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## Organic characterisation of cave drip water by LC-OCD and fluorescence analysis

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*DOI:* 10.1016/j.gca.2015.05.042

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Document Version Peer reviewed version

#### Citation for published version (Harvard):

Rutlidge, H, Andersen, MS, Baker, A, Ćhinu, KJ, Cuthbert, MO, Jex, CN, Marjo, CE, Markowska, M & Rau, GC 2015, 'Organic characterisation of cave drip water by LC-OCD and fluorescence analysis', *Geochimica et Cosmochimica Acta*, vol. 166, pp. 15-28. https://doi.org/10.1016/j.gca.2015.05.042

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Checked October 2015

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| 1  | Organic characterisation of cave drip water by LC-OCD and fluorescence analysis  |
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#### 20 Abstract

21 Cathedral Cave, Wellington, Australia, is a natural laboratory for studying water movement 22 and geochemical processes in the unsaturated zone by using artificial irrigation to activate drip sites within the cave. Water sampled from two drip sites activated by irrigations carried 23 24 out in summer 2014 was analysed for dissolved inorganic ions and fluorescent organic matter. The analysis allowed the development of a conceptual flow path model for each drip 25 26 site. DOM analysis was further complemented by liquid chromatography with organic carbon detection (LC-OCD), applied for the first time to karst drip waters, allowing the 27 characterisation of six organic matter fractions. The differences in organic matter fractions at 28 each drip site are interpreted as a signature of the proposed flow paths. LC-OCD was also 29 compared with parallel factor analysis (PARAFAC) of the fluorescence and good correlations 30 were observed for high molecular weight organic matter. Strong positive correlations were 31 32 also observed for high molecular weight matter and Cu and Ni. This is suggestive of colloidal transport of Cu and Ni by organic matter with high molecular weight, while small molecular 33 weight colloids were not efficient transporters. LC-OCD uniquely provides information on 34

non-fluorescent organic matter and can be used to further quantify drip water organic mattercomposition.

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## 38 1. Introduction

39 There has been increasing interest in dissolved organic matter (DOM) in cave drip water and the potential use of organic markers in speleothems as paleoclimate proxies (Blyth et al., 40 2008; Fairchild and Baker, 2012). While initial studies focussed on bulk organic matter 41 (Baker et al., 1997), subsequent investigations have focussed on the potential of lipid 42 biomarkers (Xie et al., 2003; Blyth et al., 2007; Rushdi et al., 2011), polycyclic aromatic 43 hydrocarbons (PAHs) (Perrette et al., 2013),  $\delta^{13}$ C of organic matter (Blyth et al., 2013a, b) 44 and trace elements associated with organic colloids (Hartland et al., 2012; 2014). Lipids such 45 as n-alkanes have been shown to record vegetation and land-use change (Blyth et al., 2007; 46 2011) and glycerol dialkyl glycerol tetraethers have shown potential as a paleotemperature 47 proxy (Blyth and Schouten, 2013; Blyth et al., 2014). 48

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50 Cave-based monitoring programs are typically utilised to understand the processes affecting 51 organic matter concentration and character as it is transported from the soil to the cave. 52 Typically cave drip water organic carbon concentrations are less than 3 mg/L (Baker et al., 1997; Ban et al., 2008), and are likely to be more variable than groundwater (Shen et al., 53 54 2014). This combined with relatively small water sample volumes that can be obtained from stalagmite drip water means that unfeasibly large water sample volumes are required for most 55 56 biomolecular characterisation techniques. Apart from one study on drip water PAHs (Perrette et al 2013) and one on LMW fatty acids (Bosle et al 2014), published drip water organic 57 58 matter monitoring has focussed on the measurement of the fluorescent organic matter fraction 59 (fDOM), as this requires about 4 mL of water sample, no pretreatment and has low detection 60 limits (Baker et al., 1997; Baker and Genty, 1999), however, this technique can only provide 61 information about the fluorescent fraction of the organic matter.

62

63 Liquid chromatography-organic carbon detection (LC-OCD) can be used to identify classes 64 of organic compounds in water samples. It gives quantitative information on natural organic 65 matter (NOM) and qualitative results regarding molecular size distribution of organic 66 impurities in water. The qualitative analysis is based on size exclusion chromatography 67 where large molecules have different degrees of interaction with the pores on the column 68 material resulting in different retention times for different molecular size fractions. 69 Quantification is completed by carbon mass determination, similar to total organic carbon (TOC) analysis, performed with both UV and organic carbon IR detectors which enable DOC 70 quantification at the sub-ppm level with only small sample volumes (10 mL). The resulting 71 chromatograms have significantly overlapping peaks, which typically require manual 72 73 processing to obtain correct peak separation, and therefore the expert knowledge of the user is important for reliable results. The technique is relatively new but has seen extensive use 74 75 characterising the efficiency of water treatment processes (Zheng et al., 2010; Li et al., 2012) as well as natural organic matter (NOM) in fresh and marine waters (Velten et al., 2011; 76 77 Rachman et al., 2014).

78

The DOC comprises hydrophobic organic carbon (HOC) that has a strong hydrophobic 79 interaction with the column material and does not elute through the column and 80 chromatographic dissolved organic carbon (CDOC). The HOC is determined by difference 81 between total organic carbon, determined by bypass of the column, and the CDOC. The five 82 different groups of CDOC that can be fractionated by the column are biopolymers, humic 83 substances, building blocks, low molecular weight (LMW) acids and neutrals, details of these 84 fractions are given in Table 1. These compounds are characterized by a UV-detector ( $\lambda = 254$ 85 nm) and quantified by IR-detection after UV oxidation in a cylindrical UV thin-film reactor 86 87 (Huber et al., 2011).

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The purpose of this study is to evaluate and compare LC-OCD and fluorescence analyses as a tool for characterisation of DOM in cave drip waters, and to understand the time evolution of organic matter fractions in the context of the routing of water between surface and drip site.

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#### 93 2. Material and methods

#### 94 <u>2.1 Study site</u>

95 Cathedral Cave at Wellington Caves, NSW, Australia (32 37'S; 148 56'E) was used as the 96 location for this study (Figure 1). The geology of the region has previously been described 97 (Johnson, 1975) and the cave is within an area of massive Devonian limestone, with a thin-98 layer of red-brown soil comprising clays, iron oxides, fine quartz sands, and calcite nodules 99 (Frank, 1971), with aeolian contributions (Hesse and McTanish, 2003). The site is within a 100 temperate semi-arid region, with mean annual precipitation of 619 mm (1956-2005) and 101 evaporation of 1825 mm (1956-2005) recorded at the nearby Wellington Research Centre (Australia Bureau of Meteorology). There is a significant seasonal temperature variation with
monthly mean maximum ranging from 15 °C in July and 32 °C in January (1956-1990,
Australia Bureau of Meteorology).

105

106 The cave continues to be a focus of long-term hydrogeological monitoring (concurrent to this study) by the investigators, commencing in 2010 and continuing, primarily using a network 107 108 of in-situ Stalagmate © drip loggers. Jex et al. (2012) described infiltration patterns and processes within the cave, and identified that infiltration only occurs after high magnitude 109 110 and long duration rainfall events, with a total precipitation of over ~60 mm within 24-48 hours, however the necessary amount of rainfall needed can vary dependant on antecedent 111 soil conditions. Such rainfall events occur very infrequently, typically 0-2 times a year, and 112 require slow-moving weather systems. In winter, this is most likely associated with westerly 113 frontal rainfall, where the associated low pressure system is slow moving, deep, and 114 relatively close to the site. In summer, atmospheric instability and associated convective 115 rainfall caused by slow moving or stationary troughs and associated upper level systems, 116 draws moist, unstable air from the north of the region. Drip water isotope monitoring 117 demonstrated that drip water  $\delta^{18}$ O is dominated by epikarst evaporation (Cuthbert et al., 118 2014a). Due to the infrequent recharge events, evaporation from near-surface karst water 119 120 stores leads to increasingly enriched cave drip waters.

121

Most recently, Cathedral Cave has been utilised for regular artificial irrigation experiments, 122 to better understand karst hydrogeochemical processes. An irrigation in the summer of 2013 123 has been used to understand drip water trace element and organic matter hydrogeochemical 124 evolution during recharge events (Rutlidge et al., 2014). An irrigation in the summer of 2014 125 has been used to understand the role of within-cave evaporation during recharge events, and 126 identified the role of evaporative cooling on drip waters for the first time (Cuthbert et al., 127 2014b). Data from both irrigation experiments has been used to quantify the processes of heat 128 transport (Rau et al., 2015). The data presented here is from water samples collected during 129 130 the summer 2014 irrigation event.

