

## Review and Hypothesis. New insights into the reaction mechanism of transhydrogenase: Swivelling the dIII component may gate the proton channel

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## Review

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***Review and Hypothesis. New insights into the reaction mechanism of transhydrogenase: swivelling the dIII component may gate the proton channel.***

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### ***Abstract***

The membrane protein transhydrogenase in animal mitochondria and bacteria couples reduction of NADP<sup>+</sup> by NADH to proton translocation. Recent X-ray data on *Thermus thermophilus* transhydrogenase indicate a significant difference in the orientations of the two dIII components of the enzyme dimer (Leung, JH, Schurig-Briccio, LA, Yamaguchi, M, Moeller, A, Speir, JA, Gennis, RB and Stout, CD, 2015, *Science*, 347, 178-181). The character of the orientation change, and a review of information on the kinetics and thermodynamics of transhydrogenase, indicate that dIII swivelling might assist in the control of proton gating by the redox state of bound NADP<sup>+</sup>/NADPH during enzyme turnover.

## Introduction

Proton-translocating transhydrogenase is an important enzyme found in the inner membranes of animal mitochondria and the cytoplasmic membranes of bacteria. In the forward direction it catalyses  $\text{NADP}^+$  reduction by NADH coupled to inward proton translocation across the membrane. Thus, the enzyme is a proton motor in the same sense as the  $\text{F}_1\text{F}_0$ -ATP synthase.

Transhydrogenase in bacteria is one of several enzymes, often operating in parallel, which supply NADPH for biosynthesis. In *Escherichia coli*, for example, about one third of the NADPH needed for biomass production can be supplied by proton-translocating transhydrogenase [1, 2]. In some bacterial species over-expression of inserted transhydrogenase genes leads to commercially significant increases in the formation of fermentation products, such as lysine for animal feed, by increasing the rate of  $\text{NADP}^+$  reduction [3].

Transhydrogenase is strongly expressed in the mitochondria of cells in a range of animal tissues [4]. Again, other enzymes participate in the reduction of  $\text{NADP}^+$  in these cells and here even approximate estimates of the contribution of transhydrogenase to NADPH production are difficult to make [5, 6]. The NADPH produced by transhydrogenase may be particularly important in pathways responsible for the removal of damaging reactive oxygen species (ROS) [7]. Knockdown of the transhydrogenase gene in a human adrenocortical cell line resulted in elevated levels of ROS, and mutations of the gene were identified in individuals with familial glucocorticoid deficiency [8]. In an interesting parallel development it was reported that mouse strains carrying deletions in their transhydrogenase genes have impaired glucose-stimulated insulin secretion and glucose tolerance [9, 10]. It was proposed [11] that loss of transhydrogenase in mice prevents removal of superoxide radicals, and hence promotes activation of the mitochondrial uncoupling-protein 2 with consequent dissipation of the proton electrochemical gradient and inhibition of the ATP synthesis required for insulin secretion. Despite difficulties in properly establishing genetic backgrounds [12], the transhydrogenase-defective mouse strains may serve as useful models for understanding human type 2 diabetes.

***The dI, dII and dIII components of transhydrogenase.***

Transhydrogenase has three structural components (Fig.1). The hydrophilic components dI and dIII protrude from the membrane on the cytoplasm side in bacteria, and dII spans the membrane. The dI binds NADH (and product  $\text{NAD}^+$ ), dIII binds  $\text{NADP}^+$  (and product NADPH), and dII conducts protons across the membrane. Transhydrogenase has two dI-dII-dIII protomers. The joining of the polypeptide chains within a protomer varies amongst species (Fig.2) but this is unlikely to significantly influence the enzyme mechanism. While dII is always covalently linked to dIII (through the “hinge region”) the dI/dII and dIII/dI components may or may not be joined together through their polypeptide chains.

High-resolution structures of isolated dI [13-15] and dIII [16-18], and that of a dI-dIII complex [19-21], have been published during the last decade, and have afforded insights particularly into nucleotide binding and the hydride transfer step. The recent structure of the membrane-spanning dII at 2.8 Å resolution, and of the holo-enzyme at 6.9 Å from *Thermus thermophilus* [22], provide clues as to how hydride transfer from NADH to  $\text{NADP}^+$  at the interface of dI and dIII is coupled to proton translocation through dII. Distances are such that coupling must be mediated by conformational changes across the protein.

