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Prospects for developing allergen-depleted food crops

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The Plant Genome 2222 6

SPECIAL ISSUE: GRAING QUALITY AND NUTRITIONAL GENOMICS FOR BREEDING NEXT GENERATION CROPS

Prospects for developing allergen-depleted food crops

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Abstract

In addition to the challenge of meeting global demand for food production, there are increasing concerns about food safety and the need to protect consumer health from the negative effects of foodborne allergies. Certain bio-molecules (usually proteins)

Abbreviations: AI, artificial intelligence; ATIs, α-amylase/trypsin inhibitors; BAT, basophile activation tests; CAGR, compound annual growth rate; CRISPR, clustered regularly interspaced short palindromic repeats; FA, food allergy; Ig, immunoglobulin; NIAID, national institute of allergy and infectious diseases; nsLTPs, nonspecific lipid transfer proteins; PR, pathogenesis-related proteins; RNAi, RNA interference; SDN, site-directed nuclease; WHO/IUIS, World Health Organization and International Union of Immunological Societies.

Lokya Vadthya and Seial Parmar contributed equally to this work.

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present in food can act as allergens that trigger unusual immunological reactions, with potentially life-threatening consequences. The relentless working lifestyles of the modern era often incorporate poor eating habits that include readymade prepackaged and processed foods, which contain additives such as peanuts, tree nuts, wheat, and soy-based products, rather than traditional home cooking. Of the predominant allergenic foods (soybean, wheat, fish, peanut, shellfish, tree nuts, eggs, and milk), peanuts (Arachis hypogaea) are the best characterized source of allergens, followed by tree nuts (Juglans regia, Prunus amygdalus, Corylus avellana, Carya illinoinensis, Anacardium occidentale, Pistacia vera, Bertholletia excels), wheat (Triticum aestivum), soybeans (Glycine max), and kidney beans (Phaseolus vulgaris). The prevalence of food allergies has risen significantly in recent years including chance of accidental exposure to such foods. In contrast, the standards of detection, diagnosis, and cure have not kept pace and unfortunately are often suboptimal. In this review, we mainly focus on the prevalence of allergies associated with peanut, tree nuts, wheat, soybean, and kidney bean, highlighting their physiological properties and functions as well as considering research directions for tailoring allergen gene expression. In particular, we discuss how recent advances in molecular breeding, genetic engineering, and genome editing can be used to develop potential low allergen food crops that protect consumer health.

1 | INTRODUCTION

Food allergy (FA) is a type of food sensitivity that has become a critical public health concern globally (Loh & Tang, 2018). FA describes the adverse immune responses to certain foods that can occur in the human body (Burks et al., 2012). The prevalence of FA has increased in recent decades, challenging both allergists and food scientists to devise rapid and accurate diagnostic tests, as well as prevention and treatment measures for vulnerable people. FA has become a global food safety issue because of the lack of reliable preventive measures, except for desensitization by immunotherapy (Du Toit et al., 2015) and the use of adrenaline injections for anaphylactic reactions. FA negatively impacts the life of sensitive individuals because of the absence of effective allergen elimination methods.

The generally accepted definition of the term "FA" is an "adverse health effect arising from a specific immune response that occurs reproducibly on exposure to a given food," as described by the National Institute of Allergy and Infectious Diseases (NIAID) guidelines (Panel, 2010). In general, allergic disease manifestations are initiated through sensitization in early life and thereafter often progress into atopic dermatitis, asthma, allergic rhinitis, and other symptoms of FA. When a susceptible individual is exposed to food allergens for the first time, the offending food protein is identified by the body as a "foreign," antigen. This results in the production of immunoglobulin E (IgE) antibodies, resulting in a "sensitization," a state which results in

no allergic symptoms. However, upon subsequent exposure, the specific allergens interact with the IgE molecules on the surface of tissue mast cells and basophils in the blood and trigger the release of a range of defense compounds, including histamine, prostaglandins, and leukotrienes, which together produce allergy symptoms (Figure 1; Sicherer & Sampson, 2014).

Food allergens are naturally occurring protein molecules that possess specific immunological characteristics and trigger inappropriate immune responses in susceptible individuals. In contrast, exposure of nonallergenic individuals to such food proteins in the gastrointestinal tract results in "oral tolerance" by producing protein-specific IgG/IgA antibodies that develop immune response (Du Toit et al., 2015). Oral tolerance is a process of active suppression of the inappropriate immune responses to the first encounter with food protein antigens in the gastrointestinal tract (Tordesillas & Berin, 2018). Oral tolerance is considered to be the general homeostatic state, while FA is the exception. The balance between oral tolerance and FA can be influenced by several factors, such as genetic disposition, the dose, route and time of antigen exposure, dietary factors, and gastrointestinal microbiota (Keet & Wood, 2011). However, the complex, precise mechanism of oral tolerance and allergic sensitization to food proteins remains to be understood.

The most common FA sources are of plant and animal origin, such as peanuts, tree nuts, fruits, wheat, soybean, kidney beans, seafood, eggs, milk, fish, honey, and other unidentified sources (Figure 2). Among different allergenic foods,

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peanuts cause the most common food allergies associated with severe anaphylaxis in children and adults and persist lifelong, which is uncommon in the case of other food allergies (Lieberman et al., 2020; Sicherer & Sampson, 2010). The allergic reaction can be unpredictable, such that the severity of health conditions ranges from mild to life threatening, the latter requiring immediate medical attention. Based on the immunological responses involved, FAs are classified as (i) IgE-mediated, also called immediate hypersensitivity, (ii) non-IgE- or cell-mediated, also called delayed hypersensitivity, and (iii) a combination of IgE- and non-IgE-mediated allergic reactions (Tordesillas et al., 2017). Of these, IgEmediated FA is the most common type of allergy that is caused by the consumption of foods such as peanuts, tree nuts, wheat, shellfish, fish, eggs, milk, and soybeans and that can cause potentially fatal anaphylaxis (Wasserman et al., 2018). This involves an immediate Type I hypersensitivity reaction that develops within minutes to >2 h of ingestion of the offending food. The clinical manifestation of anaphylaxis involves multiple organs, such as the skin, gastrointestinal tract, respiratory tract and cardiovascular system. The most common symptoms associated with anaphylactic shock are abdominal cramps, nausea, vomiting, diarrhea, hives, itching, eczema, wheezing, and coughing, which can induce death due to respiratory and cardiovascular complications (Wasserman et al., 2018). In contrast, non-IgE-mediated FA does not involve IgE antibodies. In this case, symptoms often develop after 48-72 h of ingestion of offending foods such as cow's milk, soy, and wheat proteins. Non-IgE-mediated FA is usually resolved within 1–5 years. However, the mechanisms underlying non-IgE-mediated food allergic reactions are poorly understood perhaps because it is less harmful than IgE-mediated anaphylaxis (Cianferoni, 2020).

Large numbers of allergenic food proteins have been identified and characterized, with specific details maintained in databases that are designed to provide easy access to comprehensive information. The "World Health Organization and International Union of Immunological Societies (WHO/IUIS) Allergen Nomenclature Database" was established at the millennium. The database contains a systematic nomenclature of allergenic proteins, as reviewed by the experts of the WHO/IUIS Allergen Nomenclature Sub Committee. Each entry in this database provides details on amino acid sequences, biochemical properties, and allergenicity. This reference database provides a crucial knowledge repository for the research community, clinicians, and regulatory authorities (Pomes et al., 2018). To date, on August 11, 2023, a total of 1015 (http://bioinfo. unipune.ac.in/AllerBase/PHP_codes/BrPlant.php) allergens have been experimentally validated from different plant sources and 1042 allergens from animal sources. This information is curated in the allergen database "Aller-Base" (bioinfo.unipune.ac.in/AllerBase/Home.html), which provides data on allergens and their associated basic fea-

Core Ideas

- The severity and prevalence of food allergens varies between different geographical regions.
- Scope for developing hypoallergenic crops with minimal effects on plant physiology.
- Identification of crops with reduced allergen content through selection and breeding.

tures, such as IgE binding epitopes, IgE antibodies, and cross-reactivity. Inaddition, each allergen is curated with a cross-reference to sequence and structure databases along with other allergen databases, such as the WHO/IUIS-Allergen nomenclature database (www.allergen.org), Allergome (www.allergome.org), and AllFam (www.meduniwien. ac.at/allfam/) (Kadam et al., 2017). Understanding the correlation between allergen components in the food source and clinical symptoms has gained importance in the development of more accurate diagnostic methods which further help in the management of food allergies (Maruyama, 2021). Therefore, herein we have comprehensively described the prevalence of allergies associated with major plant food sources such as peanut (Arachis hypogaea), tree nuts (Juglans regia, Prunus amygdalus, Corylus avellana, Carya illinoinensis, Anacardium occidentale, Pistacia vera, Bertholletia excels), wheat (Triticum aestivum), soybean (Glycine max), and kidney bean (Phaseolus vulgaris) and also highlighted their physicochemical properties, functions as well as proposed future research directions for developing low allergenic plant food.

2 | GLOBAL DISTRIBUTION OF FOOD ALLERGIES

The severity and prevalence of FAs vary between different geographical regions. According to WHO, the prevalence of FA has increased over the last two decades in industrialized countries, with similar trends observed in developing countries as their economic growth increases (Leung et al., 2018; Prescott et al., 2013). FA currently affects millions of people leading to restrictions in diet and daily activities, with profound emotional impacts, as well as healthcare and economic costs, lowering the overall quality of life as well as physical health and well-being (Lieberman et al., 2020). Although the exact prevalence of FA is uncertain, it is estimated that over 220 million people suffered from some sort of FA globally (Sicherer & Sampson, 2018). The general consensus is that up to 10% people are affected by food allergies and that they are more common in young children than in adults (Loh & Tang, 2018). According to FAIR Health estimates, 5.9 million American children under the age of 18

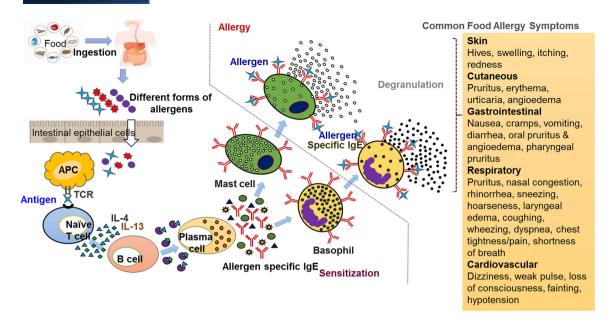


FIGURE 1 A pictorial representation of series of reactions involved in allergic immune response and most common allergic symptoms.

have FA, which accounts for one in every 13 children or two in every classroom. In the case of Australia, the pediatric allergy prevalence ranges from 3.8 to 11% in children under the age of 4 years (Peters et al., 2017). Approximately 10% of Chinese children under the age of 6 years are also prone to FA (http://www.bjreview.com.cn/nation/txt/ 2015-06/01/content 690263.htm). Similarly, the European Academy of Allergy and Clinical Immunology estimated that the allergy prevalence in European children is between 1.7 and 5%, depending on the country. It is matter of great concern that the majority of children in Australia, Italy, America, China, and Europe are predisposed to FA, followed by other nations, as illustrated in the global map (Figure 3). The incidence of peanut allergy is more prevalent in children in the United Kingdom, North America, and Australia. In these countries, the reported prevalence of peanut allergy has doubled over the last decade. The prevalence of FA is also increasing rapidly in Asian countries, correlating with increasing economic growth and westernization (Arakali et al., 2017). It is interesting to note that the prevalence of peanut allergies is lower in Asian countries; but have a prevalence of allergies to fish. The developing nations in Asia and Africa have gradually adopted a Western lifestyle and have observed increasing rates of allergic disease across age groups, particularly in younger children.

Obtaining accurate FA incidence data remains a major challenge in many parts of the world. Problems arise because of difficulties in data curation or a lack of consistency in study design, approach, specific population analysis, specific foods, and even the definition of FA, factors that together lead to a poor establishment of accurate prevalence statistics (Prescott et al., 2013). Currently, available statistical data are highly

biased not least because they rely heavily on self-reporting of FA incidence rather than challenge-confirmed FA analysis. Such factors may result in over estimation of the allergy prevalence, often because of patient's misunderstanding about wide range of symptoms that are associated with particular food allergies (Woods et al., 2002). Researchers in westernized countries have tested numerous methods to determine the precise number of people that are allergic to specific foods (Prescott et al., 2013). Hospital intake data that include healthcare burden, insurance claims, and telephone surveys is the most frequently used method of evaluation of prevalence rates (Gupta et al., 2018a). However, comprehensive studies are needed in the future to demonstrate a precise correlation between different geographic regions and specific allergy prevalence. Moreover, the instigation of common dedicated databases at the clinic/hospital level is vital for recording the incidence of food allergies.

2.1 | Specific regions and populations affected

There is a paucity of reliable data concerning FA prevalence, even in developed countries, and so it is difficult to compare prevalence rates between countries (Prescott et al., 2013). Significant variations in prevalence and causative foods have been reported. For example, in continental Europe, adults are predominantly allergic to peanuts, tree nuts, fruits, and vegetables but children are allergic to cow's milk, peanuts, and eggs (Baker, 2018). Moreover, kidney beans are the major allergic triggers of asthma and rhinitis patients in India, followed by chickpeas and peanuts (Kasera et al., 2011), whereas

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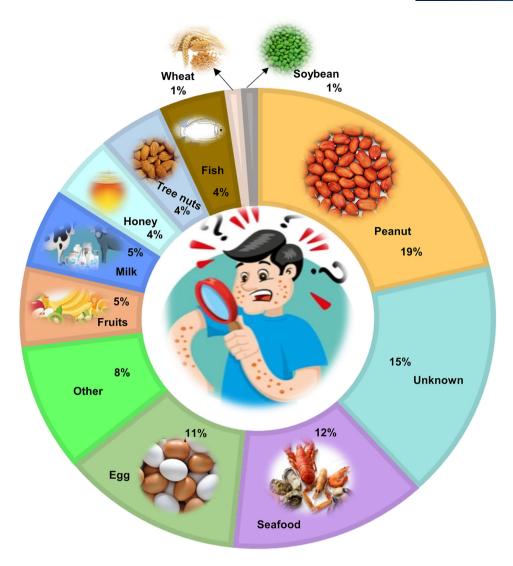


FIGURE 2 A pictorial representation of allergy composition associated with different foods of plant and animal origin and other unidentified sources.

Scandinavians have an allergic sensitivity to shellfish and cod.

The overall prevalence of self-reported FA in Europe is 6%, the most common causative foods being cow's milk, eggs, peanuts, tree nuts, wheat, soy, fish, and shellfish (Lyons et al., 2019). A systematic review of studies made between 2000 and 2012 revealed that the self-reported lifetime FA prevalence of different plant food sources, such as wheat, tree nuts, peanut, and soy, was 3.6, 2.2, 1.3, and 0.4%, respectively. In contrast, the prevalence of food-challenge-defined allergies to wheat, tree nuts, peanut, and soy was 0.1, 0.5, 0.2, and 0.3%, respectively (Nwaru et al., 2014). It has been estimated that >17 million people are currently affected by FA, of which 3.5 million are under 25 years of age (Baker, 2018). The average healthcare cost of an allergic adult is estimated to be I\$2016, compared with the cost of a healthy adult, which is I\$1089 per year (Fox et al., 2013).

Approximately 10% of the US population suffers from at least one form of IgE-mediated FA, which affects about one in 10 adults and one in 13 children (Gupta et al., 2018a, 2019; Warren et al., 2020). The most common allergic foods include peanuts and tree nuts, which are associated with severe allergic reactions in all age groups. According to the FAIR health insurance claims-based study, the incidence of severe anaphylactic reactions increased by 377% between 2007 and 2016 due to the consumption of peanuts, tree nuts/seeds, and other specific foods (Motosue et al., 2018). Peanut is a major contributor to life-threatening anaphylaxis (26%), followed by tree nuts/seeds (18%), eggs (7%), crustaceans (6%), milk products (5%), fruits and vegetables (2%), and other specific foods. A population-based survey involving 40,443 US adults (Gupta et al., 2019) suggested that 10.8% (equivalent to >26 million people) have some form of FA, while 19% of adults believed themselves to have food allergies. Shellfish allergies

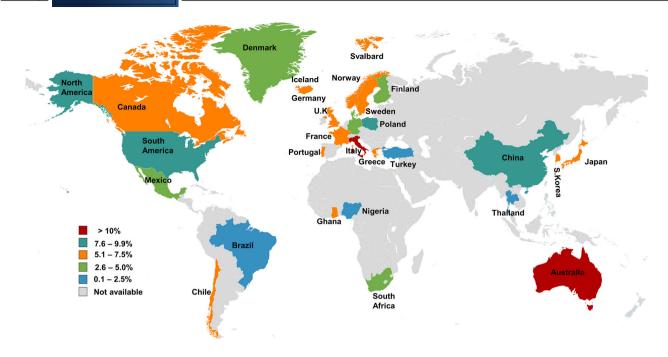


FIGURE 3 Global distribution of pediatric food allergy mostly children under the age of 5 years. *Source*: Center for Food & Asthma Research (CFAAR), Northwestern Feinberg School of Medicine, Ann & Robert H. Lurie Children Hospital of Chicago. A global map was obtained and modified to represent the current scenario of pediatric allergy.

are particularly predominant in US adults, accounting for 7.2 million people (2.9%), followed by milk (4.7 million = 1.9%), peanut (4.5 million = 1.8%), tree nuts (3.0 million = 1.2%), finfish (2.2 million = 0.9%), egg (2.0 million = 0.8%), wheat (2.0 million = 0.8%), soy (1.5 million = 0.6%), and sesame (0.5 million = 0.2%)-associated allergies. However, children appear to be more sensitive to peanuts, followed by milk, shellfish, tree nuts, eggs, finfish, wheat, soy, and sesame seeds (Gupta et al., 2011). The big eight foods and their specific allergy prevalence in children are as follows: peanut (25.2%), milk (21.1%), shellfish (17.2%), tree nuts (13.1%), egg (9.8%), finfish (6.2%), wheat (5%), and soy (4.6%) (Figure 4). The prevalence of FA in children aged 0-17 years increased by 50% from 1997 to 2011. This trend is similar to the increasing trend in increased income levels (Jackson et al., 2013). The overall economic cost of FA was estimated with be US\$24.8 billion annually, which is equivalent to \$4184 per child (Gupta et al., 2013).

