwater ecosystems are tightly linked with surface waters via a hydrological continuum with complex recharge–discharge processes (Boano et al., 2014; Boulton et al., 1998; Boulton and Hancock, 2006; Brunke and Gonser, 1997; Krause et al., 2017, 2011). However, extreme rainfall can result in the flooding of inland waters and these events are expected to become increasingly more frequent in the northern hemisphere (Taylor et al., 2012), driven by the increase of greenhouse gas concentrations in the atmosphere (Min et al., 2011). Apart from the devastating impact on the economy and human livelihoods (Munro et al., 2017), flooding has the potential to detrimentally impact on freshwater ecosystems, both, above- and below-ground (Taylor et al., 2012).

Surface water flooding can induce increases in groundwater levels with variable time lags. As water percolates from the surface, groundwater levels rise and fill sediment pore-spaces and fractures (groundwater flooding). During such extreme recharge events, infiltrating rainfall can potentially transport pollutants and nutrients, including carbon, from the surface to the aquifer (Taylor et al., 2013; Van Halem et al., 2009), affecting the delicate groundwater ecosystem and water quality. Background groundwater quality in general depends on the underlying geology of the aquifer (e.g. Weitowitz et al., 2017), however, to truly understand groundwater quality, we must recognize that groundwater is an ecosystem: in addition to the chemical and physical environment, it comprises a wide range of organisms

which have the ability to mediate biogeochemical processes and to immobilise or transform pollutants and nutrients (Griebler and Lueders, 2009).

Groundwater in Europe contains organisms that span more than 9 orders of magnitude in terms of size, from microbes such as bacteria, through microscopically small fauna (e.g. protists, rotifers and copepods) to macrofaunal crustaceans (e.g. *Niphargus* and *Proasellus* species) but we still know very little about groundwater biota and their interactions (e.g. within the food web). There is extreme endemism in this ecosystem (Gibert et al., 2009; Griebler and Lueders, 2009; Marmonier et al., 1993); most groundwater organisms are unique to this habitat (the macroscopic species are called ‘stygo-bites’) and are adapted to the low energy levels and lack of light that prevail. In mainland Britain they include the oldest inhabitants by millions of years (McInerney et al., 2014), forming distinctive truncated food webs that cannot be found in surface waters (Gibert and Deharveng, 2002).

Aquatic communities in surface waters, and especially invertebrate communities, are often described and compared to assess ecosystem health, using a range of approaches such as score systems with indicator species (e.g. Armitage et al., 1983), estimations of energy flow (e.g. Reiss and Schmid-Araya, 2010) or comparisons of community structure (e.g. Petchey & Belgrano 2010). For instance, stream invertebrate communities are strongly size-structured: small organisms are very abundant while larger ones are rare (Schmid, 2000) and the decrease in abundance from small to large organisms reflects the energy flow within the community. The size-structure of natural communities can be quantified by constructing size spectra (White et al., 2007), the frequency distribution of individual sizes within a community, which is depicted by plotting the total number of individuals occurring within ‘body size bins’ (White et al., 2007). Typically, this relationship is negative and linear on logarithmic axes and is quantified by the slope (Yvon-Durocher et al., 2011a). Since size spectra slopes provide integrated measures of trophic structure (Yvon-Durocher et al.,

2011a), they can be used to gauge ecosystem-level responses to environmental stressors (Petchey and Belgrano, 2010) such as pH (Layer et al., 2011; Mulder and Elser, 2009) and warming in fresh waters (Dossena et al., 2012; O’Gorman et al., 2012). Whether such an approach can be applied to groundwater ecosystems is unknown given that, to our knowledge, no groundwater study has estimated the body size and abundance of all groundwater groups and we do not know how these communities respond to stressors such as flooding.

We monitored abiotic and biotic conditions in a chalk aquifer following an extreme rainfall and river flooding event in the UK, by monitoring eight groundwater boreholes in two locations (Berkshire and Dorset) over the course of 25 weeks. The flooding had resulted in the elevation of water level in all boreholes by at least 10 m, and in two boreholes water levels had reached the land surface (Fig. 1). This extreme event permitted a uniquely important case study to track a pulse of energy through groundwater.

We characterized changes in the groundwater environment after the flooding by analysing its physical (groundwater levels, temperature), biogeochemical (nutrients such as carbon) and biotic (microbial biomass, metabolic activity, functional- and species richness) conditions. We then investigated the size structure of groundwaters by constructing size spectra (these included bacteria, protists and metazoans) and assessed temporal variability in the scaling of these relationships.

Our main objective was to measure the impact of this extreme flooding across the groundwater assemblage, from the energy supplied by nutrients (especially dissolved organic carbon) to prokaryotes through to macrofauna; and to track the recovery of the system after this exceptional groundwater recharge period through space and time.

We hypothesized that nutrients would have been washed into the groundwater and that this pulse would be tracked by the organisms in terms of abundance and (metabolic)



activity. We expected that the response of the community would be reflected in changes of the size distribution (e.g. because small organisms such as bacteria respond more rapidly to environmental change compared to larger ones such as macrofauna). Lastly we anticipated the deeper aquifer (>100 m deep) to be less affected by groundwater flooding, in terms of changes within the chemistry and biology, because both nutrient- and water load would have been attenuated within the underground layers above the deep groundwater.

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## 2. Methods

### 2.1 The sites

We sampled the chalk aquifer in two locations (in west Berkshire and east Dorset) in southern England from May to October 2014 (10 sampling occasions over 25 weeks, see Fig. 1, Appendix A, Tables A.1 and A.2) following a major storm period with subsequent groundwater flooding (Fig. 1). In each location, we selected 4 boreholes of similar topographic location (interfluves). In Berkshire we sampled: Bottom Barn (BB), Briff Lane (BL), Calversley Farm (CF) and Greendown Farm (GF); and in Dorset we sampled: Marley Bottom (MB; on 7 out of the 10 sampling occasions), Milborne (M), Newfield (NF) and Winterborne (WB). Boreholes enable access to the aquifer and, while faunal abundances are usually higher in boreholes than in fractures in the aquifers, the same species are found, and it is also possible to track abundance trends in the aquifer via sampling boreholes (often dubbed ‘windows to the aquifer’; Hahn and Matzke, 2005; Sorensen et al., 2013).

The water table in the boreholes was still unusually high when we started our sampling regime - from 2 to 7 meters above baseline levels, Fig. 1, Table A.1). The first sampling was on 06/05/14 (‘week 0’), followed by another 9 sampling occasions. Counting from week 0, we sampled in weeks 3, 5, 7, 10, 13, 16, 19, 22, and week 25 (see Table A.2 for dates). We used short sampling intervals to capture the expected rapid responses of the community to gradual flooding cessation. We also obtained historical data on waterlevels in all boreholes from the Environment Agency, which we combined with our measurements (Fig. 1).

Two sub-habitats were sampled in the boreholes: the open water (for abiotic parameters, bacteria and metabolic activity) and the sediment (biota). Further, sterile cotton strips were exposed in the boreholes for two weeks at a time to estimate fungal densities (90

cotton strips were evaluated but we did not detect any fungal biomass, see methods in Appendix A). Across boreholes, the sampling resulted in 77 samples for open water and sediment bacteria, 69 sediment samples for sediment protists and 77 samples for meiofauna and macrofauna (Table A.1).

