

Mass spectrometry based metabolomics comparison of liver grafts from Donors after Circulatory Death (DCD) and Donors after Brain Death (DBD) used in human orthotopic liver transplantation

Hrydziusko, Olga; Thamara, M. Thamara P R; Laing, Richard; Kirwan, Jennifer; Silva, Michael A.; Richards, Douglas A.; Murphy, Nick; Mirza, Darius F.; Viant, Mark R.

DOI:

[10.1371/journal.pone.0165884](https://doi.org/10.1371/journal.pone.0165884)

License:

Creative Commons: Attribution (CC BY)

Document Version

Publisher's PDF, also known as Version of record

Citation for published version (Harvard):

Hrydziusko, O, Thamara, MTPR, Laing, R, Kirwan, J, Silva, MA, Richards, DA, Murphy, N, Mirza, DF & Viant, MR 2016, 'Mass spectrometry based metabolomics comparison of liver grafts from Donors after Circulatory Death (DCD) and Donors after Brain Death (DBD) used in human orthotopic liver transplantation', *PLoS ONE*, vol. 11, no. 11, e0165884. <https://doi.org/10.1371/journal.pone.0165884>

[Link to publication on Research at Birmingham portal](#)

Publisher Rights Statement:

Checked for eligibility: 17/01/2017

Hrydziusko O, Perera MTPR, Laing R, Kirwan J, Silva MA, Richards DA, et al. (2016) Mass Spectrometry Based Metabolomics Comparison of Liver Grafts from Donors after Circulatory Death (DCD) and Donors after Brain Death (DBD) Used in Human Orthotopic Liver Transplantation. *PLoS ONE* 11(11): e0165884. doi:10.1371/journal.pone.0165884

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Download date: 06. May. 2024

RESEARCH ARTICLE

Mass Spectrometry Based Metabolomics Comparison of Liver Grafts from Donors after Circulatory Death (DCD) and Donors after Brain Death (DBD) Used in Human Orthotopic Liver Transplantation

Olga Hrydziusko¹✉, M. Thamara P. R. Perera²✉, Richard Laing², Jennifer Kirwan¹, Michael A. Silva², Douglas A. Richards³, Nick Murphy⁴, Darius F. Mirza¹, Mark R. Viant¹*

1 School of Biosciences, University of Birmingham, Edgbaston, Birmingham, B15 2TT, United Kingdom, **2** The Liver Unit, Queen Elizabeth Hospital Birmingham, Edgbaston, Birmingham, B15 2TH, United Kingdom, **3** The Department of Pharmacology, School of Medical and Dental Sciences, University of Birmingham, Edgbaston, Birmingham, B15 2TT, United Kingdom, **4** Department of Critical Care and Anaesthesia, Queen Elizabeth Hospital Birmingham, Edgbaston, Birmingham, B15 2TH, United Kingdom

✉ These authors contributed equally to this work.

* m.viant@bham.ac.uk



OPEN ACCESS

Citation: Hrydziusko O, Perera MTPR, Laing R, Kirwan J, Silva MA, Richards DA, et al. (2016) Mass Spectrometry Based Metabolomics Comparison of Liver Grafts from Donors after Circulatory Death (DCD) and Donors after Brain Death (DBD) Used in Human Orthotopic Liver Transplantation. PLoS ONE 11(11): e0165884. doi:10.1371/journal.pone.0165884

Editor: Francisco J. Schopfer, University of Pittsburgh, UNITED STATES

Received: November 16, 2015

Accepted: October 19, 2016

Published: November 11, 2016

Copyright: © 2016 Hrydziusko et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This study was supported by a limited study grant by Astellas Pharmaceuticals (UK) not exceeding 60000.00GBP to MTPRP, MAS and DFM for phase II study investigating biomarkers of poor allograft function following human liver transplantation. The funders had no role in study

Abstract

Use of marginal liver grafts, especially those from donors after circulatory death (DCD), has been considered as a solution to organ shortage. Inferior outcomes have been attributed to donor warm ischaemic damage in these DCD organs. Here we sought to profile the metabolic mechanisms underpinning donor warm ischaemia. Non-targeted Fourier transform ion cyclotron resonance (FT-ICR) mass spectrometry metabolomics was applied to biopsies of liver grafts from donors after brain death (DBD; $n = 27$) and DCD ($n = 10$), both during static cold storage (T_1) as well as post-reperfusion (T_2). Furthermore 6 biopsies from DBD donors prior to the organ donation (T_0) were also profiled. Considering DBD and DCD together, significant metabolic differences were discovered between T_1 and T_2 (688 peaks) that were primarily related to amino acid metabolism, meanwhile T_0 biopsies grouped together with T_2 , denoting the distinctively different metabolic activity of the perfused state. Major metabolic differences were discovered between DCD and DBD during cold-phase (T_1) primarily related to glucose, tryptophan and kynurenine metabolism, and in the post-reperfusion phase (T_2) related to amino acid and glutathione metabolism. We propose tryptophan/kynurenine and S-adenosylmethionine as possible biomarkers for the previously established higher graft failure of DCD livers, and conclude that the associated pathways should be targeted in more exhaustive and quantitative investigations.

design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: This study was supported by a limited study grant by Astella's Pharmaceuticals (UK) not exceeding 60000.00GBP to MTPRP, MAS and DFM for phase II study investigating biomarkers of poor allograft function following human liver transplantation. Astella's Pharmaceuticals (UK) did not in any way have the access to the data not involved in any stage of the scientific data analysis or write up of this manuscript. The authors did not have any relationship with Astella's Pharmaceuticals (UK) with regards to employment, consultancy, patents, products in development or marketed products. Author MTPRP has been given travel assistance to participate in two scientific conferences by Astella's Pharmaceuticals (UK) not exceeding 5000.00GBP in the last three years declared under the Association of British Pharmaceutical Industry (ABPI) guidelines, and the data presented in this manuscript has not been presented in any of these meetings. This does not alter our adherence to PLOS ONE policies on sharing data and materials.

Introduction

Small (<1,000 Da) metabolites regulate cell signalling, cell-to-cell communication and energy transfer amongst other cellular processes, and are the first biochemicals to respond to internal or external stimuli. This responsiveness makes the metabolome an informative measure of the cell's dynamic state, a property that has led to a considerable and growing interest in the application of metabolomics in the health sciences. In parallel, metabolomics has begun to be used in clinical solid organ transplantation [1,2] where it holds considerable promise for the discovery of biomarkers to predict poor graft function or patient survival, as well as to elucidate the molecular mechanisms underlying pathophysiological processes such as during graft dysfunction, injury or rejection [3]. Beginning with early studies using ¹H-NMR spectroscopy, the applications of metabolomics in transplantation have expanded in more recent publications to using mass spectrometry based profiling of poor allograft function [4]. Case studies using NMR spectroscopy identified six potential metabolic biomarkers that were distinctive of a non-functional liver [5,6]. Meanwhile a metabolomics assessment in cirrhotic patients by ¹H-NMR spectroscopy linked elevated levels of high-density lipoproteins and phosphocholine with mild chronic liver failure whereas elevated levels of lactate, pyruvate, glucose and creatinine were associated with severe chronic liver failure (CLF). This study provided new insights to hepatic functional impairment in cirrhosis as well as showed an alternative approach to evaluate the severity of CLF, a key aspect in therapeutic decision making [7]. Furthermore, analyses of human bile during liver transplantation by capillary electrophoresis showed distinct metabolic fingerprints in donors and recipients [8]. Whereas more recent publication has been focussed on much larger group of liver grafts belonging to two distinct organ donor sources, namely the cadaveric donation after circulatory death (DCD), and donation after brain death (DBD) and applied lipidomics studies identify markers of early allograft dysfunction [9].

While liver transplantation is well established as the treatment of choice for many indications, the ever growing number of patients listed for transplantation has outweighed the supply of cadaveric organs, leading to greater disparity between supply and demand. As a result, most transplant programmes turned to using marginal organs, in particular liver grafts obtained by donation after circulatory death (DCD), to supplement more traditional donation after brain death (DBD) [10]. Despite rigorous donor selection criteria and best efforts to match a DCD graft with an optimal recipient, the results of the DCD transplantations remain inferior to the standard DBD transplantations and include increased occurrence of primary non-function (PNF) and biliary complications in the immediate post-operative period as well as inferior long-term graft survival rates [11–13]. These complications have been initially associated with donor warm ischaemia (Greek; *isch*—restriction, *hema*—blood), typical of DCD grafts during the surgical withdrawal phase. This phase is of variable and unpredictable duration which each DCD donor undergoes once the life support therapy is halted to allow natural passage to circulatory death [14]. Furthermore, grafts obtained from deceased donors are typically stored on ice (at a temperature between 0–4°C) before these organs are transplanted to the recipient, and this phase of cold storage further aggravates graft injury due to ischaemia [15]. Once the blood supply is reconnected within the recipient (both DCD and DBD) to the transplanted organs (termed reperfusion), further organ injury occurs through a process known as ischaemia-reperfusion or preservation-reperfusion injury. In this multifactorial process the reactive oxygen species generated in the organ during warm ischaemia initiate a cellular cascade leading to inflammation, and in severe cases to organ failure termed as PNF [16,17].