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The European Academy of Allergy and Clinical Immunology estimated the allergy prevalence in children to be 5% in France, the United Kingdom, Germany, and the Netherlands, 4% in Italy and Spain and 1.7% in Greece. In the case of the UK, 2 million people (1–2% of adults) and 5–8% of children exhibit food allergies. In Europe, there are 14 foods, which include celery, cereals, crustaceans, eggs, fish, lupin, milk, mollusks, mustard, peanuts, sesame, soybean, sulfur dioxide, and sulfites, which have been identified as major potent and prevalent allergens. In Australia, approximately 2% of adults and 4–8% of children have food allergies and there

has been a fourfold increase in anaphylaxis hospitalizations (Tang & Mullins, 2017). The Health Nuts population-based cohort study reported the highest prevalence of challenge-confirmed FA in Australia. Overall 11% of Australian children at the age of 1 year and 3.8% at the age of 4 years had FA. Peanut was the predominant allergen at 1.9%, followed by egg (1.2%) and sesame (0.6%) in children at 4 years of age (Peters et al., 2017). Peanut allergy, which affects about 2% of the population of Western nations, has become a burden of self-management to protect against accidental exposure.

The overall prevalence of FA in Asia follows similar trends but there are significant differences in the types of reported FA. Shellfish allergy is more prevalent than peanut allergy in Asia (Lee et al., 2013). There is an increasing prevalence of FA reported in children in developing parts of Asia, including Australia, Japan, China, and Korea. Between 1 and 2% of adults and 5% of children report FA in China (Poulos et al., 2007; Prescott et al., 2013). The Population Reference Bureau, China reported that 3.8-7.7% of children have food intolerances (Hu et al., 2020). This report identified shellfish, peanuts, soybeans, wheat, tree nuts, fish, eggs, and milk as a major source of allergens. Many prepacked foods contain these constituents and they are sold with proper labeling and instructions (Baker, 2018). Most allergic diseases in India are caused by pollen grains, fungal spores, insects, and foods. About 25% of the Indian population shows sensitivities to these different forms of allergens (Bhattacharya et al., 2018). The EuroParvall-INCO survey conducted by Li et al. (2020) revealed that IgE-mediated FA prevalence in children

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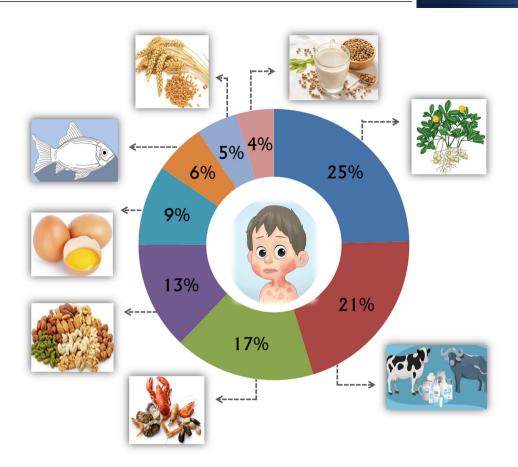


FIGURE 4 Prevalence of specific food allergy in allergic children of United States. Among the different food sources, peanut, milk, shellfish and tree nuts hold a major stake in contributing toward triggering life-threatening hypersensitivity reactions in children.

is highest in Hong Kong (1.5%), followed by Russia (0.87%), Guangzhou (0.21%) and Shaoguan (0.69%) of China, and India (0.14%). A similar study conducted on adults in Karnataka, India, revealed a high rate (26.5%) of sensitization but a low rate (1.2%) of probable FA (Mahesh et al., 2016). These studies suggest there is little correlation between foodspecific IgE sensitization and probable food allergies. This finding highlights the role of other important factors involved in the clinical manifestation of allergies.

The prevalence of food allergies varies between different ethnic groups. For example, children in Ghana (5–16 years of age) are commonly allergic to pineapple, pawpaw, orange, mango, and peanut (Obeng et al., 2011), while children in North America are allergic to peanut, milk, egg, shellfish, and soybean (Hill et al., 2016). Similarly, African-American children have a higher prevalence than their Caucasian counterparts (Gupta et al., 2018a). While Asian-American children have the lowest reported FA, Caucasians have the highest rates of diagnosed FA (Gupta et al., 2011). Additionally, the prevalence of FA varies between races, as well as rural and urbanized populations. For example, white adults in the US report lower rates of FA than their Asian, Hispanic, Black, and Multiracial counterparts (Gupta et al., 2019). Crucially,

however, the emergency admission rates of food-induced anaphylaxis were approximately three times higher in urban settings than in rural settings in New York and Florida (Gupta et al., 2012).

2.2 | Sources of food allergens

Plants are the dominant source of cause of FA in most countries (Lyons et al., 2019). Plant allergenic proteins carry IgE binding epitopes that are partially or fully resistant to digestive proteolysis. These proteins act as antigens that trigger an unusual immune response. Peanuts are the most common source of life-threatening allergens. Peanuts, tree nuts, wheat, and soybean are considered to contain a large number of allergenic proteins (Table 1 and Figure 5). Major tree nuts, including almond, walnut, cashew, hazelnut, pecan, pistachio, and brazilnut, are potent sources of IgE-induced allergens and account for a prevalence of 4.9% worldwide (McWilliam et al., 2015). The tree nut allergens are primarily characterized by relatively few protein families, particularly 2S albumin, legumins, vicilins, nonspecific lipid transfer proteins (nsLTP), and Bet v 1-homologs/profilins, which are associated with

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TABLE 1 Protein families of different types of allergens from peanut, soybean, wheat, and major six tree nuts (walnut, almond, hazelnut, pecan, cashew, pistachio, Brazil nut).

Isoallergen/variants	ıts	MW (kDa)	Uniprot Id	Allergen family	Biological function	References
Ara h 1.0101 64 P43238		P432	38	Cupin (Vicilin-type, 7S globulin)	Seed storage protein and trypsin inhibitor activity might play role in plant defense against insect	Burks et al. (1995)
Ara h 3.0101 61.0 O82580		0825	08	Cupin (Legumin-type, 11S globulin)	Seed storage protein and provide a store of amino acids for use during germination and seed growth	Rabjohn et al. (1999)
Ara h 3.0201 57.0 Q9SQH7)S6Ò	7H7			Kleber-Janke et al. (1999)
Ara h 3 61.7 Q8LKN1		Q8LK	Z.			Viquez et al. (2004)
iso-Ara h 3 60.0 Q6IWG5		WI9Q	G5			Boldt et al. (2005)
Ara h 2.0101 16.7 Q6PSU2		Q6PS	U2	Conglutin (2S albumin)	Seed storage protein and provide a store of amino acids for use during germination and seed growth	Stanley et al. (1997) and Viquez et al. (2001)
Ara h 2.0201 18.1 Q6PSU2		O6PSU	.2			Chatel et al. (2003)
Ara h 6.0101 14.5 Q647G9		Q647G	6			Kleber-Janke et al. (1999)
Ara h 7.0101 16.3 Q9SQH1		1989	11			Kleber-Janke et al. (1999)
Ara h 7.0201 17.4 B4XID4		B4XII	4			Schmidt et al. (2009)
Ara h 7.0301 17.3 Q647G8		Q647G	~			Yan et al. (2005)
Ara h 5.0101 15.0 Q9SQI9		610S60		Profilin	Cytoskeletal and membrane trafficking	Kleber-Janke et al. (1999) and Kleber-Janke et al. (2001)
Ara h 8.0101 16.9 Q6VT83		Q6VT83		Pathogenesis related (PR-10)	Unclear	Mittag et al. (2004)
Ara h 8.0201 16.3 BOYIU5		B0YIU5				Riecken et al. (2008)
Ara h 9.0101 9.1 B6CEX8		B6CEX	∞	Nonspecific lipid transfer protein (nsLTP)	Stabilization of membrane, cell wall organization, signal transduction, and plant development	Krause et al. (2010)
						(00000000)

TABLE 1 (Continued)

Allergen protein	Isoallergen/variants	MW (kDa)	Uniprot Id	Allergen family	Biological function	References
	Ara h 9.0201	0.6	B6CG41			Krause et al. (2010)
	Ara h 16	8.5	I			ı
	Ara h 17	11.0	I			1
	Ara h 10.0101	16.0	Q647G5	Oleosin	Lipid and energy metabolism	Schwager et al. (2015)
	Ara h 10.0102	16.0	Q647G4			Schwager et al. (2015)
	Ara h 11.0101	14.0	Q45W87			Schwager et al. (2015)
	Ara h 11.0102	14.0	Q45W86			Schwager et al. (2015)
	Ara h 14.0101	18.4	Q9AXII			Pons et al. (2005) and Schwager et al. (2015)
	Ara h 14.0102	18.4	Q9AXI0			Pons et al. (2005) and Schwager et al. (2015)
	Ara h 14.0103	18.4	Q6J1J8			Schwager et al. (2015)
	Ara h 15.0101	17.0	Q647G3			Schwager et al. (2015)
	Ara h 12.0101	5.2	B3EWP3		Plant defensing and antifungal activity and also showed inhibitory activity on mold strains	Petersen et al. (2015)
	Ara h 13.0101	5.5	EY396019	Defensins		Petersen et al. (2015)
	Ara h 13.0102	5.5	COHJZ1			Petersen et al. (2015)
	Ara h 18.0101	21	A0A444XS96	Cyclophilin- peptidyl-prolyl cis-trans isomerase	Unclear	Allergen.org

TABLE 1 (Continued)

Allergen protein	Isoallergen/variants	MW (kDa)	Uniprot Id	Allergen family	Biological function	References
Glycine max (8)						
Gly m 1	Gly m 1.0101 Gly m 1.0102	7	Q9S8F3 Q9S8F2		Hydrophobic protein	Gonzalez et al. (1991)
Gly m 2	Gly m 2.0101	8	I		Defensins	Codina et al. (1997)
Gly m 3	Gly m 3.0101 Gly m 3.0102	14	O65809 O65810		Profilin	Rihs et al. (1999)
Gly m 4	Gly m 4.0101	17	P26987		Pathogenesis-related protein, PR-10, Bet v 1 family member	Crowell et al. (1992)
Gly m 5	Gly m 5.0101 Gly m 5.0201 Gly m 5.0301 Gly m 5.0302	1	022120 Q9FZP9 P25974 P25974		Beta-conglycinin (vicilin, 7S globulin)	AB_P_00904 (AllerBase ID)
Gly m 6	Gly m 6.0101 Gly m 6.0201 Gly m 6.0301 Gly m 6.0401 Gly m 6.0501	1	P04776 P04405 P11828 Q9SB11 Q7GC77		Glycinin (legumin, 11S globulin)	AB_P_00906 (AllerBase ID)
Gly m 7	Gly m 7.0101	76.2	C6K8D1		Seed biotinylated protein	Riascos et al. (2010)
Gly m 8	Gly m 8.0101	28	P19594		2S albumin	Ebisawa et al. (2013) Klemans et al. (2013)
Triticum aestivum (28)						
Tri a 12	Tri a 12.0101 Tri a 12.0102 Tri a 12.0103 Tri a 12.0104	41	P49232 P49233 P49234 B6EF35		Profilin, actin binding protein	AB_P_01828 (AllerBase ID)
Tri a 14	Tri a 14. 0101 Tri a 14. 0201	6	D2T2K2		Nonspecific lipid transfer protein 1	Sander et al. (2011)
						(Continue)

TABLE 1 (Continued)

(Continues)

Tria 15.0101 Tria 15.0101 DS Tria 17 Tria 18.0101 - P9 Tria 18 Tria 18.0101 - P1 Tria 20 Tria 20.0101 35-38 A6 Tria 21 Tria 25.0101 - Q6 Tria 25 Tria 25.0101 - Q6 Tria 26 Tria 26.0201 88 P1 Tria 27 Tria 26.0201 88 P1 Tria 28 Tria 28.0101 13 Q6 Tria 29 Tria 29.0101 13 Q7 Tria 30 Tria 30.0101 - Q6 Tria 33 Tria 33.0101 - Q6 Tria 34 Tria 35.0101 - Q7 Tria 36 Tria 36.0101 - Q6 Tria 36 Tria 36.0101 - Q7 Tria 36 Tria 36.0101 - Q7 Tria 36 Tria 36.0101 - Q7 Tria 37 Tria 36.0101 - Q7	MW (kDa) Uniprot Id Allerge	Allergen family Biological function	References
Tri a 17.0101 56 Tri a 18.0101 - Tri a 20.0101 35-38 Tri a 25.0101 - Tri a 25.0101 - Tri a 26.0201 88 Tri a 26.0201 88 Tri a 26.0201 13 Tri a 27.0101 13 Tri a 29.0201 16 Tri a 39.0101 - Tri a 30.0101 - Tri a 33.0101 - Tri a 33.0101 - Tri a 35.0101 - Tri a 37.0101 - Tri a 10.0101	D2TGC3	Monomeric alpha-amylase inhibitor	Sander et al. (2011)
Tria 18.0101 – Tria 20.0101 35-38 Tria 21.0101 – Tria 25.0101 – Tria 25.0101 – Tria 26.0201 88 Tria 26.0201 13 Tria 29.0101 13 Tria 29.0201 16 Tria 30.0101 – Tria 31.0101 – Tria 33.0101 – Tria 33.0101 – Tria 35.0101 –	56 P93593	Beta-amylase	AB_P_01881 (AllerBase ID)
Tri a 19.0101 35-38 Tri a 20.0101 35-38 Tri a 25.0101 - Tri a 26.0101 88 Tri a 26.0101 88 Tri a 26.0201 77 Tri a 26.0101 13 Tri a 29.0101 16 Tri a 29.0101 16 Tri a 30.0101 - Tri a 32.0101 - Tri a 34.0101 - Tri a 35.0101 -	- P10968	Agglutinin isolectin 1	Sutton et al. (1984)
Tria 20.0101 35-38 Tria 21.0101 - Tria 25.0101 - Tria 26.0201 88 Tria 26.0201 13 Tria 28.0101 13 Tria 29.0101 146 Tria 30.0101 - Tria 31.0101 - Tria 33.0101 - Tria 33.0101 - Tria 35.0101 -	65 Q40215	Omega-5 gliadin, seed storage protein	Lehto et al. (2003)
Tri a 21.0101 - Tri a 25.0101 - Tri a 26.0201 88 Tri a 26.0201 27 Tri a 28.0101 13 Tri a 29.0201 13 Tri a 30.0101 - Tri a 30.0101 - Tri a 33.0101 - Tri a 34.0101 - Tri a 35.0101 -	35-38 A0A060N479	Gamma gliadin	Yokooji et al. (2013)
Tri a 25.0101 Tri a 26.0101 88 Tri a 26.0201 Tri a 27.0101 27 Tri a 28.0101 13 Tri a 29.0201 Tri a 30.0101 Tri a 31.0101 Tri a 33.0101 Tri a 34.0101 Tri a 35.0101 Tri a 36.0101 Tri a 35.0101	– D2T2K3	Alpha-beta-gliadin	Sander et al. (2011)
Tri a 26.0101 88 Tri a 26.0201 Tri a 27.0101 27 Tri a 28.0101 13 Tri a 29.0201 Tri a 30.0101 16 Tri a 31.0101 - Tri a 32.0101 - Tri a 33.0101 - Tri a 33.0101 - Tri a 35.0101 - Tri a 36.0101 - Tri a 35.0101 - Tri a 35.0101 - Tri a 35.0101 -	– Q9LDX4	Thioredoxin	Weichel et al. (2006)
Tri a 27.0101 27 Tri a 28.0101 13 Tri a 29.0201 13 Tri a 30.0101 16 Tri a 31.0101 - Tri a 32.0101 - Tri a 33.0101 - Tri a 33.0101 - Tri a 36.0101 - Tri a 35.0101 - Tri a 35.0101 - Tri a 35.0101 -	88 P10388 Q45R38	High-molecular-weight glutenin	AB_P_01837 (AllerBase ID)
Tri a 28.0101 13 Tri a 29.0101 13 Tri a 29.0201 Tri a 30.0101 16 Tri a 31.0101 - Tri a 32.0101 - Tri a 34.0101 - Tri a 35.0101 - Tri a 35.0101 - Tri a 35.0101 - Tri a 35.0101 1	27 Q7Y1Z2	Thiol reductase homoloque	Sander et al. (2011)
Tri a 29.0101 13 Tri a 29.0201 Tri a 30.0101 16 Tri a 31.0101 - Tri a 32.0101 - Tri a 33.0101 - Tri a 34.0101 - Tri a 35.0101 - Tri a 36.0101 - Tri a 35.0101 12	13 Q4W0V7	Alpha-amylase inhibitor	Sander et al. (2011)
Tri a 30.0101 16 Tri a 31.0101 Tri a 32.0101 Tri a 33.0101 Tri a 34.0101 Tri a 35.0101 Tri a 36.0101 40 Tri a 37.0101 12	13	Alpha-amylase inhibitor	Sander et al. (2011)
Tri a 31.0101 – Tri a 32.0101 – Tri a 33.0101 – Tri a 34.0101 – Tri a 35.0101 – Tri a 35.0101 40 Tri a 37.0101 12	16 PI7314	Alpha-amylase inhibitor	Sander et al. (2011)
Tri a 32.0101 – Tri a 33.0101 – Tri a 34.0101 – Tri a 35.0101 – Tri a 36.0101 40 Tri a 37.0101 12	– Q9FS79	Triosephosphate-isomerase	Sander et al. (2011)
Tri a 33.0101 – Tri a 34.0101 – Tri a 35.0101 – Tri a 36.0101 40 Tri a 37.0101 12	- Q6W8Q2	1-Cys-peroxiredoxin	Sander et al. (2011)
Tri a 34.0101 – Tri a 35.0101 – Tri a 36.0101 40 Tri a 37.0101 12	– Q9ST57	Serpin	Sander et al. (2011)
Tri a 35.0101 – Tri a 36.0101 40 Tri a 37.0101 12	- C7C4×1	Glyceldehyde-3-phosphate dehydrogenase	Sander et al. (2011)
Tri a 36.0101 40 Tri a 37.0101 12	– D2TE72	Dehydrin	Sander et al. (2011)
Tri a 37.0101 12	40 B2Y2Q7	Low-molecular-weight glutenin GluB3-23	AB_P_01847 (AllerBase ID)
	12 Q9T0P1	Alapa purothionin	AB_P_01848 (AllerBase ID)
Tri a 39 Tri a 39.0101 – J7/	– J7QW61	Serine protease inhibitor	Sander et al. (2015)