## 2.2 Open water sampling: chemistry and bacteria

On each sampling occasion we measured the depth of the groundwater table, dissolved oxygen (see Table A.3), pH and conductivity *in situ* with a YSI sonde. Borehole temperatures were monitored with automatically logging thermistors; temperature was stable in all boreholes and there was little inter-borehole variation ( $10.31 \pm 0.011^\circ\text{C}$  over 7 months). On every sampling occasion, groundwater was collected with a bailer from each borehole and analysed for dissolved organic carbon (DOC; 77 samples, see Table A.2) and eight other nutrients (see Appendix A) by catalyst aided combustion on a Shimadzu TOC analyser.

We further took samples of bacteria in the open water column of the borehole (270 mL of water sampled with a bailer from the top of the water column). Bacterial abundances were expressed as individuals per L of open water. To estimate the abundance of bacteria, we took a 1ml sub-sample of the borehole sample that had been brought back from the field. For open water bacteria, we subsampled the 270 ml open water bailer sample and for sediment bacteria, we sub-sampled the sediment bailer sample (sediment suspended in 270 ml filtered borehole water). The 1 ml sub-samples were frozen and roughly half of that sample (0.495 ml) was subsequently analysed by flow cytometry (Gasol and Del Giorgio, 2000) using an Accuri C6 Flow Cytometer (BD Biosciences). A threshold of 8000 on the forward scatter (FSC-H), 2000 on the side scatter (SSC-H) and slow Fluidics setting was used. Each sample was measured for 1 minute. To stain the DNA of living cells, 200  $\mu\text{L}$  of PicoGreen dye solution (Quant-iT™ PicoGreen™ dsDNA Assay Kit, Sigma–Aldrich) was added to 1 mL

filtered water and incubated at 4°C for 15 min. Sediment particles were identified through the FL1-H channel (with values < log 3.1). The list of individual events returned by the flow cytometer was extracted using the R packages flowCore and flowViz (Ellis et al., 2009; Sarkar et al., 2008).

We used Biolog EcoPlates to assess the functional capacity of the open water microbial communities (e.g. Baho et al., 2012; Christian and Lind, 2006; Korbel et al., 2013; Stefanowicz, 2006), for each borehole and sampling occasion. Water samples were filtered through a 40 µm sterile sieve and 100 µL was pipetted into each of the 96 wells on a Biolog Eco-Plate: each EcoPlate contains 31 carbon substrates plus a no-substrate control in triplicate as well as a redox dye that turns purple if it is reduced when a given carbon source is metabolized (e.g. Stefanowicz, 2006). Plates were incubated at 10 °C for 7 days in the dark, after which time colour change was quantified by measuring optical density at 595 nm using a bench-top microplate photometer (Multiskan® EX, Thermo Scientific). We estimated the functional richness of bacteria communities as the % of substrates used ( $100 * \text{number of positive substrates} / 31$ ). We scored a carbon source as positive when two out of three wells reached a predetermined optical density (after Roger et al., 2016), which we set at 0.1 after subtraction of the mean blank from all wells.

### **2.3 Open Water: Microbial Metabolic Activity**

On the second sampling occasion only (28/4/14), we took samples to characterise the metabolic activity in the boreholes (except for MB) so that we could relate these measurements to DOC levels at that time. Microbial metabolic activity was measured by applying the “smart tracer” Resazuring/Resorufin (Raz/Rru) system (see Appendix A) that was first used in ecohydrological applications by Haggerty et al. (2009).

Samples were cooled and transferred to the laboratory without filtering to preserve the microbial community. The sample (250 ml) was incubated in duplicate in 1000 ml microcosms with a concentration of 100 µg/l Raz for 24 h. Two microcosms were prepared with only deionised water to act as control treatments. A detailed overview of these methods is available in Appendix A.

#### **2.4 Sediment: Sampling of Biota, Abundance and Community Composition**

We sampled the borehole sediment with a bailer (one 270 ml sample; to sample bacteria and protists in the sediment) and a net (microscopic and macroscopic metazoans; net diameter was 21, 11 or 4 cm depending on the borehole). The bottom bailer, used for bacteria and protists (Table A.2), sampled sediment with an area of 0.11 dm<sup>2</sup> (bailer radius was 0.19 dm) and the net samples emptied the entire area of the borehole bottom (this area varied depending on the borehole from ~0.2 dm<sup>2</sup> to ~7.3 dm<sup>2</sup>; see Table A.1). With the net, we removed the sediment in the borehole (3 net hauls that were pooled in the field). All sediment samples were initially stored in 270 ml filtered borehole water for the transport back to the laboratory. This way, all subsampling in the laboratory followed the same method.

Abundances of all sediment biota were expressed as individuals per dm<sup>2</sup>. The number of bacteria was estimated using flow cytometry as described above. Protists were measured, counted and identified alive in 1 ml (subsamped from 270 ml), using a Fuchs-Rosenthal counting chamber under a light microscope using 100 times and 400 times magnification (after Reiss and Schmid-Araya, 2010). Sediment samples for hard bodied meiofauna and macrofauna were preserved in formaldehyde for identification, measurement of body size and enumeration. For meiofauna and macrofauna, we did not sub-sample but counted individuals in the entire sample. For protists and metazoans, we assigned individuals to a taxonomic group (species in most cases) and measured the size (length and width) of individuals. This

also gave a record of abundances in the (sub-) samples (Table 1). For all groups, we calculated biomass by multiplying body mass by abundance.

## 2.5 Sediment Community Size Structure

The measurements of individual sizes from bacteria to meiofauna and the estimates of abundances allowed us to construct community size spectra. Bacteria cell size was estimated using calibration beads to convert forward scatter to average diameter of bacterial cells (see Fig. A.1; after Schaum et al., 2017). Cell carbon content (C) was estimated from bacterial cell volume, assuming  $0.10 \text{ pgC} / \mu\text{m}^3$  (Norland et al., 1987) - this gives a very similar result to a different method used by Fuhrman and Azam (1980).

Body length and width (for protist and meiofauna individuals) was converted to biovolume assuming a spherical shape. Individual dry mass (mg) was estimated by assuming a density of 1.1 to convert volume to wet weight, and the carbon content was assumed to be 10 % of the wet weight (Reiss and Schmid-Araya, 2010, 2008). Taking all species found into account, the body mass range was  $\sim 10$  orders of magnitude (see Table 1).

We computed the community size spectrum (CSS after White et al., 2007) for each borehole and time point by logarithmic binning of individual body masses,  $M$ , (White et al., 2007). The total range of  $\log_{10}(M)$  values were divided up into eight bins of equal width and the total abundance,  $N$ , of all individuals within each size class were regressed against the bin centers (cf Reuman et al., 2008). The intercept of the relationships were fixed at the smallest size class (after Yvon-Durocher et al., 2011b) which reduces the correlation between the slope and the intercept of linear relationships and makes the intercept of the model,  $\log N(M_{\min})$ , equivalent to the abundance of the smallest size class (here bacteria) as opposed to an infinite small mass. Size spectra slopes and intercepts were determined from simple linear regression analysis.