Isolated dI of transhydrogenase is a stable dimer [23]. Bound  $\text{NAD}^+$  (or NADH depending on crystallization conditions) is readily observed in the X-ray structures of isolated dI and dI-dIII complexes [13, 14, 19-21]. It occupies a classical nucleotide binding site in one of the two Rossmann folds that comprise the dI monomeric unit. Interactions between the protein and the ribose group of the nucleotide AMP moiety explain why the site strongly favours  $\text{NAD}^+$ /NADH binding over  $\text{NADP}^+$ /NADPH. The nicotinamide mononucleotide moiety of the  $\text{NAD}^+$ , which carries the redox-active nicotinamide ring, can adopt more than one conformation within the binding site [24]. Two conserved polypeptide loops of dI, the “mobile loop” and the “RQD loop” (Fig.1), interact with the bound nucleotide. The former, detectable in proton NMR spectra because of its segmental mobility [25], closes down on the surface of the protein following nucleotide binding. Mutations in the mobile loop of *Rhodospirillum rubrum* dI (*Rra*<sub>1</sub>M226F, *Rra*<sub>1</sub>T231C, *Rra*<sub>1</sub>G234A, *Rra*<sub>1</sub>Y235F, *Rra*<sub>1</sub>A236G, *Rra*<sub>1</sub>K237M – see Fig.2 for polypeptide nomenclature) lead to a decrease in NADH binding affinity and in the hydride-transfer rate [26]; its function may be to exclude water from the hydride-transfer site during catalysis. Mutations in the RQD loop of *R. rubrum* dI

(*Rra*<sub>1</sub>Q132N, *Rra*<sub>1</sub>D135N, *Rra*<sub>1</sub>S138A) cause only small changes in NADH binding affinity (*Rra*<sub>1</sub>R127A has a more pronounced effect) but all lead to very strong inhibition of hydride transfer [27]. X-ray structures of these mutant dI proteins indicate that residues in the RQD loop are involved in positioning the dihydronicotinamide ring of NADH for hydride transfer to NADP<sup>+</sup> bound to dIII.

Isolated dIII proteins are monomeric and have either tightly bound NADP<sup>+</sup> or NADPH depending on purification conditions. The dIII component comprises a single, classical Rossmann fold but the NADP<sup>+</sup> (or NADPH) is bound in a non-classical manner with a reverse nucleotide orientation [16, 17]. Favoured binding of NADP<sup>+</sup>/NADPH relative to NAD<sup>+</sup>/NADH is ensured by interaction between the 2' phosphate of the AMP moiety of the former and a conserved KRS motif in loop E. The central section of this loop arches over the pyrophosphate group of the bound nucleotide forming a "lid" (Fig.1). The protruding feature designated helix D/loop D is conformationally mobile [28], and along with the interacting loop E, is thought to have a central role in the transhydrogenase energy-coupling mechanism, perhaps in the crucial transition between the open and occluded states of the enzyme (see below). Mutations of *Ec*βK424, *Ec*βR425 and *Ec*βY431 in helix E and of *Ec*βD392 in helix D/loop D in the intact *E.coli* enzyme lead to inhibition of transhydrogenation activity [29-32]. Studies on these and other mutants in loop E and helix D/loop D of isolated *E.coli* dIII reveal further consequences of interactions of the protein with NADP<sup>+</sup> and NADPH [33, 34]. In crystal structures the equivalent residues to *Ec*βD392 form hydrogen bonds with the pyrophosphate and a ribose hydroxyl group of bound NADP<sup>+</sup>/NADPH [16, 17]. Its unusually high pK<sub>a</sub>, sensitive to the redox state of the bound nucleotide [35-37], and the fact that its substitution results in a failure in NADP<sup>+</sup>/NADPH binding [33] to isolated dIII, indicate an important catalytic role for this residue.

Until recently information on the structure of the transmembrane dII component of transhydrogenase was reliant upon amino acid sequence and biochemical analysis (see ref [38]. Thus, the single-subunit enzyme from animal mitochondria was thought to have 14 transmembrane helices (TM) per protomer, the two-subunit enzyme from *E.coli*, 13, and the three subunit enzyme from *R.rubrum*, 12 TM (Fig.2). Gene fusions during evolution were thought to increase the number of TM in the *E.coli* and mitochondrial-type enzymes [39, 40]. Consistent with these views, the new X-ray structure of dII from *T.thermophilus* transhydrogenase (a three-subunit enzyme) has 12 TM per protomer, 3 in the α<sub>2</sub> subunit, and