131

#### 132 <u>2.2 Irrigation experiment</u>

For this study, an area of approximately  $5 \times 10$  m, directly above the study area in Cathedral Cave was irrigated with Wellington town supply water. The experiment was performed at the

height of the Australian summer and the antecedent soil moisture was low (initial average soil 135 moisture was measured as 14.4 wt%). Two irrigations by hand hosing were performed on 136 consecutive mornings. The irrigation on Day 1 started at 7:50 am and continued for three 137 hours until dripping was initiated in the cave at five sites below the irrigation zone, one of 138 which had been used for drip water sampling previously and designated Site 1 (Rutlidge et 139 al., 2014) (Figure 1). A total of 3400 L was applied. The irrigation on Day 2 comprised two 140 batches of irrigation water. The initial batch was 1000 L town water loaded into a tank on site 141 and spiked with 0.5 L of 99.8% deuterium (D<sub>2</sub>O), resulting in a deuterium enrichment of 142 143 6700 ‰ (VSMOW) as measured by laser cavity ring down mass spectrometry (see below). This batch was hand hosed for 1 <sup>3</sup>/<sub>4</sub> hours then followed by a second non-enriched batch of 144 1430 L of town water for 1 <sup>1</sup>/<sub>2</sub> hours. Following this irrigation, an additional site activated and 145 was designated Site 25 (Figure 1). Site 1 was equipped with a Stalagmate® drip logger 146 during the experiment. 147

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#### 149 <u>2.3 Sampling and in-cave measurements</u>

For each drip water sample collected from Sites 1 and 25, a volume of 200-300 mL was 150 collected, which took approximately 30 minutes. Alkalinity, pH, electrical conductivity (EC) 151 152 and temperature were measured immediately after sampling. Alkalinity was determined on a 10 mL sub-sample using Gran titration with a 0.16 N H<sub>2</sub>SO<sub>4</sub>. Sub-samples for cation, anion, 153 fluorescence, LC-OCD and isotopic analysis were filtered (0.45 µm) and prepared for 154 subsequent laboratory testing, with the cation samples acidified with 2% 10 N nitric acid to 155 ensure stability during storage. In addition, one sample from each batch of irrigation water 156 (i.e. Day 1, Day 2 deuterium enriched and non-enriched town water) was collected for 157 analysis as above. 158

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#### 160 <u>2.4 Trace element and anion analysis</u>

161 Trace-element analysis of the drip water and each batch of irrigation water (without dilution) 162 was carried out using a Perkin Elmer NexION 300D ICP-MS and Perkin Elmer Optima 7300 163 ICP-OES. The following elements were analysed by ICP-OES: Ca, K, Mg, Na, Si and Sr, and 164 by ICP-MS: Al, Ba, Cu, Fe, Mn, Ni and Zn (Table EA1). The method has been described in 165 detail previously (Rutlidge et al., 2014). In addition, a Dionex Ion chromatography system 166 was used to determine the concentration of the following anions, fluoride, chloride, nitrite, 167 bromine, nitrate, phosphate and sulfate. Of these only chloride, bromide and sulfate were above the detection limit (Table EA2). PHREEQC for Windows (Parkhurst & Appelo, 2003)
was used to calculate saturation indices on selected minerals from the dissolved
concentrations.

171

#### 172 <u>2.5 Fluorescence analysis</u>

Fluorescence excitation-emission matrices (EEM's) were obtained for the collected drip 173 water samples using a Horiba Aqualog fluorescence spectrometer. This spectrometer allows 174 for the collection of absorbance and fluorescence within the same instrument, with the 175 176 absorbance data used to correct for any reabsorption (or inner-filter) effects. Fluorescence EEMs were collected using an excitation range of 240 to 400 nm, with a step-size of 3 nm, 177 and emitted fluorescence detected between 210 and 600 nm with a CCD detector, at a 178 spectral resolution of 1.64 nm and integration time of 1 s. All data was inner-filter corrected, 179 scatter lines masked, and Raman normalised (to a mean Raman intensity of water in a sealed 180 water cell, excited at 380 nm, of 200 intensity units), using proprietary Aqualog software. 181 The resultant dataset of 19 EEMs was analysed using a previous calibrated parallel factor 182 analysis (PARAFAC) model for this site (see Rutlidge et al., 2014) using Eigenvector 183 Research Solo © software. These three factors were characterised, following Ishii and Boyer 184 185 (2012), as unprocessed, soil-derived humic-like and fulvic-like material (Factor 1), biogeochemically processed humic/fulvic-like material (Factor 2), and living or dead 186 187 microbial matter (tryptophan-like fluorescence) (Factor 3) which is indicative of microbiological activity (Hudson et al., 2007). 188

189

## 190 <u>2.6 LC-OCD analysis</u>

191 The LC-OCD is an automated size-exclusion chromatography system coupled to three detectors, for organic carbon, organic nitrogen and UV absorbance, respectively. Details of 192 193 the measurement procedure have been described in full by Huber et al. (2011). In this study, a Toyopearl TSK HW50S column was used with a phosphate buffer mobile phase of pH 6.4 at 194 a flow rate of 1.1 mL/min. Injection volumes were 1 mL. The chromatographic column is a 195 weak cation exchange column containing a polymethacrylate filter. The chromatography 196 subdivides into six sub-fractions, which are assigned to specific classes of compounds: that is 197 biopolymers, humics, building blocks, low molecular-weight neutrals and hydrophobic 198 organic carbon (Huber et al., 2011). 199

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## 201 <u>2.7 Deuterium analysis</u>

202 The isotopic composition of irrigation and drip water samples were determined using an LGR-100DT V2 off-axis, integrated cavity output, cavity ring-down mass-spectrometer 203 (Wassenaar et al., 2008; Cuthbert et al., 2014a) at UNSW Australia. Each sample was run 204 with a total of 10 injections and run in triplicate (total of 30 injections), with the average of 205 the replicates used as the reported value. Four internal reference standards provided by the 206 Australian Nuclear Science and Technology Organisation with deuterium values of: -174.1, -207 78.8, -6.54, 32.3 ‰ VSMOW were used. An additional independent laboratory reference 208 standard was used from Elemental Microanalysis with deuterium value of 4.93 relative to 209 210 VSMOW. The analytical precision for deuterium were ~1.5% VSMOW (1 $\sigma$ ; calculated from within run internal reference materials). This compares to an external precision for deuterium 211 of 0.6% (1 $\sigma$ , analysis of standards). The calibration curve was calculated using the reference 212 standards and was used to interpolate drip water and irrigation water samples. Samples which 213 fell outside of the range of the reference standards were extrapolated. 214

215

## 216 **3. Results**

Figure 2 shows the drip rate data for Site 1 over the course of the experiment. Dripping at 217 Site 1 activated on Day 1 and stopped on Day 3, while a secondary site, Site 25, activated on 218 219 Day 2 and continued on Day 3 and beyond. At Site 1, the two irrigations above the cave resulted in two significant pulses, with a three hour delay on Day 1 and one hour delay on 220 221 Day 2 from the start of each irrigation. The second site considered in this study (Site 25) was an unexpected activation on Day 2 in a relatively inaccessible location in the cave and no 222 223 measured drip rate data is available, however sampling for both sites was possible over the 224 course of the experiment allowing both sites to be chemically characterised. When dripping 225 had commenced at Site 25 it was observed to be consistent but at a lower frequency than the initial activation at Site 1, suggesting there are flow restrictions at this site relative to Site 1. 226 227 Site 1 shows an overall higher pH, lower alkalinity and Ca concentration than Site 25. This is reflected in the PHREEQC calculated PCO<sub>2</sub> and saturation indices for calcite that are both 228 lower for Site 1. Calculations indicate near saturation of calcite at Site 25 and a PCO<sub>2</sub> that is 229 an order of magnitude higher than at Site 1. 230