## 2.6 Statistical analysis

We used generalised additive mixed modelling (GAMM) via the *gamm* function in the *mgcv* package implemented in R (R Development Core Team, 2017) to investigate the general response of physical, chemical and biological variables in the open water and organism abundances through time. This approach allowed us to model the potential non-linear (smoothing) relationships between response variables and time (sampling week), accounting for the fact that each borehole is its own individual system (with unique baseline levels) and temporal correlations in residuals in the time series data (Zuur et al., 2009). We evaluated the significance of the smooth terms of the selected model based upon the extent to which their 95% CI intervals contained zero. A maximum number of knots of six was chosen as further required choices in model implementation: changing this value had no substantive effects on the general results. A temporal auto-correlation term was added to the models of the form  $\text{corAR1}(\text{form} = \sim 1 | \text{site})$  and the significance of the term was determined via a likelihood ratio tests of nested models with and without the auto-correlation term (Zuur et al., 2009).

For responses where relationships through time were linear, or where no significant smooth temporal trends were evident, we performed generalised least square (GLM) modelling to test our hypothesis that variation in the responses through time would be related to borehole depth. To do so we fitted Time (sampling week) and Borehole depth as continuous variables including an interaction term (Time  $\times$  Borehole depth). We fitted the same temporal auto-correlation structure as with the GAMM modelling and assessed the significance of this term as described above. We plotted all responses as histograms, as well as the residuals of the models, to identify non-normally distributed responses. Responses that were not normally distributed (concentrations, abundances and biomass density) were  $\log_{10}$ -

transformed prior to model fitting. A simple linear regression analysis was performed for the respiration data (nutrient levels explaining microbial activity). All statistical analysis was performed in R, version 3.4.1 (R Development Core Team, 2017).

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### 3. Results

#### 3.1 Open water measurements: waterlevels, DOC and bacteria

Our study captured the tail of groundwater flooding in the sampled aquifer induced by heavy rains in the winter of 2013 (Fig. 1). Groundwater flood response varied spatially, with levels reaching the surface at two locations (WB and NF in Dorset) and three locations were less affected by flooding (CF, GF and BL in Berkshire, which are also the deepest boreholes, i.e. here we sampled the deeper aquifer; Fig. 1; Table A.1). The water table in all boreholes was unusually high at the start of the sampling regime because the groundwater was still closer to the surface compared to its normal levels (at least 2 meters closer to the surface in all boreholes; Fig. 1; Table A.1).

Waterlevel, DOC, bacteria biomass and bacterial functional richness all changed over time (Fig. 2 shows the GAMM models that describe the patterns over time best, but see Fig. B.1 for raw data). The model inference procedure clearly selected the most complex GAMM models unambiguously rejecting both the null model (Table 2) and models without a temporal auto-correlation term (Appendix B, Table B.1) for all four responses. Examination of the P-values of the individual smooth terms (Table 2) and their confidence intervals (Fig. 2) provided further support for nonlinear temporal trends through time.

Over the 25 weeks study period, waterlevel decreased in an almost linear fashion (Fig. 2a), back to baseline levels (Fig. 1). DOC concentrations were highest at the beginning of the sampling regime and decreased until week 10 before gently rising towards week 16 and then dropping again (Fig. 2b); i.e. we observed the highest DOC values when waterlevel was high and the lowest DOC values when water level was low, but whilst waterlevel and DOC appear related, the pattern is not a simple, linear one (Fig. 2). Bacterial biomass in the open water increased over the sampling period, as waterlevel and nutrients were declining (Fig. 2c, Table

B.1), but peaked around week 16 before declining. Thus, DOC pulses (Fig. 2b) were not immediately tracked by the open water bacteria (Fig. 2c). Bacterial functional richness was generally high with 45 to 100% of carbon substrates utilised. Functional richness was initially high at the beginning of the sampling regime and decreased until week 5 before rising towards week 10 and dropping after week 20 (Fig. 2d).

Groundwater samples from May 2014 showed microbial metabolic activity above control conditions, indicating recharge driven nutrient fertilisation of groundwater microbial communities. The analysis of Raz/Rru conversion rates as indicators of microbial metabolic activity revealed that increased DOC levels correlated with high microbial metabolic activity and respiration ( $R^2 = 0.66$ , Fig. B.2).

### 3.2 Sediment biota: community composition and size structure

In the sediment, we identified 67 different protist taxa, 7 microscopic metazoan taxa and 9 macrofaunal taxa (Table B.2). Very small ciliates and flagellates were the most abundant component of the protists (Table B.2), while copepods dominated the meiofauna in all boreholes (Table B.2), and *Niphargus kochianus* was the most frequently found and abundant stygobite within the macrofauna (in Calversley Farm the total counted exceeded 1000 individuals over the 25 weeks).

There was no general relationship between abundance and time for the four organism groups. This was the case irrespective of whether GLS or GAMM models were used (Tables B.3 and B.4), indicating that the responses of organism abundance through time were borehole specific and could not be captured using either non-linear or linear models or by the addition of borehole depth as an explanatory variable. Although bacteria and protists increased in abundance over time in most boreholes (Fig. B.3), there was considerable difference between boreholes regarding abundance peaks and (as a consequence) the

abundance patterns were unrelated to borehole depth (Table B.3) or any other variables we investigated. Bacterial counts exceeded  $10^9$  ind.  $\text{dm}^{-2}$  and protists showed an average abundance of  $\sim 70,000$  ind.  $\text{dm}^{-2}$ . While bacteria and protists were present in all boreholes and on all sampling occasions, meiofauna and macrofauna were sometimes absent on particular sampling occasions (Fig. B.3).

Meiofauna were always present in BB, BL, CF and MB and macrofauna were always present in BB, CF and MB. In two of the boreholes, Bottom Barn and Calversley Farm, macrofaunal abundance increased steadily from under 10 to over 40 individuals per  $\text{dm}^2$  (mainly *N. kochianus* and *P. cavaticus*) over the 25 weeks (Fig. B.3). Mean abundance of meiofauna and macrofauna in the boreholes was  $\sim 5$  ind.  $\text{dm}^{-2}$  and  $\sim 7$  ind.  $\text{dm}^{-2}$  respectively.

We used mass-abundance spectra to estimate the response of the entire community. As expected, over 10 orders of magnitude in body mass from bacteria to macrofauna (Figs. 3 and B.4), abundance declined linearly. The average slope of the size spectrum, across sites and sampling occasions was  $-1.12$  (95% CI:  $-1.16$  to  $-1.07$ , Fig. 3). Size spectra slopes did not vary systematically through time or with borehole depth (Table 3). However, a significant time  $\times$  depth interaction was evident (Table 3) with slopes becoming steeper (more negative) through time for 'shallower' boreholes (Fig. 3). This pattern can be explained by the fact that, whilst not individually significant, bacteria and protozoan abundances tended to increase through time in the 'shallower' boreholes, whilst meiofauna and macrofauna abundances were relatively constant (Fig. B.3). This is also supported by the marginally significant p-value for the time  $\times$  depth interaction for size spectra intercepts which were fixed at the minimum body size class (Table 3).

#### 4. Discussion

This study was the first to analyse the impacts of groundwater flooding induced intensive recharge using an extensive analysis of both abiotic and biotic factors. We showed that nutrient concentrations increased in response to elevated groundwater levels, with more nutrients being available for groundwater organisms in consequence. Bacteria in the open water (i.e. bacteria not associated with sediment) became more abundant as groundwater levels and DOC concentrations decreased, indicating that they incorporated the additional energy. The measures of bacterial functional richness show that bacterial richness was high when DOC was high, demonstrating the link between nutrients and functional richness and bacterial activity. This finding was further supported by the respiration study that revealed a positive correlation between DOC concentrations and microbial metabolic activity in boreholes (see 4.1 Open water chemistry and biota). In the sediment, the change over time back to baseline levels was reflected in changes of the size distribution (measurements of individual groups were less conclusive) of the community towards steeper M-N slopes because, overall, small organisms (bacteria and protozoans) increased in abundance while the larger ones did not. This was however not the case in the deeper aquifer which was also much less affected by the flooding (see 4.2 Sediment biota).