9 in the  $\beta$  subunit [22]. The most highly conserved of the transmembrane helices are TM2, TM3, TM4, TM9, TM10, and especially, TM13 and TM14 (for the TM numbering system, see Fig.2). The proton channel through dII (one per protomer) is thought to comprise the hexagram of TM3, TM4, TM9, TM10, TM13 and TM14 seen in the *T.thermophilus* crystal structure. Within the hexagram TM3, TM9 and TM13 form a central three-helix bundle enclosing a “tunnel”, perhaps the proton-translocation pathway itself, and the bundle is surrounded by TM4, TM10 and TM14. Six layers of amino acid residues are distinguished along the channel. Probably necessary for  $H^+$  transport, pairs of residues in layers 3 and 4 are hydrogen bonded. There are no ordered water molecules within the proton channel. The two protomers of the dII dimer interact through the respective TM2. In the holoenzyme crystal structure one dII protomer is slightly displaced relative to the other. Conservative single-site substitutions of only *Eca*H450 (in TM3), *Ec* $\beta$ H91 (TM9), *Ec* $\beta$ S139 (TM10), *Ec* $\beta$ D213 (in the cytoplasmic loop between TM12 and 13), *Ec* $\beta$ N222 (TM13), *Ec* $\beta$ G252 (TM14) and in a run of residues from *Ec* $\beta$ K261- $\beta$ R265 (in the hinge at the C-terminus of TM14 which connects dII to dIII – see Fig.1) of many examined in *E.coli* dII lead to strong inhibition of transhydrogenase activity [30, 31, 41-48]. *Ec* $\beta$ H91 has often provoked interest in that it is the only conserved protonatable residue close to the centre of the membrane dielectric whose substitution significantly deactivates transhydrogenation. However, in many species (including *T.thermophilus*) the His is replaced by an Asn (*Tt* $\beta$ N89); in some of these species the lost His is accompanied by the appearance of another in TM3 (as in *Tt* $\beta$ H42) though this is not always the case (we may speculate that transhydrogenases in this last category do not translocate  $H^+$  but perhaps  $Na^+$ ). Low concentrations of  $Zn^{++}$  block proton translocation through the *E.coli* dII channel; both FTIR [49] and X-ray absorption studies [50] implicate a His residue in the metal-ion binding but the role of  $\beta$ His91 in the process is not yet clear. The conserved salt bridge between *Tt* $\beta$ D202 and *Tt* $\beta$ R254 in the hinge, which very likely has an important role in catalysis [44], is located next to the proton channel at the membrane surface [22]. Differences in electron density of the hinge region suggest different conformations in the two protomers. Despite extensive mutagenesis experiments on the dII sequence the roles of individual amino acid residues in proton translocation and energy transduction have remained unclear. In Fig.3 the amino acid residue(s) responsible for proton binding in the dII channel, and the subsequent coupling to hydride transfer (see below), is labelled “X”.

### ***The dIII component of transhydrogenase swivels during turnover***



A most unexpected feature of the holo-enzyme X-ray structure confirmed by cross-linking inserted cysteine residues is that the dIII component of one protomer has a “face-up” orientation, in which its bound  $\text{NADP}^+$  is located close to the  $\text{NAD}^+/\text{NADH}$  binding site of dI, but the other dIII has a “face-down” orientation in which the bound  $\text{NADP}^+$  would be brought into the short cytoplasmic loops linking the TM of dII (Fig.1) [22]. Transhydrogenase is thus asymmetric, and its dIII components evidently swivel between face-up and face-down orientations during turnover. Modelling studies show that neither the holo-enzyme [22] nor dI-dIII complexes [19] can simultaneously accommodate two dIII components in the face-up orientation. The emerging view is that during operation transhydrogenase undergoes an alternation of sites: the reaction sequence in one protomer runs approximately  $180^\circ$  out-of-phase with that in the other (Fig.3).

Below we discuss the function of the dIII swivel in the context of the transhydrogenation mechanism. In accordance with earlier suggestions dIII in the face-up orientation is positioned for the enzyme to catalyse hydride transfer from  $\text{NADH}$  to  $\text{NADP}^+$  but in the light of the the new holo-enzyme structure we now propose that in its face-down orientation dIII is responsible for gating the proton channel in dII to give  $\text{H}^+$  access to one side of the membrane or the other.

