

Transcriptional profiling of *Pseudomonas aeruginosa* mature single- and dual-species biofilms in response to meropenem

Alam, Farhana; Blair, Jessica M A; Hall, Rebecca A

DOI:

[10.1099/mic.0.001271](https://doi.org/10.1099/mic.0.001271)

License:

Creative Commons: Attribution (CC BY)

Document Version

Publisher's PDF, also known as Version of record

Citation for published version (Harvard):

Alam, F, Blair, JMA & Hall, RA 2023, 'Transcriptional profiling of *Pseudomonas aeruginosa* mature single- and dual-species biofilms in response to meropenem', *Microbiology*, vol. 169, no. 1, 001271.
<https://doi.org/10.1099/mic.0.001271>

[Link to publication on Research at Birmingham portal](#)

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Transcriptional profiling of *Pseudomonas aeruginosa* mature single- and dual-species biofilms in response to meropenem

Farhana Alam¹, Jessica M. A. Blair² and Rebecca A. Hall^{3,*}

Abstract

Pseudomonas aeruginosa is a Gram-negative opportunistic pathogen frequently isolated from chronic infections of the cystic fibrosis lung and burn wounds, and is a major cause of antimicrobial-resistant nosocomial infections. *P. aeruginosa* is frequently co-isolated with the opportunistic fungal pathogen *Candida albicans*, with the presence of *C. albicans* in dual-species biofilms promoting tolerance to meropenem. Here, transcription profiling of mature *P. aeruginosa* single- or dual-species biofilms was carried out to understand the molecular mechanism(s) by which *C. albicans* enhances meropenem tolerance. *C. albicans* appeared to have a mild impact on the transcriptome of *P. aeruginosa* mature biofilms, with most differentially regulated genes being involved in interkingdom interactions (i.e. quorum sensing and phenazine biosynthesis). The addition of meropenem to mature single- or dual-species biofilms resulted in a significant bacterial transcriptional response, including the induction of the beta-lactamase, *ampC*, genes involved in biofilm formation. *P. aeruginosa* elicited a similar transcriptional response to meropenem in the presence of *C. albicans*, but *C. albicans* promoted the expression of additional efflux pumps, which could play roles in increasing the tolerance of *P. aeruginosa* to meropenem.

INTRODUCTION

Pseudomonas aeruginosa is a Gram-negative bacterial pathogen associated with chronic infections in the cystic fibrosis (CF) lung [1] and burn wounds [2], and is a major contributor to nosocomial infections [3]. The ability of the bacterium to form biofilms is critical to the pathogenicity of *P. aeruginosa*, with biofilms being the predominant mode of growth in both the CF lung and wounds [4, 5]. Biofilms are of medical importance as they protect the microbes from the host's innate immune system [4, 6], and significantly enhance antimicrobial resistance, with biofilms being 100–1000 times more resistant to antimicrobial therapies than their planktonic counterparts [7]. Furthermore, cells dispersed from biofilms exhibit a unique transcription profile [8], suggesting that dispersed cells represent a distinct stage that may enhance dissemination and infection progression.

P. aeruginosa is frequently co-isolated from sites of infection with the fungal pathogen *Candida albicans* [9]. These two microbes undergo complex interactions including direct cell–cell interactions, and through the secretion of signalling molecules and metabolites [10], the outcome of which is dependent on environmental factors. During biofilm formation, *P. aeruginosa* use the fungal hyphae as a scaffold to enhance the structure, composition and complexity of the biofilm [11, 12]. The presence of *C. albicans* in biofilms enhances the antibiotic tolerance of several important bacterial pathogens, including *P. aeruginosa* [13–15]. Enhanced antimicrobial resistance is hypothesized to result from increased extracellular matrix production in dual-species biofilms, as a result of the fungus contributing cell wall carbohydrates such as glucans and mannans to the matrix [13–16]. These fungal polysaccharides are thought to provide protection against antimicrobial agents through either limiting the diffusion of the antimicrobial through the biofilm (i.e. vancomycin) [15] or binding and sequestering the drugs (i.e. azoles) [17, 18].

Received 29 July 2022; Accepted 09 November 2022; Published 23 January 2023

Author affiliations: ¹Institute of Microbiology and Infection, School of Biosciences, University of Birmingham, Birmingham B15 2TT, UK; ²Institute of Microbiology and Infection, College of Medical and Dental Sciences, University of Birmingham, Birmingham B15 2TT, UK; ³Kent Fungal Group, Division of Natural Sciences, School of Biosciences, University of Kent, Canterbury CT2 7NJ, UK.