Identifying the metabolic differences between DCD and DBD liver grafts could significantly improve current clinical practise by defining biomarkers that are predictive of poor graft

function prior to transplantation. Selection of grafts from donors that exhibit such metabolic biomarkers could assist in clinical decision making and the exclusion of those organs from transplantation, thereby preventing the adverse clinical sequelae after transplantation. This would also help those transplant programs that are reliant on cadaveric donor organs for transplantation by preventing the necessity to perform re-transplant operations on those who had failed liver grafts, minimising the burden and demand for organs. Furthermore, identifying the metabolic differences between DCD and DBD liver grafts could help to identify the metabolic modifications of livers prior to and after the organ procurement from the donor that would improve the organ quality, an approach called metabolic therapy. This approach is justified by the nature of current organ donation practice. In the United Kingdom alone there has been a steep rise in DCD donations in recent years, however only 27% of liver grafts from these donors are used in clinical transplantation [18] with approximately 8% of these being excluded due to the high risk of PNF. In addition, the majority of grafts are not even procured due to increased time elapsed between the treatment withdrawal phase and circulatory death, which is beyond the currently accepted criteria of donor warm ischaemia time (30 minutes). Relating the metabolic profiles of DCD and DBD liver grafts to the outcome of the transplantation could supplement and expand the traditional methods to predict organ function, in particular early on during the transplantation procedure.

Previously we reported a pilot study that demonstrated the potential of Fourier transform ion cyclotron resonance (FT-ICR) mass spectrometry to detect a few thousand metabolic features (or peaks) in biopsies obtained from liver grafts in the cold and post-reperfusion phases of orthotopic liver transplantation (OLT)[19]. We observed and characterized changes in multiple metabolic pathways showing a rapid resumption of biochemical function within the grafts following reperfusion [19]. Here, we expand considerably upon this initial investigation, specifically with the aim to investigate and characterize the metabolic differences between DCD and DBD liver grafts at two key phases of the liver transplantation, the cold storage phase (T_1) and post-reperfusion phase (T_2). We seek to reveal the underlying metabolic pathways associated with the clinical observation of reduced success of the DCD grafts.

Materials and Methods

This study was approved by the South Birmingham Regional and National Research Ethics committee. The study was aimed at discovery of biomarker differences in the liver grafts used in the clinical orthotopic liver (OLT) setting with possibility of identifying those that may predict poor graft function and primary non-function. Considering the inferior clinical outcomes reported from DCD liver transplantation, we specifically focussed on identifying whether metabolite features differed between DCD and DBD liver grafts. Biopsies were obtained while the grafts were in an ice bath after a variable period of cold ischaemia had elapsed, and again in the post reperfusion phase. In a limited number of patients biopsies were also obtained in the pre-donation setting. All recipients involved were adequately informed and signed a consent form.

Clinical data

Overall, a total of 37 (DBD; $n = 27$ and DCD; $n = 10$) liver grafts were studied and the clinical courses of the recipients were followed. In DBD donors, the brain stem death criteria had been confirmed prior to the donor operation, which involved dissection and isolation of graft blood vessels while the circulatory function was intact. In DCD donors ($n = 10$), death was confirmed according to the Institute of Medicine guidelines after obligatory 5 minutes standoff time from the circulatory arrest, after which the donor operation was performed as rapidly as possible

[14]. The technical aspects of the organ procurement were otherwise similar. Both in the DBD and DCD procedures, grafts were perfused with Hyper-Osmolar Citrate (HOS, via cannula inserted to the aorta) and University of Wisconsin (UW, via portal vein) preservation fluids. Following donor operations, the liver grafts were packed on ice and transported to the location of the recipient operation, where the grafts were prepared for implantation while immersed in an ice bath. A wide spectrum of procedure related parameters as well as patient demographics and outcome data were recorded for comparison. These included cold ischaemia time (CIT) elapsed prior to the bench biopsy, overall CIT, donor warm ischaemia time (dWIT) in DCD, immediate post operative outcomes, acute physiological status of the recipient while in the intensive care unit, basic liver graft functions, episodes of graft rejection, graft failures and survival outcomes.

Patients underwent OLT for a variety of indications including alcoholic liver disease, primary biliary cirrhosis, primary sclerosing cholangitis, viral hepatitis (HBV/HCV), non-alcoholic steatohepatitis, polycystic disease, Cryptogenic/Autoimmune and Wilson's disease. The median age of recipient was 53 and 56 years for DCD and DBD respectively. The median (range) model for end-stage liver disease scores of DCD and DBD graft recipients were 12 (8–22) and 16 (6–22) respectively (Table 1).

Liver samples were obtained by Menghini biopsy needle [11520–19, 19swg (1.0mm) x 70mm; Dixons Surgical Instruments Ltd, Wickford, Essex, UK] for all 37 liver grafts at two stages of OLT: T_1 –after organ retrieval and transportation to the implanting centre while the liver was prepared on the bench while immersed in an ice bath at a temperature of 0–4°C T_1 and T_2 (post reperfusion biopsy) once the graft had been reconnected and the patient haemodynamics stabilised, and usually towards the end of the recipient procedure, after warm ischaemic period and reperfusion injury. In addition, for six DBD grafts an additional biopsy was taken at T_0 , during the donor surgical phase while the liver was still in the donor's body cavity. There were 80 liver allograft biopsies and these were snap frozen in liquid nitrogen and stored at -80°C until sample preparation for the direct infusion FT-ICR mass spectrometry based metabolomics.

Direct infusion FT-ICR mass spectrometry based metabolomics

Samples were prepared for the metabolomics analysis as described previously [19]. Briefly, biopsies were extracted using a methanol:chloroform:water method, separating the extracts into polar and non-polar fractions [20]. In total, 80 samples liver biopsy samples were extracted and from these one quality control (QC) sample was prepared by pooling a fraction of each of the 80 extracts (which was then aliquoted into 11 identical fractions). The polar metabolite fraction of each sample was analysed by USA; LTQ FT Ultra) from m/z 70 to 590, in positive ion mode, using the SIM-stitching approach [21]. Each sample was analysed in triplicate. To minimise false positive metabolites in the data matrix (due to noise), only peaks present in at least 2 of the 3 replicate measurements of each sample were retained, and then only peaks present in at least 75% of all the samples were retained for further analysis [22]. This data processing also served to exclude any peaks in the mass spectra that arose from the drugs that were known to be administered to the donors and recipients. The final data matrix consisted of 1260 reproducibly detected peaks (rows) and 91 variables (80 biopsies and 11 quality control samples; columns). The matrix contained 9.29% of missing data which was imputed using a weighted k-nearest neighbours algorithm ($k = 5$) [23]. Data were then normalized using the probabilistic quotient method [24] and subjected to a generalised log transformation (prior multivariate analysis) to stabilise the technical variance across the peaks and hence to avoid the highest abundance peaks from dominating the multivariate analysis [25]. Putative

Table 1. Donor and recipient demographics and surgical data.

	DBD (n = 27)	DCD (n = 10)	p-value
Donor gender	•	•	0.440
• Male	• 11 (41%)	• 2 (20%)	
• Female	• 16 (59%)	• 8 (80%)	
Donor age	50.0 (14.2)	49.7 (12.8)	0.948
Donor BMI	26.0 (5.4)	25.4 (3.0)	0.738
Donor cause of death	• 18 (67%)	• 4 (40%)	0.294
• ICH	• 3 (11%)	• 1 (10%)	
• Head Injury	• 1 (3%)	• 2 (20%)	
• Cardiac Arrest	• 5 (19%)	• 3 (30%)	
• Other			
Graft CIT	484.5 (143.6)	461.0 (117)	0.646
dWIT	-	19.9 (5.7)	
Graft microsteatosis	• None (4)	• None (2)	
	• Mild (22)	• Mild (8)	
	• Moderate (1)		
Graft macrosteatosis	• None (2)	• None (1)	
	• Mild (19)	• Mild (6)	
	• Moderate (3)	• Severe (3)	
	• Severe (3)		
Recipient gender	• 17 (63%)	• 5 (50%)	0.708
• Male	• 10 (37%)	• 5 (50%)	
• Female			
Recipient age	53.7 (9.1)	52.9 (7.3)	0.812
Recipient aetiology	• 9 (33%)	• 5 (50%)	0.681
• ALD	• 4 (15%)	• 2 (20%)	
• HCV	• 2 (7%)	• 1 (10%)	
• PSC	• 4 (15%)	• 1 (10%)	
• PBC	• 2 (7%)	• 1 (10%)	
• PCLD	• 6 (23%)	• 0	
• Other			
Presence of HCC	3 (11%)	3 (30%)	0.166
Recipient MELD	16.2 (5.1)	12.3 (4.1)	0.036
Graft Implantation time (minutes)	46.9 (15.9)	40.7 (15.8)	0.296
Operating time (minutes)	276.7 (71.7)	320.2 (98.3)	0.148
Peak AST	1783.5 (2088.2)	3374.7 (2641.3)	0.064
Days on ITU	2.5 (3.0)	5.4 (9.5)	0.160
Length of hospital stay (days)	13.9 (8.5)	14.5 (5.5)	0.825

Values expressed as mean (standard deviation) or number (percentage) as appropriate. Bold values indicate a significant difference ($p < 0.05$) between donor types by ANOVA, t-test or Chi-square as appropriate. ALD = Alcoholic liver disease; AST = aspartate aminotransferase; BMI = Body mass index; CIT = Cold ischaemic time; DBD = Donation after brain death; DCD = Donation after circulatory death; HBV = Hepatitis B; HCC = Hepatocellular carcinoma; HCV = Hepatitis C; ICH = Intracranial haemorrhage; MELD = Model for end-stage liver disease; PBC = Primary biliary cirrhosis; PCLD = Polycystic liver disease; PSC = Primary sclerosis cholangitis; dWIT = Donor warm ischaemic time (DCD only)

doi:10.1371/journal.pone.0165884.t001

metabolite names were assigned to the peaks based on their mass-to-charge ratio and taking into account commonly detected ions forms, including $[M-e]^+$, $[M+H]^+$, $[M+Na]^+$, $[M+^{39}K]^+$, $[M+^{41}K]^+$, $[M+2Na-H]^+$, $[M+2^{39}K-H]^+$ and $[M+NH_4]^+$.