##### 4.1 Open water chemistry and biota

Groundwater that is enriched with nutrients (e.g. in catchments with intensive agriculture) has previously been shown to exhibit high bacterial biodiversity values (Stein et al., 2010), which is atypical for pristine groundwater of comparable systems (Griebler et al., 2010). In accordance with these studies, we found that temporally variable DOC levels can produce similar differences in bacterial richness over time (however, in our case measured as functional richness not as operational taxonomic units used by these authors). Intriguingly,

the DOC pattern observed over time shows a ‘new pulse’ of nutrients after some weeks of sampling. We can only speculate about the reasons behind this pattern but two scenarios are possible: the new pulse could be the result of a new input from the surface or be the result of food web responses: the pattern resembles nutrient dynamics in chemostats, where bacteria can first utilise the nutrients, then crash and subsequently recover (e.g. Behrends et al., 2014).

#### **4.2 Sediment biota**

Intriguingly, with exception of the macrofauna in two of the boreholes, the sediment biota (bacteria, protists, microscopic metazoans and macrofauna) did not show any statistically significant response to the changes in nutrient concentrations or water table (although trends of increasing abundance of bacteria and protists in the shallow aquifer were obvious). The non-significance is largely due to two methodological issues: firstly, boreholes were used as replicates, which ‘worked’ for very robust patterns such as water level decline, DOC levels and size spectra, but for abundance patterns of individual groups each borehole represents its own unique system (e.g. in some boreholes groups such as meiofauna were almost absent despite the fact that the borehole connects to the same aquifer as a near-by borehole with meiofauna; see Fig. B.3). Secondly, GAMM is a useful method to show trends but it is not possible to fit linear correlations (such as fitting borehole depth as a predictor).

For all boreholes, species richness of macrofauna was similar to that reported elsewhere in the UK (Robertson et al 2009) but lower than reported in Europe (Griebler et al., 2010). Protozoan species richness was comparable to other studies outside the UK (Loquay et al., 2009), possibly because protozoans can colonise ‘extreme’ habitats more easily than metazoans. Previous studies on 5 of the 8 boreholes found the same macrofauna species, and that, like this study, these species were more abundant than meiofaunal copepods (they represent the majority of the meiofauna) (Johns et al., 2015; Maurice et al., 2016). This

pattern seems to be unique for groundwater habitats, for example, in streams, and rivers, meiofauna are always more abundant than larger invertebrates (e.g. Stead et al. 2002) and indeed, we would expect higher abundances of small invertebrates compared to larger ones from metabolic theory (Brown et al., 2004).

We observed systematic changes in the size spectrum over the 25 weeks monitoring period, with slopes of the size abundance relationship (the spectra) becoming steeper (more negative) in the ‘shallower’ boreholes. This pattern is consistent with the notion that the community dynamics at the bottom of shallower boreholes are more closely coupled with conditions in the open water compared to deep boreholes. Shallow boreholes received increased carbon resources and exhibited an increase in microbial biomass and metabolism over the sampling period. In other words, compared to the shallow aquifer, the deeper aquifer receives much less nutrients and water load when groundwater is flooded. These findings also highlight that size spectra analysis can provide key insights into the responses of ecosystems to environmental variability (Petchey and Belgrano, 2010) and understanding energy flow through groundwater food webs.

The size spectra analysis also revealed that groundwater communities were strongly size-structured with the slope of the relationships ( $N \sim M^{-1.1}$ ) similar to the negatively isometric pattern often reported for aquatic food webs (Kerr and Dickie, 2001). For any given size spectrum, the slope depends on the community-wide mean predator–prey mass ratio (PPMR: the mean size of predators relative to prey), and the trophic transfer efficiency (TE: the proportion of prey production converted to predator production) (Kerr and Dickie, 2001). The model of Brown and Gillooly (2003) incorporates general allometric scaling principles to predict the scaling of abundance with body mass as:  $N = M^\lambda \times M^{\log(\text{TE}) / + \log(\text{PPMR})}$ , where  $\lambda$  is the  $M$ - $N$  scaling coefficient within trophic levels (typically  $-3/4$ ), the reciprocal of the mass-dependence of metabolic rate for many multicellular taxa (Brown et al., 2004). The trophic

structure of groundwater food webs is largely unknown but assuming a TE range of between 10-20% (Jennings and Mackinson, 2003) suggests that the PPMR ratio in these systems is somewhere between 90:1 and 600:1, similar to other benthic freshwater ecosystems (Perkins et al., in press, Woodward and Warren 2007).

Abundances were generally high compared with other aquifer studies (Griebler et al., 2010), which is explained by the fact that we sampled boreholes (Hahn and Matzke, 2005; Sorensen et al., 2013) and did not pump the aquifer water (see references in Griebler and Lueders, 2009 and Korbel et al., 2017). Collecting groundwater from boreholes is the most common method for groundwater sampling because they provide the only suitable sampling 'window' into deeper aquifers. Previous studies have shown that boreholes have higher abundance but the same species composition compared to the rest of the aquifer (Hahn and Matzke, 2005, Sorensen et al., 2013). However, boreholes do still represent groundwater community patterns over time (our focus) and species composition in the aquifer.

#### **4.3 The wider context and future directions**

Ecological assessment of groundwater is a very recent discipline (see reviews by Danielopol et al., 2003; Gregory et al., 2014; Griebler et al., 2014; Steube et al., 2009). However, we now have methods, such as cytometry (Bayer et al., 2016), that help us to monitor stygobite populations and also to assess microscopically small organisms in groundwater (see literature for bacteria, protists and meiofauna reviewed in Griebler and Lueders (2009) and Novarino et al. (1997) for protists).

It will be crucial to determine if groundwater biota have the potential to buffer inputs of nutrients through increased secondary production. Although their environment is stable, the relatively low temperatures might inhibit the incorporation of these additional nutrients into biomass. We believe a more rigorous analysis of the groundwater food web is needed to

show how the composition and activity of bacteria changes when nutrient availability changes and whether the bacterial response is coupled with increased secondary production of eukaryotes. Further, it is vital to include hydrological knowledge in these studies because they determine how stable the habitat is. Menció et al. (2014) found that steadier groundwater head levels and lower nitrate concentrations promote a more diverse and abundant stygofauna community. Historical hydrological data from UK chalk aquifers, and our study, showed that the deep aquifer is less affected by flooding and it could therefore represent a very stable environment for groundwater organisms. However, shallower aquifers (down to 100 m) seem to respond rapidly to flooding and we do not know to what extent the groundwater community is perturbed in these important underground layers that hold clean fresh water. As extreme rainfall events are predicted to become more frequent in the Northern hemisphere, it is likely that groundwater quality will ultimately be affected.