231

A previous study by the authors at this location established that certain elements were derived predominantly from either the bedrock or the soil using a principal component analysis (PCA) model (Rutlidge et al., 2014). Ba, Cu and Ni were soil-derived elements and Ca and Sr were limestone bedrock-derived elements. A soil source for both Ni and Cu is likely as they 236 are both organo-colloid associated trace elements (Hartland et al., 2012). Ba was not detected in limestone bedrock samples and since it is strongly absorbed to clay minerals, the clay-rich 237 soils likely provide the dominant source of the Ba. Table 2 shows that at Site 1 the bedrock-238 derived Ca concentration was lower than at Site 25, while the soil-derived elements of Ba, Cu 239 and Ni were higher. A relatively small difference was observed for Sr between the two sites. 240 While the initial concentrations of elements in irrigation water were comparable to the drip 241 water concentrations, irrigation water does not make a significant contribution to the 242 measured drip water concentrations due to dilution with existing stored water, based on the 243 244 deuterium measurements (see Table 2). A minimum dilution down to 3.5% can be estimated based on the initial deuterium isotope ratio in the irrigation tank (6700% VSMOW) and the 245 maximum deuterium ratio measured at Site 1 of 220% VSMOW against the background 246 deuterium ratio of -13‰ VSMOW. The deuterium ratio measured at Site 25 (38‰ VSMOW 247 on average) is above the background level (assumed to be same as Site 1) but relatively 248 consistent and significantly lower than the maximum value for Site 1. 249

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- 252

253 Figure 3 shows the behaviour of the PARAFAC factors derived from the analysis of the fluorescent EEM data over the course of the study. The three factors characterised were 254 255 unprocessed, soil-derived humic-like and fulvic-like material (Factor 1), biogeochemically processed humic/fulvic-like material (Factor 2), and living or dead microbial matter 256 257 (tryptophan-like fluorescence) (Factor 3). These three factors are the same as those observed in the 2013 irrigation experiments (Rutlidge et al., 2013). Overall the drip water from Site 1 258 259 contains more organic matter than Site 25. Factor 1 and Factor 2 show a relatively small decrease between the two irrigations at Site 1. At Site 25 these factors are relatively constant. 260 Factor 3 (microbial-derived) shows initial high values for the first irrigation and then 261 decreasing for the second irrigation at Site 1 compared to Site 25 where it is relatively 262 unchanged. 263

264

Figure 4 shows the time-series concentration profiles for the organic fractions measured by LC-OCD for both sites. Overall the DOC concentration at Site 1, as measured by LC-OCD, decreased over the course of the study, while at Site 25 the DOC concentration was lower and remained relatively unchanged. The DOC fraction comprises both HOC (hydrophobic) and CDOC (hydrophilic) organic carbon. The CDOC fraction can be further broken down into chemical fractions of bio-polymers, humics, building blocks and LMW neutrals. For both sites the concentration of CDOC (Site 1 average was 4565  $\mu$ g/L and Site 25 average was 2150  $\mu$ g/L) was significantly higher than the HOC concentrations (the average values for Site 1 and Site 25 were 1213  $\mu$ g/L and 736  $\mu$ g/L respectively). In terms of individual fractions, the bio-polymers decreased over time at Site 1 but were close to the detection limit at Site 25. Building blocks were fairly consistent each day but showed a decrease from day to day at Site 1.

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Table EA3 shows the Kendall tau correlations between selected elements and the LC-OCD
fractions. Ca (bedrock-derived) shows negative correlations with DOC (-0.61), CDOC (-0.61)
and humics (-0.66). While Cu and Ni (soil-derived) show positive correlations with all LCOCD fractions except building blocks and LMW neutrals. Ba shows positive correlations
with DOC (0.62) and humics (0.62).

283

Overall the irrigation water contains the highest proportion of CDOC (hydrophilic) followed 284 by Site 1, with Site 25 the lowest. Conversely, HOC (hydrophobic) showed the lowest 285 proportion in the irrigation water, increasing in Site 1 and again in Site 25. There are clear 286 287 differences in the relative proportions of the different hydrophilic fractions measured by LC-OCD in the DOC of the irrigation water and the drip water from the two sites as shown in 288 289 Figure 5. Site 1 has higher proportion of humics, building blocks & bio-polymers while Site 25 has a higher proportion of LMW neutrals. Site 25 shows the most difference from the 290 291 irrigation water.

292

#### 293 4. Discussion

#### 294 <u>4.1 Flow path Model</u>

295 After the first irrigation dripping at Site 1 responded within 3 hours, with an initial high drip rate decreasing over 20 hours. This 'flashy' flow response, in combination with a short arrival 296 time for the deuterium tracer, suggests a short flow path for Site 1. This is supported by an 297 excavation that revealed a soil filled void in the limestone directly above Site 1, and a 298 measured maximum bedrock thickness of only ~ 1.5 m. In comparison, Site 25 showed a 299 300 lagged and damped flow response and deuterium tracer breakthrough, suggesting a combination of more unsaturated zone storage water, tortuous/longer pathways through the 301 epikarst and lower fracture permeability. Deuterium measurements at both sites showed 302 significant dilution by existing stores of water. Site 25 showed a higher level of dilution of 303

the tracer (down to 0.9%) compared to Site 1 (down to 3.5%), suggesting that the Site 25
store contained more water. This increased dilution can partly explain the decreased total
DOC observed at Site 25 compared to Site 1.

307

308 Since cave drip water carries a chemical signature indicative of its flow path, the analysis provided here can be used to reveal characteristics of flow routing between surface and cave. 309 Our interpretation benefits from the long-term drip water isotope monitoring program at the 310 cave (Cuthbert et al 2014a). That study demonstrated the ubiquitous presence of enriched drip 311 312 water isotopes, that could only be explained by evaporation from water stored in fractures or voids the shallow vadose zone. Site 1 has PCO<sub>2</sub> values (Figure 2) only slightly elevated 313 above atmospheric values that could indicate poor CO<sub>2</sub> production in the soil, loss of 314 produced CO<sub>2</sub> by ventilation, or degassing as the water flowed down over the exposed 315 flowstone surface between the exfiltration point and drip site. Site 25 on the other hand had 316 PCO<sub>2</sub> values about one order of magnitude above those of Site 1 due to longer residence 317 times in the bedrock and a more restricted pore space minimising CO<sub>2</sub> loss to the atmosphere. 318 The recorded pH values largely reflect the calculated PCO<sub>2</sub> values, but may be tempered by 319 320 calcite dissolution (hence the near neutral pH values). Calcium, alkalinity and the saturation 321 index (SI) for calcite indicate that water sampled at Site 25 is more 'evolved' since it is closer to equilibrium with calcite. It further indicates that there was a larger volume of stored water 322 323 in the flow path connecting to Site 25. Conversely at Site 1, the inorganic carbon observations suggest that there is lower volume of stored water and that the store is in the soil 324 325 above the limestone. This is supported by higher concentrations of trace elements at Site 1 previously shown to be associated with soil at this study location, specifically Ba, Cu and Ni 326 327 (Rutlidge et al., 2014).

328

Figure 6 shows a proposed schematic of flow paths for Sites 1 and 25 derived from the 329 observations above. In summary, Site 1 drip water travels through a void in the bedrock that 330 was filled with partially saturated soil, followed by further passage through a relatively thin 331 (<1.5m) amount of limestone, before entering the cave near the cave roof. The water 332 continues to flow over the cave wall, forming a flowstone deposit, before dripping from a 333 stalactite at Site 1. . Site 25 is fed by a long thin bedrock fracture, or a series of fractures, 334 335 which requires more water to enter the epikarst and/or more time than Site 1 before activating the drip point. 336

337

#### 338 4.2 Correlations between PARAFAC, LC-OCD and trace elements

The site had experienced a total of only 88 mm of rainfall over six events in the three months 339 prior to the irrigation and as a result the volume of water required to overcome the soil 340 moisture deficit and activate dripping was at least 1500 L. This volume of water was only 341 available from the local town water supply where DOM is present at comparable levels to 342 those measured in the drip water. Hence, DOM in the drip water will be derived from existing 343 DOM in town water and DOM extracted from soil organic matter in the epikarst. However, 344 as with the trace elements the DOM in the irrigation water would be diluted by the existing 345 346 water stored in the epikarst.