Allergen profein	Isoalleroen/variants	MW (kDa)	Unincot Id	Alleroen family	Biological function	References
Tri a 40	Tri a 40.0101	15.96		0	Alpha-amylase inhibitor	Sander et al. (2016)
Tri a 41	Tri a 41.0101	1	A0A0G3F2P1		Mitochondrial ubiquitin ligase activator of NFKB 1	AB_P_01852 (AllerBase ID)
Tri a 42	Tri a 42.0101	I	A0A0G3F2F5		Hypothetical protein	AB_P_01853 (AllerBase ID)
Tri a 43	Tri a 43.0101	1	A0A0G3F5F7		Hypothetical protein	AB_P_01854 (AllerBase ID)
Tri a 44	Tri a 44.0101	I	A0A0G3F720		Endosperm transfer cell specific PR60 precursor	AB_P_01855 (AllerBase ID)
Tri a 45	Tri a 45.0101	1	A0A0G3F715		Elongation factor 1	AB_P_01856 (AllerBase ID)
Juglans regia (8)						
Jug r 1	Jug r 1.0101	15-16	P93198		2S albumin seed storage protein	Teuber et al. (1998)
Jug r 2	Jug r 1.0101 Jug r 2.0101	44	Q9SEW4 Q9SEW4		Vicilin seed storage protein	Teuber et al. (1998)
Jug r 3	Jug r 3.0101	6	C5H617		Nonspecific lipid transfer protein 1 (nsLTP1)	Pastorello et al. (2004)
Jug r 4	Jug r 4.0101	58.1	Q2TPW5		11S globulin seed storage protein	Wallowitz et al. (2006)
Jugr5	Jug r 5.0101	20	AOA1JORET5		PR-10	AB_P_01052 (AllerBase ID)
Jug r 6	Jug r 6.0101	47	AOA214E5L6		Vicilin-like cupin	Dubiela et al. (2018)
Jug r 7	Jug r 7.0101	13	AOA214DNN6		Profilin	AB_P_01054 (AllerBase ID)
Jug r 8	Jug r 8.0101 Jug r 8.0101	6	AOA214EB91 AOA214GT96		nsLTP-2	AB_P_02015 (AllerBase ID)
Prunus amygdalus (6)						
Pru du 3	Pru du 3.0101	6	C0L0I5		nsLTP1	AB_P_01557 (AllerBase ID)
Pru du 4	Pru du 4.0101 Pru du 4.0102	14	Q8GSL5 Q8GSL5		Profilin	Tawde et al. (2006)

AB_P_00555 (AllerBase ID)

AB_P_00554 (AllerBase ID)

Lauer et al. (2004)

7S seed storage globulin

Q8S4P9

48

Cor a 11.0101

Cor a 11

Q84T21

17

Cor a 12.0101

Cor a 12

Q84T91

14-16

Cor a 13.0101

Cor a 13

Oleosin

Oleosin

AB_P_01559 (AllerBase ID) AB_P_00558 (AllerBase ID) Pastorello et al. (2002) Pastorello et al. (2002) Kabasser et al. (2021) Kabasser et al. (2021) Gruehn et al. (2003) Lauer et al. (2004) Beyer et al. (2002) Chw et al. (2019) Antimicrobial seed storage protein PR-10, Bet v 1 family member 60s acidic ribosomal protein 2 11S seed storage globulin Luminal binding protein Amandin, 11S globulin legumin-like protein Mandelonitrile lyase 2 Isoflavone reductase **Biological function** nsLTP type 1 Profilin Allergen family A0A0U1VZC8 40A516F3L2 Uniprot Id Q9SWR4 29AXH5 Q9FPK4 Q9FPK3 Q9FPK2 **Q9AXH4** 09АТН2 Q8W1C2 E3SH28 E3SH29 Q9FSY7 Q8H2B9 Q945K2 Q39454 Q08407 Q08407 Q39453 008407 Q08407 MW (kDa) 360 10 31 9 70 17 4 35 40 6 Isoallergen/variants Pru du 10.0101 Pru du 5.0101 Pru du 8.0101 Pru du 6.0101 Pru du 6.0201 Cor a 10.0101 Cor a 1.0102 Cor a 1.0103 Cor a 1.0104 Cor a 1.0402 Cor a 1.0403 Cor a 1.0404 Cor a 1.0101 Cor a 1.0301 Cor a 1.0401 Cor a 8.0101 Cor a 1.0201 Cor a 2.0101 Cor a 6.0101 Cor a 9.0101 Corylus avellana (11) Allergen protein Pru du 8 Pru du 10 Cor a 10 Pru du 5 Pru du 6 Cor a 2 Cor a 6 Cor a 8 Cor a 9 Cor a 1

TABLE 1 (Continued)

TABLE 1 (Continued)

Allergen protein	Isoallergen/variants	MW (kDa)	Uniprot Id	Allergen family	Biological function	References
Cor a 14	Cor a 14.0101	10	D0PWG2		2S albumin	Garino et al. (2010)
Cor a 15	Cor a 15.0101	17	ı		Oleosin	AB_P_02079 (AllerBase ID)
Carya illinoinensis (3)						
Cari1	Cari i 1.0101	16	Q84XA9		2S albumin seed storage protein	Sharma et al. (2011)
Car i 2	Cari i 2.0101	55	B3STU4		Vicilin-like protein	Zhang et al. (2016)
Car i 4	Cari i 4.0101	55.4	B5KVH4		Legumin seed storage protein	Sharma et al. (2011)
Anacardium occidentale (3)						
Ana o 1	Ana o 1.0101 Ana o 1.0102	50	Q8L5L5 Q8L5L6		Vicilin-like protein	Wang et al. (2002)
Ana o 2	Ana o 2.0101	55	Q8GZP6		Legumin-like protein	Wang et al. (2003)
Ana o 3	Ana o 3.0101	14	Q8H2B8		2S albumin	Robotham et al. (2005)
Pistacia vera (5)						
Pis v 1	Pis v 1.0101	7	B7P072		2S albumin	Ahn et al. (2009)
Pis v 2	Pis v 2.0101 Pis v 2.0201	32	B7P073 B7P074		11S globulin	Ahn et al. (2009)
Pis v 3	Pis v 3.0101	55	$B4 \times 640$		Vicilin	Willison et al. (2008)
Pis v 4	Pis v 4.0101	25.7	B2BDZ8		Manganese superoxide dismutase	Noorbakhsh et al. (2010)
Pis v 5	Pis v 5.0101	36	B7SLJ1		11S globulin	AB_P_01464 (AllerBase ID)
Bertholletia excels (2)						
Ber e 1	Ber e 1.0101	6	P04403		2S sulfur rich seed storage albumin	Alcocer et al. (2012)
Ber e 2	Ber e 2.0101	29	Q84ND2		11S globulin seed storage protein	AB_P_00284 (AllerBase ID)

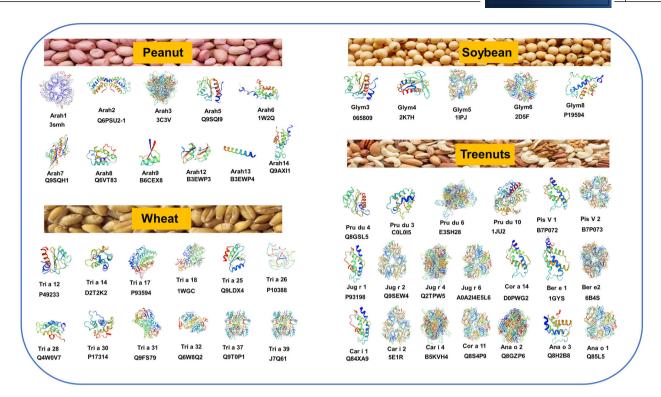


FIGURE 5 Different allergen protein structure of peanut, wheat, soybean, and treenuts. (*Source*: https://fermi.utmb.edu/cgi-bin/SDAP/sdap_01).

pollen tree nut allergies (Geiselhart et al., 2018). To date, 963 allergens have been isolated and characterized from different plant sources. Details concerning these allergens are maintained in the dedicated allergen database "AllerBase" (http://bioinfo.unipune.ac.in/AllerBase/Home.html).

Many food plants contain proteins that are referred to as lectins based on their specific carbohydrate-binding properties. Lectins are major antinutritional factors in seeds. For example, the most abundant lectin family proteins are arcelin, phytohemagglutinin, and a-amylase inhibitor—APA proteins) in common bean (*P. vulgaris*). The major antinutritional effects of these proteins are caused by their low digestibility and high toxicity in the intestinal tract (Bardocz et al., 1995). Four allergen proteins in kidney beans (alpha-amylase inhibitor precursor, phaseolin, and group 3 late embryogenesis protein) showed significant matches with the common bean lectins (PHAs) (Kasera et al., 2011).

2.3 | Labeling regulations

People who have food allergies should read labels and avoid foods to which they are allergic. Legislation regulating the packaging and labeling of prepackaged foods containing allergens at the level of food service establishments is essential to ensure food safety. The US government's Food Allergen Labeling and Consumer Protection Act mandates the

proper labeling of prepacked foods containing the big eight derived ingredients and these labeling requirements extend to retail and food service establishments that offer products for human consumption (Messina & Venter, 2020). The allergen's food source must be mentioned on the food label at least once in one of two ways, either "flour (wheat)," and "whey (milk)" or "Contains wheat, milk, and soy". Further, the Food Allergy Safety, Treatment, Education, and Research Act was signed into law on April 23, 2021, including sesame as the 9th significant food allergen recognized by the United States. The new policy has become effective from January 1, 2023 and all United States Food and Drug Administration requirements of labeling and manufacturing a new major food allergen sesame (https://www.fda.gov/food/food-labelingnutrition/food-allergies). In Europe, 14 major food products are identified as a major source of food allergies. The EU Food Information Regulation and Food Information for Consumers Regulation have several amendments to ensure that allergen labeling laws apply to prepacked and nonprepacked foods (Baker, 2018). Globally, the Codex Alimentarius Commission developed the Codex General Standards for the Labeling of Prepacked foods, listing peanuts, tree nuts, soy, milk, eggs, fish, crustaceans, as well as cereals containing gluten and sulfites. This sets the limit of >10 mg/kg above which products should contain a precautionary labeling statement. The Codex standards guide helps legislative bodies in developing the regulatory frameworks of Codex member countries (FAO, WHO, n.d.). Several national and international organizations, such as Food Allergy Research and Education, aim to enhance food allergen labeling regulations around the world (https://www.foodallergy.org/). The food safety regulatory body of India, the Food Safety and Standard Authority of India, published regulations that comply with Codex Standards specifying packaging and labeling principles for packaged foods containing allergenic constituents ("Food Safety & Standards [Labelling & Display]", 2020). The food service establishments should provide a declaration on the package regarding food allergens specifying the name of allergy causing ingredients. In the case of packaged foods with cross contaminated ingredients that are known to cause allergies declared "may contain particular allergy causing ingredients." Whereas, allergen labeling requirements, that is, the declaration is not required in the case of oils and alcoholic beverages derived from these ingredients and raw agricultural (https://www.fssai.gov.in/upload/uploadfiles/ commodities files/Compendium_Labelling_Display_23_09_2021.pdf). Further, evaluation of global food allergen labeling laws pertaining to foods and allergens labeling were comprehensively reviewed at the level of county and region, and also

emphasize implementation of other protective measures by

2.4 | Impact of global trade

Chang et al. (2023).

Due to the globalization of the market, peanut-based food product industries have huge export potential. There are no general estimates of direct economic losses in the global food market due to the allergenicity of food commodities. However, the indirect economic costs of FA concerns in global trade can be calculated from the rise in the FA and food intolerance product market. Increasing healthcare awareness has led to a growing preference for allergen-free, gluten-free, and lactose-free diets, which have accelerated the growth of the global FA product market. According to Strategy R (https://www.strategyr.com/market-reportfood-allergy-and-intolerance-products-forecasts-globalindustry-analysts-inc.asp), which is a trademark for global industry analysis, the global market of FA and intolerant products will reach US\$32 billion by 2027. It was estimated to be US\$22.4 billion in 2020, with a projected compound annual growth rate (CAGR) of 5.2% between 2020 and 2027. The lactose-free food product segment alone is projected to have a CAGR of 4.6%, reaching US\$15.1 billion by the end of 2027. The FA and intolerance products market of the United States was estimated to be US\$6.6 billion in 2020, followed. The forecast for China was US\$5.7 billion in 2027 at a CAGR of 4.9%. The North American market is growing fast due to a rapid increase in the rate of food allergies and sensitivity. Subsequently, the European and Asia Pacific markets have followed a similar trend due to innovations in food processing industries and a rise in consumer awareness about food safety (https://www.coherentmarketinsights.com/ongoing-insight/food-allergy-and-intolerance-products-market-788).

3 | THE FUNCTIONAL BIOLOGY OF ALLERGENS

Humans are constantly exposed to thousands of plant food proteins through ingestion. However, only a limited group of proteins can trigger an allergic response in certain individuals. This intrinsic property of allergenicity is attributed to the biochemical and structural makeup complementary to IgE antibodies. Other factors such as biochemical and molecular properties, such as size, solubility, stability to acidic pH and enzymatic hydrolysis, disulfide bond-stabilized conformational IgE epitopes, oligomerization, post-translational modifications, interactions of the protein food matrix and ligand binding, also influence allergenicity (Pekar et al., 2018).

3.1 | The nature and functions of allergens

Most plant food allergens are seed storage and defense-related proteins (Breiteneder & Radauer, 2004). Seed storage proteins, which account for approximately 50% of the total protein content, act as the seed nutrient store and are required for germination and seedling growth. Of these, legumins, 2S albumin and vicilins contribute 50-70, 20-60, and 20% of the total protein fraction, respectively (Monsalve et al., 2007). However, there is no relationship between the abundance of seed storage proteins and allergic sensitization. Factors such as the chemical composition of allergens, processing methods, routes of exposure, structural stability, interactions with lipid molecules, aggregation, cross-reactivity, and patient factors determine the potency of allergenicity (Breiteneder & Mills, 2005; Smits et al., 2018). Plant defense proteins such as pathogenesis-related (PR) proteins, nsLTPs and profilins protect against invasion by pathogenic microorganisms and herbivory by insect pests, as well as preventing the adverse effects of abiotic stresses (Sinha et al., 2014). PR proteins, such as PR-10, are induced in response to exposure to biotic and abiotic stresses (Sinha et al., 2014). In addition to their defensive role, nonspecific nsLTPs are required for plant growth and development, cuticle formation, suberin biosynthesis, pollen development, seed maturation and germination, fruit ripening, and defense signaling (D'Agostino et al., 2019; Liu et al., 2015). The profilin superfamily proteins are known to be involved in the reorganization of the actin

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cytoskeleton and signal transduction by regulating intracellular calcium levels (Asturias et al., 2002).

3.2 | Classification of plant food allergens

In the AllFam database (http://www.meduniwien.ac.at/allfam/), allergens are classified into protein families based on the data available from the WHO/IUIS Allergen Nomenclature Database and Allergen Online and Pfam databases (Radauer et al., 2008). An AllFam search for plant food allergens with ingestion as a route of exposure revealed 233 allergens that are primarily distributed in allergen super families, such as Prolamin (75), Cupin (36), Profilin (26), PR-10 protein (21), Thaumatin-like protein (10), Oleosin (8), Defensins (2) and others (Costa et al., 2020).

3.2.1 | The prolamin superfamily

A prolamin superfamily is a major group of allergens, such as 2S albumin, nsLTPs, and cereal alpha-amylase/protease inhibitors. These allergens are primarily found in rice, wheat, peanuts, brazilnut, and fruits with peaches. They are low-molecular-weight proteins that are rich in proline and glutamine. They contain α-helical globular domains with conserved intramolecular disulfide bonds that are resistant to thermal processing and proteolytic digestion (Breiteneder & Radauer, 2004; Mills et al., 2004). The prolamin superfamily includes cereal prolamin (soluble gliadins and insoluble glutenins), 2S albumin, nsLTPs, bifunctional alphaamylase/proteinase inhibitors, soybean hydrophobic proteins related to nsLTP, and indolines, which are cereal antimicrobial proteins that contribute to grain softness. 2S albumin is a predominant seed storage protein. It is widely distributed in the seeds and nuts of dicotyledonous plants. Examples of 2S albumins include Ara h 2, Ara h 6, Ara h 7 from peanut, Gly m 8 from soybean, and the Jug r 1, Cor a 14, Car i 1, Ana o 3, Pis v 1, and Ber e 1 proteins from different tree nuts (Table 1 and Figure 5). In the case of peanut allergens, Ara h 1, 2, 3, and 6 show predominant expression in seeds.

nsLTPs are small (6.5–10.5 kDa), basic proteins that are widely distributed in higher plants and serve many important physiological processes, including defense against bacteria and fungi (D'Agostino et al., 2019). While nsLTPs contain an eight-cysteine motif backbone, the type 1 and type 2 nsLTPs differ in size and disulfide bonding patterns (Liu et al., 2015; Salminen et al., 2016). nsLTPs bind phospholipids, fatty acids and a variety of hydrophobic molecules. They are commonly found in nuts, celery tubers, seeds, fruits, vegetables, pollen, and latex.

Bifunctional α -amylase/protease inhibitors are proteinaceous enzyme inhibitors that are primarily found in storage

organs such as seeds and tubers. They function in protection against phytophagous insect pests by inhibiting insect gut enzyme activity, thereby hindering the digestion of plant food starch and protein. These proteins possess a small, compact structure rich in disulfide bonds and act as a storage reserve during seed germination. They include Ara h 1, Ara h 2 from peanut, Tri a 28, Tri a 29, Tri a 30, Tri a 33, Tri a 39, and Tri a 40 from wheat (Table 1; Maleki et al., 2003; Figure 5).

3.2.2 | The cupin superfamily

The allergenic cupins superfamily are globulin-type seed storage proteins (Radauer & Breiteneder, 2007). They have a β -barrel core domain structure and exist as single domain-cupins and bicupins. They elicit life-threatening allergic reactions in individuals sensitive to peanuts, soybean, almond, hazelnut, and walnut and are grouped into the legumin and vicilin families (Costa et al., 2020).

Legumins (11S globulin) are abundant (50–70%) seed storage proteins (Mills et al., 2002). They have hexameric structures (360 kDa each) linked by noncovalent interactions. They are composed of six monomeric units derived from the respective gene products. Members of 11S globulin allergens include peanut Ara h 3, soybean Gly m 6, walnut Jug r 4, almond Pru du 6, hazelnut Cor a 9, pecan Car i 4, cashew Ana o 2, pistachio Pis v 2, Pis v 5, and brazil nut Ber e 2 (Table 1 and Figure 5).

Vicilins (7S globulin) are abundant seed storage proteins (up to 20%) that are often found in legumes and tree nuts. They exist as trimeric proteins (150–190 kDa) that can aggregate into hexamers. They differ from legumins because they lack disulfide bonds. Examples include the peanut Ara h 1, soybean Gly m 5, walnut Jug r 2, Jug r 6, hazelnut Cor an 11, pecan Car i 2, cashew Ana o 1, and pistachio Pis v 3 (Table 1; Geiselhart et al., 2018).