### ***The binding-change mechanism of coupling to proton translocation in transhydrogenase***

The strongest evidence on the mechanism of action of transhydrogenase relates to the hydride-transfer step. This takes place with the dIII component in an “occluded state” in which both the binding and release of  $\text{NADP}^+$  and  $\text{NADPH}$  are extremely slow relative to enzyme turnover [51-53]. Crystal structures show that with dIII in the occluded state the *pro-R* hydrogen atom on C4 of the dihydronicotinamide ring of  $\text{NADH}$  (on dI) can be brought into close apposition with the *si* face of C4 of the nicotinamide ring of  $\text{NADP}^+$  (on dIII) to effect direct, stereo-specific and rapid hydride transfer [21]. The dIII component can also adopt an “open state”, where  $\text{NADP}^+$  and  $\text{NADPH}$  can rapidly bind and dissociate, respectively, but where hydride transfer between bound  $\text{NADP}^+$  and  $\text{NADH}$  is blocked [36, 54]. X-ray structures reveal nucleotide-binding conformations in which the block may be achieved; essentially the nicotinamide and dihydronicotinamide rings are held apart by the enzyme [24]. The pH dependences of transhydrogenation reactions in bacterial membranes



suggest that protonation of the enzyme converts open dIII into its occluded state and, subsequent to the hydride transfer step, deprotonation then converts occluded dIII back into its open state [51, 54, 55]. Thus, the basic mechanism is (i) substrate nucleotides,  $\text{NADP}^+$  and  $\text{NADH}$ , associate with their binding sites in an enzyme protomer having both dIII and dI in open states, (ii) protonation of dII from the outside aqueous phase converts the open dIII to occluded, (iii) hydride-ion equivalents are transferred from bound  $\text{NADH}$  to  $\text{NADP}^+$ , (iv) deprotonation of dII on the cytoplasmic side regenerates the open state of dIII, and (v) the nucleotide products,  $\text{NADPH}$  and  $\text{NAD}^+$ , are released. The binding affinities of  $\text{NAD}^+$  and  $\text{NADH}$  to dI are not significantly altered during the catalytic cycle; this enzyme component remains open except perhaps during the brief ( $< 10^{-3}$  s) period of hydride transfer [23, 56]. The proposed mechanism affords two important features. First, by confining hydride transfer to a form of the enzyme in which the nucleotides  $\text{NADP}^+$  and  $\text{NADPH}$  remain tightly bound, it prevents the redox step (potentially a facile reaction) from taking place without proton translocation (“slip”). Second, it permits the enzyme to adjust the binding properties of dIII for  $\text{NADP}^+$  and  $\text{NADPH}$  differently in the open and occluded states: in the open state to encourage substrate binding and product release, and in the occluded state to favour a large equilibrium constant for hydride transfer [57]. The consequent change in  $\text{NADP}^+/\text{NADPH}$  binding energy necessitates compensating changes in the equilibrium constants of steps associated with proton binding and proton release.

### ***Gating of the proton channel***

To provide directionality, for example to enable the enzyme to utilize a transmembrane proton electrochemical gradient and drive nucleotides to an elevated ratio of  $[\text{NADPH}][\text{NAD}^+]/[\text{NADP}^+][\text{NADH}]$ , the  $\text{H}^+$ -translocation reaction has to be appropriately gated: proton binding/release steps on either side of the membrane must be coordinated with the progress of the reaction [54]. In principle, the redox state of either  $\text{NAD}^+/\text{NADH}$  or  $\text{NADP}^+/\text{NADPH}$  bound to reaction intermediates could provide the trigger to allow proton access to or from one side of the membrane or the other. However, the dI component remains essentially in an open state during turnover of the enzyme (see above). Thus,  $\text{NAD}^+$  and  $\text{NADH}$  can rapidly bind to and be released from dI throughout the transhydrogenase reaction – they would be a poor trigger for controlling proton gating.  $\text{NADP}^+/\text{NADPH}$  are a much better proposition. In the framework of the proposed mechanism, gating would be effectively achieved if elements in the proton channel were to sense the redox state of

NADP<sup>+</sup>/NADPH bound to dIII, and adjust the local conformation to allow or disallow the passage of protons to or from the solvent. When NADP<sup>+</sup> is bound, the channel is able to access protons only from the outside, and when NADPH is bound, only from the cytoplasmic side. Thus, with NADP<sup>+</sup> in its dIII site H<sup>+</sup> binds into the channel from the outside (at relatively high proton electrochemical potential in respiring bacteria) generating the occluded state and, following reduction to NADPH, H<sup>+</sup> is released from the channel into the bacterial cytoplasm (at relatively low proton potential) regenerating the open state. A difficulty with this view was that previous predictions of the holo-enzyme structure based only on X-ray data on dI-dIII complexes had dIII in a face-up orientation [19, 20]. Thus, bound NADP<sup>+</sup> and NADPH were thought to be held some 20 Å or more from the proton channel. This arrangement is essential for direct hydride transfer from NADH (see above) but is not so well suited for gating: the “signal” relaying information on the redox state of the dIII nucleotide would have to be transmitted a considerable distance to the proton channel to control the gating.