347

Figure 7 shows plots of selected LC-OCD derived classes of compounds that can be related 348 to the three corresponding PARAFAC factors from all samples taken over the course of the 349 study. Factor 1, unprocessed humic and fulvic acids, shows a strong correlation with the LC-350 OCD humic fraction (Kendall tau coefficient of 0.61), as does Factor 2 (Kendall tau 351 coefficient of 0.79), which is processed humic and fulvic acids. The LC-OCD humic fraction 352 therefore correlates with the two PARAFAC factors which are also associated with humic-353 like material, with the PARAFAC analysis suggesting that the LC-OCD humic fraction 354 355 comprises two different optical components. Factor 3 shows the highest correlation with biopolymers (Kendall tau coefficient of 0.69). Factor 3 is characterised as tryptophan-like 356 357 fluorescence, typically derived from living or dead microbial matter, and the correlation with bio-polymers suggests a microbial biopolymer source. In addition to the plots shown in Figure 358 359 6, Factor 1 and 2 are positively correlated with DOC measured by LC-OCD, and the HOC (hydrophobic) and CDOC (hydrophilic) LC-OCD fractions (see Table EA3). Factor 1 is also 360 361 positively correlated with the derived aromaticity index and Factor 2 with the building block fraction. In most cases the LC-OCD derived indices, aromaticity, molecular weight, inorganic 362 colloids and specific UV absorbance (SUVA) did not show strong correlations with any 363 PARAFAC factors or LC-OCD fractions. The correlations observed in Figure 7 demonstrate 364 that LC-OCD agrees with the PARAFAC factor assignments from the fluorescence analysis. 365

366

367 LC-OCD uniquely provides information on non-fluorescent organic matter and can be used to 368 identify and quantify changes in organic matter composition. Figure 8 shows individual 369 scatter plots of the LC-OCD fractions and HOC (hydrophobic) expressed in terms of percent 370 of total DOC, to minimise the changes due to dilution. LMW neutrals are relatively higher at 371 Site 25 than Site 1 indicating that they are highly mobile and eluted rapidly as expected for uncharged low molecular weight DOM (Shen et al., 2014). In contrast building blocks are
relatively lower at Site 25 compared to Site 1 which suggests that they are interacting with
the calcite surface in the epikarst (Alipour Tabrizy et al. 2011; Suess, 1970 and Suess, 1973).

375

Drip water from Site 25 consistently contained low levels of biopolymer and were often below detection. Along with the correlation with PARAFAC Factor 3, this suggests that there was an initial flush of biopolymer at Site 1 that would be expected to have accumulated in the partially saturated lower soil layer prior to the first irrigation. The LC-OCD biopolymer fraction is defined as being >20 kDa and hydrophilic (Huber et al 2011) organic compounds such as polysaccharides. We hypothesise that the longer flow path at Site 25 leads to greater loss of this fraction, for example by microbial degradation, compared to Site 1.

383

The relative decrease in the humic substances fraction at Site 25 can be explained by the 384 preferential adsorption of the negatively charged humic fraction at the positively charged 385 calcite surface during flow through the bedrock fracture. We observe ongoing calcite 386 formation in the form of active flowstone deposits (Site 1) and stalactite formation (Sites 1 387 and 25). Ongoing calcite formation from solution provides a continual source of fresh 388 389 surfaces that are positively charged at the drip water pH (approximately 8), below the point of zero charge of calcite,  $pH_{PZC} = 9.5$  (Appelo and Postma, 2005). This preferential adsorption 390 391 is supported by numerous laboratory studies of organic matter interaction with calcite (Alipour Tabrizy et al. 2011; Suess, 1970 and Suess, 1973). LC-OCD therefore helps identify 392 393 changes in organic matter composition, with loss of different organic matter fractions due to processes which include sorption and microbial processing. Such changes in composition can 394 395 be conceived as a smaller-scale manifestation of the "regional scale chromatography" proposed for groundwater (Hedges et al., 1986; Shen et al., 2014). 396

397

Strong positive correlations were observed for Cu and Ni with the higher molecular weight 398 LC-OCD fractions, for example biopolymers-Cu (0.76) and humics-Ni (0.80). This supports 399 a prior study by Hartland et al (2012) who used flow field-flow fractionation of karst drip 400 401 waters that showed colloidal transport of Cu and Ni by organic matter with high molecular weight. This same study observed that small molecular weight colloids were not efficient 402 403 transporters of Cu and Ni, and we further confirm this observation here as we see low correlations observed for these elements and the LMW neutrals (0.40 and 0.24, respectively). 404 We also considered the relationship between the Cu/Ni ratio and LC-OCD fractions and 405

406 derived indices (Table EA3 and Figure 9). Hartland et al (2012) have previously hypothesised 407 that the Cu/Ni ratio in cave drip waters is indicative of a quality change in DOM, as Cu has an increased affinity to binding to aromatic binding sites than Ni. For the whole dataset 408 (Table EA2), our results show that the only statistically significant correlation between Cu/Ni 409 and LC-OCD fractions is a negative correlation with humics (-0.49) and we see no significant 410 correlations between Cu/Ni and the derived indices of molecular weight or aromaticity. 411 However, at a site-by-site basis (Figure 9), we observe that these global correlations are 412 masking site-specific patterns. At Site 1, there are positive correlations between Cu/Ni and 413 414 aromaticity and molecular weight, as predicted by Hartland et al (2012), and also with building blocks and LMW neutrals. In contrast, correlations between Cu/Ni and LC-OCD 415 fractions and derived indices are very weak at Site 25, but the site as a whole has high Cu/Ni 416 than Site 1, which explains the global negative correlations observed (Table EA2). Our LC-417 OCD results suggest a complex relationship between organic matter character as determined 418 by LC-OCD and Cu/Ni over the event timescale which requires further investigation. 419

420

421 Compared with fluorescence analysis LC-OCD provides more information on the types of colloid-forming DOM involved in metal transport. Our results that demonstrate correlation 422 423 between Cu and Ni concentration and DOM concentration for all data, but only with the largest size fractions (humics and biopolymers). The correlation between Cu/Ni and 424 425 aromaticity varies between the two sites. It is strongest at Site 1, despite the lack of correlation between humic fraction concentration and Cu/Ni, confirming an aromaticity 426 427 control on Cu/Ni (Hartland et al. 2012). However, variations in the correlation with LC-OCD fractions and indices between flow paths and within recharge events suggest that flow 428 429 switching and discharge variability may be preserved in speleothem records of these organictransported metals. 430

## 431 5. Conclusions

Trace element composition measured by ICP and organic matter analysis by fluorescence has been applied to cave drip water samples collected after a series of artificial irrigations. This revealed an understanding of the flow paths feeding two drip sites. We propose that Site 1 is fed by a short and permeable bedrock fracture via a soil store which retains relatively high moisture content during periods of no rainfall. In comparison the data suggest that Site 25 is fed by an area of thinner, drier soil through a thicker epikarst with greater unsaturated storage and/or lower permeability.

439

440 DOM analysis was also performed by LC-OCD, applied for the first time to karst drip waters. Good correlations were observed between LC-OCD humic and bio-polymer fractions and the 441 corresponding PARAFAC factors from the fluorescent analysis. LC-OCD enabled a more 442 detailed characterisation of the non-fluorescent fractions of DOM present in the drip water 443 sample. LC-OCD identified that as the drip water moves through bedrock fractures there are 444 changes in dissolved organic matter composition which will depend on the path-length of the 445 water. Locations nearer the surface with shorter path-length and higher discharge have higher 446 DOC, as there is less time for organic matter sorption and microbial processing. At Site 25, 447 448 with the longer path-length, this leads to the preferential adsorption of the humic substance fraction, resulting in a relative decease in the hydrophilic portion of DOM compared to Site 449 1, while the uncharged low molecular weight neutrals pass through unaffected. 450

451

LC-OCD provides insights into vadose zone processes affecting dissolved organic matter, 452 such as sorption and microbial processing, which occur between different flow paths, within 453 discharge events and between discharge events. The greater level of chemical detail, unique 454 to LC-OCD, demonstrates its potential for providing insight into the evolution of organic 455 matter from soil to cave, through different flow paths in the unsaturated zone. An improved 456 457 understanding of these processes could have implications for selecting speleothem samples suitable for biomarker research, such as when using lipids for paleoclimate reconstructions 458 459 (Blyth et al., 2007) or fluorescent organic matter to construct fluorescence lamina growth chronologies (Baker et al., 1993). Research presented here, focusing on just two drip waters 460 461 of contrasting flow path-length, needs to be extended for a wider range of drip sites and caves 462 using LC-OCD.

463

#### 464 **5. Acknowledgements**

We thank the staff at Wellington Caves for their support. Funding for this research was provided by the National Centre for Groundwater Research and Training, an Australian Government initiative, supported by the Australian Research Council and the National Water Commission and Mark Wainwright Analytical Centre at UNSW Australia. Mark Cuthbert was supported by Marie Curie Research Fellowship funding from the European Community's Seventh Framework Programme [FP7/2007-2013] under grant agreement n°299091. We also acknowledge the anonymous reviewers that provided helpful feedback.