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## References

- Armitage, P.D., Moss, D., Wright, J.F., Furse, M.T., 1983. The performance of a new biological water quality score system based on macroinvertebrates over a wide range of unpolluted running-water sites. *Water Res.* 17, 333–347.
- Baho, D.L., Peter, H., Tranvik, L.J., 2012. Resistance and resilience of microbial communities—temporal and spatial insurance against perturbations. *Environ. Microbiol.* 14, 2283–2292.
- Bayer, A., Drexel, R., Weber, N., Griebler, C., 2016. Quantification of aquatic sediment prokaryotes—A multiple-steps optimization testing sands from pristine and contaminated aquifers. *Limnologica* 56, 6–13. <https://doi.org/10.1016/j.limno.2015.11.003>
- Behrends, V., Maharjan, R.P., Ryall, B., Feng, L., Liu, B., Wang, L., Bundy, J.G., Ferenci, T., 2014. A metabolic trade-off between phosphate and glucose utilization in *Escherichia coli*. *Mol. Biosyst.* 10, 2820–2822. <https://doi.org/10.1039/c4mb00313f>
- Boano, F., Harvey, J.W., Marion, A., Packman, A.I., Revelli, R., Ridolfi, L., Wörman, A., 2014. Hyporheic flow and transport processes: Mechanisms, models, and biogeochemical implications. *Rev. Geophys.* 52, 603–679.
- Boulton, A.J., Findlay, S., Marmonier, P., Stanley, E.H., Valett, H.M., 1998. The functional significance of the hyporheic zone in streams and rivers. *Annu. Rev. Ecol. Syst.* 29, 59–81.
- Boulton, A.J., Hancock, P.J., 2006. Rivers as groundwater-dependent ecosystems: a review of degrees of dependency, riverine processes and management implications. *Aust. J. Bot.* 54, 133–144.
- Brown, J., Gillooly, J., Allen, A., Savage, V., West, G., 2004. Toward a metabolic theory of ecology. *Ecology* 85, 1771–1789.
- Brown, J.H., Gillooly, J.F., 2003. Ecological food webs: high-quality data facilitate theoretical unification. *Proc. Natl. Acad. Sci.* 100, 1467–1468.
- Brunke, M., Gonser, T.O.M., 1997. The ecological significance of exchange processes between rivers and groundwater. *Freshw. Biol.* 37, 1–33.
- Christian, B.W., Lind, O.T., 2006. Key issues concerning Biolog use for aerobic and anaerobic freshwater bacterial community-level physiological profiling. *Int. Rev. Hydrobiol.* 91, 257–268. <https://doi.org/10.1002/iroh.200510838>
- Danielopol, D.L., Griebler, C., Gunatilaka, A., Notenboom, J., 2003. Present state and future prospects for groundwater ecosystems. *Environ. Conserv.* 30, 104–130.
- Dossena, M., Yvon-Durocher, G., Grey, J., Montoya, J.M., Perkins, D.M., Trimmer, M., Woodward, G., 2012. Warming alters community size structure and ecosystem functioning. *Proc. R. Soc. London B Biol. Sci.* 279, 3011–3019.
- Ellis, B., Haaland, P., Hahne, F., Le Meur, N., Gopalakrishnan, N., Spidlen, J., Jiang, M., 2009. flowCore: basic structures for flow cytometry data. R Packag. version 1.
- Fuhrman, J.A., Azam, F., 1980. Bacterio Plankton Secondary Production Estimates for

- Coastal Waters of British-Columbia Canada Antarctica and California Usa. *Appl. Environ. Microbiol.* 39, 1085–1095.
- Gasol, J.M., Del Giorgio, P.A., 2000. Using flow cytometry for counting natural planktonic bacteria and understanding the structure of planktonic bacterial communities. *Sci. Mar.* 64, 197–224.
- Gibert, J., Culver, D.C., Dole-Olivier, M.J., Malard, F., Christman, M.C., Deharveng, L., 2009. Assessing and conserving groundwater biodiversity: Synthesis and perspectives. *Freshw. Biol.* <https://doi.org/10.1111/j.1365-2427.2009.02201.x>
- Gibert, J., Deharveng, L., 2002. Subterranean Ecosystems: A Truncated Functional Biodiversity: This article emphasizes the truncated nature of subterranean biodiversity at both the bottom (no primary producers) and the top (very few strict predators) of food webs and discusses the implic. *AIBS Bull.* 52, 473–481.
- Gregory, S.P., Maurice, L.D., West, J.M., Goody, D.C., 2014. Microbial communities in UK aquifers: current understanding and future research needs. *Q. J. Eng. Geol. Hydrogeol.* 47, 145–157.
- Griebler, C., Avramov, M., 2014. Groundwater ecosystem services: a review. *Freshw. Sci.* 34, 355–367.
- Griebler, C., Lueders, T., 2009. Microbial biodiversity in groundwater ecosystems. *Freshw. Biol.* <https://doi.org/10.1111/j.1365-2427.2008.02013.x>
- Griebler, C., Malard, F., Lefébure, T., 2014. Current developments in groundwater ecology—from biodiversity to ecosystem function and services. *Curr. Opin. Biotechnol.* 27, 159–167.
- Griebler, C., Stein, H., Kellermann, C., Berkhoff, S., Brielmann, H., Schmidt, S., Selesi, D., Steube, C., Fuchs, A., Hahn, H.J., 2010. Ecological assessment of groundwater ecosystems - Vision or illusion? *Ecol. Eng.* 36, 1174–1190. <https://doi.org/10.1016/j.ecoleng.2010.01.010>
- Haggerty, R., Martí, E., Argerich, A., Von Schiller, D., Grimm, N.B., 2009. Resazurin as a “smart” tracer for quantifying metabolically active transient storage in stream ecosystems. *J. Geophys. Res. Biogeosciences* 114.
- Hahn, H.J., Matzke, D., 2005. A comparison of stygofauna communities inside and outside groundwater bores. *Limnol. Manag. Inl. Waters* 35, 31–44.
- Jennings, S., Mackinson, S., 2003. Abundance–body mass relationships in size-structured food webs. *Ecol. Lett.* 6, 971–974.
- Johns, T., Jones, J.I., Knight, L., Maurice, L., Wood, P., Robertson, A., 2015. Regional-scale drivers of groundwater faunal distributions. *Freshw. Sci.* 34, 316–328. <https://doi.org/10.1086/678460>
- Kerr, S.R., Dickie, L.M., 2001. *The biomass spectrum: a predator-prey theory of aquatic production.* Columbia University Press.
- Korbel, K.L., Hancock, P.J., Serov, P., Lim, R.P., Hose, G.C., 2013. Groundwater ecosystems vary with land use across a mixed agricultural landscape. *J. Environ. Qual.*

42, 380–390.