Statistical analyses

Potential clinical differences between DBD and DCD grafts were evaluated by testing each OLT variable, including cold ischaemia time, warm ischaemia time, hours in ITU, number of days-in hospital following OLT, peak aspartate transaminase (AST), and incidence of primary

non-function. Non-parametric 2-sample Wilcoxon rank-sum test was applied for continuous numerical variables (e.g., CIT) and Fisher's exact tests were used for binary variables (e.g., the occurrence of primary non-function). The obtained *p* values were adjusted for multiple hypothesis testing using the Benjamini and Hochberg method to control the false discovery rate [26].

Principal component analysis (PCA) was used to represent the multivariate FT-ICR mass spectral metabolomics data in 2-dimensional space in terms of principal components PC1 and PC2. Univariate statistical analysis, on a per peak basis, was used to discover if any metabolites differed significantly a) between donor (T_0), cold (T_1) and post-reperfusion (T_2) phases across all patients, and b) between DCD and DBD grafts in the cold phase (T_1) and, separately, post-reperfusion (T_2) phase. Here, the Anderson-Darling test was used to evaluate normality assumptions, and since ca. 40% of the peaks did not follow a normal distribution, non-parametric statistical methods were used, specifically a two-sample Wilcoxon rank-sum test (DCD and DBD comparison) and its extension to more groups, Kruskal-Wallis one-way analysis of variance (T_0 , T_1 and T_2 comparison), both with a Benjamini and Hochberg false discovery correction. In addition, Gain Scores Analysis (Kruskal-Wallis on gain scores) was used to discover those metabolites that changed in a significantly different manner from cold-phase to post-reperfusion between the DCD and DBD grafts. FDR of 5% was used as cut-off values to identify statistically significant features. All statistical analyses were carried out using R version 3.0.2, a free programming language and software for statistical computing and graphics. Putatively identified metabolites were assigned to KEGG metabolic pathways, as defined in the KEGG database [27,28].

Results and Discussion

Clinical outcomes

The liver transplantation procedures were carried out in a similar manner for DBD and DCD grafts and we did not notice any significant differences neither at the procedure level nor with the short-term outcomes. The mean CIT was 484.52 ± 143.59 minutes (DBD) and 461.00 ± 116.97 minutes (DCD), whereas the mean implantation time, when grafts were exposed to further warm ischaemia until the circulation was restored, was 41.85 ± 6.94 minutes (DBD) and 41.80 ± 9.77 minutes (DCD). The recipients spent on average 95.04 ± 120.80 (DBD) and 123.22 ± 174.48 (DCD) hours in the intensive care unit. The majority of OLTs were successful; three patients in the entire study group had perioperative mortality ($n = 2$ in the DBD group). The causes of death were related to PNF in two patients (one patient each in the DCD and DBD groups) and related to hepatic artery thrombosis in the third patient (DBD) (Table 1).

Changes in hepatic metabolism during transplantation

FT-ICR mass spectra of the extracted biopsies contained 1260 reproducibly detected peaks of which 448 (35.56%) were putatively annotated based upon accurate mass measurements and the Kyoto Encyclopaedia of Genes and Genomes database (S1 Table). Principal component analysis verified the high technical reproducibility of the mass spectra, evidenced by the clustering of the measurements of the QC sample on the PCA scores plot (Fig 1). Furthermore, the PCA scores showed a clear separation between the biopsies from the cold phase (T_1) and post reperfusion (T_2). The clustering of the donor biopsies (T_0 ; obtained from six DBD grafts while the organs were still perfused with warm circulation) close to the post-reperfusion biopsies (T_2), with both groups having very distinctive metabolic profiles compared to the biopsies originating from the cold phase sampling is a striking result (Fig 1). This signifies the

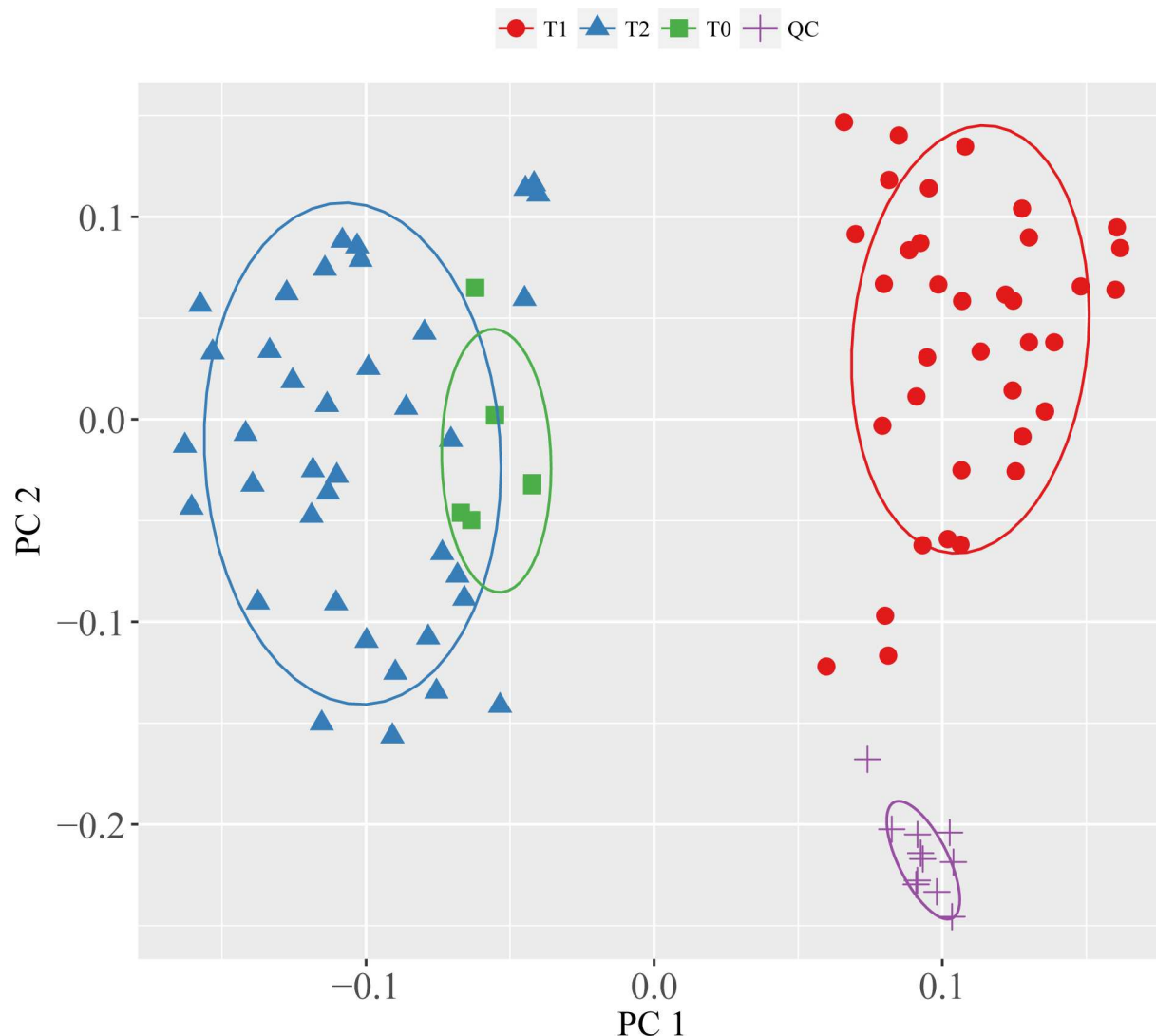


Fig 1. Principal component analysis scores plot showing the similarities and differences between the metabolic profiles of the grafts in donor (T_0), cold (T_1) and post-reperfusion phases (T_2). The variance explained by PC1 = 39.82% and PC2 = 29.58. The close grouping of the donor and post-reperfusion biopsies along with their clear separation from the cold-phase biopsies (along PC1) is further supportive of the rapid resumption of the biochemical functions in the reperfused grafts and shows the direction of metabolic changes through the patient journey and the OLT procedure—from ‘healthy’ donor grafts through cold-phase to almost fully functional grafts post reperfusion.

doi:10.1371/journal.pone.0165884.g001

distinctively different metabolism of hepatocytes in the perfused state compared to those in cold storage. This metabolic separation was confirmed by univariate testing that detected 688 (54.60%) significantly different peaks between T_1 and T_2 , 293 peaks (23.25%) between T_0 and T_1 , and only 124 peaks (9.84%) between T_0 and T_2 (Kruskal-Wallis test, $p < 0.05$) (Table 2). In our previous proof-of-principle OLT study, we identified a plethora of metabolic responses in the post-reperfused grafts compared to their cold-phase state and concluded that these changes reflected the rapid resumption of the biochemical functions of hepatocytes following reperfusion, including increased urea production, bile acid synthesis and clearance of the preservation solution. Here, in addition to verifying these expected metabolic responses, we observed additional key metabolic changes including, amongst others, putatively annotated essential

Table 2. Top putatively annotated metabolic fold-changes (FC) in the liver grafts between donor phase (T₀), cold phase (T₁) and post-reperfusion phase (T₂), considering the DBD and DCD biopsies as one group. The average absolute ppm error was 0.3631, range: 0.0029–0.9828.