472

#### 473 **6. References**

- 474
- Alipour Tabrizy, V., Denoyel, R. and Hamouda, A.A. (2011). Characterization of wettability
  alteration of calcite, quartz and kaolinite: Surface energy analysis. *Colloids and Surfaces*
- 477 *A: Physicochemical and Engineering Aspects* **384**, 98-108.
- 478 Appelo, C.A.J. and Postma, D. (2005). *Geochemistry, Groundwater, and Pollution*, second
  479 ed. A.A. Balkema, Rotterdam, 649 pp.
- Baker, A., Smart, P. L., Edwards, R. L., and Richards, D. A. (1993). Annual growth banding
  in a cave stalagmite. *Nature*, 364, 518-520.
- Baker, A., Barnes, W.L. and Smart, P.L. (1997). Stalagmite Drip Discharge and Organic
  Matter Fluxes in Lower Cave, Bristol. *Hydrological Processes* 11, 1541-1555.
- Baker, A. and Genty, D. (1999). Fluorescence wavelength and intensity variations of cave
  waters. *Journal of Hydrology* 217, 19-34.
- 486 Ban, F., Pan, G., Zhu, J., Cai, B., and Tan, M. (2008). Temporal and spatial variations in the
- discharge and dissolved organic carbon of drip waters in Beijing Shihua Cave, China. *Hydrological Processes* 22, 3749-3758.
- Blyth, A.J., Asrat, A., Baker, A., Gulliver, P., Leng, M.J. and Genty, D. (2007). A new
  approach to detecting vegetation and land-use change using high-resolution lipid
  biomarker records in stalagmites. *Quaternary Research* 68, 314-324.
- Blyth, A.J., Baker, A., Collins, M.J., Penkman, K.E.H., Gilmour, M.A., Moss, J.S., Genty, D.
- and Drysdale, R.N. (2008). Molecular organic matter in speleothems and its potential as an
  environmental proxy. *Quaternary Science Reviews* 27, 905-921.
- Blyth, A.J., Thomas, L.E., Calsteren, P.V. and Baker, A. (2011). A 2000 year lipid biomarker
  record preserved in a stalagmite from NW Scotland. *Journal of Quaternary Science* 26,
  326-334.
- 498Blyth, A.J., Shutova, Y. and Smith, C.I. (2013).  $\delta^{13}$ C analysis of bulk organic matter in499speleothems using liquid chromatography–isotope ratio mass spectrometry. *Organic*
- 500 *Geochemistry* **55**, 22-25.
- 501 Blyth, A.J., Smith, C.I. and Drysdale, R.N. (2013). A new perspective on the  $\delta^{13}$ C signal
- preserved in speleothems using LC-IRMS analysis of bulk organic matter and compound
  specific stable isotope analysis. *Quaternary Science Reviews* 75, 143-149.
- 504 Blyth, A.J. and Schouten, S. (2013). Calibrating the glycerol dialkyl glycerol tetraether
- temperature signal in speleothems. *Geochimica Cosmochimica Acta* **109**, 312-328.

- 506 Blyth, A.J., Jex, C., Baker, A., Khan, S.J. and Schouten, S. (2014). Contrasting distributions
- 507 of glycerol dialkyl glycerol tetraethers (GDGTs) in speleothems and associated soils.
  508 *Organic Geochemistry* 69, 1-10.
- Bosle, J.M., Mischel, S.A., Schulze A.-L., Scholz, D. and Hoffman, T. (2014) Quantification
  of low molecular weight fatty acids in cave drip water and speleothems using HPLC-ESI-
- 511 IT/MS development and validation of a selectiv emethod. Anal. Bioanal. Chem., 406,
- 512 3167-3177.
- Cuthbert, M.O., Baker, A., Jex, C.N., Graham, P.W., Treble, P.C., Andersen, M.S. and
  Acworth, R.I. (2014a). Drip water isotopes in semi-arid karst: Implications for speleothem
  paleoclimatology, *Earth and Planetary Science Letters* 395, 194-204.
- 516 Cuthbert, M.O., Rau, G.C., Andersen, M.S., Roshan, H., Rutlidge, H., Marjo, C.E.,
- 517 Markowska, M., Jex, C.N., Graham, P.W., Mariethoz, G. Acworth, R.I. and Baker, A.
- 518 (2014b). Evaporative cooling of speleothem drip water. *Scientific Reports* **4**, .
- 519 Fairchild I. J. and Baker A. (2012) Speleothem Science. Wiley-Blackwell, 432 pp.
- Frank, R. (1971). The clastic sediments of the Wellington Caves, New South Wales. *Helictite*9, 3-26.
- 522 Hartland, A., Fairchild, I.J., Lead, J.R., Borsato, A., Baker, A., Frisia, S. and Baalousha, M.
- 523 (2012). From soil to cave: transport of trace metals by natural organic matter in cave
- dripwaters. *Chemical Geology* **304-305**, 68-82.
- 525 Hartland, A., Fairchild, I.J., Müller, W. and Dominguez-Villar, D. (2014). Preservation of
- NOM-metal complexes in a modern hyperalkaline stalagmite: Implications for speleothem
  trace element geochemistry. *Geochimica et Cosmochimica Acta* 128, 29-43.
- 528 Hedges, J.I., Ertel, J.L., Quay, P.D., Grootes, P.M., Richey, J.E., Devol A.H., Farewell G.W.,
- Schmidt, F.W., and Salati, E. (1986). Organic carbon-14 in the Amazon River system. *Science* 231, 1129-1131.
- Hesse P.P. and McTanish, G.H. (2003). Australian dust deposits: modern processes and the
  Quaternary record. *Quaternary Sciences Reviews* 22, 2007-2035.
- 533 Huber, S.A., Balz, A., Abert, M. and Pronk, W. (2011). Characterisation of aquatic humic
- and non-humic matter with size-exclusion chromatography organic carbon detection –
   organic nitrogen detection (LC-OCD-OND). *Water Research* 45, 879-885.
- 536 Hudson, N., Baker, A., Ward, D., Reynolds, D.M., Brunsdon, C., Carliell-Marquet, C. and
- 537 Browning, S. (2008). Can fluorescence spectrometry be used as a surrogate for the
- Biochemical Oxygen Demand (BOD) test in water quality assessment? An example from
- 539 South West England. *Science of the Total Environment* **391**, 149-158.

- 540 Ishii S. K. and Boyer T. H. (2012). Behavior of reoccurring PARAFAC components in
- 541 fluorescent dissolved organic matter in natural and engineered systems: A critical review.
- 542 *Environmental Science and Technology* **46**, 2006–2017.
- Jex C. N., Mariethoz G., Baker A., Graham P., Andersen M. S., Acworth I., Edwards N. and
   Azcurra C. (2012). Spatially dense drip hydrological monitoring and infiltration behaviour
- at the Wellington Caves, South East Australia. *International Journal of Speleology*. **41**,
- 546 285–298.
- Johnson, B.D. (1975). The Garra Formation (early Devonian) at Wellington, N.S.W. *Journal and Proceedings of The Royal Society of New South Wales* 108, 111-118.
- Li, S., Heijman, S.G.J., Verberk, J.Q.J.C., Verliefde, A.R.D., Amy, G.L. and van Dijk, J.C.
- (2012). Removal of different fractions of NOM foulants during demineralized water
  backwashing. *Separation and Purification Technology* **98**, 186-192.
- 552 Parkhurst, D. L. and Appelo C. A. J. (2003). PHREEQC for Windows: A Computer Program
- for Speciation, Batch-Reaction, One-Dimensional Transport, and Inverse Geochemical
  Calculations; [2.8. 03]. USGC, Washington, DC.
- Perrette, Y., Poulenard, J., Durand, A., quiers, M., Malet, E., Fanget, B and Naffrechoux E.
  (2013) Atmopsheric sources and soil filtering of PAH content in karst seepage waters. *Organic Geochemistry* 65, 37-45
- 558 Rachman, R.M., Li, S., Missimer, T.M. (2014). SWRO feed water quality improvement using
- subsurface intakes in Oman, Spain, Turks and Caicos Islands, and Saudi Arabia.
- 560 *Desalination*, **351**, 88-100.
- 561 Rau, G.C., Cuthbert , M.O., Andersen, M.S., Baker, A., Rutlidge, H., Markowska, M.,
- Roshan, H., Marjo, C.E., Graham, P.W. and Acworth, R.I (2015). Controls on cave drip
- water temperature and implications for speleothem based paleoclimate reconstructions.
- 564 *Quaternary Science Reviews*, in press.
- 565 Rushdi, A.I., Clark, P.U., Mix, A.C., Ersek, V., Simoneit, B.R.T., Cheng, H. and Edwards,
- 566 R.L. (2011). Composition and sources of lipid compounds in speleothem calcite from
- southwestern Oregon and their paleoenvironmental implications. *Environmental Earth Sciences* 62, 1245-1261.
- 569 Rutlidge, H., Baker, A., Marjo, C.E., Andersen, M.S., Graham, P.W., Cuthbert, M.O., Rau,
- 570 G.C., Roshan, H., Markowska, M., Mariethoz, G. and Jex, C.N. (2014). Dripwater organic
- 571 matter and trace element geochemistry in a semi-arid karst environment: Implications for
- 572 speleothem paleoclimatology. *Geochimica et Cosmochimica Acta* **135**, 217-230.