The new structure of the *T.thermophilus* holo-enzyme reveals that the dIII component of one protomer is in the face-up orientation appropriate for hydride transfer; the other has a face-down orientation in which bound NADP<sup>+</sup> or NADPH would be brought into the short cytoplasmic loops linking the TM of dII [22]. In this latter conformation nucleotide influence on the structure of the proton channel is much easier to envisage. The (dihydro-) nicotinamide end of the bound nucleotide is proximal to the inter-TM loops. The positive charge on the nicotinamide ring of NADP<sup>+</sup> relative to the uncharged dihydronicotinamide ring of NADPH might thus be sufficient to switch the channel gates.

In Fig.3 the view that dIII swivelling regulates proton gating has been incorporated into the basic and well-supported transhydrogenase mechanism outlined above. At the end of catalytic turnover in a single protomer, dIII is face-down and in its open state (intermediate 1, either red or blue). Release of products NAD<sup>+</sup> and NADPH are followed by substrate NADH and NADP<sup>+</sup> binding. NADP<sup>+</sup> newly-bound to the face-down dIII (intermediate 2) triggers access of the proton channel to the outside aqueous phase. Protonation of X from the outside causes the dIII to become occluded and swivel into the face-up orientation to enable hydride transfer to its bound NADP<sup>+</sup> from NADH bound to dI (intermediate 3). The still-occluded dIII, now bearing NADPH (intermediates 4 and 5), then swivels back into the face-down orientation (intermediate 6). The bound NADPH in its face-down dIII triggers access of the

proton channel to the inside aqueous phase where  $H^+$  is released from X, and the dIII is converted back into the open state.

### *Alternation of sites*

It is recalled that the *T.thermophilus* holo-enzyme structure has one dIII in the face-up orientation and the other face-down (Fig.1), and that there is strong evidence that isolated dI-dIII complexes from *R.rubrum* have their dIII fixed in the face-up orientation, are occluded and are capable of rapid hydride transfer between bound nucleotides [19, 57, 58]. The events taking place during operation of the dimer then fall into place (Fig.3). The two protomers run approximately  $180^\circ$  out of phase: while one is completing the hydride-transfer step with dIII in an occluded state (intermediate 3  $\rightarrow$  4), the other is preparing for the replacement of bound product NADPH with substrate  $NADP^+$  in dIII in an open state (intermediate 6  $\rightarrow$  1). During the period in which proton binding from the outside aqueous phase is converting the open to the occluded state of dIII in one protomer (intermediate 2), proton release to the cytoplasm is converting dIII in the other from the occluded to the open state (intermediate 6).

The available experimental evidence on transhydrogenase would be satisfied if the two protomers in the figure operate independently of one another, only occasionally and briefly arrested to prevent both dIII components simultaneously adopting the clashing face-up orientation. However, this independent operation seems unlikely since then the enzyme would function more effectively as isolated protomers. It is more probable that events in the two protomers are conformationally linked to secure a kinetic advantage in the operation of the dimer. A conformational linkage between the protomers is most likely at steps 2  $\rightarrow$  3 and 5  $\rightarrow$  6 where proton binding/release, interconversion of the open/occluded states, and dIII swivelling all take place. Thus, the swivelling of face-up  $\rightarrow$  face-down orientations in one protomer could be compulsorily linked to face-down  $\rightarrow$  face-up swivelling in the other; and/or conformational changes resulting from proton binding in one protomer could be linked to conformational changes resulting from proton release in the other. These possibilities are not distinguished in the figure. In general, however, there are clear indications that structural changes in one protomer are linked to structural changes in the other – differences in the orientation of dIII, in the structure of the dII proton channel, and in the conformations of the hinge region, of the two protomers were all referred to above. Even in the isolated dI dimer the two  $NAD^+/NADH$  binding sites of the monomers adopt slightly different conformations,

one appropriately poised for hydride transfer, the other not [13]. It seems likely that cooperative interactions between the two protomers of transhydrogenase are central to its mechanism of action. It may also be noted that it has not been ruled out that the dimer could accommodate both dIII components in the face-down orientation, which might simplify our understanding of the transition, 5  $\rightarrow$  6, and that protonation/deprotonation of dIII itself is the cause of the occluded/open transition of this component.