Putative metabolite	m/z (observed)	Empirical formula	Ion	FC: T ₂ /T ₁	FC: T ₂ /T ₀	FC: T ₁ /T ₀	Significance	Univariate rank	PC1 rank
Histidine*	178.05876	C ₆ H ₉ N ₃ O ₂	Na, H, ³⁹ K	3.88	1.43	0.37	T ₁ vs. T ₂ ; T ₁ vs. T ₂	1, 117, 181	1, 130, 198
Malate	157.01079	C ₄ H ₆ O ₅	Na, 2K-H	4.61	1.64	0.36	T ₁ vs. T ₀ ; T ₁ vs. T ₂	3, 259	58, 116
Glutamate*	170.04244	C ₅ H ₉ NO ₄	Na, 2Na-H	5.14	1.69	0.33	T ₁ vs. T ₀ ; T ₁ vs. T ₂	13, 37	15, 20
Serine	128.03181	C ₃ H ₇ NO ₃	Na, 2Na-H, K (39), H	2.95	1.77	0.60	T ₁ vs. T ₀ ; T ₁ vs. T ₂	15, 19, 143, 203	101, 105, 111, 205
Glutamine	169.05842	C ₅ H ₁₀ N ₂ O ₃	Na, ³⁹ K, 2K-H	4.51	1.41	0.31	T ₁ vs. T ₀ ; T ₁ vs. T ₂	16, 161, 233	12, 128, 172
N-Acetyl-L-glutamate	212.05305	C ₇ H ₁₁ NO ₅	Na	3.60	2.81	0.78	T ₁ vs. T ₂ ; T ₀ vs. T ₂	18	30
O-Phospho-L-serine	207.99835	C ₃ H ₈ NO ₆ P	Na, H, ³⁹ K	6.14	1.37	0.22	T ₁ vs. T ₀ ; T ₁ vs. T ₂	20, 84, 146	23, 56, 152
Tyrosine	204.06321	C ₉ H ₁₁ NO ₃	Na, H	3.46	1.32	0.38	T ₁ vs. T ₀ ; T ₁ vs. T ₂	23, 175	25, 90
ATOP*	472.00083	C ₁₀ H ₁₅ N ₅ O ₁₀ P ₂	2Na-H, Na	5.46	1.11	0.20	T ₁ vs. T ₀ ; T ₁ vs. T ₂	26, 51	45, 84
CTOP-choline	489.11491	C ₁₄ H ₂₆ N ₄ O ₁₁ P ₂	H	0.42	1.16	2.73	T ₁ vs. T ₀ ; T ₁ vs. T ₂	49	41
Mannitol*	223.04042	C ₆ H ₁₄ O ₆	⁴¹ K	0.03	1.34	46.97	T ₁ vs. T ₀ ; T ₁ vs. T ₂	61	4
Taurine	148.00390	C ₂ H ₇ NO ₃ S	Na	2.11	1.15	0.55	T ₁ vs. T ₀ ; T ₁ vs. T ₂	88	109
Citrate	193.03435	C ₆ H ₈ O ₇	H	0.20	2.44	12.14	T ₁ vs. T ₀ ; T ₁ vs. T ₂	97	92
Threonine	142.04748	C ₄ H ₉ NO ₃	Na	2.76	1.37	0.50	T ₁ vs. T ₂	110	66
T2oline	138.05256	C ₅ H ₉ NO ₂	Na, H	2.40	2.75	1.14	T ₁ vs. T ₂ ; T ₀ vs. T ₂	115, 221	235, 331
GMP	386.04730	C ₁₀ H ₁₄ N ₅ O ₈ P	Na	3.03	0.98	0.32	T ₁ vs. T ₀ ; T ₁ vs. T ₂	140	147
Glucose	221.02478	C ₆ H ₁₂ O ₆	⁴¹ K	0.38	0.85	2.25	T ₁ vs. T ₀ ; T ₁ vs. T ₂	153	114
Glycocholate*	488.29844	C ₂₆ H ₄₃ NO ₆	Na	4.49	3.27	0.73	T ₁ vs. T ₂	154	170
Succinate	141.01584	C ₄ H ₆ O ₄	Na	0.49	1.06	2.17	T ₁ vs. T ₀ ; T ₁ vs. T ₂	185	106
Valine	140.06820	C ₅ H ₁₁ NO ₂	Na, ³⁹ K, H	2.63	1.44	0.55	T ₁ vs. T ₂	186, 218, 253	260, 272, 281
Choline	145.06889	C ₅ H ₁₄ NO	⁴¹ K	2.01	2.73	1.36	T ₁ vs. T ₂ ; T ₀ vs. T ₂	199	377
Formate	90.97661	CH ₂ O ₂	2Na-H	5.37	3.47	0.65	T ₁ vs. T ₂	205	582
O-Phospho-L-homoserine	200.03205	C ₄ H ₁₀ NO ₆ P	H	2.87	1.35	0.47	T ₁ vs. T ₂	209	83
Kynurenine	209.09221	C ₁₀ H ₁₂ N ₂ O ₃	H	1.78	2.76	1.55	T ₁ vs. T ₂ ; T ₀ vs. T ₂	232	338
Aspartate	134.04479	C ₄ H ₇ NO ₄	H	2.56	1.17	0.46	T ₁ vs. T ₀ ; T ₁ vs. T ₂	243	139
Urea*	98.99550	CH ₄ N ₂ O	³⁹ K	2.48	1.31	0.53	T ₁ vs. T ₂	268	308

*, metabolic changes observed, verifying those reported in our proof-of-principle study[19]

doi:10.1371/journal.pone.0165884.t002

(threonine and valine) and non-essential (tyrosine, serine and proline) amino acids, taurine (a major constituent of bile), and kynurenine (a central compound in the tryptophan metabolism pathway).

Hepatic metabolism in DBD compared to DCD grafts

We identified a small subset of peaks that distinguished the DCD and DBD grafts at the metabolic level. In particular, we detected 50 peaks including 11 putatively annotated compounds that differed between DCD and DBD in the cold phase (T_1), 64 peaks (10 putatively annotated) that differed between DCD and DBD following reperfusion (T_2), and 72 peaks (10 putatively annotated) that changed from T_1 to T_2 in a significantly different manner between DCD and DBD grafts (Table 3). The PCA scores plots, based only on these sub-selections of peaks, provide a visualisation of the clear separation of the DCD vs. DBD grafts along PC1 for the cold-phase biopsies and almost as clear separation for the post-reperfusion biopsies (Fig 2)

The key metabolic differences between DBD and DCD grafts in the cold phase (T_1) included increased levels (in DCD) of the putatively annotated metabolites tryptophan,

Table 3. Top putatively annotated metabolic fold-changes between the DCD and DBD grafts: combined results for the comparison, (i) in the cold phase (T_1), (ii) following reperfusion (T_2), and (iii) in response from going from T_1 to T_2 . The average absolute ppm error was 0.2758, range: 0.0069–0.6835.

Putative metabolite	m/z (observed)	Empirical formula	Ion	Fold-change DCD/DBD ¹	Univariate Rank ²
Tryptophan	205.09725	C ₁₁ H ₁₂ N ₂ O ₂	H	1.88 ^{T₁} , 1.10 ^{T₂}	3 ^{T₁} , 28 ^G
Adenylosuccinate	464.08155	C ₁₄ H ₁₈ N ₅ O ₁₁ P	H	0.40 ^{T₁}	6 ^{T₁}
GMP	402.02124	C ₁₀ H ₁₄ N ₅ O ₈ P	³⁹ K	0.47 ^{T₁}	15 ^{T₁}
Malate	210.94066	C ₄ H ₆ O ₅	2K-H	0.44 ^{T₁}	18 ^{T₁}
ADP	465.99284	C ₁₀ H ₁₅ N ₅ O ₁₀ P ₂	³⁹ K	0.41 ^{T₁}	20 ^{T₁}
D-Glucose	203.05261	C ₆ H ₁₂ O ₆	Na	1.72 ^{T₁} , 1.22 ^{T₂}	22 ^{T₁} , 9 ^G
ADP	450.01895	C ₁₀ H ₁₅ N ₅ O ₁₀ P ₂	Na	0.53 ^{T₁}	25 ^{T₁}
O-Acetyl-L-carnitine	204.12305	C ₉ H ₁₈ NO ₄	-e	0.45 ^{T₁}	28 ^{T₁}
Kynurenine	209.09221	C ₁₀ H ₁₂ N ₂ O ₃	H	1.80 ^{T₁}	43 ^{T₁}
Leucine	132.10190	C ₆ H ₁₃ NO ₂	H	1.49 ^{T₁}	47 ^{T₁}
Pantothenate	220.11806	C ₉ H ₁₇ NO ₅	H	0.60 ^{T₁}	49 ^{T₁}
Glutathione	352.05497	C ₁₀ H ₁₇ N ₃ O ₆ S	2Na-H	1.13 ^{T₁} , 1.88 ^{T₂}	3 ^{T₂} , 38 ^G
Threonine	142.04748	C ₄ H ₉ NO ₃	Na	1.37 ^{T₁} , 2.07 ^{T₂}	4 ^{T₂} , 70 ^G
Leucine	154.08388	C ₆ H ₁₃ NO ₂	Na	1.37 ^{T₂}	16 ^{T₂}
Glutamate	170.04244	C ₅ H ₉ NO ₄	Na	0.99 ^{T₁} , 1.61 ^{T₂}	25 ^{T₂} , 72 ^G
Creatine	154.05872	C ₄ H ₉ N ₃ O ₂	Na	1.45 ^{T₂}	32 ^{T₂}
Glutamate	192.02440	C ₅ H ₉ NO ₄	2Na-H	1.01 ^{T₁} , 2.78 ^{T₂}	38 ^{T₂} , 45 ^G
Threonine	120.06551	C ₄ H ₉ NO ₃	H	1.37 ^{T₂}	41 ^{T₂}
T2oline	138.05256	C ₅ H ₉ NO ₂	Na	1.59 ^{T₂}	53 ^{T₂}
Pantothenate	220.11806	C ₉ H ₁₇ NO ₅	H	0.79 ^{T₂}	62 ^{T₂}
Leucine	132.10190	C ₆ H ₁₃ NO ₂	H	1.32 ^{T₂}	64 ^{T₂}
Ornithine	133.09716	C ₅ H ₁₂ N ₂ O ₂	H	1.98 ^{T₁} , 1.05 ^{T₂}	42 ^G
Serine	150.01378	C ₃ H ₇ NO ₃	2Na-H	0.78 ^{T₁} , 3.01 ^{T₂}	54 ^G
SAM	399.14460	C ₁₅ H ₂₂ N ₆ O ₅ S	H	1.23 ^{T₁} , 0.68 ^{T₂}	51 ^G
Glucose	221.02478	C ₆ H ₁₂ O ₆	⁴¹ K	1.40 ^{T₁} , 1.05 ^{T₂}	59 ^G