| 573 | Shen, Y., Chapelle, F.H., Strom, E.W., and Benner R. (2014). Origins and bioavailablity of       |
|-----|--|
| 574 | dissolved organic matter in groundwater. Biogeochemistry DOI 10.1007/s10533-014-                 |
| 575 | 0029-4.  |
| 576 | Suess, E. (1970). Interaction of organic compounds with calcium carbonate—I. Association         |
| 577 | phenomena and geochemical implications. Geochimica et Cosmochimica Acta 34, 157-                 |
| 578 | 168.   |
| 579 | Suess, E. (1973). Interaction of organic compounds with calcium carbonate-II. Organo-            |
| 580 | carbonate association in recent sediments. Geochimica et Cosmochimica Acta 37, 2435-             |
| 581 | 2447.  |
| 582 | Velten, S., Knappe, D.R.U., Traber, J., Kaiser, H-P., von Gunten, U., Boller, M. and Meylan,     |
| 583 | S. (2011). Characterization of natural organic matter adsorption in granular activated           |
| 584 | carbon adsorbers. Water Research 45, 3951-3959.  |
| 585 | Xie, S., Yi, Y., Huang, J., Hu, C., Cai, Y., Collins, M. and Baker A. (2003). Lipid distribution |
| 586 | in a subtropical southern China stalagmite as a record of soil ecosystem response to             |
| 587 | paleoclimate change. Quaternary Research 60, 340-347.  |
| 588 | Zheng, X.; Ernst, M., Jekel, M. (2010). Pilot-scale investigation on the removal of organic      |
| 589 | foulants in secondary effluent by slow sand filtration prior to ultrafiltration. Water           |
| 590 | Research 44, 3203-3213.  |
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| 595 | Table 2: Average trace element concentrations for the irrigation water and Site 1 and 25.        |
| 596 | Irrigation contribution is the estimated concentration of elements from the irrigation water to  |
| 597 | the cave drip water after dilution by existing water stored in the vadose zone based on          |
| 598 | deuterium measurements.  |

599

Figure 1 : Plan view of the study site at Cathedral Cave, Wellington Caves (top), with a

boxed area indicating the surface irrigation area and cave locations of the drip sites (adapted

from Sydney University Speleology Society survey map 2006-2007). The study site is in the

state of New South Wales in south eastern Australia (bottom).

604

| 605 | Figure 2: Drip rate data (Site 1 only), deuterium, pH, alkalinity, Ca, LogPCO <sub>2</sub> and saturation |
|-----|---|
| 606 | index for calcite (Sites 1 and 25) for the irrigation water and drip water samples. Grey                  |
| 607 | vertical bars indicate the irrigation periods. Lines between datapoints indicate periods of               |
| 608 | continuous dripping.  |
| 609 |   |
| 610 | Figure 3: PARAFAC factors 1-3 over time from the analysis of the fluorescence EEM data.                   |
| 611 | Lines between datapoints indicate periods of continuous dripping.   |
| 612 |   |
| 613 | Figure 4: Time series of LC-OCD fractions for both sites. Lines between datapoints indicate               |
| 614 | periods of continuous dripping.   |
| 615 |   |
| 616 | Figure 5: Average composition of the DOC fraction for the irrigation water and both sites.                |
| 617 |   |
| 618 | Figure 6: Simplified schematic of the different flow paths (prior to irrigation) derived from             |
| 619 | the chemical signature of the drip water. The width of markers on each axis corresponds to                |
| 620 | one metre.  |
| 621 |   |
| 622 | Figure 7: PARAFAC factors versus selected LC-OCD fractions.   |
| 623 |   |
| 624 | Figure 8: Matrix scatter plot of the percent of DOC of the LC-OCD fractions.                              |
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| 626 | Figure 9: Matrix scatter plot of Cu/Ni against the LC-OCD fractions and indices.                          |
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| 606 | Xie, S., Yi, Y., Huang, J., Hu, C., Cai, Y., Collins, M. and Baker A. (2003). Lipid distribution          |
|-----|---|
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| 636 |   |
| 637 | Figure 5: Average composition of the DOC fraction for the irrigation water and both sites.                |
| 638 |   |

|          |  | Description   |  |  |  |
|----------|--|---|--|--|--|
| Fraction | Biopolymers                            | Organic matter with high molecular weight,<br>including polysaccharides, proteins and<br>aminosugers. (Molecular weight > 20 kDa)   |  |  |  |
|          | Humics                                 | Mixture of acids containing carboxyl and<br>phenolate groups produced by biodegradation of<br>dead organic matter. (Molecular weight ~ 1000<br>Da)  |  |  |  |
|          | Building blocks                        | Molecular chains of polyphenolics/polyaromatic<br>acids that have deaggregated, due to breakage of<br>hydrogen bonding and electrostatic interactions.<br>(Molecular weight 300 – 500 Da) |  |  |  |
|          | Low molecular weight<br>(LMW) acids    | Representing protic organic acids. (Molecular weight < 350 Da)  |  |  |  |
|          | Low molecular weight<br>(LMW) neutrals | Uncharged small organics, including LMW alcohols, aldehydes, ketones, sugars and LMW amino acids. (Molecular weight < 350 Da)   |  |  |  |
|          | Hydrophobic organic carbon<br>(HOC)    | Fraction of DOC remaining in the column,<br>implying a strong hydrophobic interaction with<br>the column material, comprising longer chain<br>aliphatic and polycyclic aromatic material. |  |  |  |
| Index    | Aromaticity                            | Aromaticity provides an estimation of the degree<br>of aromatic and unsaturated structures of the<br>humic fraction.  |  |  |  |
|          | Molecular weight                       | A derived value of average molecular mass of the humic fraction.  |  |  |  |
|          | Inorganic colloids                     | Negatively charged inorganic polyelectrolytes,<br>polyhydroxides and oxidhydrates of Fe, Al, S or<br>Si, detected by UV light-scattering.   |  |  |  |
|          | SUVA                                   | An additional parameter derived from the ratio of DOC and spectral absorption coefficient.  |  |  |  |

Table2

Table 2: Average trace element concentrations for the irrigation water and Site 1 and 25. Irrigation contribution is the estimated concentration of elements from the irrigation water to the cave drip water after dilution by existing water stored in the vadose zone based on deuterium measurements.

|              | Ca     | Sr    | Ва    | Cu   | Ni   |
|--------------|--------|-------|-------|------|------|
|              |        |       |       |      |      |
|              | mg/L   | mg/L  | μg/L  | µg/L | µg/L |
| Irrigation   | 26.93  | 0.18  | 59.71 | 1.32 | 0.67 |
| Irrigation   |        |       |       |      |      |
| contribution | 0.90   | 0.006 | 2.00  | 0.04 | 0.02 |
| Site 1       | 97.68  | 0.07  | 10.77 | 0.90 | 0.26 |
| Site 25      | 116.73 | 0.06  | 8.77  | 0.21 | 0.05 |