***Correspondence:** Rebecca A. Hall, r.a.hall@kent.ac.uk

Keywords: *Candida albicans*; *Pseudomonas aeruginosa*; meropenem; transcriptional profiling; dual-species biofilms.

Abbreviations: CA, *Candida albicans*; CF, cystic fibrosis; DEGs, differentially expressed genes; FDR, false discovery rate; GO, gene ontology; KEGG, Kyoto encyclopaedia of genes and genomes; PA, *Pseudomonas aeruginosa*.

Sequencing reads and gene hit count tables are available at the Gene Expression Omnibus (GEO) database (<https://www.ncbi.nlm.nih.gov/geo/>), under accession number GSE167137.

Two supplementary figures are available with the online version of this article.

001271 © 2023 The Authors



This is an open-access article distributed under the terms of the Creative Commons Attribution License.

Meropenem is a carbapenem antibiotic that is frequently used to treat chronic *P. aeruginosa* infections. Previously we have shown that in dual-species biofilms *C. albicans* increases the tolerance of *P. aeruginosa* to meropenem, a process dependent on fungal mannan [13]. To provide more detail on the molecular mechanism(s) underlying this phenomenon, we analysed the transcriptional response of mature *P. aeruginosa* biofilms, and dual-species biofilms in the absence and presence of meropenem. The transcriptional profile identifies key *P. aeruginosa* genes and biological processes required for resistance to meropenem. Similar genes and processes were unregulated by *P. aeruginosa* in dual-species biofilms, indicating that the presence of *C. albicans* in mature biofilms does not perturb the *P. aeruginosa* transcriptional response to meropenem.

METHODS

Strains and media

C. albicans SC5314 was grown and maintained on yeast peptone dextrose (YPD; Sigma) media, while *P. aeruginosa* PAO1 (ATCC15692) was grown and maintained on Luria-Bertani (LB) medium. All biofilm assays were performed in Muller-Hinton broth (MHB).

Biofilm assay

Biofilm assays were based on previously described methodology [13], but scaled up to six-well plates. In brief, overnight cultures of *C. albicans* and *P. aeruginosa* were washed in phosphate-buffered saline (PBS), and *C. albicans* resuspended at 1×10^6 cells ml⁻¹ and *P. aeruginosa* to an OD₆₀₀ of 0.2 [$\sim 2 \times 10^8$ colony-forming units (c.f.u.) ml⁻¹] in Mueller-Hinton broth. Each well contained 3 ml *C. albicans* and 300 μ l of *P. aeruginosa* in a total of 6 ml. Plates were incubated at 37 °C for 2 h to allow cells to adhere, at which point the media were replaced with fresh sterile media, and plates were incubated statically at 37 °C for 24 h. Cells not part of the biofilm were removed, media were replaced with fresh MHB containing 0 or 5 μ g ml⁻¹ meropenem, and plates were incubated for 4 h. Media were replaced with 2 ml PBS containing 50 μ g ml⁻¹ DNase I and plates were incubated at 37 °C for 1 h to degrade the extracellular matrix. Biofilms were detached from the plate by scraping, serially diluted and plated onto selective agar (YPD agar supplemented with 100 μ g ml⁻¹ tetracycline to determine viable *C. albicans* c.f.u. and cetrimide agar to determine viable *P. aeruginosa* c.f.u.).

Preparation of samples for RNA extraction

Biofilms were formed as described above and triplicate biofilms were pooled, 50 μ l serially diluted and plated on cetrimide agar or YPD supplemented with 100 μ g ml⁻¹ tetracycline to check for contamination. Remaining biofilm cells were centrifuged at 3500 r.p.m at 4 °C for 5 min and pellets snap frozen in liquid nitrogen. Four biological replicates were shipped to GeneWiz, UK, for RNA extraction sequencing and basic bioinformatic analysis.

RNA extraction and sequencing

RNA was extracted from the biofilms by GeneWiz using the Qiagen RNeasy Plus mini kit. Library preparation was done in the following stages: (a) ribosomal RNA depletion; (b) RNA fragmentation and random priming; (c) first and second strand cDNA synthesis; (d) end repair, 5' phosphorylation and dA-tailing; (e) adapter ligation, PCR enrichment and sequencing. Paired-end sequencing was performed using Illumina HiSeq 4000 (2 \times 150bp configuration, single index per lane).