¹ Fold-change calculated for the corresponding phase, cold phase (T_1) or post reperfusion (T_2)

² Ranking carried out separately for the three comparisons: in the cold phase (T_1), post-reperfusion (T_2) or based on the Gain Scores Analysis (G): capturing the metabolic responses between DCD and DBD grafts from the T_1 to T_2

doi:10.1371/journal.pone.0165884.t003

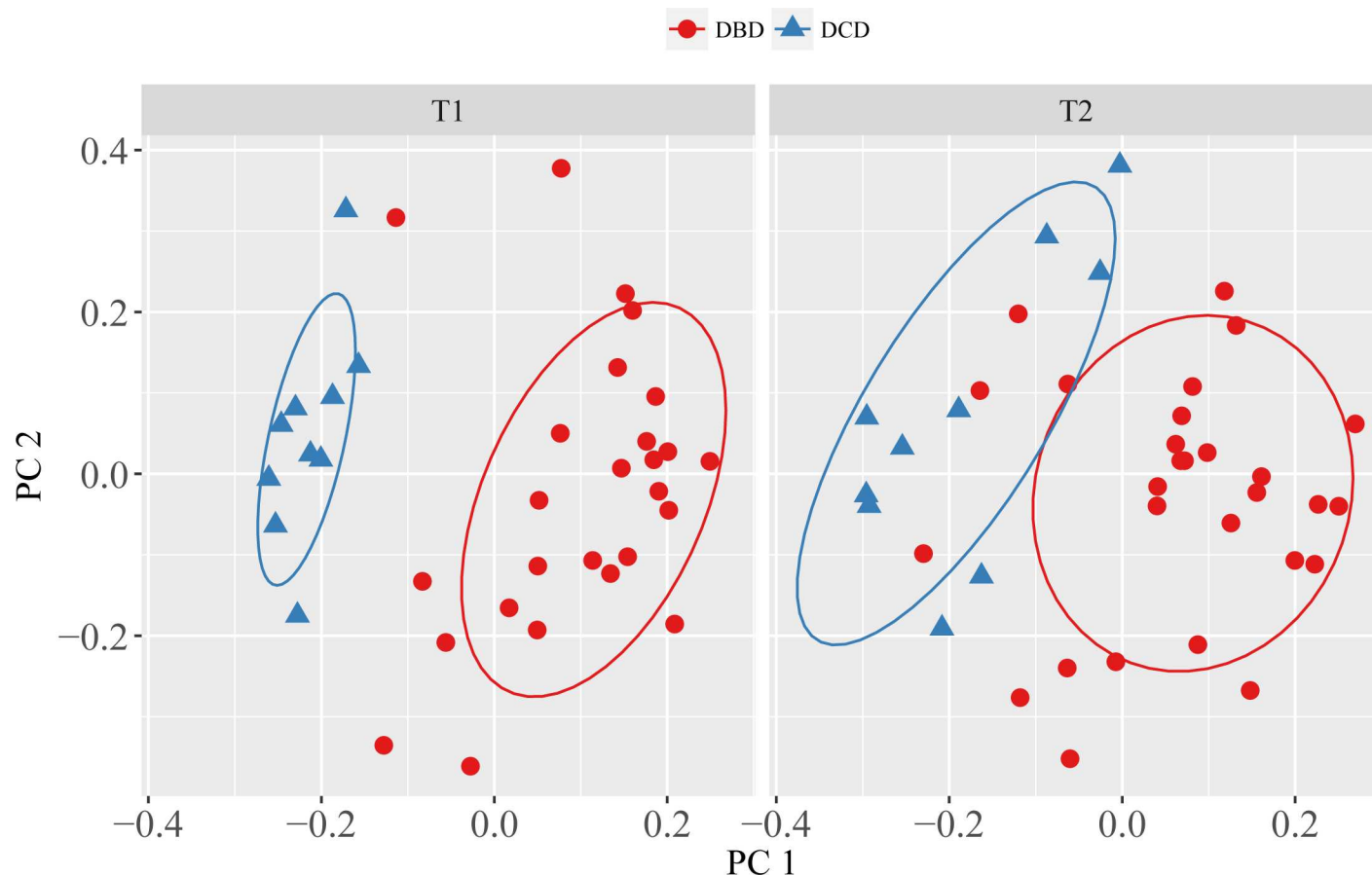


Fig 2. Principal component analysis scores plots highlighting the metabolic separation of the DBD and DCD grafts in the cold phase (T₁) and separately the post-reperfusion phase (T₂), based on analyses of just the 50 and 64 peaks identified as being significantly different (between DBD and DCD) for the T₁ and T₂ groups, respectively. Variance explained for T₁, PC1 = 36.75% and PC2 = 19.39% and for T₂, PC1 = 25.22% and PC2 = 17.41%.

doi:10.1371/journal.pone.0165884.g002

kynurenine, glucose and leucine and decreased levels of adenylosuccinate, GMP, ADP, malate, O-acetyl carnitine and pantothenate (Table 3). The observed putative metabolites are involved mainly in tryptophan metabolism, purine metabolism, oxidative phosphorylation and a set of carbohydrate metabolic pathways, including the TCA cycle, pyruvate metabolism, glycolysis/gluconeogenesis and the pentose phosphate pathway (Table 4). Among these findings, tryptophan and its metabolism have received earlier attention in the liver transplantation field. Tryptophan is an essential amino acid that, amongst other roles, serves as a precursor of the neurotransmitter serotonin and vitamin B₃. Histidine-tryptophan-ketoglutarate (HTK) solution, which contains tryptophan to prevent membrane injury, was proposed as an alternative liver preservation solution to the current gold standard, University of Wisconsin solution. Interestingly the organ preservation solutions used in this study cohort (UW solution and Hyper-osmolar citrate/Marshall's solution) do not have added tryptophan. The systematic review to compare the efficacy and safety of these two solutions did not show overall significant differences, yet in some cases HTK was believed to perform better, especially in terms of biliary tract flush and prevention of biliary complications [29]. Furthermore, tryptophan can be catabolised either via the kynurenine or serotonin pathways, and hence kynurenine was studied previously to investigate tryptophan metabolism in potential cirrhotic liver transplant

Table 4. Metabolic pathways discovered to differ significantly between DCD and DBD grafts, including the associated putatively annotated metabolites in those pathways.