3.2.3 | Profilin superfamily

Profilins are highly conserved, cross-reactive cytosolic panallergens (12–15 kDa) that are primarily found in pollen and latex. Sensitization occurs because of cross-reaction with IgE antibodies (Asero et al., 2003). Major allergens of the profiling family were reported in the peanut Ara h 5, soybean Gly m 3, wheat Tri a 12, walnut Jug r 7, almond Pru du 4, and hazelnut Cor a 2 (Table 1 and Figure 5).

3.2.4 | Pathogenesis-related (PR-10) proteins

PR-10 proteins (15-17 kDa) are mostly found in fruits and vegetables but they are also the cause of birch

pollen-associated allergies (Sinha et al., 2014). They are structurally distinct from other PR proteins because they contain highly conserved seven-stranded antiparallel β -sheets surrounding α -helix at the C-terminus (Fernandes et al., 2013). Many birch pollen allergic individuals are also sensitive to multiple fruits and vegetable allergens because of Bet v 1 cross-reactive IgE antibodies. Symptoms ranging from mild to potentially life-threatening conditions have been reported following the consumption of raw foods (Costa et al., 2020).

4 | APPROACHES TO REDUCE ALLERGEN CONTENT

Most plant food allergens are water-soluble glycoproteins that are relatively stable with regard to enzymatic hydrolysis, heat, and chemical treatments (Sicherer & Sampson, 2010). However, different food processing methods can modify, at least in part, immunogenic reactivity (Bhalla & Singh, 2008). Such methods have both limitations and advantages in reducing the allergenic effect but may decrease the nutritional value of food (Fu et al., 2019). Most studies used to evaluate processing methods depend on in vitro IgE binding assays rather than more relevant *ex vivo* basophile activation tests (BAT) and mast cell activation tests (Shah et al., 2019).

4.1 | The genomics of allergens

In the last decade, more than 35 different species of legumes have been sequenced for developing reference genome and transcriptome assemblies. This genomic resource information accelerates the development of cultivars of superior grain legume crops by genomic-assisted breeding and precision breeding (Bauchet et al., 2019; Varshney et al., 2019). Similarly, genetic approaches are being used increasingly to develop low allergen food crops (Zhou et al., 2013). For example, a lower Gly m Bd 30K (P34) allergen was identified in soybean (Jeong et al., 2013). The high-quality RefSeq (reference sequence) v1.0 reference genome from the International Wheat Genome Sequencing Consortium was used to detect and properly discriminate allergens and antigens in wheat proteins linked to or involved in human disease (Juhász et al., 2018). While relatively few quantitative trait locus or association studies have been conducted in other species, the expression levels of allergens can be altered through molecular breeding and/or genetic engineering using RNA interference (RNAi) or clustered regularly interspaced short palindromic repeats (CRISPR)/Cas9 mediated gene editing (Jouanin et al., 2018; Saurabh et al., 2014). A hypoallergenic peanut variety that was produced by gene silencing showed a 25% reduction in the expression of the most potent allergen Ara h 2, thereby significantly decreasing allergenicity (Dodo et al., 2008). Studies involving the silencing of the

Ara h 1. Ara h 2. Ara h 3. and Ara h 6 genes resulted in a significant reduction in IgE binding. Crucially, no changes in seed weight or germination were observed between the transgenic and wildtype plants (Chu et al., 2008). Several peanut lines with lower levels of the major allergens (Ara h 1, Ara h 2, Ara h 3, Ara h 6, and Ara h 8) were identified at ICRISAT using a large-scale phenotyping approach (Pandey et al., 2019). Such studies can lead to the identification of functional variations that may be useful in molecular breeding approaches involving marker-assisted selection and markerassisted backcrossing (Janila et al., 2016; Varshney et al., 2014). Similarly, RNAi approaches have been successfully used to mitigate the expression of the allergen Gly m Bd 30k protein in soybean (Herman, 2003) and rice (Ogo et al., 2014) and apple (Dubois et al., 2015). In addition, CRISPR/Cas9mediated site-directed mutagenesis was successfully used to eliminate two major allergen genes, Gly m Bd 28k and Gly m Bd 30k and thus generate hypoallergenic soybeans (Sugano et al., 2020). The gene silencing studies of Barro et al. (2016) selectively targeted the gliadin and glutenin genes using RNAi technology. An absence of epitopes related to coeliac disease related to immunogenic gliadins was reported in the wheat lines. Similarly, a low-gluten wheat variety that showed an 85% decrease in immunoreactivity was produced (Sánchez-León et al., 2018). Such lines could be used as source material for further introgression studies. Efforts have been made in durum wheat to edit the genes encoding α-amylase/trypsin inhibitors (ATIs) that are involved in wheat allergy and nonceliac wheat sensitivity (Camerlengo et al., 2020). Such studies illustrate the enormous potential of molecular genetic approaches to produce hypoallergenic food crops. However, the removal of allergenic seed storage proteins on a large scale may result in large decreases in nutritive value and taste.

4.2 | Food processing methods

4.2.1 | Physical processing methods

Thermal processing, such as frying, roasting, curing, and various types of cooking, can result in a variety of nonenzymatic, biochemical events in meals. Many foods brown due to a phenomenon known as the Maillard reaction, which is one of the key processes that occur during food cooking or browning (e.g., roasting, frying, and curing). The Maillard reaction is an important in the development of flavour and colour in foods such as peanuts and tree nuts during roasting, enhances flavours in beverages such as beer and coffee, and involves a process similar to caramelization in which amino groups of proteins are modified via nonenzymatic condensation with reducing sugars. Each method has a varied effect on the allergenic potency of different foods.

Processing approaches involve include the use of physical methods (heat, mechanical, electric, and magnetic energy) that disrupt protein structure and induce aggregation but without disrupting the primary structure. The most common physical methods are thermal processing, irradiation, ultrasound, ultrahigh pressure, and microwaves (Cabanillas et al., 2018; Vanga et al., 2017; Verhoeckx et al., 2015). Such methods have been widely applied to reduce allergenicity. They have advantages over chemical and enzymatic methods in terms of cost, time, side effects, and nutritional quality. Boiling, roasting, frying, and autoclaving are the most common methods of household food preparation. While boiling can decrease the immunoreactivity of allergenic protein, roasting increases allergenicity (Kopper et al., 2005; Turner et al., 2014). In the case of peanuts, for example, roasting enhances IgE binding activity by 90-fold and makes the allergens Ara h 1 and Ara h 2 resistant to digestive enzyme proteolysis because of Maillard reactions (Maleki et al., 2000). Thermal processing can therefore reduce immunogenicity to a certain degree but it may also destroy nutrients and bioactive ingredients (Gupta et al., 2018b).

Structural denaturation, unfolding, glycation, and aggregation occur during the physical processing of food proteins. This affects solubility and digestibility, which might lead to the elimination of conformational IgE epitopes or the formation of new allergenic linear epitopes that can increase the risk of allergy (Shah et al., 2019; Verhoeckx et al., 2015). The application of ultrasound to reduce the allergenicity of certain food products has proved to be a useful pretreatment before food processing (Corzo-Martínez et al., 2017). In addition, the application of a pulsed ultraviolet light was able to decrease the levels of glycinin and β-conglycinin allergens in soybean (Yang et al., 2010). Unfortunately, there are no inconsistent food processing methods that reduce allergenicity in different food materials. Moreover, the conventional thermal processing methods applied in the reduction of food immunoreactivity significantly destroy nutritional components present in food sources. Therefore, the use of novel nonthermal processing techniques including high-pressure processing, ultrasound, pulsed light, cold plasma, fermentation, pulsed electric field, and enzymatic hydrolysis generally have better performance in retaining the original characteristics of food and improving the efficiency of eliminating allergens (Dong et al., 2021).

4.2.2 | Chemical and enzymatic methods

The physical methods discussed for food processing affect the physicochemical properties of food proteins in diverse ways and influences their gastrointestinal digestion, bioavailability, and allergenicity. However, the application of nonthermal including chemical and enzymatic methods can induce min-

imal changes to food quality attributes and can extend the shelf-life of food (Dong et al., 2021; Ekezie et al., 2018). Chemical and enzyme treatments can reduce or destroy immunogenic determinants in food by disrupting the allergen structure that is stabilized by covalent and noncovalent bonds (Ekezie et al., 2018; Wang et al., 2022). Acid hydrolysis is widely used to treat wheat flour and destroy gluten allergen proteins to produce low allergenic products (Fu et al., 2019). Major covalent modifications, such as acylation, reduction, and alkylation, show remarkably reduced immunogenicity by altering the solubility and digestibility of allergens in the gastrointestinal tract (Apostolovic et al., 2013). In addition, non-covalent modifications that involve binding with compounds such as phytic acid, phenolic compounds, and tannic acid to form insoluble complexes have been shown to decrease allergic potency in peanuts by hindering proteolytic digestion (Chung & Champagne, 2008; Chung & Reed, 2012). Furthermore, polyphenol-enriched peanut matrices were shown to significantly minimize allergen interactions with IgE and decrease ex vivo basophil degranulation and mast cell degranulation in a mouse model (Plundrich et al., 2017). This phenomenon involved excessive proteolytic digestion of the polyphenol-allergen complex, which facilitated alterations in conformational epitopes and the simultaneous masking of linear epitopes. However, the addition of phenolic compounds and polyphenols can cause stomach discomfort and it also obstructs nutrient absorption in the intestine.

Enzymatic hydrolysis has shown promising results with regard to reducing allergenicity. Cross-linking of enzymatic proteins with allergens masks antibody-specific epitopes. In contrast, proteolysis with food-grade enzymes, such as trypsin, chymotrypsin, papain, ficin, bromelain, and so on, disrupts the native structure and physiochemical characteristics, as well as IgE-specific conformational and linear epitopes, which ultimately reduces allergenicity (Meng et al., 2020; Zhou et al., 2013). The roasted peanut allergens Ara h 1 and Ara h 2 are completely hydrolyzed by treatment with trypsin (0.15%) and chymotrypsin (0.1%) for 3 h (Yu et al., 2011). Similarly, Ara h 1, Ara h 2, and Ara h 3 are effectively eliminated by hydrolysis using alcalase and flavorzyme (Cabanillas et al., 2012).

Enzymatic hydrolysis followed by physical processing methods, such as irradiation, pulsed ultraviolet light, pulsed electric field, high-pressure processing, and high-intensity ultrasound have been proven to be effective in reducing allergenicity (Shah et al., 2019). However, the combination of autoclaving and fermentation of raw peanut pulp with *Bacillus natto* effectively diminishes the allergens in raw peanuts (Pi et al., 2021). Such effects have also been demonstrated in wheat for gluten-free bread (Diowksz & Leszczyńska, 2014).

The combination of physical methods and enzymatic hydrolysis (hurdle technologies) has the advantage of facilitating efficient enzyme penetration and proteolysis. Similarly, the chemical reduction of disulfide bonds in allergenic proteins destabilizes the three-dimensional structure and increases the efficiency of enzymatic proteolysis (Mikiashvili & Yu, 2018). Unfortunately, however, the majority of food processing methods also alter the texture and flavor of food and this can significantly affect consumer acceptance.

4.3 | Common methods for allergy diagnosis

There are various diagnostic methods available for testing allergies which involve skin or blood. Allergy testing assesses the body's reaction to specific allergens and the test must be chosen by a trained health professional called an allergist based on symptoms, age, hobbies, exposures and patient medical history. Such testing reveals allergens that might cause allergies, such as plant pollens, molds, dust mites, animal dander, insect stings, and various foods such as peanuts, eggs, wheat, shellfish, and milk and also includes some medicines like penicillin. Once the allergens have been identified through proper diagnostic methods, the specific treatments can be possible through medications, allergen immunotherapy, and/or environmental control measures to achieve long-term sustainable outcomes (Ansotegui et al., 2020; Dreborg, 2001; Heinzerling et al., 2013; Maruyama, 2021).

4.3.1 | Skin prick/scratch test (SPT)

It uses a thin needle to prick the skin on your forearm or back with a possible number of different potential allergens or the allergist may place droplets of potential allergens onto your skin and use a device to scratch and lightly puncture the area. It helps the liquid to enter into the skin and is observed for the body's reactions might be a rash or round spots which are generally used for the detection of airborne allergies, food allergies and penicillin allergies. The skin prick test represents the most reliable and cost-effective tool for the diagnosis and management of IgE-mediated allergy.

4.3.2 | Intradermal skin test

If skin prick test results turn inconclusive, a small amount of the allergen is injected into the epidermis and records the observations. This test is used for the diagnosis of allergies to airborne irritants, insect stings and medications.

4.3.3 | Patch test

The purpose of this test is to determine the cause of contact dermatitis in which a patch of allergen-containing bandage is applied on to the skin. After 2–3 days the allergist records the observation of allergic reactions.

4.3.4 | Blood (IgE) test

When skin tests are inconvenient for a particular patient, an allergist can proceed with the blood test. This test measures levels of allergen-responsive antibody IgE in the serum by the addition of different potential allergens to the blood.

4.3.5 | Basophile activation tests

This laboratory test measures the activation of basophils, a type of red blood cells in response to a specific allergen. Further, the BAT is specific which allows better defining the IgE profile of the patient but it is complex to perform.

4.3.6 | Challenge tests

This test is particularly used to identify the source of food allergies. Under the supervision of a health professional, the person with suspected FA ingests a small amount of an allergen and the allergist observes the symptoms of allergic reaction. This test is highly risky for the individual sensitive to anaphylaxis which required immediate epinephrine injection to stop the reaction.

Among all the allergy testing methods, the skin test is the gold standard and is used along with a person's medical history to identify the source of FA. While blood tests generally have a higher rate of false-positive results, in addition to the pain and chances of bleeding.

5 | SCOPE FOR DEVELOPING HYPOALLERGENIC CROPS WITH MINIMAL EFFECTS ON PLANT PHYSIOLOGY

Most allergens are seed storage proteins that play a key role in plant biology, with functions ranging from seed germination to defense against biotic and abiotic stresses (Zhou et al., 2013). Therefore, the selection of target genes of allergens in particular crop plants requires a comprehensive understanding of gene function in plant growth and development.

Of the 18 recognized allergens in peanuts, Ara h 1, Ara h 2, Ara h 3, and Ara h 6 (Shah et al., 2019; Wu et al., 2016), Ara h 2 is recognized in most peanut-allergic individuals. Therefore, a reduction in peanut allergy by eliminating the Ara h 2 genes might be a preferable choice that might not result in major alterations to plant growth and

development. Efforts have been made to silence Ara h 2 and Ara h 6 in peanuts using RNAi technology. Such approaches have resulted in a significant decrease in IgE binding with no significant effects on seed germination or defenses against fungal infection (Chu et al., 2008). Other studies have also sought to decrease peanut allergy by silencing Ara h 2 using a specific RNAi gene silencing (Dodo et al., 2008). Transgenic peanut lines with suppressed Ara h 2 and Ara h 6 protein expression remained stable for several generations (Chandran et al., 2015). In the case of wheat, silencing of gluten synthesis led to the production of a low gluten wheat variety, which is safe for many gluten allergy-sensitive individuals (Wen et al., 2012). Similarly, the ω –5 gliadin-free wheat line 1BS-18 had low efficiency in inducing allergy symptoms in guinea pigs (Kohno et al., 2016).

Soybeans contain two major allergens in the form of 7S globulin (β-conglycinin) and 11S globulin (glycinin). These together make up >50% of the total seed protein. Suppression of globulin and conglycinin expression using RNAi did not affect seed size, weight or developmental ontogeny. However, these soybean lines undergo were found to express other seed proteins (Schmidt et al., 2015). Another study using microR-NAs specific to 7S globulin had no adverse effects on seed lipid, carbon and nitrogen contents (Yamada et al., 2014). CRISPR/Cas9 gene editing was used to create a double mutant (Gly m Bd 28 K and Gly m Bd 30 K) which resulted in the loss of both proteins from Japanese soybean seeds (Sugano et al., 2020). Taken together, such studies provide substantial evidence that plants show a compensatory response to the suppression or elimination of allergenic seed proteins. The success of such studies demonstrates the potential of such current gene technologies for the creation of hypoallergenic plant foods.

6 | PHENOMICS AND OMICS APPROACHES TO REDUCING ALLERGENS

6.1 | Identification of crops with reduced allergen content through selection and breeding

Conventional plant breeding plays a significant role in developing new plant varieties with desired plant traits/features. Unfortunately, due to lack of information on low allergen content lines, there is not much effort on breeding varieties with reduced allergen content using conventional breeding approaches (Pandey et al., 2019; Riascos et al., 2010). Limited conventional breeding efforts are reported to improve the nutritional content of crops, which indirectly contributes to reducing allergenicity (Lemke et al., 2022). By increasing the nutritional value of crops, individuals with food allergies may have access to a wider range of nutrients and alterna-

tive food sources (Kaiser et al., 2020). Plant food allergens are not always a strict selection criterion comparable to other plant toxins, especially considering that food allergens are always unique proteins of large protein families with complex inheritance in plant breeding. Although screening germplasm to identify individuals with decreased allergen content is time-consuming, traditional breeding attempts toward hypoallergenic variants in peanut (Pandey et al., 2019; Perkins et al., 2006), wheat (Yamada et al., 2022), and soybean (Gao et al., 2012) have been attempted. Gluten in hexaploid bread wheat is made up of numerous distinct proteins, the most prominent of which are glutenin and gliadin. Glutenins are essential for baking quality, but gliadins include the majority of celiac disease-associated pieces (epitopes). Although old hexaploid bread and tetraploid durum wheat varieties have been identified with few epitopes connected to gluten intolerance, generating favourable combinations of gluten genes to satisfy baking quality standards in a polyploid is difficult (Gilissen et al., 2014; Lemke et al., 2022).

Little information is available concerning the major allergen contents of peanut germplasm lines that are commercially grown around the world. Recently, Pandey et al. (2019) identified hypo-allergenic lines for Ara h 1, Ara h 2, Ara h 3, Ara h 6, and Ara h 8. These and other studies have used monoclonal antibodies to screen peanut-based products (Filep et al., 2018). Earlier, 34 peanut genotypes were screened using patient sera but no substantial differences in allergen content were identified (Dodo et al., 2002). The analysis of a "Reference set" consisting of 300 genotypes representing 48 countries has also been reported (Upadhyaya et al., 2003, 2010). Little variation was observed in 53 Chinese peanut cultivars (Wu et al., 2016) using human sera to assess the allergen content in their cultivars. However, the Spanish bunch varieties had lower peanut allergen contents than the other agronomic types. These authors also reported that Xinxiandahuasheng of the Virginia type, Bangjihonghuasheng of the Valencia type, Mangdou of the Spanish type, and Yaoshangxiao make of the Peruvian type had lower peanut allergen contents. A study of 35 US peanut cultivars using antisera from allergic patients also found no significant variation (Dodo et al., 2002; Isleib & Wynne., 1992). However, rapid and easy phenotyping methods for different allergens are required to increase the efficiency of breeding reduced allergen crops (Liu et al., 2023).