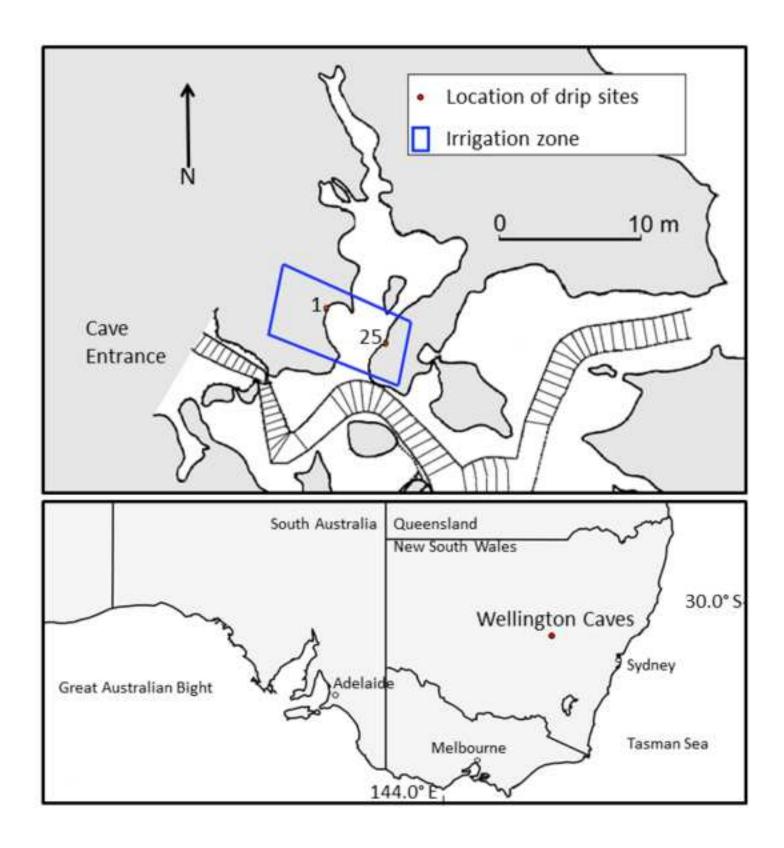


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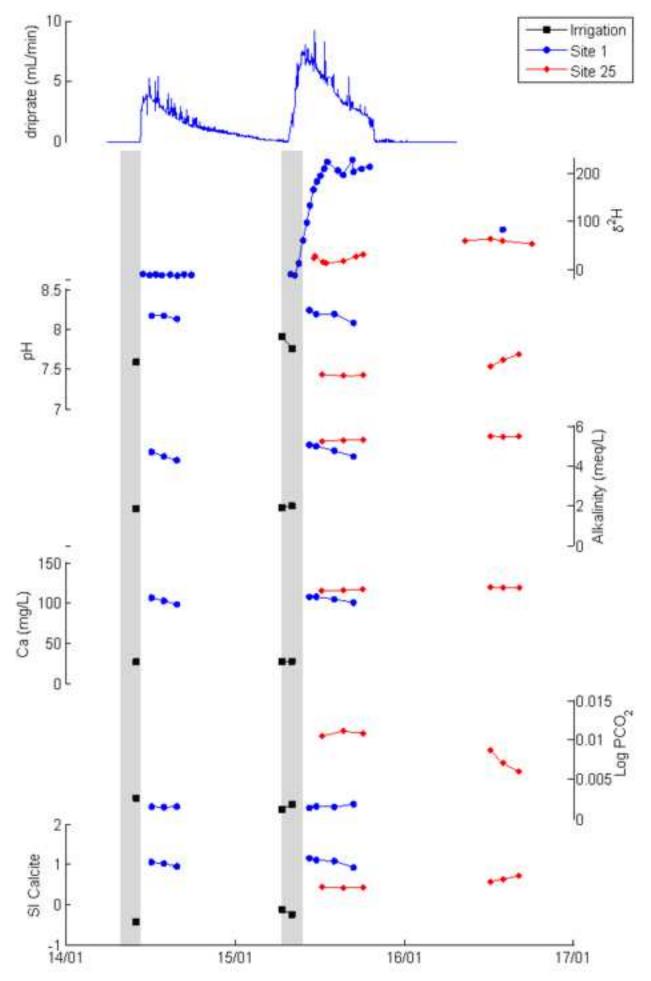


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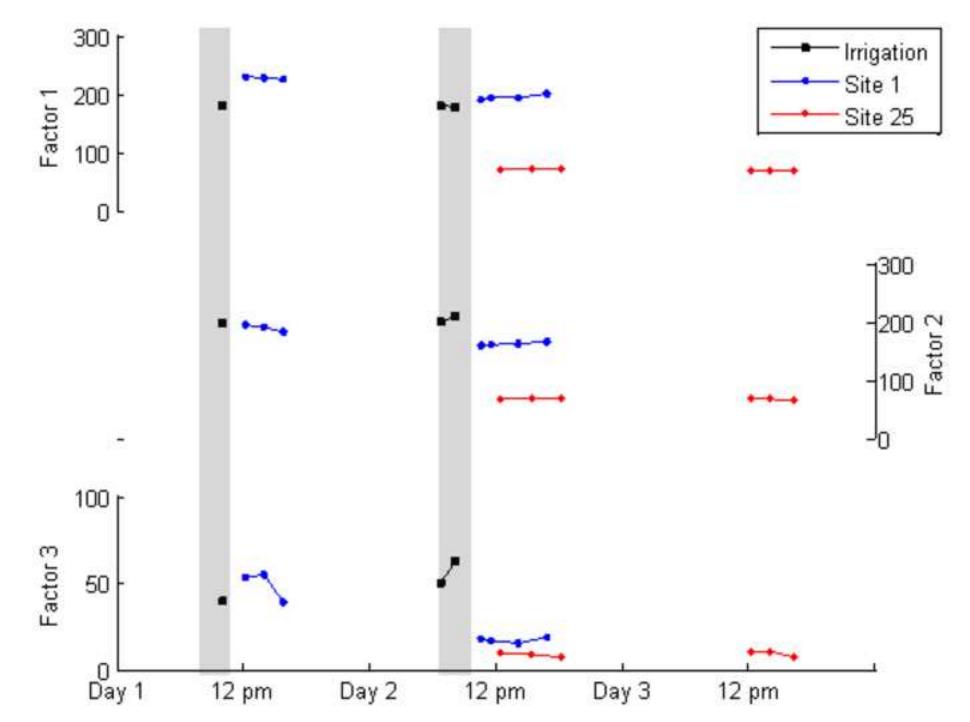
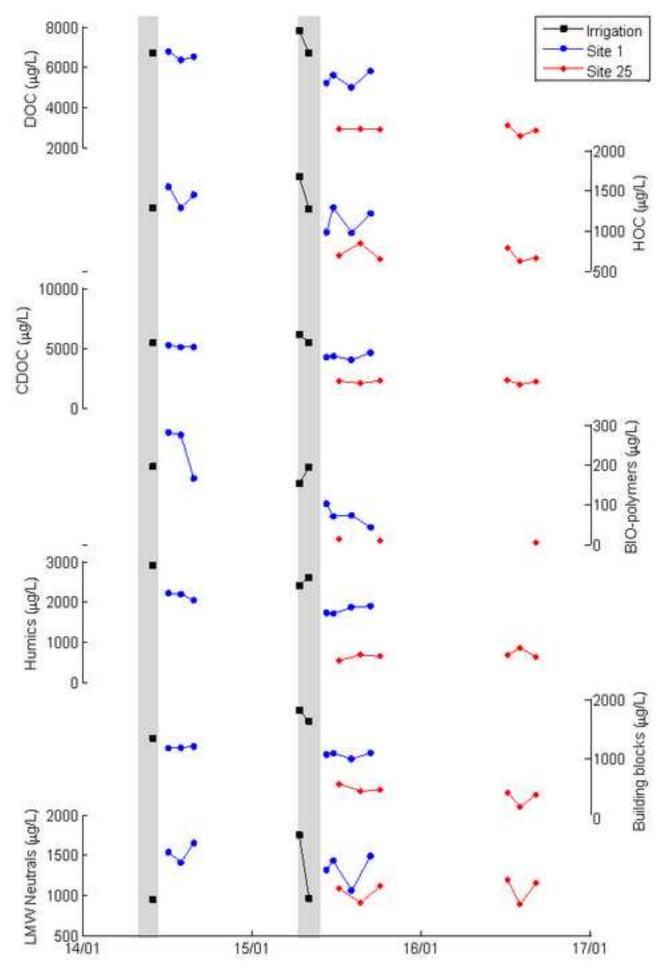
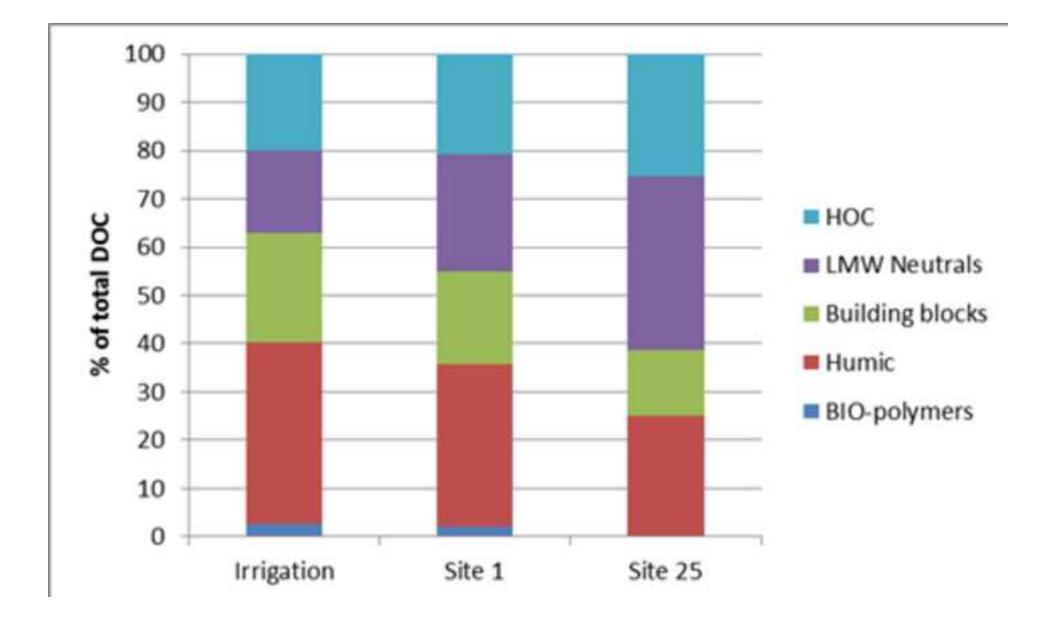
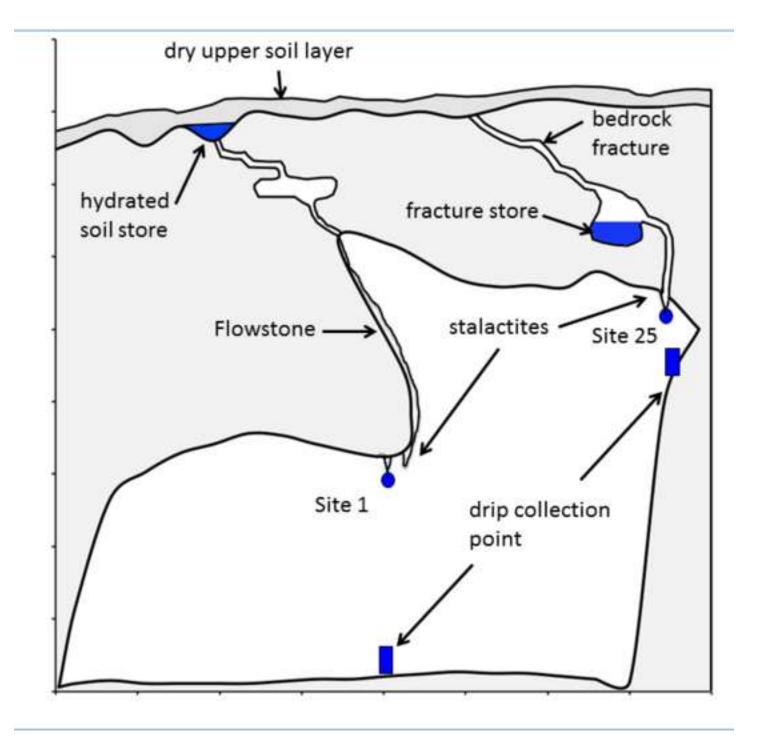