The development of procedures for purifying and crystallizing the holoenzyme and its isolated components from the *T.thermophilus* transhydrogenase opens up ways to test predictions of the above model. Effort will be directed towards improving the X-ray resolution of the holoenzyme. Crucially, it is predicted that the open state of dIII will be stabilized in the face-down orientation of this component, and this will be reflected in changes in the structure of the NADP<sup>+</sup>/NADPH binding pocket. Interactions of the (dihydro)nicotinamide group of the bound NADP<sup>+</sup>/NADPH with amino acid residues in dII will test the prediction that the redox state of the nucleotide is involved in gating the proton channel. Changes in the structure of the hinge region are expected to have a central role in the swivelling of dIII. It is anticipated that working with low concentrations of Zn<sup>++</sup> and the construction of mutants in the hinge, in the dII proton channel and in the conformationally mobile region of dIII (for example at the equivalent of *Ec*βD392 – see above) might be required to lock the conformational state of the enzyme. The possibility of introducing specific mutations into just one protomer of the dimer should help to refine our understanding of the alternating-sites mechanism and identify points of conformational linkage. Changes in the binding of NADP<sup>+</sup>/NADPH are at the heart of the proposed mechanism; methods for the fluorescence detection of these changes have been developed for isolated dIII and should be adaptable for use in the holoenzyme.

### ***The different mechanisms of coupling to the proton electrochemical gradient in bioenergetic machines***

Although many features of the mechanism of coupling the proton electrochemical gradient to the chemical reaction catalyzed by transhydrogenase are far from clear, it is evident that the enzyme utilizes mechanisms unlike those of the other bioenergetic machines present in the mitochondrial inner membrane or in the bacterial membrane: NADH:quinone oxidoreductase (Complex I); quinol:cytochrome *c* oxidoreductase (Complex III or the *bc*<sub>1</sub>

Complex); cytochrome *c* oxidase (Complex IV); and the  $F_1F_0$ -ATP synthase (Complex V). The  $F_1F_0$ -ATP synthase couples proton flux across the membrane through two half-channels to conformational changes linked to rotatory motion and a three-fold alternating sites mechanism of ATP synthesis [59]. Complex I also conveys protons across the membrane via four sets of half-channels whose opening/closing conformations are gated by the chemistry occurring at the single distant quinone binding site [60]. The chemistry of oxygen reduction by cytochrome *c* oxidase is used to gate proton channels and alter proton affinities resulting in proton pumping, but conformational changes must be subtle compared to either Complex I or the ATP synthase [61]. Complex III has a unique “Q-cycle” to move charges across the membrane and a large conformational change of the Rieske Fe-S subunit to assure the two electrons from quinol are bifurcated, i.e., directed to different electron acceptors [62]. The enzymology of each of these systems is distinct from the others and each is a remarkable testament to the different ways that evolution has solved the problem of both generating and utilizing the proton motive force across a biological membrane. We can now add the transhydrogenase to this list of amazing molecular machines at the heart of bioenergetics.

### Figure Legends

#### **Fig.1. The three-dimensional structure of transhydrogenase from *T.thermophilus*.**

Modified from ref [22]. The gray bars indicate the position of the membrane. The dashed lines delineate the short polypeptide hinges of presently unknown structure that link dII and dIII. NAD(H) and NADP(H) correspond to nucleotides bound in the  $NAD^+/NADH$  and  $NADP^+/NADPH$  sites, respectively. See text for comments on secondary-structure features and on the orientation change of dIII.

#### **Fig.2. The four types of polypeptide organization of transhydrogenases from representative species.**

*Tt*, *Thermus thermophilus*; *Rr*, *Rhodospirillum rubrum*; *Ec*, *Escherichia coli*; *Bt*, *Bos taurus*; *Eh*, *Entamoeba histolytica*. The horizontal lines correspond to polypeptide chains. The labels  $\alpha$ ,  $\alpha_1$ ,  $\alpha_2$  and  $\beta$  are names of the polypeptide chains (but sometimes also called PntA, PntAA, PntAB and PntB, respectively). The components dI, dII and dIII, and the

hinge region, are common to all 4 types of polypeptide organization. The small black rectangles represent transmembrane helices (TM) numbered according to predictions on bovine transhydrogenase but to date only visualised in X-ray structures of the *T.thermophilus* enzyme [22]: question marks adjacent to the number of TM in other species show that estimation is based only on prediction and limited biochemical experiments.