Metabolic pathway	Putative metabolite	Description ¹
Tryptophan metabolism	Tryptophan, Kynurenine	Amino acid metabolism; T ₁ ^H
Purine metabolism	Adenylosuccinate, GMP, ADP	Nucleotide metabolism; T ₁ ^L
Oxidative phosphorylation	ADP	Energy metabolism; T ₁ ^L
TCA cycle	Malate	Carbohydrate metabolism; T ₁ ^L
Pyruvate metabolism	Malate	Carbohydrate metabolism; T ₁ ^L
Glycolysis / Gluconeogenesis	Glucose	Carbohydrate metabolism; T ₁ ^H
Pentose phosphate pathway	Glucose	Carbohydrate metabolism; T ₁ ^H
Alanine, aspartate and glutamate metabolism	Adenylosuccinate, Glutamate	Amino acid metabolism; T ₁ ^L , PR ^H
Glycine, serine and threonine metabolism	Tryptophan, Threonine, Creatine, Serine	Amino acid metabolism; T ₁ ^H , PR ^H
Cysteine and methionine metabolism	Glutathione, Serine, SAM	Amino acid metabolism; T ₂ ^H with exception of lower levels of SAM
Arginine and proline metabolism	Glutamate, Creatine, Proline, Ornithine, SAM	Amino acid metabolism; T ₂ ^H with exception of lower levels of SAM
Valine, leucine and isoleucine degradation & biosynthesis	Leucine, Threonine	Amino acid metabolism; T ₁ ^H , PR ^H
Glutathione metabolism	Glutathione, Glutamate, Ornithine	Metabolism of other amino acids; PR ^H
Taurine and hypotaurine metabolism	Glutamate	Metabolism of other amino acids; PR ^H
D-Glutamine and D-glutamate metabolism	Glutamate	Metabolism of other amino acids; PR ^H
Aminoacyl-tRNA biosynthesis	Tryptophan, Leucine, Threonine, Glutamate, Proline, Serine	Translation; T ₁ ^H , PR ^H
ABC transporters	Glucose, Leucine, Glutathione, Glutamate, Proline, Ornithine, Serine	Membrane transport; T ₁ ^H , PR ^H
Pantothenate and CoA biosynthesis	Pantothenate	Metabolism of cofactors and vitamins; T ₁ ^L , PR ^L
Vitamin digestion and absorption	Pantothenate	Digestive system; T ₁ ^L , PR ^L
Bile secretion	Glucose, Glutathione	Digestive system; T ₁ ^H , PR ^H

¹, H, higher levels and L, lower levels of putative metabolites in DCD in the corresponding OLT stage (T₁ or T₂)

doi:10.1371/journal.pone.0165884.t004

recipients. The pre-transplant serum levels of kynurenine as well as the kynurenine/tryptophan ratios were positively correlated with the disease severity, while serum levels of tryptophan and serotonin showed no correlation[30]. The significantly higher levels of tryptophan and kynurenine in DCD grafts (vs. DBD grafts) during the cold phase in our study appears to support the previous studies that identified tryptophan metabolism via kynurenine pathways as a key metabolic change in liver transplantation. Although it was not a key objective to analyse biomarkers related to primary non-function in the present study given the small sample size, the two failed allografts due to primary non-function both had abundantly higher levels of tryptophan and kynurenine (Fig 3).

The putatively annotated metabolites that significantly differed between DCD and DBD grafts following reperfusion (T₂) included increased levels (in DCD) of glutathione, threonine, leucine, glutamate, creatine, glutamate, proline and decreased levels of pantothenate (Table 2). In addition, four of these metabolites including glutathione, threonine, glutamate and glutamate were changed in a significantly different manner in DCD and DBD grafts while they were removed from cold storage (T₁) and following reperfusion (T₂). The remaining six metabolites identified as different in the Gain Scores Analyses included tryptophan and glucose (previously observed as significantly different in the cold-phase) as well as ornithine, serine, S-adenosyl methionine (SAM) and glucose (Table 2 and Fig 4). All of these putatively annotated metabolites are primarily involved in amino acid metabolism and translation

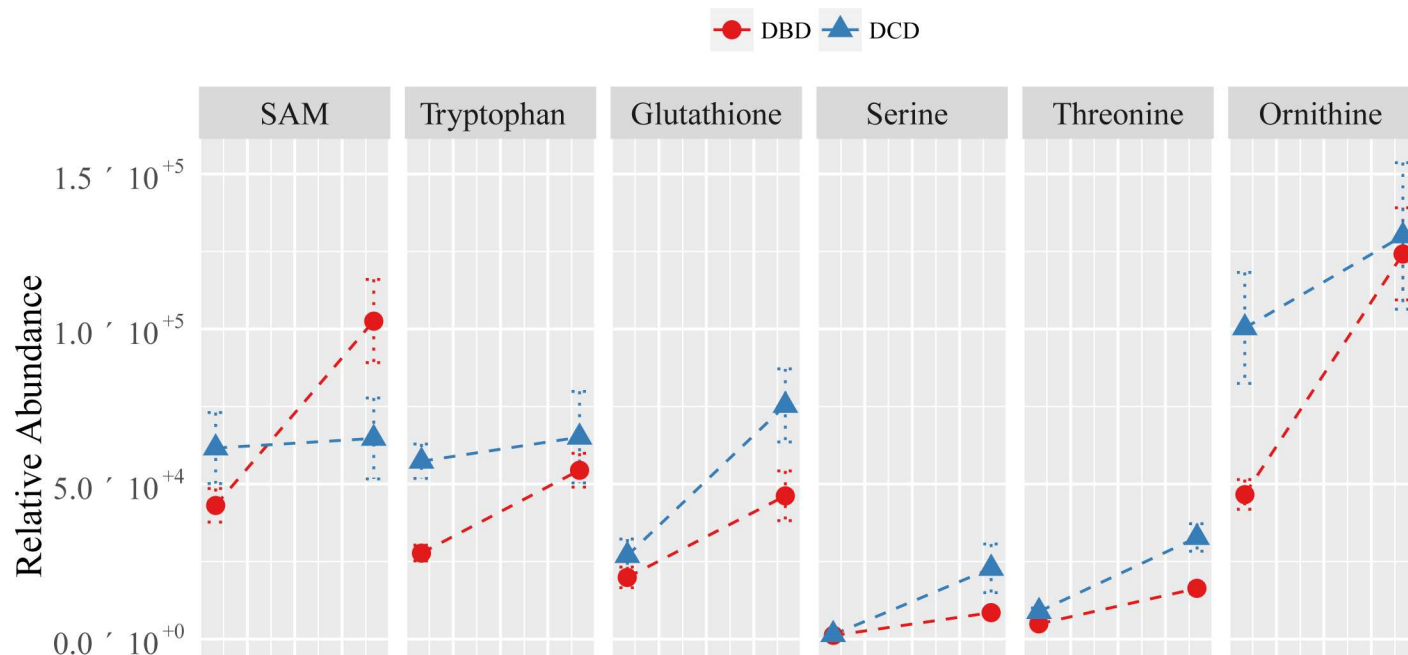


Fig 3. Selected top putatively identified metabolites changed in a significantly different way from the cold phase (T₁) to post-reperfusion (T₂) between DCD and DBD grafts. Gain Scores Analysis (Kruskal-Wallis on gain scores) was used to discover those metabolites that changed in a significantly different manner from cold-phase to post-reperfusion between the DCD and DBD grafts.

doi:10.1371/journal.pone.0165884.g003

(aminoacyl-tRNA biosynthesis) and to a lesser extent in metabolism of cofactors and vitamins and bile secretion (Table 3). Interestingly, in DBD grafts, there is a large increase in SAM between the cold phase and post-reperfusion, which does not occur in DCD grafts. While the biochemical implications of this lack of recovery are unknown, we hypothesise that this could affect methylation reactions including DNA methylation, given the importance of this metabolite as a methyl donor [31]. Further targeted investigations of the SAM (and related S-adenosylhomocysteine) metabolic pathway are therefore warranted.

The increased level of glutathione in DCD grafts, which occurs in the cold phase (1.13 times higher in DCD compared to DBD grafts) but is considerably more pronounced (1.88 times higher) following reperfusion, is also an intriguing finding. Glutathione, owing to its thiol group, is known as one of the most effective antioxidants preventing cellular damage caused by reactive oxygen species, as occurs in ischaemia/reperfusion injury. One of its precursors, acetylcysteine, has been studied as a protective molecule in the perioperative treatment of patients undergoing liver transplantation [32]. Our findings show that not only were glutathione levels increased but so were other intermediates of glutathione metabolism such as glutamate and ornithine, indicative of disruption to the glutathione pathway. While glutathione and glutamate shared similar responses at the post-reperfusion time point, i.e. a rapid increase of levels in DCD compared to DBD, ornithine levels were higher in the cold-phase and similar following reperfusion (Fig 4). This could be due to ornithine being used up for the biosynthesis of glutathione via the intermediate by-product of glutamate. These changes may explain the increased oxidative stress incurred by the DCD grafts, owing to the increased ischaemic injury through warm ischaemic damage.

Although metabolomics studies have caused initial enthusiasm in the transplant community as a potential investigative tool to differentiate poorly functioning grafts from those that perform well after transplantation certain limitations have halted the wider applicability. Only