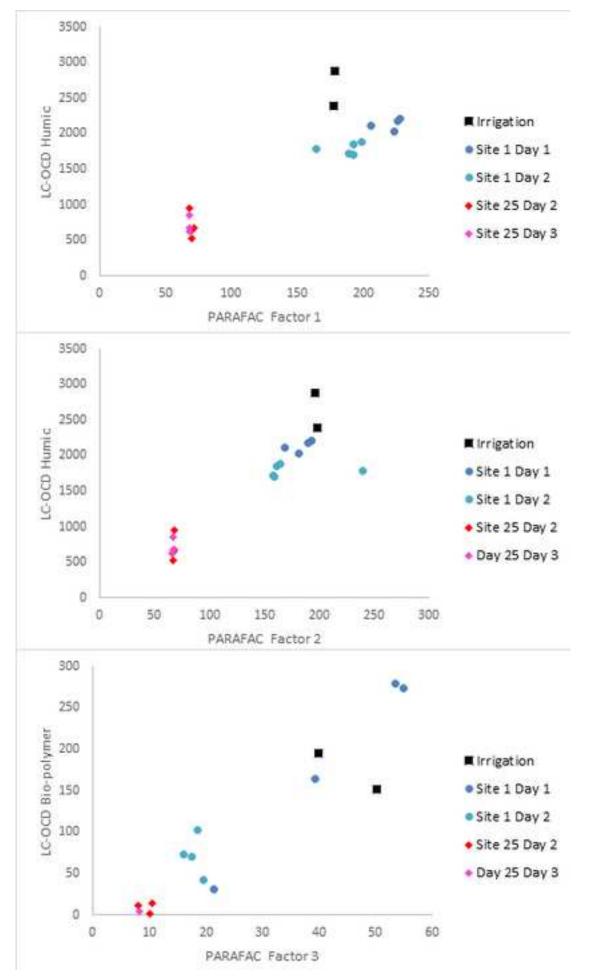
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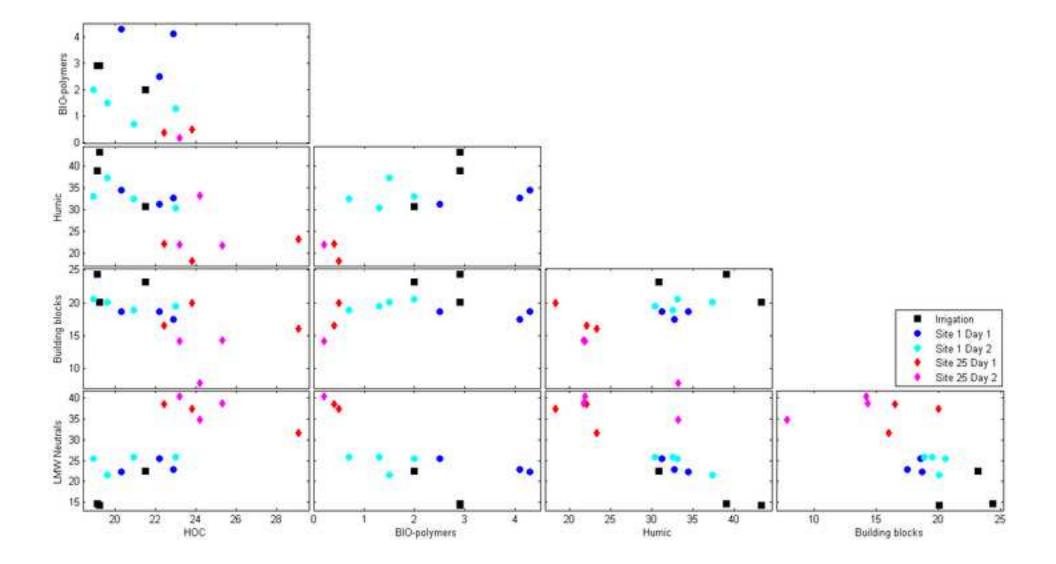


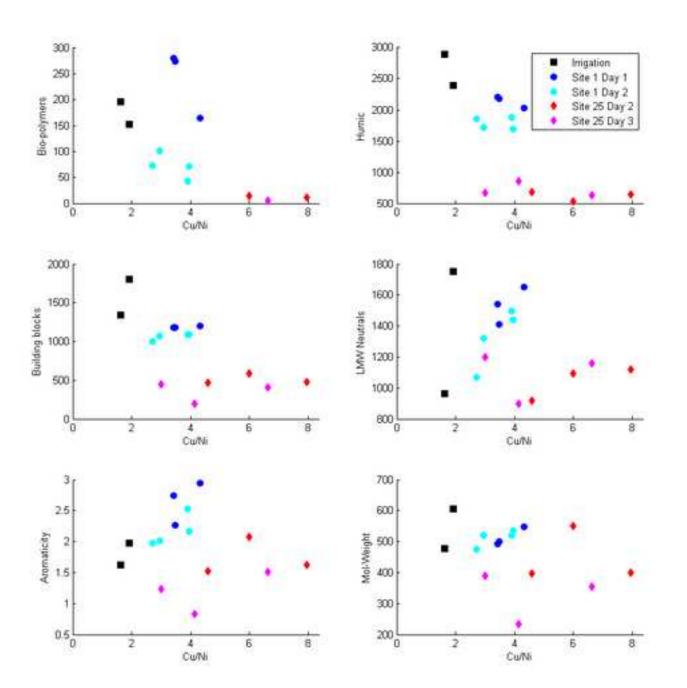












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