**Fig.3. Proposed mechanism of coupling to proton translocation in transhydrogenase.**

Each panel diagrammatically shows an intermediate in the transhydrogenase dimer as it runs in the forward direction. One protomer of the dimer is coloured red, the other blue. The relative positions of the dI, dII and dIII components correspond to those seen in the X-ray structure of the *T.thermophilus* holo-enzyme (PDB, 4O9U) – see Fig.1. The dI component is shown as a rectangle, dII as a rounded rectangle spanning the membrane, and dIII as a trapezoid with its NADP<sup>+</sup>/NADPH binding site located at the broader end. In the face-up orientation of dIII (see text) the NADP<sup>+</sup>/NADPH binding site is located next to the NAD<sup>+</sup>/NADH binding site of dI; in the face-down orientation the NADP<sup>+</sup>/NADPH binding site is directed into the cytoplasmic loops between the TM of dII. The dI and dIII components and X, a proton binding site in the dII channel (see text), are shown bounded by dashed lines in open states, and solid lines in occluded states. The pairs of parallel dashed lines in dII show allowed proton access along the channel between X and the aqueous phase on one side of the membrane or the other. The six reaction intermediates (1-6) run approximately 180° out-of-phase in the two protomers. Thus, intermediates 1, 2, 3, 4, 5 and 6 in one protomer are paired with intermediates 4, 5, 6, 1, 2 and 3, respectively, in the other. Hydride transfer from NADH to NADP<sup>+</sup>, only possible with dIII in its occluded face-up orientation, is highlighted with a green arrow. For simplicity, NAD<sup>+</sup> is shown as dissociating, and NADH as binding, only to intermediate 1; in fact, NAD<sup>+</sup> and NADH can probably reach equilibrium binding with all intermediates on the reaction path (see text). The reverse transhydrogenation reaction, oxidation of NADPH by NAD<sup>+</sup> coupled to H<sup>+</sup> efflux, is described essentially by a simple reversal of the reaction sequence shown in this figure.