The pattern of sensitization to peanut allergens varies in different geographical regions (Vereda et al., 2011). For example, Ara h 1, Ara h 2, and Ara h 3 were identified in allergic reactions in the United States. Similarly, Ara h 1, Ara h 2, and Ara h 3 11 were found in European nations (Ballmer-Weber & Beyer, 2018). Nine soybean allergens were identified in three soybean varieties developed at nine locations in three states in the same climate zone in North America: Illinois, Iowa and the United States (Mcclain et al., 2018).

Mutation breeding has also been shown to be effective in a wide range of crop species, such as tomato, rapeseed, cotton, barley, sunflower, peanut, cassava, and can be successfully used to improve plant varieties (Xia et al., 2022). Mutation breeding has been used to increase the yield and oil and protein contents of peanuts (Hamid et al., 2006). A Targeting Induced Local Lesions IN Genomes approach was used to create gene-specific primers for each of the two Ara h 1 and two Ara h 2 genes to find mutations in peanuts (Knoll et al., 2011). Similarly, gamma irradiation mutagenesis was used to identify the mutant alleles of the Gly m Bd 28 K and Gly m Bd 30 K genes in soybean.

6.2 | Role of artificial intelligence and bioinformatics in reducing allergencity

Traditional methods for identifying food allergens mostly rely on in vivo and in vitro experiments, which can be time consuming and uneconomical. However, artificial intelligence (AI) and bioinformatics have the potential to significantly reduce plant food allergens by aiding in the identification of specific allergenic proteins in plants as well as the development of novel methods to modify or remove allergens from food products (Liu et al., 2023). AI in allergy and immunology has various potential therapeutic applications ranging from disease diagnosis to multidimensional data reduction in electronic health records or immunologic datasets (Khoury et al., 2022). The applications of AI and bioinformatics can be deployed in the prediction of allergenicity by analysing available genomic and proteomic data. By analyzing the genetic information of various plant species, AI algorithms can identify potential allergenic proteins and predict their structure and function. One of the study proves quick food allergen identification approach powered by AI is now a useful auxiliary tool for the prediction of allergenicity of food proteins using deep learning models by overcoming the limitations of low accuracy traditional machine learning models (Wang et al., 2021). A novel chemometric method for analysing and investigating the allergenic properties of dietary proteins has been developed by using machine learning. The approach is based on rating descriptors and evaluating their classification performance. It is necessary to create a reliable and effective protein categorization system in order to overcome the issue of food allergies (Nedyalkova et al., 2023). This information can then be used to guide the selection of plant varieties and to design breeding strategies that reduce the expression of allergenic proteins or to develop targeted approaches for modifying allergenic proteins. Further, bioinformatics can potentially analyze large datasets of allergen information and identify common allergenic epitopes across different plant species. This can help researchers identify potential targets for developing plant varieties with

reduced allergen content. This information can be used to design novel food processing techniques that can reduce the allergenicity of plant foods. For instance, AI algorithms can be used to model the effects of different processing conditions on the structure and function of allergenic proteins, allowing researchers to identify conditions that reduce allergenicity without compromising the nutritional quality or taste of the food. Further, AI can help researchers optimize the production process of hypoallergenic plant products by analyzing factors such as temperature, humidity, and nutrient levels. By analyzing the structure and function of allergenic proteins, AI algorithms can identify potential targets for genetic engineering or protein modification that can reduce allergenicity while maintaining the nutritional value of the food. The scope of AI and bioinformatics for reducing plant food allergens is vast and includes applications in genetic analysis, allergen identification, and production optimization (MacMath et al., 2023; Solanki et al., 2020). These technologies have the potential to revolutionize the field of plant food allergen reduction, leading to safer and more accessible food for individuals with allergies for their health and well-being.

6.3 | Developing varieties with reduced allergen content using genetic engineering and gene editing technologies

The genetic engineering approach RNAi is a powerful tool that can be used to improve various traits in crops, including nutrient value, disease or pathogen resistance, and crop allergenicity reduction. RNAi is a natural biological process that regulates gene expression by suppressing the activity of specific genes. Further, integrating RNAi technology with conventional breeding approaches contributes to the development of improved crop varieties that address nutritional, health, and environmental challenges (Rajam, 2020). By using RNAi, the expression of allergen proteins can be reduced or eliminated, resulting in crops with reduced allergenic potential and this approach holds promise for improving the safety of food products for individuals with food allergies. A significant progress has been made in gene silencing approaches to suppress the immunodominant allergen Ara h 2 in peanuts (Dodo et al., 2008). Decreased allergenicity to the immune dominant Ara h 2 protein was achieved in peanuts using RNAi technology (Dodo et al., 2008). In addition, RNAi approaches have been successfully used to suppress the expression of Ara h 2 and Ara h 6 allergens without adverse effects on seed germination and plant growth and development (Chandran et al., 2015; Chu et al., 2008). Similarly, the silencing of gluten synthesis in wheat led to the production of a low-gluten wheat variety (Wen et al., 2012). Moreover, the ω -5 gliadin-free 1BS-18 wheat line had low efficiency in

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inducing allergy symptoms in guinea pigs (Kohno et al., 2016).

In addition to RNAi, the use of technological advancement, the discovery and characterization of plant food allergen genes offers a significant opportunity for successful genetic modifications (Brackett et al., 2022). Biotechnological techniques, such as gene editing or genome editing, are widely employed to produce designer crops with desired traits (Wang et al., 2016; Fernie and Yan, 2019; Awasthi et al., 2022). Site-directed nuclease (SDN) methods have also recently been employed to achieve genetic alterations by a precise cleavage in the intended target region of the genome. A few SDN tools, such as zinc-finger nucleases (ZFN) (Urnov et al., 2010), transcription activator-like effector nucleases (TALEN) (Joung & Sander, 2013), and CRISPR (Wang et al., 2016) are currently used for gene editing (Camerlengo et al., 2020; Kaur et al., 2020; Lakhani et al., 2022; Singh et al., 2023).

CRISPR/Cas9 is a well-proven technology useful for creating desirable mutations at specific genetic locations that can also be applied for key polyploid crops, such as peanut and wheat, and diploid crops, such as soybean, kidney beans, and mustard (Assou et al., 2022; Bortesi & Fischer, 2015; Gao, 2021; Steinward & Ronald, 2020; Weeks, 2017). CRISPR/Cas9 has been used successfully to edit the Ara h 2 genes in peanuts (Biswas et al., 2022). This technology has also been successfully used to edit fatty acid desaturase (AhFAD2) in peanuts to increase the oleic content (Yuan et al., 2019). Similarly, the roles of nod factor receptors (AhNFRs) were verified by using CRISPR/Cas9 (Shu et al., 2020). The research group of Rustgi et al. (2022) has developed low-gluten producing wheat by expressing glutenases through genetic engineering. They have also screened selected genotypes of the USDA and ICRISAT mini-core of peanuts for low allergenic content and targeted the major allergen genes by CRISPR/Cas approaches.

The successful genome editing of GmDcl4a and GmDcl4b genes was reported in hairy roots (Curtin et al., 2011). Recently, two major soybean allergens, Gly m Bd 28 k and Gly m Bd 30 k, were removed from seeds using CRISPR/Cas9mediated site-directed mutagenesis (Sugano et al., 2020). The CRISPR/Cas9 system was also used to knock out the TaMLO locus in wheat (Shan et al., 2013), including TaPDS and TalNOX (Upadhyay et al., 2013). The three TaMLO alleles were silenced together for resistance to powdery mildew in bread wheat (Wang et al., 2014). The wheat TaLOX2 gene was silenced by expressing sgRNA under the transcriptional control of the TaU6 promoter (Shan et al., 2013). The major allergenic proteins ATIs were recently silenced using the CRISPR/Cas9 approach, in order to decrease wheat allergies such as Baker's asthma and nonceliac sensitivity (Camerlengo et al., 2020).

7 | SUMMARY AND FUTURE PROSPECTS

Legume food crops are the major source of essential amino acids and plant-based proteins besides their natural antinutrients and allergenic substances that hampers digestibility and trigger an abnormal immune response which makes them undesirable for human consumption. The implementation of novel approaches like high throughput phenotyping, target gene identification assisted by AI, genomics-assisted selection, and precision breeding by gene editing can help to address the challenges of food safety and security. In the post-genomic era, researchers have made enormous progress in genome sequencing and structural analysis of legume genomes by the intervention of NGS and bioinformatics. These genome information resources are facilitating gene discovery and development of molecular markers of complex traits toward the generation of superior legume crops. Particularly, in the matter of food safety, allergies have become a serious health concern in both developed and developing countries in the era of modernization. The increasing prevalence of FA presents a formidable challenge to researchers seeking to find accurate, rapid diagnostic methods, as well as prevention and treatment measures for vulnerable people.

Although, a variety of thermal and nonthermal food processing methods have been applied by the food service establishments, however, removal of allergenic substances from the food source material is still a challenge for packaged foods. Therefore, the combined use of technological advancement in AI, bioinformatics, molecular breeding, RNAi, and CRISPR/Cas-based gene editing can help to develop varieties of legume crops with reduced allergen. Realizing the strength of currently available tools and technologies, developing allergen-free crop varieties seems very difficult; nevertheless, the available genetic variation among diverse germplasm can be exploited to accumulate superior alleles promoting production of low allergen content in seeds for ensuring food safety. On the other hand, the best possible cure needs to be explored to treat affected patients in addition to developing tolerance from childhood through mini-exposure to diverse food including causing allergenicity.

AUTHOR CONTRIBUTION

Vadthya Lokya: Data curation; investigation; methodology; writing—original draft. Sejal Parmar: Data curation; investigation; methodology; writing—original draft. Arun K. Pandey: Data curation; writing—original draft. Hari K. Sudini: Investigation; methodology; writing—review and editing. Dongxin Huai: Investigation; methodology; writing—review and editing. Peggy Ozias-Akins: Data curation; investigation; methodology; writing—review and editing. Christine H. Foyer: Investigation; methodology; writing—review and editing; Chogozie Victor Nwosu:

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investigation; methodology; writing—review and editing. Barbara Karpinska: Investigation; methodology; writing review and editing. Alison Baker: Investigation; methodology; writing—review and editing. **Pei Xu**: Data curation; methodology; writing—review and editing. Boshou Liao: Data curation; methodology; writing—review and editing. Reyazul Rouf Mir: Data curation; investigation; methodology; writing—review and editing. Xiaoping Chen: Data curation; methodology; writing—review and editing. Baozhu Guo: Data curation; methodology; writing—review and editing. Henry T. Nguyen: Data curation; investigation; methodology; writing—review and editing. Rakesh Kumar: Data curation; methodology; writing—review and editing. Sandeep K. Bera: Data curation; investigation; methodology; writing—review and editing. Prashant Singam: Data curation; methodology; writing—review and editing. Anirudh Kumar: Data curation; methodology; writing—review and editing. Rajeev K. Varshney: Investigation; methodology; writing-review and editing. Manish K. Pandey: Conceptualization; data curation; funding acquisition; investigation; project administration; resources; supervision; writingreview and editing.

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CONFLICT OF INTEREST STATEMENT

The authors declare there is no conflict of interest.

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REFERENCES

Ahn, K., Bardina, L., Grishina, G., Beyer, K., & Sampson, H. A. (2009). Identification of two pistachio allergens, Pis v 1 and Pis v 2, belonging

- to the 2S albumin and 11S globulin family. *Clinical & Experimental Allergy*, 39(6), 926–934.
- Alcocer, M., Rundqvist, L., & Larsson, G. (2012). Ber e 1 protein The versatile major allergen from Brazil nut seeds. *Biotechnology Letters*, 34, 597–610.
- Ansotegui, I. J., Melioli, G., Canonica, G. W., Caraballo, L., Villa, E., Ebisawa, M., & Zuberbier, T. (2020). IgE allergy diagnostics and other relevant tests in allergy, a World Allergy Organization position paper. *The World Allergy Organization Journal*, 13(2), 100080. https://doi.org/10.1016/j.waojou.2019.100080
- Apostolovic, D., Luykx, D., Warmenhoven, H., Verbart, D., Stanic-Vucinic, D., de Jong, G. A., Velickovic, T. C., & Koppelman, S. J. (2013). Reduction and alkylation of peanut allergen isoforms Ara h 2 and Ara h 6; characterization of intermediate-and end products. Biochimica et Biophysica Acta, 1834(12), 2832–2842. https://doi.org/10.1016/j.bbapap.2013.10.004
- Arakali, S. R., Green, T. D., & Dinakar, C. (2017). Prevalence of food allergies in South Asia. Annals of Allergy, Asthma & Immunology, 118(1), 16–20.
- Asero, R., Mistrello, G., Roncarolo, D., Amato, S., Zanoni, D., Barocci, F., & Caldironi, G. (2003). Detection of clinical markers of sensitization to profilin in patients allergic to plant-derived foods. *Journal of Allergy and Clinical Immunology*, 112(2), 427–432. https://doi.org/10.1067/mai.2003.1611
- Assou, J., Zhang, D., Roth, K. D., Steinke, S., Hust, M., Reinard, T., & Boch, J. (2022). Removing the major allergen Bra j I from brown mustard (Brassica juncea) by CRISPR/Cas9. *The Plant Journal*, 109(3), 649–663. https://doi.org/10.1111/tpj.15584
- Asturias, J. A., Ibarrola, I., Bartolome, B., Ojeda, I., Malet, A., & Martinez, A. (2002). Purification and characterization of Pla a 1, a major allergen from *Platanus acerifolia* pollen. *Allergy*, *57*(3), 221–227. https://doi.org/10.1034/j.1398-9995.2002.03406.x
- Awasthi, P., Khan, S., Lakhani, H., Chaturvedi, S., Kaur, N., Singh, J., Kesarwani, A. K., & Tiwari, S. (2022). Transgene-free genome editing supports the role of carotenoid cleavage dioxygenase 4 as a negative regulator of β-carotene in banana. *Journal of Experimental Botany*, 73(11), 3401–3416.
- Baker, R. (2018). The global status of food allergen labeling laws. *California Western Law Review*, 54(2), 4.
- Ballmer-Weber, B. K., & Beyer, K. (2018). Food challenges. *Journal of Allergy and Clinical Immunology*, 141(1), 69–71. https://doi.org/10.1016/j.jaci.2017.06.038
- Bardocz, S., Grant, G., Ewen, S. W., Duguid, T. J., Brown, D. S., Englyst, K., & Pusztai, A. (1995). Reversible effect of phytohaemagglutinin on the growth and metabolism of rat gastrointestinal tract. *Gut*, *37*(3), 353–360. https://doi.org/10.1136/gut.37.3.353
- Barro, F., Iehisa, J. C., Giménez, M. J., García-Molina, M. D., Ozuna, C. V., Comino, I., Sousa, C., & Gil-Humanes, J. (2016). Targeting of prolamins by RNAi in bread wheat: Effectiveness of seven silencing-fragment combinations for obtaining lines devoid of coeliac disease epitopes from highly immunogenic gliadins. *Plant Biotechnology Journal*, 14(3), 986–996. https://doi.org/10.1111/pbi.12455
- Bauchet, G. J., Bett, K. E., Cameron, C. T., Campbell, J. D., Cannon, E. K., Cannon, S. B., & Zhao, P. X. (2019). The future of legume genetic data resources: Challenges, opportunities, and priorities. *Legume Science*, 1(1), 16. https://doi.org/10.1002/leg3.16
- Beyer, K., Grishina, G., Bardina, L., Grishin, A., & Sampson, H. A. (2002). Identification of an 11S globulin as a major hazelnut food

VADTHYA ET AL. The Plant Genome 25 of 32

allergen in hazelnut-induced systemic reactions. *Journal of Allergy and Clinical Immunology*, 110(3), 517–523.