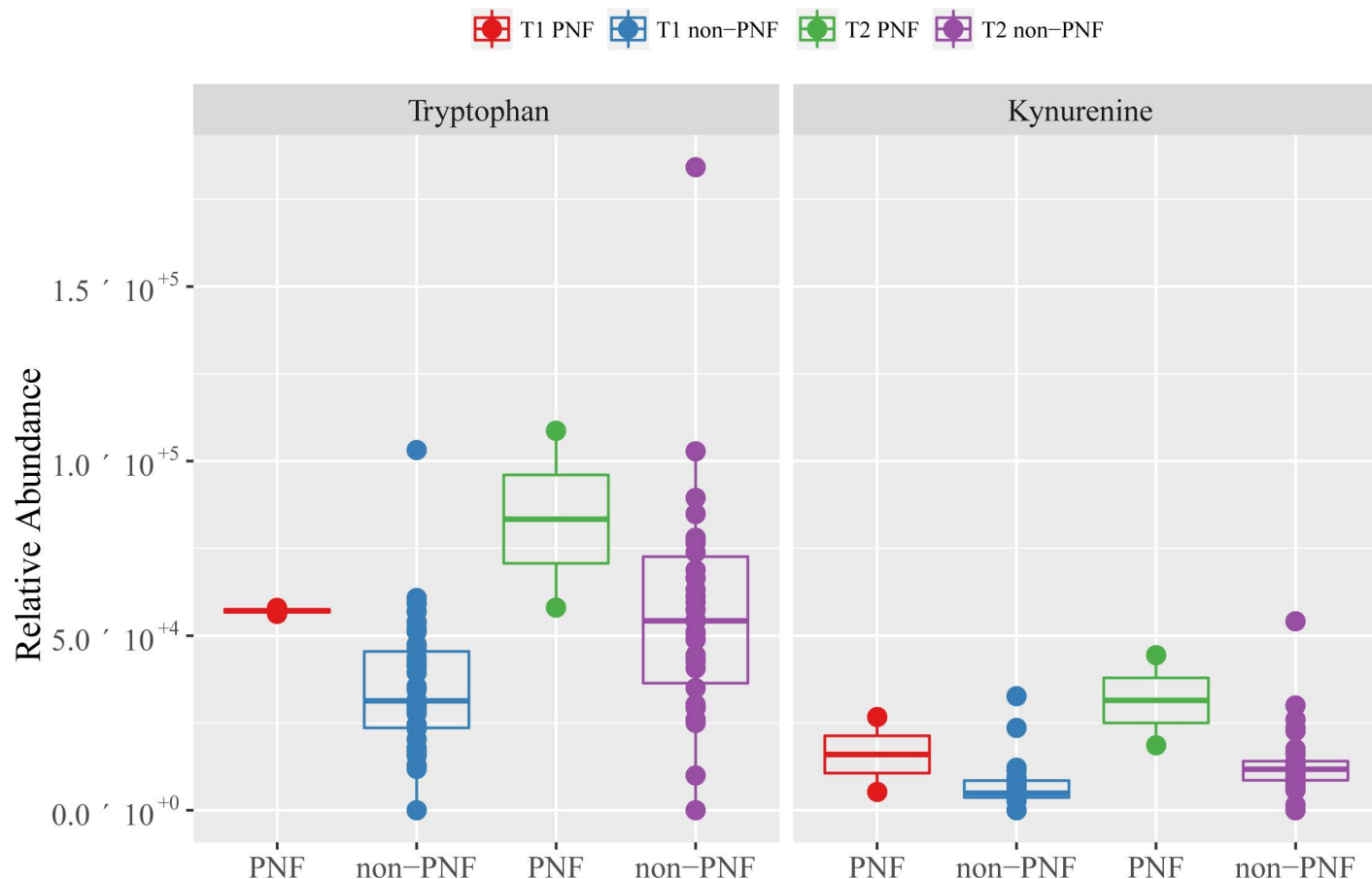


Fig 4. The differences between tryptophan and kynurenine in failed allografts due to Primary non-function/PNF (n = 2) vs. non-PNF (n = 36) in the cold phase and post reperfusion. The data show the relative abundances of the metabolites with 95% confidence intervals (statistics not applied due to limited sample size).

doi:10.1371/journal.pone.0165884.g004

one recent publication proposes a metabolic biosignature, based on combination of metabolites altered in liver allografts that perform unsatisfactorily following liver transplantation [4] and this by far the most explicit application of metabolomics in the field of solid organ transplantation. These metabolites responsible for early allograft dysfunction and graft failure include phospholipid metabolism, bile production, ammonia and urea cycles as well as glutathione metabolism. Interestingly some of the metabolites have also been identified in our study, the key metabolite changes related to tryptophan and kynurenine metabolism have not been identified that could be explained by differences in allograft types and analytical techniques involved.

Conclusions

Previous studies on larger cohorts (n > 300) of DBD and DCD liver transplantations provide evidence for the inferiority of the DCD donations, including increased incidence of primary non-function, biliary complications as well as lower graft- and patient survival [12,33]. Our study has for the first time identified, in un-targeted manner, key metabolic differences between DCD and DBD liver grafts. Since we did not observe any significant differences between the DCD and DBD procedures and outcomes in our study, we believe that these metabolic differences are reflective of the inherent molecular dissimilarities between DCD and

DBD grafts. Some of the identified metabolic alterations correlate with our current understanding of the physiological changes surrounding DCD organ donation, including an impact on glucose metabolism by donor warm ischaemia in DCD grafts. However, our observed changes to the tryptophan/kynurenine axis in the DCD grafts are novel findings. Both of these metabolites were observed at ca. 2-fold higher concentration in the DCD grafts (compared to DBD grafts) in the cold phase, suggesting the possibility that these metabolites are responsible for, or at least could be indicators of, the reported higher incidences of increased graft failures in DCD grafts in the literature. In fact, we observed increased levels of tryptophan/kynurenine in allografts with PNF in our study; however, due to few cases we were not able to verify statistically verify this association. Given the role of metabolomics as a hypothesis generating tool, and not to determine whether this metabolic pathway is indeed the cause of graft failure, we conclude that the subsequent clinical investigations of DCD versus DBD transplantations should employ a targeted analytical approach to robustly quantify the metabolites in the tryptophan/kynurenine pathway in the pursuit of more reliable biomarkers of graft function.

Supporting Information

S1 Table. Minimal dataset—FTICR mass spectrometry analysis.xlsx.
(XLSX)

Author Contributions

Conceptualization: MTPRP MAS NM DFM MRV.

Data curation: OH MTPRP.

Formal analysis: OH JK MRV.

Funding acquisition: MTPRP MAS DFM.

Investigation: OH JK MRV.

Methodology: MTPRP OH JK DAR MRV.

Project administration: MTPRP.

Resources: MTPRP MAS DAR NM DFM MRV.

Software: OH JK MRV.

Supervision: DFM MRV.

Validation: OH JK MRV.

Writing – original draft: OH MTPRP.

Writing – review & editing: MAS JK DAR NM DFM RL MRV.

References

1. Wishart DS (2005) Metabolomics: the principles and potential applications to transplantation. *American journal of transplantation* 5: 2814–2820. doi: [10.1111/j.1600-6143.2005.01119.x](https://doi.org/10.1111/j.1600-6143.2005.01119.x) PMID: [16302993](https://pubmed.ncbi.nlm.nih.gov/16302993/)
2. Bohra R, Klepacki J, Klawitter J, Klawitter J, Thurman JM, Christians U (2013) Proteomics and metabolomics in renal transplantation-quo vadis? *Transplant International* 26: 225–241. doi: [10.1111/tri.12003](https://doi.org/10.1111/tri.12003) PMID: [23350848](https://pubmed.ncbi.nlm.nih.gov/23350848/)
3. Naesens M, Sarwal MM (2010) Molecular diagnostics in transplantation. *Nature Reviews Nephrology* 6: 614–628. doi: [10.1038/nrneph.2010.113](https://doi.org/10.1038/nrneph.2010.113) PMID: [20736923](https://pubmed.ncbi.nlm.nih.gov/20736923/)