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Figure 1

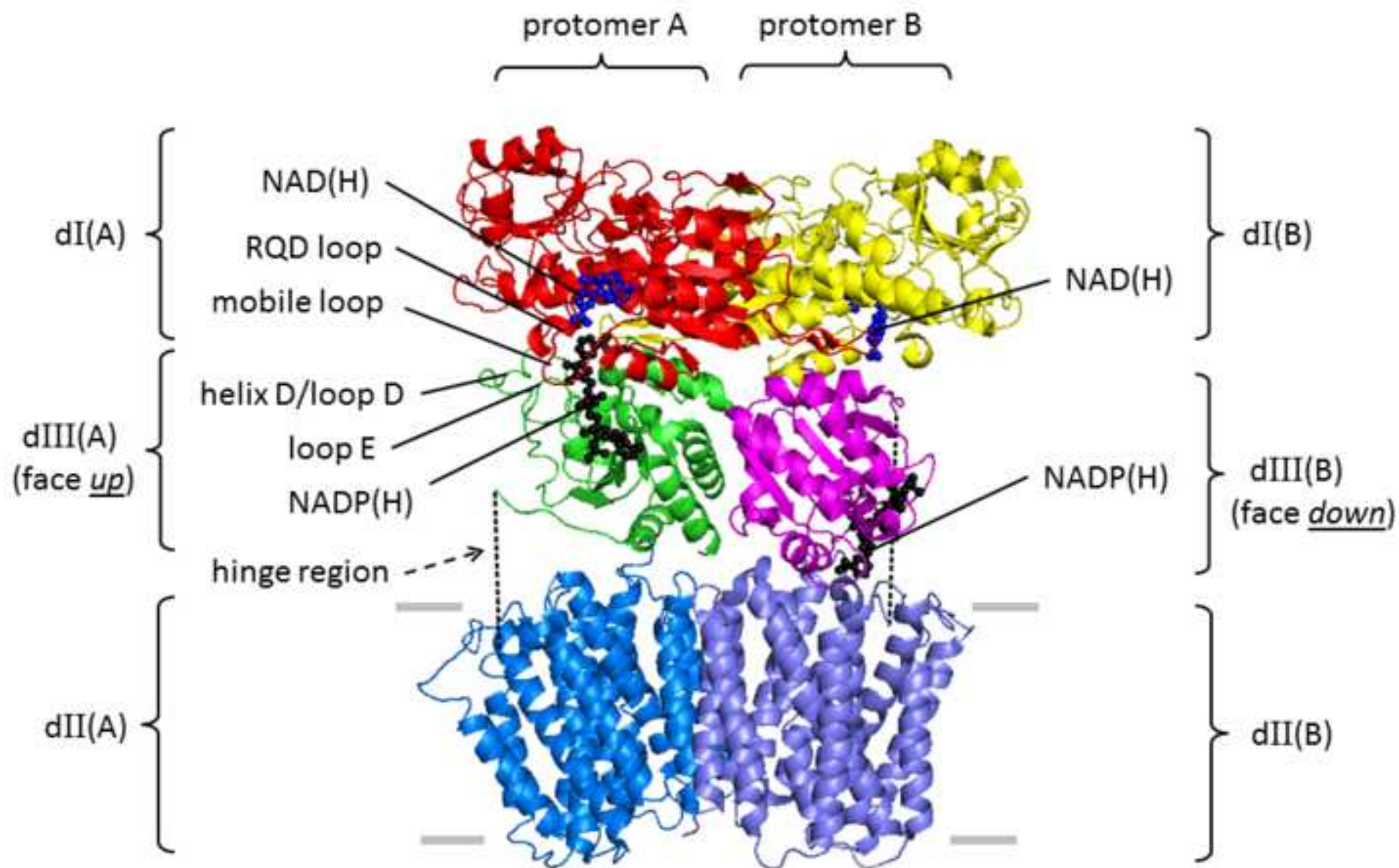


Figure 2

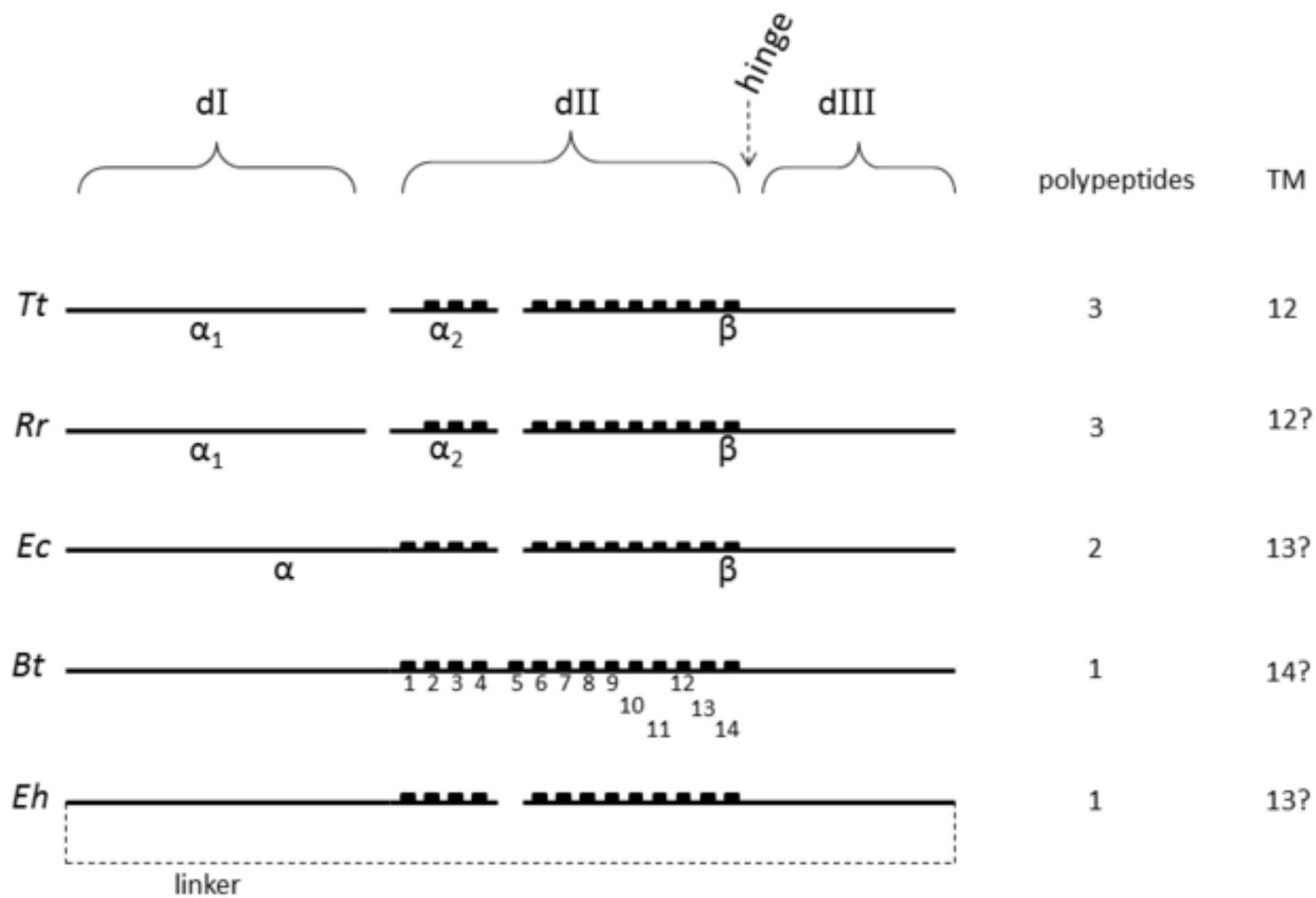


Figure 3