- Bhalla, P. L., & Singh, M. B. (2008). Biotechnology-based allergy diagnosis and vaccination. *Trends in Biotechnology*, 26(3), 153–161. https://doi.org/10.1016/j.tibtech.2007.11.010
- Bhattacharya, K., Sircar, G., Dasgupta, A., & Bhattacharya, S. G. (2018).
 Spectrum of allergens and allergen biology in India. *International Archives of Allergy and Immunology*, 177(3), 219–237. https://doi.org/10.1159/000490805
- Boldt, A., Fortunato, D., Conti, A., Petersen, A., Ballmer-Weber, B., Lepp, U., & Becker, W. M. (2005). Analysis of the composition of an immunoglobulin E reactive high molecular weight protein complex of peanut extract containing Ara h 1 and Ara h 34. *Proteomics*, 5(3), 675–686.
- Biswas, S., Wahl, N. J., Thomson, M. J., Cason, J. M., McCutchen, B. F., & Septiningsih, E. M. (2022). Optimization of protoplast isolation and transformation for a pilot study of genome editing in peanut by targeting the allergen gene *Arah 2. International Journal of Molecular Sciences*, 23(2), 837. https://doi.org/10.3390/ijms23020837
- Bortesi, L., & Fischer, R. (2015). The CRISPR/Cas9 system for plant genome editing and beyond. *Biotechnology Advance*, 33(1), 41–52. https://doi.org/10.1016/j.biotechadv.2014.12.006
- Brackett, N. F., Pomés, A., & Chapman, M. D. (2022). New frontiers: Precise editing of allergen genes using CRISPR. Frontiers in Allergy, 2, 109. https://doi.org/10.3389/falgy.2021.821107
- Breiteneder, H., & Mills, E. C. (2005). Plant food allergens—structural and functional aspects of allergenicity. *Biotechnology Advance*, 23(6), 395–399. https://doi.org/10.1016/j.biotechadv.2005.05.004
- Breiteneder, H., & Radauer, C. (2004). A classification of plant food allergens. *The Journal of Allergy and Clinical Immunology*, *113*(5), 821–830. https://doi.org/10.1016/j.jaci.2004.01.779
- Burks, A. W., Cockrell, G., Stanley, J. S., Helm, R. M., & Bannon, G. A. (1995). Recombinant peanut allergen Ara h I expression and IgE binding in patients with peanut hypersensitivity. *The Journal of Clinical Investigation*, 96(4), 1715–1721.
- Burks, A. W., Tang, M., Sicherer, S., Muraro, A., Eigenmann, P. A., Ebisawa, M., Fiocchi, A., Chiang, W., Beyer, K., Wood, R., & Hourihane, J. (2012). ICON: Food allergy. *Journal of Allergy and Clinical Immunology*, 129(4), 906–920. https://doi.org/10.1016/j.jaci. 2012.02.001
- Cabanillas, B., Jappe, U., & Novak, N. (2018). Allergy to peanut, soybean, and other legumes: recent advances in allergen characterization, stability to processing and IgE cross-reactivity. *Molecular Nutrition & Food Research*, 62(1), 1700446. https://doi.org/10.1002/mnfr. 201700446
- Cabanillas, B., Pedrosa, M. M., Rodriguez, J., Muzquiz, M., Maleki, S. J., Cuadrado, C., Burbano, C., & Crespo, J. F. (2012). Influence of enzymatic hydrolysis on the allergenicity of roasted peanut protein extract. *International Archives of Allergy and Immunology*, 157(1), 41–50. https://doi.org/10.1159/000324681
- Camerlengo, F., Frittelli, A., Sparks, C., Doherty, A., Martignago, D., Larré, C., Lupi, R., Sestili, F., & Masci, S. (2020). CRISPR-Cas9 multiplex editing of the α-amylase/trypsin inhibitor genes to reduce allergen proteins in durum wheat. Frontiers in Sustainable Food Systems, 4, 104. https://doi.org/10.3389/fsufs.2020.00104
- Chandran, M., Chu, Y., Maleki, S. J., & Ozias-Akins, P. (2015). Stability of transgene expression in reduced allergen peanut (*Arachis hypogaea* L.) across multiple generations and at different soil sulfur levels. *Jour-*

- nal of Agricultural and Food Chemistry, 63(6), 1788–1797. https://doi.org/10.1021/jf504892f
- Chang, F., Eng, L., & Chang, C. (2023). Food allergy labeling laws: International Guidelines for Residents and Travelers. *Clinical Reviews in Allergy & Immunology*, 1–18. Advance online publication. https://doi.org/10.1007/s12016-023-08960-6
- Chatel, J. M., Bernard, H., & Orson, F. M. (2003). Isolation and characterization of two complete Ara h 2 isoforms cDNA. *International Archives of Allergy and Immunology*, 131(1), 14–18.
- Chu, Y., Faustinelli, P., Ramos, M. L., Hajduch, M., Stevenson, S., Thelen, J. J., Maleki, S. J., Cheng, H., & Ozias-Akins, P. (2008). Reduction of IgE binding and nonpromotion of *Aspergillus flavus* fungal growth by simultaneously silencing Ara h 2 and Ara h 6 in peanut. *Journal of Agricultural and Food Chemistry*, 56(23), 11225–11233. https://doi.org/10.1021/jf802600r
- Chung, S., & Champagne, E. T. (2008). Using phenolic compounds to reduce the allergenic properties of peanut extracts and peanut butter slurries. *The Journal of Allergy and Clinical Immunology*, *121*(2), 249. https://doi.org/10.1016/j.jaci.2007.12.985
- Chung, S. Y., & Reed, S. (2012). Removing peanut allergens by tannic acid. Food Chemistry, 134(3), 1468–1473. https://doi.org/10.1016/j. foodchem.2012.03.057
- Cianferoni, A. (2020). Eosinophilic esophagitis as a side effect of food oral immunotherapy. *Medicina*, 56(11), 618. https://doi.org/10.3390/ medicina56110618
- Codina, R., Lockey, R. F., Eernández-Caldas, E., & Rama, R. (1997).
 Purification and characterization of a soybean hull allergen responsible for the Barcelona asthma outbreaks. II. Purification and sequencing of the Gly m 2 allergen. *Clinical & Experimental Allergy*, 27(4), 424–430.
- Corzo-Martínez, M., Villamiel, M., & Moreno, F. J. (2017). Impact of high-intensity ultrasound on protein structure and functionality during food processing. In M. Villamiel, A. Montilla, J. V. García-Pérez, J. A. Cárcel, J. Benedito (Eds), *Ultrasound in food processing: Recent* advances (pp. 417–436). Wiley.
- Costa, C., Coimbra, A., Vítor, A., Aguiar, R., Ferreira, A. L., & Todo-Bom, A. (2020). Food allergy—from food avoidance to active treatment. Scandinavian Journal of Immunology, 91(1),12824. https://doi.org/10.1111/sji.12824
- Crowell, D. N., John, M. E., Russell, D., & Amasino, R. M. (1992). Characterization of a stress-induced, developmentally regulated gene family from soybean. *Plant Molecular Biology*, 18, 459–466.
- Curtin, S. J., Zhang, F., Sander, J. D., Haun, W. J., Starker, C., Baltes, N. J., Reyon, D., Dahlborg, E. J., Goodwin, M. J., Coffman, A. P., & Dobbs, D. (2011). Targeted mutagenesis of duplicated genes in soybean with zinc-finger nucleases. *Plant Physiology*, 156(2), 466–473. https://doi.org/10.1104/pp.111.172981
- D'Agostino, N., Buonanno, M., Ayoub, J., Barone, A., Monti, S. M., & Rigano, M. M. (2019). Identification of non-specific Lipid Transfer Protein gene family members in *Solanum lycopersicum* and insights into the features of Sola 1 3 protein. *Scientific Report*, 9(1), 1–16.
- Diowksz, A., & Leszczyńska, J. (2014). Hypoallergenic wheat bread: Response to an emerging issue. *Food and Agriculture Immunology*, 25(4), 535–544. https://doi.org/10.1080/09540105.2013.848184
- Dodo, H., Marsic, D., Callender, M., Cebert, E., & Viquez, O. (2002). Screening 34 peanut introductions for allergen content using ELISA. Food and Agriculture Immunology, 14(2), 147–154. https://doi.org/ 10.1080/09540100220145179

26 of 32

- Dong, X., Wang, J., & Raghavan, V. (2021). Critical reviews and recent advances of novel non-thermal processing techniques on the modification of food allergens. *Critical Reviews in Food Science and Nutrition*, 61(2), 196–210. https://doi.org/10.1080/10408398.2020. 1722942
- Dreborg, S. (2001). Diagnosis of food allergy: Tests in vivo and in vitro. *Pediatric Allergy and Immunology*, 12(14), 24–30. https://doi.org/10. 1034/j.1399-3038.2001.121406.x
- Dubiela, P., Kabasser, S., Smargiasso, N., Geiselhart, S., Bublin, M., Hafner, C., & Hoffmann-Sommergruber, K. (2018). Jug r 6 is the allergenic vicilin present in walnut responsible for IgE crossreactivities to other tree nuts and seeds. *Scientific Reports*, 8(1), 11366.
- Dubois, A. E., Pagliarani, G., Brouwer, R. M., Kollen, B. J., Dragsted, L. O., Eriksen, F. D., Callesen, O., Gilissen, L. J., Krens, F. A., Visser, R. G., & Smulders, M. J. M. (2015). First successful reduction of clinical allergenicity of food by genetic modification: Mald1-silenced apples cause fewer allergy symptoms than the wild-type cultivar. *Allergy*, 70(11), 1406–1412. https://doi.org/10.1111/all.12684
- Du Toit, G., Roberts, G., Sayre, P. H., Bahnson, H. T., Radulovic, S., Santos, A. F., Brough, H. A., Phippard, D., Basting, M., Feeney, M., & Turcanu, V. (2015). Randomized trial of peanut consumption in infants at risk for peanut allergy. *The New England Journal of Medicine*, 372, 803–813. https://doi.org/10.1056/NEJMoa1414850
- Ekezie, F. G. C., Cheng, J. H., & Sun, D. W. (2018). Effects of non-thermal food processing technologies on food allergens: A review of recent research advances. *Trends in Food Science & Technology*, 74, 12–25.
- FAO, WHO. (n.d.). Codex Alimentarius International Food Standards ("CAIFS") About Codex. FAO, WHO. https://www.fao.orgfao-whocodexalimentariusabout-codexenc453333
- Fernandes, H., Michalska, K., Sikorski, M., & Jaskolski, M. (2013).
 Structural and functional aspects of PR-10 proteins. The Federation of European Biochemical Societies Journal, 280(5), 1169–1199.
- Filep, S., Block, D. S., Smith, B. R., King, E. M., Commins, S., Kulis, M., Vickery, B. P., & Chapman, M. D. (2018). Specific allergen profiles of peanut foods and diagnostic or therapeutic allergenic products. *The Journal of Allergy and Clinical Immunology*, 141(2), 626–631. https://doi.org/10.1016/j.jaci.2017.05.049
- Food Safety and Standards (Labelling and Display) Regulations. (2020). https://www.fssai.gov.in/upload/uploadfiles/files/Compendium_Labelling_Display_23_09_2021.pdf
- Fox, M., Mugford, M., Voordouw, J., Cornelisse-Vermaat, J., Antonides, G., de la Hoz Caballer, B., & Clark, A. B. (2013). Health sector costs of self-reported food allergy in Europe: A patient-based cost of illness study. *European Journal of Public Health*, 23(5), 757–762. https://doi.org/10.1093/eurpub/ckt010
- Fu, G., Zhao, K., Chen, H., Wang, Y., Nie, L., Wei, H., & Wan, C. (2019). Effect of 3 lactobacilli on immunoregulation and intestinal microbiota in a β-lactoglobulin–induced allergic mouse model. *Journal of Dairy Science*, 102(3), 1943–1958. https://doi.org/10.3168/jds.2018-15683

- Gao, C. (2021). Genome engineering for crop improvement and future agriculture. Cell, 184(6), 1621–1635. https://doi.org/10.1016/j.cell. 2021.01.005
- Gao, Z. S., Zheng, M., Gilissen, L. J., Shen, H. H., Frewer, L. J., Guan, R. X., & Qiu, L. J. (2012). Hypoallergenic soybean, from genes to cultivar. *Multidisciplinary Approaches to Allergies*, 347–358.
- Garino, C., Zuidmeer, L., Marsh, J., Lovegrove, A., Morati, M., Versteeg, S., & van Ree, R. (2010). Isolation, cloning, and characterization of the 2S albumin a new allergen from hazelnut. *Molecular Nutrition & Food Research*, 54(9), 1257–1265.
- Geiselhart, S., Hoffmann-Sommer, G. K., & Bublin, M. (2018). Tree nut allergens. *Molecular Immunology*, 100, 71–81. https://doi.org/10. 1016/j.molimm.2018.03.011
- Gilissen, L. J., van der Meer, I. M., & Smulders, M. J. (2014). Reducing the incidence of allergy and intolerance to cereals. *Journal of Cereal Science*, 59(3), 337–353. https://doi.org/10.1016/j.jcs.2014.01.005
- González, R., Zapatero, L., Caravaca, F., & Carreira, J. (1991). Identification of soybean proteins responsible for respiratory allergies. International Archives of Allergy and Applied Immunology, 95(1), 53–57.
- Gruehn, S., Suphioglu, C., O'Hehir, R. E., & Volkmann, D. (2003).
 Molecular cloning and characterization of hazel pollen protein (70 kD) as a luminal binding protein (BiP) A novel cross-reactive plant allergen. *International Archives of Allergy and Immunology*, 131(2), 91–100.
- Gupta, R., Holdford, D., Bilaver, L., Dyer, A., Holl, J. L., & Meltzer, D. (2013). The economic impact of childhood food allergy in the United States. *Journal of the American Medical Association Pediatrics*, 167(11), 1026–1031.
- Gupta, R. K., Gupta, K., Sharma, A., Das, M., Ansari, I. A., & Dwivedi, P. D. (2018b). Maillard reaction in food allergy: Pros and cons. *Critical Reviews in Food Science and Nutrition*, 58(2), 208–226. https://doi.org/10.1080/10408398.2016.1152949
- Gupta, R. S., Springston, E. E., Smith, B., Warrier, M. R., Pongracic, J., & Holl, J. L. (2012). Geographic variability of childhood food allergy in the United States. *Clinical Pediatrics*, 51(9), 856–861. https://doi. org/10.1177/0009922812448526
- Gupta, R. S., Springston, E. E., Warrier, M. R., Smith, B., Kumar, R., Pongracic, J., & Holl, J. L. (2011). The prevalence, severity, and distribution of childhood food allergy in the United States. *Pediatrics*, 128(1), 9–17. https://doi.org/10.1542/peds.2011-0204
- Gupta, R. S., Warren, C. M., Smith, B. M., Blumenstock, J. A., Jiang, J., Davis, M. M., & Nadeau, K. C. (2018a). The public health impact of parent-reported childhood food allergies in the United States. *Pediatrics*, 142(6), e20181235. https://doi.org/10.1542/peds. 2018-1235
- Gupta, R. S., Warren, C. M., Smith, B. M., Jiang, J., Blumenstock, J. A., Davis, M. M., Schleimer, R. P., & Nadeau, K. C. (2019). Prevalence and severity of food allergies among US adults. *JAMA Network Open*, 2(1), 185630–185630. https://doi.org/10.1001/jamanetworkopen.2018.5630
- Hamid, M. A., Azad, M. A. K., & Howelider, M. A. R. (2006). Development of three peanut varieties with improved quantitative and qualitative traits through induced mutation. *Plant Mutation Reports*, *1*(2), 14–16.
- Heinzerling, L., Mari, A., Bergmann, K. C., Bresciani, M., Burbach, G., Darsow, U., Durham, S., Fokkens, W., Gjomarkaj, M., Haahtela, T., Bom, A. T., Wöhrl, S., Maibach, H., & Lockey, R. (2013). The skin

VADTHYA ET AL. The Plant Genome 27 of 32

prick test - European standards. *Clinical and Translational Allergy*, 3(1), 3. https://doi.org/10.1186/2045-7022-3-3

- Herman, E. M. (2003). Genetically modified soybeans and food allergies. *Journal of Experimental Botany*, 54(386), 317–1319. https://doi.org/ 10.1093/jxb/erg164
- Hill, D. A., Grundmeier, R. W., Ram, G., & Spergel, J. M. (2016). The epidemiologic characteristics of healthcare provider-diagnosed eczema, asthma, allergic rhinitis, and food allergy in children: A retrospective cohort study. *BMC Pediatrics*, 16, 133. https://doi.org/10. 1186/s12887-016-0673-z
- Hu, Y., Xu, Z., Jiang, F., Li, S., Liu, S., Wu, M., Yan, C., Tan, J., Yu, G., Hu, Y., & Yin, Y. (2020). Relative impact of meteorological factors and air pollutants on childhood allergic diseases in Shanghai, China. Science of the Total Environment, 706, 135975. https://doi.org/10.1016/j.scitotenv.2019.135975
- Isleib, T. G., & Wynne, J. C. (1992). Use of plant introductions in peanut improvement. Use of Plant Introductions in Cultivar Development Part 2, 20, 75–116.
- Jackson, K. D., Howie, L. D., & Akinbami, O. J. (2013). Trends in allergic conditions among children: United States, 1997–2011 (No. 121).
 US Department of Health and Human Services, Centers for Disease Control and Prevention, National Center for Health Statistics.
- Janila, P., Variath, M. T., Pandey, M. K., Desmae, H., Motagi, B. N., Okori, P., & Varshney, R. K. (2016). Genomic tools in groundnut breeding program: Status and perspectives. *Frontiers in Plant Science*, 7, 289. https://doi.org/10.3389/fpls.2016.00289
- Jeong, K. H., Choi, M. S., Lee, S. K., Seo, M. J., Hwang, T. Y., Yun, H. T., Kim, H. S., Kim, J. T., Kwon, Y. U., & Kim, Y. H. (2013). Development of low Gly m Bd 30K (P34) allergen breeding lines using molecular marker in soybean [Glycine max L.]. *Plant Breeding and Biotechnology*, 1(3), 298–306. https://doi.org/10.9787/PBB. 2013.1.3.298
- Jouanin, A., Boyd, L., Visser, R. G., & Smulders, M. J. (2018). Development of wheat with hypo immunogenic gluten obstructed by the gene editing policy in Europe. *Frontiers of Plant Science*, 9, 1523. https://doi.org/10.3389/fpls.2018.01523
- Joung, J. K., & Sander, J. D. (2013). TALENs: A widely applicable technology for targeted genome editing. *Nature Reviews Molecular Cell Biology*, 14(1), 49–55. https://doi.org/10.1038/nrm3486
- Juhász, A., Belova, T., Florides, C. G., Maulis, C., Fischer, I., Gell, G., Birinyi, Z., Ong, J., Keeble-Gagnère, G., Maharajan, A., & Ma, W. (2018). Genome mapping of seed-borne allergens and immunore-sponsive proteins in wheat. *Science Advances*, 4(8), 8602. https://doi.org/10.1126/sciadv.aar8602
- Kabasser, S., Hafner, C., Chinthrajah, S., Sindher, S. B., Kumar, D., Kost, L. E., & Bublin, M. (2021). Identification of Pru du 6 as a potential marker allergen for almond allergy. *Allergy*, 76(5), 1463–1472.
- Kadam, K., Karbhal, R., Jayaraman, V. K., Sawant, S., & Kulkarni-Kale, U. (2017). AllerBase: A comprehensive allergen knowledgebase. *Database*, 2017, bax066. https://doi.org/10.1093/database/bax066
- Kaiser, N., Douches, D., Dhingra, A., Glenn, K. C., Herzig, P. R., Stowe, E. C., & Swarup, S. (2020). The role of conventional plant breeding in ensuring safe levels of naturally occurring toxins in food crops. *Trends* in Food Science & Technology, 100, 51–66.
- Kasera, R., Singh, B. P., Lavasa, S., Prasad, K. N., Sahoo, R. C., & Singh, A. B. (2011). Kidney bean: A major sensitizer among legumes in asthma and rhinitis patients from India. *PLoS ONE*, 6, 27193. https://doi.org/10.1371/journal.pone.0027193