4. Cortes M, Pareja E, Garcia-Canaveras JC, Donato MT, Montero S, Mir J et al.(2014) Metabolomics discloses donor liver biomarkers associated with early allograft dysfunction. *Journal of hepatology* 61: 564–574. doi: [10.1016/j.jhep.2014.04.023](https://doi.org/10.1016/j.jhep.2014.04.023) PMID: [24798621](https://pubmed.ncbi.nlm.nih.gov/24798621/)
5. Serkova NJ, Zhang Y, Coatney JL, Hunter L, Wachs ME, Niemann CU et al.(2007) Early detection of graft failure using the blood metabolic profile of a liver recipient. *Transplantation* 83: 517. doi: [10.1097/01.tp.0000251649.01148.f8](https://doi.org/10.1097/01.tp.0000251649.01148.f8) PMID: [17318087](https://pubmed.ncbi.nlm.nih.gov/17318087/)
6. Serkova NJ, Niemann CU (2006) Pattern recognition and biomarker validation using quantitative ¹H-NMR-based metabolomics 6:717–731 doi: [10.1586/14737159.6.5.717](https://doi.org/10.1586/14737159.6.5.717) PMID: [17009906](https://pubmed.ncbi.nlm.nih.gov/17009906/)
7. Amathieu R, Nahon P, Triba M, Bouchemal N, Trinchet JC, Beaugrand M et al. (2011) Metabolomic approach by ¹H NMR spectroscopy of serum for the assessment of chronic liver failure in patients with cirrhosis. *Journal of proteome research* 10: 3239–3245. doi: [10.1021/pr200265z](https://doi.org/10.1021/pr200265z) PMID: [21568267](https://pubmed.ncbi.nlm.nih.gov/21568267/)
8. Papaspyridonos K, Garcia-Perez I, Angulo S, Domann PJ, Vilca-Melendez H, Heaton N et al.(2008) Fingerprinting of human bile during liver transplantation by capillary electrophoresis. *Journal of separation science* 31: 3058–3064. doi: [10.1002/jssc.200800194](https://doi.org/10.1002/jssc.200800194) PMID: [18693311](https://pubmed.ncbi.nlm.nih.gov/18693311/)
9. Xu J, Casas-Ferreira AM, Ma Y, Sen A, Kim M, Proitsi P et al.(2015) Lipidomics comparing DCD and DBD liver allografts uncovers lysophospholipids elevated in recipients undergoing early allograft dysfunction. *Sci Rep* 5: 17737. srep17737 [pii]; doi: [10.1038/srep17737](https://doi.org/10.1038/srep17737) PMID: [26635289](https://pubmed.ncbi.nlm.nih.gov/26635289/)
10. Monbaliu D, Pirenne J, Talbot D (2012) Liver transplantation using donation after cardiac death donors. *Journal of hepatology* 56: 474–485. doi: [10.1016/j.jhep.2011.07.004](https://doi.org/10.1016/j.jhep.2011.07.004) PMID: [21782762](https://pubmed.ncbi.nlm.nih.gov/21782762/)
11. Foley DP, Fernandez LA, Levenson G, Chin LT, Krieger N, Cooper JT et al.(2005) Donation after cardiac death: the University of Wisconsin experience with liver transplantation. *Annals of surgery* 242: 724. doi: [10.1097/01.sla.0000186178.07110.92](https://doi.org/10.1097/01.sla.0000186178.07110.92) PMID: [16244547](https://pubmed.ncbi.nlm.nih.gov/16244547/)
12. De Vera ME, Lopez-Solis R, Dvorchik I, Campos S, Morris W, Demetris AJ et al.(2009) Liver Transplantation Using Donation After Cardiac Death Donors: Long-Term Follow-Up from a Single Center. *American journal of transplantation* 9: 773–781. doi: [10.1111/j.1600-6143.2009.02560.x](https://doi.org/10.1111/j.1600-6143.2009.02560.x) PMID: [19344466](https://pubmed.ncbi.nlm.nih.gov/19344466/)
13. Grewal HP, Willingham DL, Nguyen J, Hewitt WR, Taner BC, Cornell D et al.(2009) Liver transplantation using controlled donation after cardiac death donors: An analysis of a large single-center experience. *Liver Transplantation* 15: 1028–1035. doi: [10.1002/lt.21811](https://doi.org/10.1002/lt.21811) PMID: [19718636](https://pubmed.ncbi.nlm.nih.gov/19718636/)
14. Reich DJ, Mulligan DC, Abt PL, Pruett TL, Abecassis MMI, D'Alessandro A et al.(2009) ASTS recommended practice guidelines for controlled donation after cardiac death organ procurement and transplantation. *American Journal of Transplantation* 9: 2004–2011. doi: [10.1111/j.1600-6143.2009.02739.x](https://doi.org/10.1111/j.1600-6143.2009.02739.x) PMID: [19624569](https://pubmed.ncbi.nlm.nih.gov/19624569/)
15. Mathur AK, Heimbach J, Steffick DE, Sonnenday CJ, Goodrich NP, Merion RM (2010) Donation after cardiac death liver transplantation: predictors of outcome. *American Journal of Transplantation* 10: 2512–2519. doi: [10.1111/j.1600-6143.2010.03293.x](https://doi.org/10.1111/j.1600-6143.2010.03293.x) PMID: [20977642](https://pubmed.ncbi.nlm.nih.gov/20977642/)
16. Totsuka E, Fung JJ, Hakamada K, Ohashi M, Takahashi K, Nakai M et al.(2004) Synergistic effect of cold and warm ischemia time on postoperative graft function and outcome in human liver transplantation. *Transplantation proceedings* 36: 1955–1958. doi: [10.1016/j.transproceed.2004.08.068](https://doi.org/10.1016/j.transproceed.2004.08.068) PMID: [15518710](https://pubmed.ncbi.nlm.nih.gov/15518710/)
17. Kupiec-Weglinski JW, Busuttil RW (2005) Ischemia and reperfusion injury in liver transplantation. *Transplantation proceedings* 37: 1653–1656. doi: [10.1016/j.transproceed.2005.03.134](https://doi.org/10.1016/j.transproceed.2005.03.134) PMID: [15919422](https://pubmed.ncbi.nlm.nih.gov/15919422/)
18. Callaghan CJ, Charman SC, Muiesan P, Powell JJ, Gimson AE, van der Meulen JH (2013) Outcomes of transplantation of livers from donation after circulatory death donors in the UK: a cohort study. *BMJ open* 3: e003287. doi: [10.1136/bmjopen-2013-003287](https://doi.org/10.1136/bmjopen-2013-003287) PMID: [24002984](https://pubmed.ncbi.nlm.nih.gov/24002984/)
19. Hrydziusko O, Silva MA, Perera MT, Richards DA, Murphy N, Mirza D et al.(2010) Application of metabolomics to investigate the process of human orthotopic liver transplantation: a proof-of-principle study. *OMICS A Journal of Integrative Biology* 14: 143–150. doi: [10.1089/omi.2009.0139](https://doi.org/10.1089/omi.2009.0139) PMID: [20210660](https://pubmed.ncbi.nlm.nih.gov/20210660/)
20. Wu H, Southam AD, Hines A, Viant MR (2008) High-throughput tissue extraction protocol for NMR- and MS-based metabolomics. *Analytical biochemistry* 372: 204–212. doi: [10.1016/j.ab.2007.10.002](https://doi.org/10.1016/j.ab.2007.10.002) PMID: [17963684](https://pubmed.ncbi.nlm.nih.gov/17963684/)
21. Southam AD, Payne TG, Cooper HJ, Arvanitis TN, Viant MR (2007) Dynamic range and mass accuracy of wide-scan direct infusion nanoelectrospray fourier transform ion cyclotron resonance mass spectrometry-based metabolomics increased by the spectral stitching method. *Anal Chem* 79: 4595–4602. doi: [10.1021/ac062446p](https://doi.org/10.1021/ac062446p) PMID: [17511421](https://pubmed.ncbi.nlm.nih.gov/17511421/)
22. Payne TG, Southam AD, Arvanitis TN, Viant MR (2009) A signal filtering method for improved quantification and noise discrimination in fourier transform ion cyclotron resonance mass spectrometry-based metabolomics data. *J Am Soc Mass Spectrom* 20: 1087–1095. doi: [10.1016/j.jasms.2009.02.001](https://doi.org/10.1016/j.jasms.2009.02.001) PMID: [19269189](https://pubmed.ncbi.nlm.nih.gov/19269189/)

23. Hrydziusko O, Viant MR (2012) Missing values in mass spectrometry based metabolomics: an under-valued step in the data processing pipeline. *Metabolomics* 8: 161–174.
24. Dieterle F, Ross A, Schlotterbeck G, Senn H (2006) Probabilistic quotient normalization as robust method to account for dilution of complex biological mixtures. Application in ¹H NMR metabonomics. *Analytical Chemistry* 78: 4281–4290. doi: [10.1021/ac051632c](https://doi.org/10.1021/ac051632c) PMID: [16808434](https://pubmed.ncbi.nlm.nih.gov/16808434/)
25. Parsons HM, Ludwig C, Gunther UL, Viant MR (2007) Improved classification accuracy in 1-and 2-dimensional NMR metabolomics data using the variance stabilising generalised logarithm transformation. *BMC bioinformatics* 8: 234. doi: [10.1186/1471-2105-8-234](https://doi.org/10.1186/1471-2105-8-234) PMID: [17605789](https://pubmed.ncbi.nlm.nih.gov/17605789/)
26. Benjamini Y, Hochberg Y (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B (Methodological)* 289–300.
27. Kanehisa M, Araki M, Goto S, Hattori M, Hirakawa M, Itoh M et al. (2008) KEGG for linking genomes to life and the environment. *Nucleic acids research* 36: D480–D484. doi: [10.1093/nar/gkm882](https://doi.org/10.1093/nar/gkm882) PMID: [18077471](https://pubmed.ncbi.nlm.nih.gov/18077471/)
28. Kanehisa M, Goto S (2000) KEGG: kyoto encyclopedia of genes and genomes. *Nucleic acids research* 28: 27–30. PMID: [10592173](https://pubmed.ncbi.nlm.nih.gov/10592173/)
29. Feng L, Zhao N, Yao X, Sun X, Du L, Diao X et al. (2007) Histidine-tryptophan-ketoglutarate solution vs. University of Wisconsin solution for liver transplantation: A systematic review. *Liver Transplantation* 13: 1125–1136. doi: [10.1002/lt.21208](https://doi.org/10.1002/lt.21208) PMID: [17665493](https://pubmed.ncbi.nlm.nih.gov/17665493/)
30. Rossouw JE, Labadarios D, Davis M, Williams R (1977) The degradation of tryptophan in severe liver disease. *International journal for vitamin and nutrition research* 48: 281–289.
31. Mato JM, Lu SC (2007) Role of S-adenosyl-L-methionine in liver health and injury. *Hepatology* 45: 1306–1312. doi: [10.1002/hep.21650](https://doi.org/10.1002/hep.21650) PMID: [17464973](https://pubmed.ncbi.nlm.nih.gov/17464973/)
32. Clavien PA, Petrowsky H, DeOliveira ML, Graf R (2007) Strategies for safer liver surgery and partial liver transplantation. *New England Journal of Medicine* 356: 1545–1559. doi: [10.1056/NEJMra065156](https://doi.org/10.1056/NEJMra065156) PMID: [17429086](https://pubmed.ncbi.nlm.nih.gov/17429086/)
33. Mateo R, Cho Y, Singh G, Stapfer M, Donovan J, Kahn J et al. (2006) Risk factors for graft survival after liver transplantation from donation after cardiac death donors: an analysis of OPTN/UNOS data. *American journal of transplantation* 6: 791–796. doi: [10.1111/j.1600-6143.2006.01243.x](https://doi.org/10.1111/j.1600-6143.2006.01243.x) PMID: [16539637](https://pubmed.ncbi.nlm.nih.gov/16539637/)