- Korbel, K., Chariton, A., Stephenson, S., Greenfield, P. and Hose, G.C., 2017. Wells provide a distorted view of life in the aquifer: implications for sampling, monitoring and assessment of groundwater ecosystems. *Scientific Reports* 7, 40702.
- Krause, S., Hannah, D.M., Fleckenstein, J.H., Heppell, C.M., Kaeser, D., Pickup, R., Pinay, G., Robertson, A.L., Wood, P.J., 2011. Inter-disciplinary perspectives on processes in the hyporheic zone. *Ecohydrology* 4, 481–499.
- Krause, S., Lewandowski, J., Grimm, N.B., Hannah, D.M., Pinay, G., McDonald, K., Martí, E., Argerich, A., Pfister, L., Klaus, J., 2017. Ecohydrological interfaces as hot spots of ecosystem processes. *Water Resour. Res.* 53, 6359–6376.
- Layer, K., Hildrew, A.G., Jenkins, G.B., Riede, J.O., Rossiter, S.J., Townsend, C.R., Woodward, G., 2011. Long-Term Dynamics of a Well-Characterised Food Web. Four Decades of Acidification and Recovery in the Broadstone Stream Model System, *Advances in Ecological Research*. <https://doi.org/10.1016/B978-0-12-374794-5.00002-X>
- Loquay, N., Wylezich, C., Arndt, H., 2009. Composition of groundwater nanoprotist communities in different aquifers based on aliquot cultivation and genotype assessment of cercozoans. *Fundam. Appl. Limnol.* 174, 261–269. <https://doi.org/10.1127/1863-9135/2009/0174-0261>
- Marmonier, P., Vervier, P., Giber, J., Dole-Olivier, M.-J., 1993. Biodiversity in ground waters. *Trends Ecol. Evol.* 8, 392–395.
- Maurice, L., Robertson, A.L., White, D., Knight, L., Johns, T., Edwards, F., Arietti, M., Sorensen, J.P.R., Weitowitz, D., Marchant, B.P., Bloomfield, J.P., 2016. The invertebrate ecology of Chalk groundwaters. *Hydrogeol. J.* 24, 459–474.
- McInerney, C.E., Maurice, L., Robertson, A.L., Knight, L.R.F.D., Arnscheidt, J., Venditti, C., Dooley, J.S.G., Mathers, T., Matthijs, S., Eriksson, K., 2014. The ancient Britons: groundwater fauna survived extreme climate change over tens of millions of years across NW Europe. *Mol. Ecol.* 23, 1153–1166.
- Menció, A., Korbel, K.L., Hose, G.C., 2014. River-aquifer interactions and their relationship to stygofauna assemblages: A case study of the Gwydir River alluvial aquifer (New South Wales, Australia). *Sci. Total Environ.* 479–480, 292–305. <https://doi.org/10.1016/j.scitotenv.2014.02.009>
- Min, S.-K., Zhang, X., Zwiers, F.W., Hegerl, G.C., 2011. Human contribution to more-intense precipitation extremes. *Nature* 470, 378–381.
- Mulder, C., Elser, J.J., 2009. Soil acidity, ecological stoichiometry and allometric scaling in grassland food webs. *Glob. Chang. Biol.* 15, 2730–2738. <https://doi.org/10.1111/j.1365-2486.2009.01899.x>
- Munro, A., Kovats, R.S., Rubin, G.J., Waite, T.D., Bone, A., Armstrong, B., Beck, C.R., Amlôt, R., Leonardi, G., Oliver, I., 2017. Effect of evacuation and displacement on the association between flooding and mental health outcomes: a cross-sectional analysis of UK survey data. *Lancet Planet. Heal.* 1, e134–e141.

- Norland, S., Heldal, M., Tumyr, O., 1987. On the relation between dry matter and volume of bacteria. *Microb. Ecol.* 13, 95–101. <https://doi.org/10.1007/BF02011246>
- Novarino, G., Warren, A., Butler, H., Lambourne, G., Boxshall, A., Bateman, J., Kinner, N.E., Harvey, R.W., Mosse, R.A., Teltsch, B., 1997. Protistan communities in aquifers: A review, in: *FEMS Microbiology Reviews*. pp. 261–275. [https://doi.org/10.1016/S0168-6445\(97\)00046-6](https://doi.org/10.1016/S0168-6445(97)00046-6)
- O’Gorman, E.J., Pichler, D.E., Adams, G., Benstead, J.P., Cohen, H., Craig, N., Cross, W.F., Demars, B.O.L., Friberg, N., Gislason, G.M., 2012. Impacts of warming on the structure and functioning of aquatic communities: individual-to ecosystem-level responses, in: *Advances in Ecological Research*. Elsevier, pp. 81–176.
- Perkins, D.M., Durance, I., Edwards, F.K., Grey, J., Hildrew, A.G., Jackson, M., Jones, J.I., Lauridsen, R., Layer-Dobra, K., Thompson, M., Woodward, G., n.d. Bending the rules: exploitation of allochthonous resources by a top-predator modifies size-abundance scaling in stream food webs. *Ecol. Lett.*
- Petchey, O.L., Belgrano, A., 2010. Body-size distributions and size-spectra: universal indicators of ecological status?
- R Development Core Team, 2017. A language and environment for statistical computing.
- Reiss, J., Schmid-Araya, J.M., 2010. Life history allometries and production of small fauna. *Ecology* 91, 497–507. <https://doi.org/10.1890/08-1248.1>
- Reiss, J., Schmid-Araya, J.M., 2008. Existing in plenty: Abundance, biomass and diversity of ciliates and meiofauna in small streams. *Freshw. Biol.* 53, 652–668. <https://doi.org/10.1111/j.1365-2427.2007.01907.x>
- Reuman, D.C., Mulder, C., Raffaelli, D., Cohen, J.E., 2008. Three allometric relations of population density to body mass: theoretical integration and empirical tests in 149 food webs. *Ecol. Lett.* 11, 1216–28. <https://doi.org/10.1111/j.1461-0248.2008.01236.x>
- Robertson, A.L., Smith, J.W.N., Johns, T., Proudlove, G.S., 2009. The distribution and diversity of stygobites in Great Britain: an analysis to inform groundwater management. Geological Society of London.
- Roger, F., Bertilsson, S., Langenheder, S., Osman, O.A., Gamfeldt, L., 2016. Effects of multiple dimensions of bacterial diversity on functioning, stability and multifunctionality. *Ecology* 97, 2716–2728.
- Sarkar, D., Le Meur, N., Gentleman, R., 2008. Using flowViz to visualize flow cytometry data. *Bioinformatics* 24, 878–879.
- Schaum, C.-E., Barton, S., Bestion, E., Buckling, A., Garcia-Carreras, B., Lopez, P., Lowe, C., Pawar, S., Smirnov, N., Trimmer, M., 2017. Adaptation of phytoplankton to a decade of experimental warming linked to increased photosynthesis. *Nat. Ecol. Evol.* 1, 94.
- Schmid, P.E., 2000. Relation Between Population Density and Body Size in Stream Communities. *Science* (80-. ). 289, 1557–1560. <https://doi.org/10.1126/science.289.5484.1557>