- Kaur, N., Alok, A., Kumar, P., Kaur, N., Awasthi, P., Chaturvedi, S., & Tiwari, S. (2020). CRISPR/Cas9 directed editing of lycopene epsilon-cyclase modulates metabolic flux for β-carotene biosynthesis in banana fruit. *Metabolic Engineering*, 59, 76–86. https://doi.org/10.1016/j.ymben.2020.01.008
- Kaur, N., Awasthi, P., & Tiwari, S. (2020). Fruit crops improvement using CRISPR/Cas9 system. In *Genome Engineering via CRISPR-Cas9 System*. (pp. 131–145) Academic Press.
- Keet, C., & Wood, R. (2011). Overview of mucosal immunity and development of oral tolerance. Food Allergy, 1–14.
- Khoury, P., Srinivasan, R., Kakumanu, S., Ochoa, S., Keswani, A., Sparks, R., & Rider, N. L. (2022). A framework for augmented intelligence in allergy and immunology practice and research—A Work Group Report of the AAAAI Health Informatics, Technology, and Education Committee. *The Journal of Allergy and Clinical Immunology. In practice*, 10(5), 1178–1188. https://doi.org/10.1016/j.jaip.2022.01.047
- Kleber-Janke, T., Crameri, R., Appenzeller, U., Schlaak, M., & Becker, W. M. (1999). Selective cloning of peanut allergens, including profilin and 2S albumins, by phage display technology. *International Archives of Allergy and Immunology*, 119(4), 265–274.
- Knoll, J. E., Ramos, M. L., Zeng, Y., Holbrook, C. C., Chow, M., Chen, S., Maleki, S., Bhattacharya, A., & Ozias-Akins, P. (2011). TILLING for allergen reduction and improvement of quality traits in peanut (*Arachis hypogaea* L.). *BMC Plant Biology*, 11(1), 1–13. https://doi.org/10.1186/1471-2229-11-81
- Kohno, K., Takahashi, H., Endo, T. R., Matsuo, H., Shiwaku, K., & Morita, E. (2016). Characterization of a hypoallergenic wheat line lacking ω-5 gliadin. *Allergology International*, 65(4), 400–405. https://doi.org/10.1016/j.alit.2016.03.002
- Kopper, R. A., Odum, N. J., Sen, M., Helm, R. M., Stanley, J. S., & Burks, A. W. (2005). Peanut protein allergens: The effect of roasting on solubility and allergenicity. *International Archives of Allergy and Immunology*, 136(1), 16–22. https://doi.org/10.1159/000082580
- Krause, S., Latendorf, T., Schmidt, H., Darcan-Nicolaisen, Y., Reese, G., Petersen, A., Janssen, O., & Becker, W. M. (2010). Peanut varieties with reduced Ara h 1 content indicating no reduced allergenicity. *Molecular Nutrition & Food Research*, 54(3), 381–7.
- Lakhani, H., Thakur, N., & Tiwari, S. (2022). Genome editing for vegetatively propagated crops improvement: A new horizon of possibilities. *Journal of Plant Biochemistry and Biotechnology*, 1–12.
- Lauer, I., Foetisch, K., Kolarich, D., Ballmer-Weber, B. K., Conti, A., Altmann, F., & Scheurer, S. (2004). Hazelnut (Corylus avellana) vicilin Cor a 11 molecular characterization of a glycoprotein and its allergenic activity. *Biochemical Journal*, 383(2), 327–334.
- Lee, A. J., Thalayasingam, M., & Lee, B. W. (2013). Food allergy in Asia: How does it compare? *Asia Pacific Allergy*, *3*, 3–14. https://doi.org/10.5415/apallergy.2013.3.1.3
- Lehto, M., Palosuo, K., Varjonen, E., Majuri, M. L., Andersson, U., Reunala, T., & Alenius, H. (2003). Humoral and cellular responses to gliadin in wheat-dependent, exercise-induced anaphylaxis. *Clinical & Experimental Allergy*, 33(1), 90–95.
- Lemke, S., Tao, X., & Kushner, G. J. (2022). Assuring the food safety of crops developed through breeding. ACS Agricultural Science & Technology, 2(6), 1151–1165.
- Leung, A. S. Y., Leung, N. Y. H., Wai, C. Y. Y., Leung, T. F., & Wong, G. W. K. (2018). Allergen immunotherapy for food allergy from the Asian perspective: Key challenges and opportunities. *Expert*

Review of Clinical Immunology, 15(2), 153–164. https://doi.org/10.1080/1744666X

28 of 32

- Li, J., Ogorodova, L. M., Mahesh, P. A., Wang, M. H., Fedorova, O. S., Leung, T. F., Fernandez-Rivas, M., Mills, E. C., Potts, J., Kummeling, I., & Versteeg, S. A. (2020). Comparative study of food allergies in children from China, India, and Russia: The EuroPrevall-INCO surveys. *The Journal of Allergy and Clinical Immunology*, 8(4), 1349–1358.
- Lieberman, J. A., Bingemann, T. A., & Wang, J. (2020). Diagnostic challenges in anaphylaxis. *The Journal of Allergy and Clinical Immunology*, 8(4), 1177–1184.
- Liu, B., Lee, J. B., Chen, C. Y., Hershey, G. K. K., & Wang, Y. H. (2015).
 Collaborative interactions between type 2 innate lymphoid cells and antigen-specific CD4+ Th2 cells exacerbate murine allergic airway diseases with prominent eosinophilia. *The Journal of Immunology*, 194(8), 3583–3593. https://doi.org/10.4049/jimmunol.1400951
- Liu, Z., Wang, S., Zhang, Y., Feng, Y., Liu, J., & Zhu, H. (2023). Artificial intelligence in food safety: A decade review and bibliometric analysis. Foods (Basel, Switzerland), 12(6), 1242.
- Loh, W., & Tang, M. (2018). Adjuvant therapies in food immunotherapy. Immunology and Allergy Clinics of North America, 38(1), 89–101. https://doi.org/10.1016/j.iac.2017.09.008
- Lyons, S. A., Burney, P. G., Ballmer-Weber, B. K., Fernandez-Rivas, M., Barreales, L., Clausen, M., & Kowalski, M. L. (2019). Food allergy in adults: Substantial variation in prevalence and causative foods across Europe. *The Journal of Allergy and Clinical Immunology*, 7(6), 1920– 1928.
- MacMath, D., Chen, M., & Khoury, P. (2023). Artificial intelligence: Exploring the future of innovation in allergy immunology. *Current Allergy and Asthma Reports*, 23(6), 351–362. https://doi.org/10.1007/s11882-023-01084-z
- Mahesh, P. A., Wong, G. W., Ogorodova, L., Potts, J., Leung, T. F., Fedorova, O., & Versteeg, S. A. (2016). Prevalence of food sensitization and probable food allergy among adults in India: The EuroPrevall INCO study. *Allergy*, 71(7), 1010–1019. https://doi.org/10.1111/all.12868
- Maleki, S. J., Kopper, R. A., Shin, D. S., Park, C.-W., Compadre, C. M., Sampson, H., Burks, A. W., & Bannon, G. A. (2000). Structure of the major peanut allergen Ara h 1 may protect IgE-binding epitopes from degradation. *The Journal of Immunology*, 164(11), 5844–5849. https://doi.org/10.4049/jimmunol.164.11.5844
- Maleki, S. J., Viquez, O., Jacks, T., Dodo, H., Champagne, E. T., Chung, S. Y., & Landry, S. J. (2003). The major peanut allergen, Ara h 2, functions as a trypsin inhibitor, and roasting enhances this function. *Journal of Allergy and Clinical Immunology*, 112(1), 190–195. https://doi.org/10.1067/mai.2003.1551
- Maruyama, N. (2021). Components of plant-derived food allergens: Structure, diagnostics, and immunotherapy. Allergology International, 70(3), 291–302. https://doi.org/10.1016/j.alit.2021.05.001
- McClain, S., Stevenson, S. E., Brownie, C., Herouet-Guicheney, C., Herman, R. A., Ladics, G. S., Privalle, L., Ward, J. M., Doerrer, N., & Thelen, J. J. (2018). Variation in seed allergen content from three varieties of soybean cultivated in nine different locations in Iowa, Illinois, and Indiana. Frontiers of Plant Science, 9, 1025. https://doi.org/ 10.3389/fpls.2018.01025
- McWilliam, V., Koplin, J., Lodge, C., Tang, M., Dharmage, S., & Allen, K. (2015). The prevalence of tree nut allergy: A systematic review. *Current Allergy and Asthma Reports*, 15(9), 1–13. https://doi.org/10.1007/s11882-015-0555-8

- Meng, S., Tan, Y., Chang, S., Li, J., Maleki, S., & Puppala, N. (2020).
 Peanut allergen reduction and functional property improvement by means of enzymatic hydrolysis and transglutaminase crosslinking.
 Food Chemistry, 302, 125186. https://doi.org/10.1016/j.foodchem.
 2019 125186
- Messina, M., & Venter, C. (2020). Recent surveys on food allergy prevalence. *Nutrition Today*, 55(1), 22–29. https://doi.org/10.1097/ NT.00000000000000389
- Mikiashvili, N., & Yu, J. (2018). Changes in immunoreactivity of allergen-reduced peanuts due to post-enzyme treatment roasting. *Food Chemistry*, 256, 188–194. https://doi.org/10.1016/j.foodchem. 2018.02.119
- Mills, E. C., Jenkins, J. A., Alcocer, M. J., & Shewry, P. R. (2004). Structural, biological, and evolutionary relationships of plant food allergens sensitizing via the gastrointestinal tract. *Critical Reviews in Food Science and Nutrition*, 44(5), 379–407. https://doi.org/10.1080/ 10408690490489224
- Mills, E. N. C., Jenkins, J., Marigheto, N., Belton, P. S., Gunning, A. P., & Morris, V. J. (2002). Allergens of the cupin superfamily. *Biochemical Society Transactions*, 30(6), 925–929. https://doi.org/10.1042/bst0300925
- Mittag, D., Akkerdaas, J., Ballmer-Weber, B. K., Vogel, L., Wensing, M., Becker, W. M., & Vieths, S. (2004). Ara h 8, a Bet v 1-homologous allergen from peanut, is a major allergen in patients with combined birch pollen and peanut allergy. *Journal of Allergy and Clinical Immunology*, 114(6), 1410–1417.
- Monsalve, R., Villalba, M., Rico, M., Shewry, P. R., & Rodríguez, R. (2007). The 2S albumin proteins. In *Plant food allergens* (pp. 42–56).
- Motosue, M. S., Bellolio, M. F., Van Houten, H. K., Shah, N. D., & Campbell, R. L. (2018). National trends in emergency department visits and hospitalizations for food-induced anaphylaxis in US children. Pediatric Allergy and Immunology, 29(5), 538–544. https://doi.org/10.1111/pai.12908
- Nedyalkova, M., Vasighi, M., Azmoon, A., Naneva, L., & Simeonov, V. (2023). Sequence-based prediction of plant allergenic proteins: Machine learning classification approach. ACS Omega, 8(4), 3698–3704. https://doi.org/10.1021/acsomega.2c02842
- Noorbakhsh, R., Mortazavi, S. A., Sankian, M., Shahidi, F., Maleki, S. J., Nasiraii, L. R., & Varasteh, A. (2010). Influence of processing on the allergenic properties of pistachio nut assessed in vitro. *Journal of Agricultural and Food Chemistry*, 58(18), 10231–10235.
- Nwaru, B. I., Takkinen, H. M., Kaila, M., Erkkola, M., Ahonen, S., Pekkanen, J., & Knip, M. (2014). Food diversity in infancy and the risk of childhood asthma and allergies. *The Journal of Allergy and Clinical Immunology*, 133(4), 1084–1091. https://doi.org/10.1016/j. jaci.2013.12.1069
- Obeng, B. B., Amoah, A. S., Larbi, I. A., Yazdanbakhsh, M., Van Ree, R., Boakye, D. A., & Hartgers, F. C. (2011). Food allergy in Ghanaian schoolchildren: Data on sensitization and reported food allergy. *International Archives of Allergy and Immunology*, 155(1), 63–73. https://doi.org/10.1159/000318704
- Ogo, Y., Wakasa, Y., Hirano, K., Urisu, A., Matsuda, T., & Takaiwa, F. (2014). Generation of transgenic rice with reduced content of major and novel high molecular weight allergens. *Rice*, 7(1), 1–9. https://doi.org/10.1186/s12284-014-0019-0
- Pandey, A. K., Sudini, H. K., Upadhyaya, H. D., Varshney, R. K., & Pandey, M. K. (2019). Hypoallergen peanut lines identified through large-Scale phenotyping of global diversity panel: Providing hope toward addressing one of the major global food safety concerns.

Frontiers in Genetics, 10, 1177. https://doi.org/10.3389/fgene.2019.

- Panel, N. S. E. (2010). Guidelines for the diagnosis and management of food allergy in the United States: Report of the NIAID-sponsored expert panel. *The Journal of Allergy and Clinical Immunology*, 126(6), 1–58.
- Pastorello, E. A., Farioli, L., Pravettoni, V., Robino, A. M., Scibilia, J., Fortunato, D., & Ortolani, C. (2004). Lipid transfer protein and vicilin are important walnut allergens in patients not allergic to pollen. *Journal of Allergy and Clinical Immunology*, 114(4), 908–914.
- Pastorello, E. A., Vieths, S., Pravettoni, V., Farioli, L., Trambaioli, C., Fortunato, D., & Conti, A. (2002). Identification of hazelnut major allergens in sensitive patients with positive double-blind, placebo-controlled food challenge results. *Journal of Allergy and Clinical Immunology*, 109(3), 563–570.
- Pekar, J., Ret, D., & Untersmayr, E. (2018). Stability of allergens. Molecular Immunology, 100, 14–20. https://doi.org/10.1016/j.molimm. 2018.03.017
- Perkins, T., Schmitt, D. A., Isleib, T. G., Cheng, H., & Maleki, S. J. (2006). Breeding a hypoallergenic peanut. *Journal of Allergy and Clinical Immunology*, 117(2), 328. https://doi.org/10.1016/j.jaci. 2005.12.1293
- Peters, R. L., Koplin, J. J., Gurrin, L. C., Dharmage, S. C., Wake, M., Ponsonby, A. L., Tang, M. L., Lowe, A. J., Matheson, M., Dwyer, T., & Allen, K. J (2017). The prevalence of food allergy and other allergic diseases in early childhood in a population-based study: HealthNuts age 4-year follow-up. *The Journal of Allergy and Clinical Immunology*, 140(1), 145–153. https://doi.org/10.1016/j.jaci. 2017.02.019
- Petersen, A., Kull, S., Rennert, S., Becker, W. M., Krause, S., Ernst, M., & Jappe, U. (2015). Peanut defensins Novel allergens isolated from lipophilic peanut extract. *Journal of Allergy and Clinical Immunology*, 136(5), 1295–1301.
- Pi, X., Fu, G., Dong, B., Yang, Y., Wan, Y., & Xie, M. (2021). Effects of fermentation with Bacillus natto on the allergenicity of peanut. *Lwt*, *141*, 110862. https://doi.org/10.1016/j.lwt.2021.110862
- Plundrich, N. J., Bansode, R. R., Foegeding, E. A., Williams, L. L., & Lila, M. A. (2017). Protein-bound Vaccinium fruit polyphenols decrease IgE binding to peanut allergens and RBL-2H3 mast cell degranulation in vitro. Food & Function, 8(4), 1611–1621.
- Pomes, A., Davies, J. M., Gadermaier, G., Hilger, C., Holzhauser, T., Lidholm, J., Lopata, A. L., Mueller, G. A., Nandy, A., Radauer, C., & Chan, S. K. (2018). WHO/IUIS allergen nomenclature: Providing a common language. *Molecular Immunology*, 100, 3–13. https://doi. org/10.1016/j.molimm.2018.03.003
- Pons, L., Palmer, K., & Burks, W. (2005). Towards immunotherapy for peanut allergy. *Current Opinion in Allergy and Clinical Immunology*, 5(6), 558–562.
- Poulos, L. M., Waters, A. M., Correll, P. K., Loblay, R. H., & Marks, G. B. (2007). Trends in hospitalizations for anaphylaxis, angioedema, and urticaria in Australia, 1993–1994 to 2004–2005. *The Journal of Allergy and Clinical Immunology*, 120(4), 878–884. https://doi.org/10.1016/j.jaci.2007.07.040
- Prescott, S. L., Pawankar, R., Allen, K. J., Campbell, D. E., Sinn, J. K., Fiocchi, A., & Lee, B. W. (2013). A global survey of changing patterns of food allergy burden in children. *World Allergy Organization Journal*, 6(1), 1–12.
- Rabjohn, P., Helm, E. M., Stanley, J. S., West, C. M., Sampson, H. A., Burks, A. W., & Bannon, G. A. (1999). Molecular cloning and epi-

- tope analysis of the peanut allergen Ara h 3. *The Journal of Clinical Investigation*, 103(4), 535–542.
- Radauer, C., & Breiteneder, H. (2007). Evolutionary biology of plant food allergens. *Journal of Allergy and Clinical Immunology*, 120(3), 518–525. https://doi.org/10.1016/j.jaci.2007.07.024
- Radauer, C., Bublin, M., Wagner, S., Mari, A., & Breiteneder, H. (2008).
 Allergens are distributed into few protein families and possess a restricted number of biochemical functions. *The Journal of Allergy and Clinical Immunology*, 121(4), 847–852. https://doi.org/10.1016/j.jaci.2008.01.025
- Rajam, M. V. (2020). RNA silencing technology: A boon for crop improvement. *Journal of Biosciences*, 45, 1–5. https://doi.org/10. 1007/s12038-020-00082-x
- Riascos, J. J., Weissinger, A. K., Weissinger, S. M., & Burks, A. W. (2010). Hypoallergenic legume crops and food allergy: Factors affecting feasibility and risk. *Journal of Agricultural and Food Chemistry*, 58(1), 20–27. https://doi.org/10.1021/jf902526y
- Riecken, S., Lindner, B., Petersen, A., Jappe, U., & Becker, W. M. (2008).
 Purification and characterization of natural Ara h 8, the Bet v 1 homologous allergen from peanut, provides a novel isoform. *Biological Chemistry*, 389(4), 415–423.
- Rihs, H. P., Chen, Z., Ruëff, F., Petersen, A., Rozynek, P., Heimanna, H., & Baur, X. (1999). IgE binding of the recombinant allergen soybean profilin (rGly m 3) is mediated by conformational epitopes. *Journal of Allergy and Clinical Immunology*, 104(6), 1293–1301.
- Robotham, J. M., Wang, F., Seamon, V., Teuber, S. S., Sathe, S. K., Sampson, H. A., & Roux, K. H. (2005). Ana o 3, an important cashew nut (*Anacardium occidentale* L.) allergen of the 2S albumin family. *Journal of Allergy and Clinical Immunology*, 115(6), 1284–1290.
- Rustgi, S., Alam, T., Jones, Z. T., Brar, A. K., & Kashyap, S. (2022). Reduced-immunogenicity wheat and peanut lines for people with foodborne disorders. *Chemistry Proceedings*, 10, 67.
- Salminen, S., Endo, A., Isolauri, E., & Scalabrin, D. (2016). Early gut colonization with lactobacilli and staphylococcus in infants: The hygiene hypothesis extended. *Journal of Pediatric Gastroenterology and Nutrition*, 62(1), 80–86. https://doi.org/10.1097/MPG. 000000000000000925
- Sánchez-León, S., Gil-Humanes, J., Ozuna, C. V., Giménez, M. J., Sousa, C., Voytas, D. F., & Barro, F. (2018). Low-gluten, nontransgenic wheat engineered with CRISPR/Cas9. *Plant Biotechnology Journal*, 16(4), 902–910. https://doi.org/10.1111/pbi.12837
- Sander, I., Rihs, H. P., Brüning, T., & Raulf, M. (2016). A further wheat allergen for baker's asthma Tri a 40. *Journal of Allergy and Clinical Immunology*, 137(4), 1286.
- Sander, I., Rihs, H. P., Doekes, G., Quirce, S., Krop, E., Rozynek, P., & Raulf, M. (2015). Component-resolved diagnosis of baker's allergy based on specific IgE to recombinant wheat flour proteins. *Journal of Allergy and Clinical Immunology*, 135(6), 1529–1537.
- Sander, I., Rozynek, P., Rihs, H. P., Van Kampen, V., Chew, F. T., Lee, W. S., & Raulf-Heimsoth, M. (2011). Multiple wheat flour allergens and cross-reactive carbohydrate determinants bind IgE in baker's asthma. *Allergy*, 66(9), 1208–1215.
- Saurabh, S., Vidyarthi, A. S., & Prasad, D. (2014). RNA interference: Concept to reality in crop improvement. *Planta*, 239(3), 543–564. https://doi.org/10.1007/s00425-013-2019-5
- Schmidt, H., Gelhaus, C., Latendorf, T., Nebendahl, M., Petersen, A., Krause, S., & Janssen, O. (2009). 2-D DIGE analysis of the proteome of extracts from peanut variants reveals striking differences in major allergen contents. *Proteomics*, 9(13), 3507–3521.