- Sorensen, J.P.R., Maurice, L., Edwards, F.K., Lapworth, D.J., Read, D.S., Allen, D., Butcher, A.S., Newbold, L.K., Townsend, B.R., Williams, P.J., 2013. Using boreholes as windows into groundwater ecosystems. *PLoS One* 8, e70264.
- Stefanowicz, a, 2006. The Biolog Plates Technique as a tool in ecological studies of microbial communities. *Polish J. Environ. Stud.* 15, 669–676.  
<https://doi.org/http://dx.doi.org/10.3390/s120303253>
- Stein, H., Kellermann, C., Schmidt, S.I., Brielmann, H., Steube, C., Berkhoff, S.E., Fuchs, A., Hahn, H.J., Thulin, B., Griebler, C., 2010. The potential use of fauna and bacteria as ecological indicators for the assessment of groundwater quality. *J. Environ. Monit.* 12, 242–254.
- Steube, C., Richter, S., Griebler, C., 2009. First attempts towards an integrative concept for the ecological assessment of groundwater ecosystems. *Hydrogeol. J.* 17, 23–35.
- Taylor, R.G., Scanlon, B., Döll, P., Rodell, M., Van Beek, R., Wada, Y., Longuevergne, L., Leblanc, M., Famiglietti, J.S., Edmunds, M., 2013. Ground water and climate change. *Nat. Clim. Chang.* 3, 322–329.
- Taylor, R.G., Scanlon, B., Döll, P., Rodell, M., van Beek, R., Wada, Y., Longuevergne, L., Leblanc, M., Famiglietti, J.S., Edmunds, M., Konikow, L., Green, T.R., Chen, J., Taniguchi, M., Bierkens, M.F.P., MacDonald, A., Fan, Y., Maxwell, R.M., Yechieli, Y., Gurdak, J.J., Allen, D.M., Shamsudduha, M., Hiscock, K., Yeh, P.J.-F., Holman, I., Treidel, H., 2012. Ground water and climate change. *Nat. Clim. Chang.* 3, 322–329.  
<https://doi.org/10.1038/nclimate1744>
- Van Halem, D., Bakker, S.A., Amy, G.L., Van Dijk, J.C., 2009. Arsenic in drinking water: a worldwide water quality concern for water supply companies. *Drink. Water Eng. Sci.* 2, 2009.
- Weitowitz D., Maurice L., Lewis M., Bloomfield J., Reiss J. Robertson A.L., 2017. Defining geo-habitats for groundwater ecosystem assessments: An example from England and Wales (UK). *Hydrogeology Journal*. DOI:10.1007/s10040-017-1629-6
- White, E.P., Ernest, S.K.M., Kerkhoff, A.J., Enquist, B.J., 2007. Relationships between body size and abundance in ecology. *Trends Ecol. Evol.* 22, 323–30.  
<https://doi.org/10.1016/j.tree.2007.03.007>
- Woodward, G., Warren, P.H., 2007. Body size and predatory interactions in freshwaters: scaling from individuals to communities. *Body size Struct. Funct. Aquat. Ecosyst.* 98–117.
- Yvon-Durocher, G., Reiss, J., Blanchard, J., Ebenman, B., Perkins, D.M., Reuman, D.C., Thierry, A., Woodward, G., Petchey, O.L., 2011a. Across ecosystem comparisons of size structure: Methods, approaches and prospects. *Oikos* 120, 550–563.  
<https://doi.org/10.1111/j.1600-0706.2010.18863.x>
- Yvon-Durocher, G., Montoya, J.M., Trimmer, M., Woodward, G.U.Y., 2011b. Warming alters the size spectrum and shifts the distribution of biomass in freshwater ecosystems. *Glob. Chang. Biol.* 17, 1681–1694.
- Zuur, A.F., Ieno, E.N., Walker, N.J., Saveliev, A.A., Smith, G.M., 2009. Mixed effects

models and extensions in ecology with R. Springer, New York.

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**Figure legends**

**Fig. 1:** 10 yrs time series of the groundwater table (measured as metres below the surface) for eight boreholes in the UK Chalk aquifer. The arrow shows the start of the flooding period (induced by heavy rains in Winter 2013) and the area shaded in grey indicates the sampling period (10 sampling occasions over 25 weeks). The water level on the y-axis is given as mAOD (Metres Above Ordnance Datum); a value of zero indicates that the waterlevel had reached the land surface.

**Fig. 2:** Water levels (a), DOC dynamics (b), bacterial biomass (c) and bacterial functional richness (d) over a 25 weeks study period following the tail of groundwater flooding in a chalk aquifer (two locations [Berkshire and Dorset] and a total of 8 boreholes). The plots are the results of GLMM and the patterns over time are significant in all cases. Note that peaks and dips in DOC correspond to those observed for dissolved organic carbon and functional richness of bacteria.

**Fig. 3:** Upper panel: Body mass and abundance scaling in groundwater communities. The average slope of the size spectrum, estimated from mixed-effects modelling, was - 1.12 (95% CI: -1.16 to -1.07) across boreholes and sampling occasions. Lower panel: Variation in size spectrum slopes in relation to sampling occasion and total borehole depth (here represented by shading of data points). The size spectrum slope steepened with each sampling occasion in the shallow boreholes (light grey data points).



**Table 1**

Overview of the biota found in eight boreholes showing sampling effort, body mass, number of individuals measured and the total of functional groups/taxa recorded.

Group	Total no of samples scanned	Min DWC in g	Max DWC in g	Total no of individuals measured and identified	Total functional groups/taxa
Open water bacteria	80	0.0000000001	0.000001	>4Mio	31 functional groups
Sediment bacteria	77	0.0000000001	0.000001	>4Mio	na
Protists	69	0.0000000428	0.000052	>2000	67
Meiofauna	77	0.0000321386	0.008484	>700	7
Macrofauna	77	0.0000681359	6.395189	>3000	9

Note: Across boreholes, this sampling resulted in 77 samples for open water bacteria (see S1), 69 sediment samples for protists, 77 samples for meiofauna and macrofauna (see S1); and 90 cotton strips were evaluated for fungal biomass (no ergosterol, i.e. fungal biomass, was detected). DWC= dry weight carbon.

**Table 2**

Summary output from generalised additive mixed modelling (GAMM) of open water parameters through time (sampling week).

Response	Smoother term	edf	F	p-value
Waterlevel	<b>s(time, k = 6)</b>	4.14	3.83	0.007
DOC (log10)	<b>s(time, k = 6)</b>	1.93	10.22	<.0001
Bacteria biomass (log10)	<b>s(time, k = 6)</b>	3.85	5.51	<.0001
Functional diversity	<b>s(time, k = 6)</b>	4.95	40.75	<.0001

Note: The results provide support for nonlinear temporal trends in response to flooding. Significant terms are highlighted in bold, k refers to the number of knots in the smoother term.

**Table 3**

Summary output from generalised least squares modelling (GLS) of size spectra parameters.

Term	DF	Size-spectrum slope		Size-spectrum intercept	
		F-value	p-value	F-value	p-value
Time	1,66	0.25	0.619	0.95	0.334
Depth	1,66	0	0.979	0.35	0.557
Time:Depth	1,66	4.98	<b>0.029</b>	2.89	0.094

Note: Significant terms are highlighted in bold.