Schmidt, M. A., Hymowitz, T., & Herman, E. M. (2015). Breeding and characterization of soybean triple null; a stack of recessive alleles of Kunitz trypsin inhibitor, soybean agglutinin, and P34 allergen nulls. *Plant Breeding*, 134(3), 310–315. https://doi.org/10.1111/pbr.12265

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- Schwager, C., Kull, S., Krause, S., Schocker, F., Petersen, A., Becker, W. M., & Jappe, U. (2015). Development of a novel strategy to isolate lipophilic allergens (oleosins) from peanuts. *Plos One*, 10(4), 0123419.
- Shah, F., Shi, A., Ashley, J., Kronfel, C., Wang, Q., Maleki, S. J., Adhikari, B., & Zhang, J. (2019). Peanut allergy: Characteristics and approaches for mitigation. *Comprehensive Reviews in Food Science* and Food Safety, 18(5), 1361–1387. https://doi.org/10.1111/1541-4337.12472
- Shan, Q., Wang, Y., Li, J., Zhang, Y., Chen, K., Liang, Z., Zhang, K., Liu, J., Xi, J. J., Qiu, J. L., & Gao, C. (2013). Targeted genome modification of crop plants using a CRISPR-Cas system. Nature Biotechnology, 31(8), 686–688. https://doi.org/10.1038/nbt. 2650
- Sharma, G. M., Irsigler, A., Dhanarajan, P., Ayuso, R., Bardina, L., Sampson, H. A., & Sathe, S. K. (2011). Cloning and characterization of an 11S legumin, Car i 4, a major allergen in pecan. *Journal of Agricultural and Food Chemistry*, 59(17), 9542–9552.
- Shu, H., Luo, Z., Peng, Z., & Wang, J. (2020). The application of CRISPR/Cas9 in hairy roots to explore the functions of *AhNFR1* and *AhNFR5* genes during peanut nodulation. *BMC Plant Biology*, 20, 417. https://doi.org/10.1186/s12870-020-02614-x
- Sicherer, S. H., & Sampson, H. A. (2010). Food allergy. The Journal of Allergy and Clinical Immunology, 125(2), 116–125. https://doi.org/ 10.1016/j.jaci.2009.08.028
- Sicherer, S. H., & Sampson, H. A. (2014). Food allergy: Epidemiology, pathogenesis, diagnosis, and treatment. *The Journal of Allergy and Clinical Immunology*, 133(2), 291–307. https://doi.org/10.1016/j.jaci. 2013.11.020
- Sicherer, S. H., & Sampson, H. A. (2018). Food allergy: A review and update on epidemiology, pathogenesis, diagnosis, prevention, and management. *Journal of Allergy and Clinical Immunology*, 141(1), 41–58. https://doi.org/10.1016/j.jaci.2017.11.003
- Singh, S., Chaudhary, R., Deshmukh, R., & Tiwari, S. (2023). Opportunities and challenges with CRISPR-Cas mediated homologous recombination based precise editing in plants and animals. *Plant Molecular Biology*, 111(1-2), 1–20. https://doi.org/10.1007/s11103-022-01321-5
- Sinha, M., Singh, R. P., Kushwaha, G. S., Iqbal, N., Singh, A., Kaushik, S., Kaur, P., & Sharma, S., & Singh, T. P. (2014). Current overview of allergens of plant pathogenesis related protein families. *Scientific World Journal*, 2014, 1–19. https://doi.org/10.1155/2014/543195
- Smits, M., Le, T. M., Welsing, P., Houben, G., Knulst, A., & Verhoeckx, K. (2018). Legume protein consumption and the prevalence of legume sensitization. *Nutrients*, 10(10), 1545. https://doi.org/10. 3390/nu10101545
- Stanley, J. S., King, N., Burks, A. W., Huang, S. K., Sampson, H., Cockrell, G., & Bannon, G. A. (1997). Identification and mutational analysis of the immunodominant IgE binding epitopes of the major peanut AllergenAra h 2. Archives of Biochemistry and Biophysics, 342(2), 244–253.
- Solanki, D., Mandaliya, V., & Georrge, J. J. (2020). Allergen bioinformatics: Repositories and tools to predict allergic proteins. *Recent Trends in Science and Technology*-2020 (ISBN: 9788192952154), 162–172. https://doi.org/10.6084/m9.figshare.13491423

- Steinwand, M. A., & Ronald, P. C. (2020). Crop biotechnology and the future of food. *Nature Food*, 1(5), 273–283. https://doi.org/10.1038/ s43016-020-0072-3
- Sugano, S., Hirose, A., Kanazashi, Y., Adachi, K., Hibara, M., Itoh, T., Mikami, M., Endo, M., Hirose, S., Maruyama, N., & Abe, J. (2020). Simultaneous induction of mutant alleles of two allergenic genes in soybean by using site-directed mutagenesis. *BMC Plant Biology*, 20(1), 1–15. https://doi.org/10.1186/s12870-020-02708-6
- Sutton, R., Skerritt, J. H., Baldo, B. A., & Wrigley, C. W. (1984).
 The diversity of allergens involved in bakers' asthma. *Clinical & Experimental Allergy*, 14(1), 93–107.
- Tang, M. L., & Mullins, R. J. (2017). Food allergy: Is prevalence increasing? *Internal Medicine Journal*, 47(3), 256–261. https://doi.org/10.1111/imj.13362
- Tawde, P., Venkatesh, Y. P., Wang, F., Teuber, S. S., Sathe, S. K., & Roux, K. H. (2006). Cloning and characterization of profilin (Pru du 4), a cross-reactive almond (Prunus dulcis) allergen. *Journal of Allergy and Clinical Immunology*, 118(4), 915–922.
- Teuber, S. S., Dandekar, A. M., Peterson, W. R., & Sellers, C. L. (1998).
 Cloning and sequencing of a gene encoding a 2S albumin seed storage protein precursor from English walnut (Juglans regia), a major food allergen. *Journal of Allergy and Clinical Immunology*, 101(6), 807–814.
- Tordesillas, L., & Berin, M. C. (2018). Mechanisms of oral tolerance. Clinical Reviews in Allergy & Immunology, 55(2), 107–117.
- Tordesillas, L., Berin, M. C., & Sampson, H. A. (2017). Immunology of food allergy. *Immunity*, 47(1), 32–50. https://doi.org/10.1016/j. immuni.2017.07.004
- Turner, P. J., Mehr, S., Sayers, R., Wong, M., Shamji, M. H., Campbell, D. E., & Mills, E. C. (2014). Loss of allergenic proteins during boiling explains tolerance to boiled peanut in peanut allergy. *The Journal of Allergy and Clinical Immunology*, 134(3), 751–753. https://doi.org/10.1016/j.jaci.2014.06.016
- Upadhyay, S. K., Kumar, J., Alok, A., & Tuli, R. (2013). RNA-guided genome editing for target gene mutations in wheat. *G3: Genes, Genomes, Genetics*, 3(12), 2233–2238. https://doi.org/10.1534/g3. 113.008847
- Upadhyaya, H. D., Ortiz, R., Bramel, P. J., & Singh, S. (2003). Development of a peanut core collection using taxonomical, geographical and morphological descriptors. *Genetic Resources and Crop Evolution*, 50(2), 139–148. https://doi.org/10.1023/A:1022945715628
- Upadhyaya, H. D., Yadav, D., Dronavalli, N., Gowda, C. L. L., & Singh, S. (2010). Mini core germplasm collections for infusing genetic diversity in plant breeding programs. *Electron Journal of Plant Breeding*, 1(4), 1294–1309.
- Urnov, F. D., Rebar, E. J., Holmes, M. C., Zhang, H. S., & Gregory, P. D. (2010). Genome editing with engineered zinc finger nucleases. *Nature Reviews Genetics*, 11(9), 636–646. https://doi.org/10.1038/nrg2842
- Vanga, S. K., Singh, A., & Raghavan, V. (2017). Review of conventional and novel food processing methods on foosd allergens. *Critical Reviews in Food Science and Nutrition*, 57(10), 2077–2094. https://doi.org/10.1080/10408398.2015.1045965
- Varshney, R. K., Pandey, M. K., Bohra, A., Singh, V. K., Thudi, M., & Saxena, R. K. (2019). Toward the sequence-based breeding in legumes in the post-genome sequencing era. *Theoretical and Applied Genetics*, 132(3), 797–816. https://doi.org/10.1007/s00122-018-3252-x
- Varshney, R. K., Pandey, M. K., Janila, P., Nigam, S. N., Sudini, H., Gowda, M. V. C., & Nagesh, P. (2014). Marker-assisted introgression

of a QTL region to improve rust resistance in three elite and popular varieties of peanut (*Arachis hypogaea* L.). *Theoretical and Applied Genetics*, 127, 1771–1781.

- Vereda, A., van Hage, M., Ahlstedt, S., Ibañez, M. D., Cuesta-Herranz, J., van Odijk, J., & Sampson, H. A. (2011). Peanut allergy: Clinical and immunologic differences among patients from 3 different geographic regions. *The Journal of Allergy and Clinical Immunology*, 127(3), 603–607. https://doi.org/10.1016/j.jaci.2010.09.010
- Verhoeckx, K. C. M., Vissers, Y. M., Baumert, J. L., Faludi, R., Feys, M., Flanagan, S., & Kimber, I. (2015). Food processing and allergenicity. Food and Chemical Toxicology, 80, 223–240. https://doi.org/10.1016/ j.fct.2015.03.005
- Viquez, O. M., Summer, C. G., & Dodo, H. W. (2001). Isolation and molecular characterization of the first genomic clone of a major peanut allergen, Ara h 2. *Journal of Allergy and Clinical Immunology*, 107(4), 713–717.
- Viquez, O. M., Konan, K. N., & Dodo, H. W. (2004). Genomic organization of peanut allergen gene, Ara h 3. *Molecular Immunology*, 41(12), 1235–1240.
- Wallowitz, M., Peterson, W. R., Uratsu, S., Comstock, S. S., Dandekar, A. M., & Teuber, S. S. (2006). Jug r 4, a legumin group food allergen from walnut (Juglans regia Cv. Chandler). *Journal of Agricultural and Food Chemistry*, 54(21), 8369–8375.
- Wang, F., Robotham, J. M., Teuber, S. S., Sathe, S. K., & Roux, K. H. (2003). Ana o 2, a major cashew (*Anacardium occidentale L.*) nut allergen of the legumin family. *International Archives of Allergy and Immunology*, 132(1), 27–39.
- Wang, F., Robotham, J. M., Teuber, S. S., Tawde, P., Sathe, S. K., & Roux, K. H. (2002). Ana o 1, a cashew (Anacardium occidental) allergen of the vicilin seed storage protein family. *Journal of Allergy and Clinical Immunology*, 110(1), 160–166.
- Wang, H., La Russa, M., & Qi, L. S. (2016). CRISPR/Cas9 in genome editing and beyond. *Annual Review of Biochemistry*, 85, 227–264. https://doi.org/10.1146/annurev-biochem-060815-014607
- Wang, J., He, Z., & Raghavan, V. (2022). Soybean allergy: Characteristics, mechanisms, detection and its reduction through novel food processing techniques. *Critical Reviews in Food Science and Nutrition*, 1–14. Advance online publication. https://doi.org/10.1080/10408398.2022.2029345
- Wang, L., Niu, D., Zhao, X., Wang, X., Hao, M., & Che, H. (2021).
 A comparative analysis of novel deep learning and ensemble learning models to predict the allergenicity of food proteins. *Foods*, 10(4), 809. https://doi.org/10.3390/foods10040809
- Wang, Y., Cheng, X., Shan, Q., Zhang, Y., Liu, J., Gao, C., & Qiu, J. L. (2014). Simultaneous editing of three homoeoalleles in hexaploid bread wheat confers heritable resistance to powdery mildew. *Nature Biotechnology*, 32(9), 947–951. https://doi.org/10.1038/nbt.2969
- Warren, C. M., Jiang, J., & Gupta, R. S. (2020). Epidemiology and burden of food allergy. *Current Allergy and Asthma Reports*, 20(2), 1–9. https://doi.org/10.1007/s11882-020-0898-7
- Wasserman, R. L., Jones, D. H., & Windom, H. H (2018). Oral immunotherapy for food allergy: The FAST perspective. *Annals of Allergy*, *Asthma & Immunology*, 121(3), 272–275.
- Weeks, D. P. (2017). Gene editing in polyploid crops: Wheat, camelina, canola, potato, cotton, peanut, sugar cane, and citrus. *Progress in Molecular Biology and Translational Science*, 149, 65–80. https://doi.org/10.1016/bs.pmbts.2017.05.002
- Wen, S., Wen, N., Pang, J., Langen, G., Brew-Appiah, R. A., Mejias, J. H., Osorio, C., Yang, M., Gemini, R., Moehs, C. P., & Zemetra,

- R. S. (2012). Structural genes of wheat and barley 5-methylcytosine DNA glycosylases and their potential applications for human health. *Proceedings of the National Academy of Sciences of the United States of America*, 109(50), 20543–20548. https://doi.org/10.1073/pnas.1217927109
- Willison, L. N., Tawde, P., Robotham, J. M., Penney, IV. R. M., Teuber, S. S., Sathe, S. K., & Roux, K. H. (2008). Pistachio vicilin, Pis v 3, is immunoglobulin E-reactive and cross-reacts with the homologous cashew allergen, Ana o 1. Clinical & Experimental Allergy, 38(7), 1229–1238.
- Woods, R. K., Stoney, R. M., Raven, J., Walters, E. H., Abramson, M., & Thien, F. C. K. (2002). Reported adverse food reactions overestimate true food allergy in the community. *European Journal of Clinical Nutrition*, 56(1), 31–36. https://doi.org/10.1038/sj.ejcn.1601306
- Wu, Z., Lian, J., Han, Y., Zhou, N., Li, X., Yang, A., & Chen, H. (2016). Crosslinking of peanut allergen Ara h 2 by polyphenol oxidase: Digestibility and potential allergenicity assessment. *Journal of the Science of Food and Agriculture*, 96(10), 3567–3574. https://doi.org/10.1002/jsfa.7542
- Wu, Z., Zhou, N., Xiong, F., Li, X., Yang, A., Tong, P., & Chen, H. (2016). Allergen composition analysis and allergenicity assessment of Chinese peanut cultivars. *Food Chemistry*, 196, 459–465. https:// doi.org/10.1016/j.foodchem.2015.09.070
- Xia, W. J., Zhang, P. P., Wu, X. Y., Li, M. X., Sun, T., Fang, P. P., Pandey, A. K., & Xu, P. (2022). Mutant library resources for legume crops and the emerging new screening technologies. *Euphytica*, 218(3). https://doi.org/10.1007/s10681-022-02979-0
- Yamada, T., Mori, Y., Yasue, K., Maruyama, N., Kitamura, K., & Abe, J. (2014). Knockdown of the 7S globulin subunits shifts distribution of nitrogen sources to the residual protein fraction in transgenic soybean seeds. *Plant Cell Reports*, *33*(12), 1963–1976. https://doi.org/10.1007/s00299-014-1671-y
- Yamada, Y., Yokooji, T., Kunimoto, K., Inoguchi, K., Ogino, R., Taogoshi, T., & Matsuo, H. (2022). Hypoallergenic wheat line (1BS-18H) lacking ω5-gliadin induces oral tolerance to wheat gluten proteins in a rat model of wheat allergy. *Foods*, 11(15), 2181. https://doi.org/10.3390/foods11152181
- Yan, D. C., Ou, L. S., Tsai, T. L., Wu, W. F., & Huang, J. L. (2005). Prevalence and severity of symptoms of asthma, rhinitis, and eczema in 13-to 14-year-old children in Taipei, *Taiwan. Annals of Allergy*, *Asthma & Immunology*, 95(6), 579–585.
- Yang, W. W., Chung, S. Y., Ajayi, O., Krishnamurthy, K., Konan, K., & Goodrich-Schneider, R. (2010). Use of pulsed ultraviolet light to reduce the allergenic potency of soybean extracts. *International Jour*nal of Food Engineering, 6(3), https://doi.org/10.2202/1556-3758. 1876
- Yokooji, T., Kurihara, S., Murakami, T., Chinuki, Y., Takahashi, H., Morita, E., & Matsuo, H. (2013). Characterization of causative allergens for wheat-dependent exercise-induced anaphylaxis sensitized with hydrolyzed wheat proteins in facial soap. *Allergology International*, 62(4), 435–445.
- Yu, J., Ahmedna, M., Goktepe, I., Cheng, H., & Maleki, S. (2011). Enzymatic treatment of peanut kernels to reduce allergen levels. *Food Chemistry*, 127(3), 1014–1022. https://doi.org/10.1016/j.foodchem. 2011.01.074
- Yuan, M., Zhu, J., Gong, L., He, L., Lee, C., Han, S., Chen, C., & He, G. (2019). Mutagenesis of FAD2 genes in peanut with CRISPR/Cas9 based gene editing. BMC Biotechnology, 19(1), 1–7. https://doi.org/10.1186/s12896-019-0516-8

Zhou, Y., Wang, J. S., Yang, X. J., Lin, D. H., Gao, Y. F., Su, Y. J., & Zheng, J. J. (2013). Peanut allergy, allergen composition, and methods of reducing allergeni.city. A Review International Journal of Food Sciences and Nutrition, 2013, 1–8. https://doi.org/10.1155/2013/909140

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