**HIGHLIGHTS**

- Groundwater that had been flooded after an extreme rainfall event was sampled
- The flooding had resulted in high DOC levels that were tracked by bacteria
- Overall, small organisms increased in abundance while larger ones did not
- This altered the size distribution of the community towards steeper M-N slopes
- The deeper aquifer was less affected by the flooding

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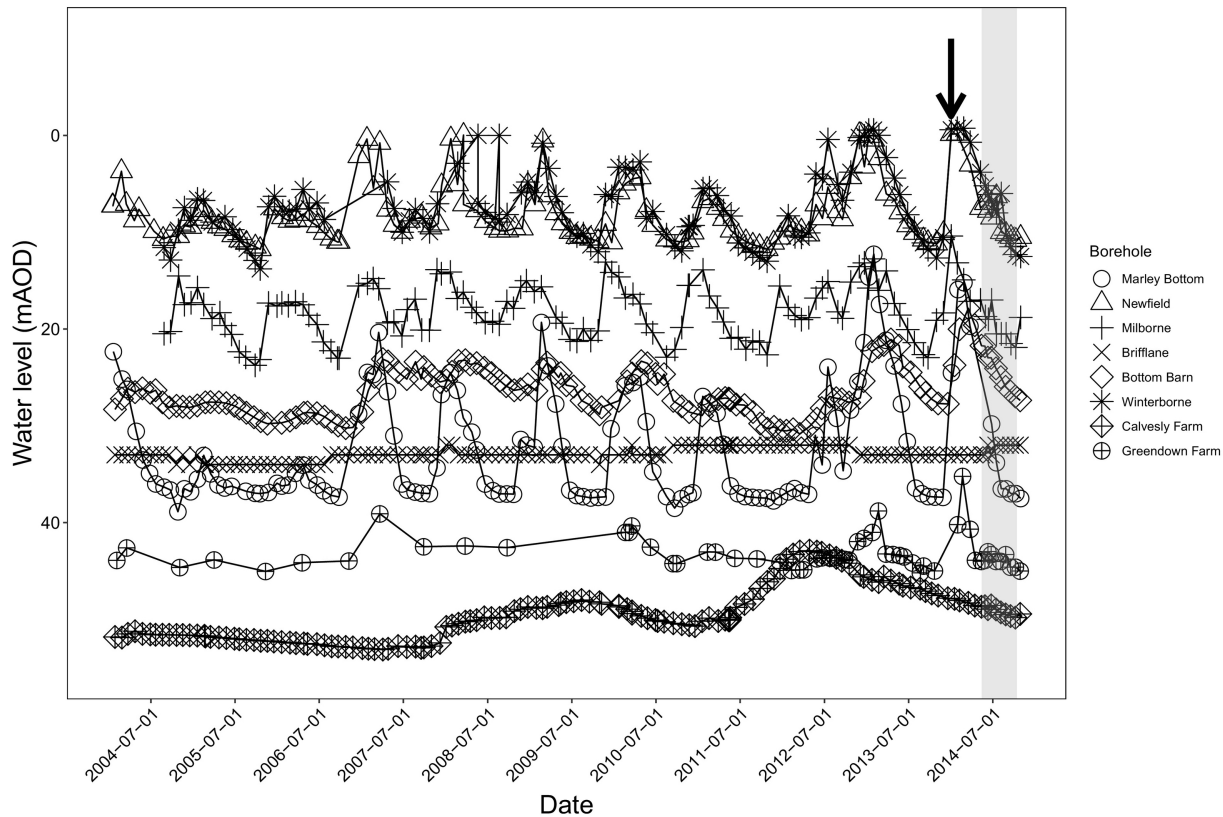


Figure 1

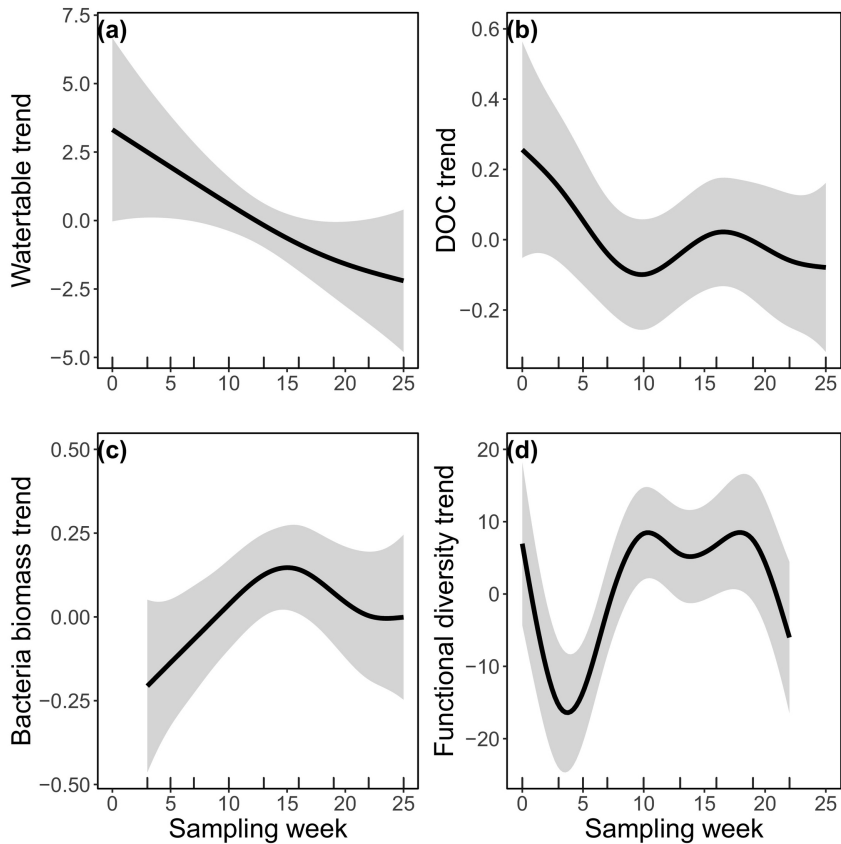


Figure 2

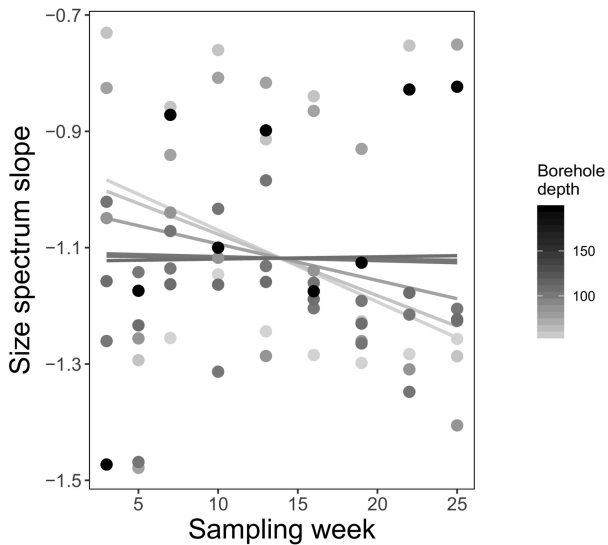
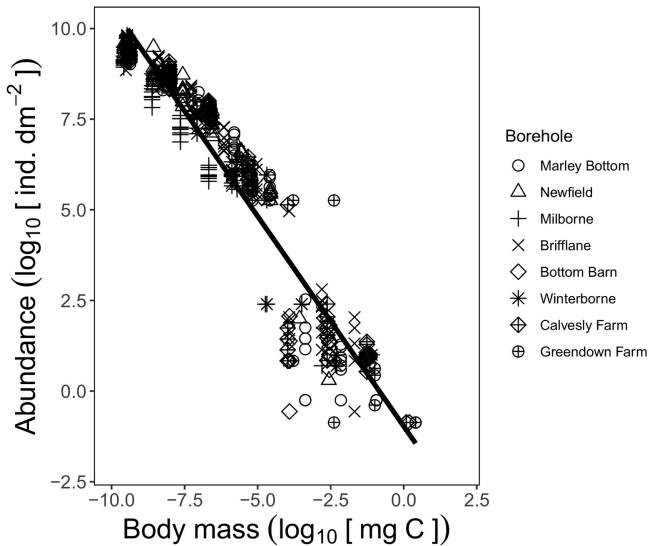


Figure 3