Bioinformatic analysis

The sequence quality of each sample was evaluated by determining the number of reads, the yield (Mbases), the mean quality score and the percentage of reads over 30 bases in length. FastQC software was used to determine per-base-sequence quality and per-sequence GC content. Sequence reads were trimmed to remove adapter sequences and nucleotides with poor quality using Trimmomatic v.0.36. The trimmed reads were mapped to the *P. aeruginosa* reference genome, available on ENSEMBL, using the STAR aligner v.2.5.2b. For dual-species biofilms, samples were treated the same as single-species biofilms. Reads were mapped to the PAO1 genome, and non-mapped reads were discarded and later aligned to the *C. albicans* reference genome. Unique gene hit counts were calculated using featureCounts from the Subread package v.1.5.2. Only unique reads that fell within exonic regions were counted (for unique genes, the number of hits per read was set to 10 as default, with reads that mapped to <10 distinct places assigned to one place by the EM algorithm, and reads that mapped to >10 distinct places being discarded). For the analysis, only read counts for genes in the *P. aeruginosa* genome were used and the TPM values were calculated as follows: each read count was divided by the length of each gene in kb to generate reads per kb (RPK), then all the RPK values in the sample were counted and divided by 1 000 000 to generate the scaling factor, and finally the RPK values were divided by the scaling factor to generate the TPM value. Differential gene expression analysis was performed using DESeq2 and the comparisons listed in Table 1. Principal component analysis (PCA) was performed to reveal the similarities within and between groups, with PCA plots included in the output (Fig. S1, available with the online version of this article). As expected, dual-species biofilms exhibited greater biological variation than single-species biofilms

Table 1. Differential expression analysis carried out by DESeq2

Comparison	Abbreviation	Output file
Untreated <i>P. aeruginosa</i> mono-species biofilm vs <i>P. aeruginosa</i> mono-species treated with 5 µg ml ⁻¹ meropenem	PA_0M vs PA_5M	P_aeruginosa_expression_PA_0M_vs_PA_5M.xlsx
Untreated <i>P. aeruginosa</i> mono-species biofilm vs dual-species biofilms	PA_0M vs PACA_0M	P_aeruginosa_expression_PA_0M_vs_PACA_0M.xlsx
Untreated <i>P. aeruginosa</i> mono-species biofilms vs dual-species biofilms treated with 5 µg ml ⁻¹ meropenem	PACA_0M vs PACA_5M	P_aeruginosa_expression_PACA_0M_vs_PACA_5M.xlsx

(Fig. S1), which likely reflects the heterogeneity of the biofilm structure. Meropenem treatment had the greatest effect on the bacteria transcriptome, with samples clustering into distinct groups in the PCA plots, while *C. albicans* had a reduced impact on the *P. aeruginosa* transcriptome, resulting in great spread of the data. DESeq2 output files for each comparison, and files containing summary raw and normalized reads and TPM values for each gene are available at the Gene Expression Omnibus (GEO) database (<https://www.ncbi.nlm.nih.gov/geo/>), under accession number GSE167137.

Enrichment analysis

For *P. aeruginosa* transcriptomic analysis, differential expression of genes between conditions was considered significant if the adjusted *P*-value (Padj) was ≤0.05. Gene ontology (GO) analysis was performed using KOBAS 3.0 software [Kyoto Encyclopaedia of Genes and Genomes (KEGG) Orthology Based Annotation System] [19].

Statistical analysis

Biofilm data were analysed in GraphPad Prism (version 9.1.0) using two-way analysis of variance (ANOVA) and Holm–Sidak's multiple comparisons test.

RESULTS

C. albicans enhances the tolerance of *P. aeruginosa* biofilms to meropenem even at early timepoints

Previously we have observed that when *P. aeruginosa* is grown in a dual-species biofilm with the fungal pathogen *C. albicans*, the tolerance of *P. aeruginosa* to meropenem is increased [13]. To understand the molecular mechanism(s) behind this increased tolerance, transcriptional profiling was performed. To avoid the transcriptional profile focusing on genes related to cell death, we analysed the transcriptome after the antibiotic had been added to mature biofilms for 4 h. Plating of mono- and dual-species biofilms in the absence and presence of meropenem confirmed that the majority of the cell population was viable at this time point. Furthermore, the tolerance of *P. aeruginosa* to meropenem in the presence of *C. albicans* was still enhanced even at this early timepoint (Fig. 1).

Meropenem enhances *ampC* expression in *P. aeruginosa* single-species biofilms

Addition of meropenem to *P. aeruginosa* mono-species biofilms resulted in the significant upregulation of 354 genes, while 509 genes were downregulated (Figs 2 and S2a, Table 2). Of the significantly differentially regulated genes, 45 (159/354) and 43% (217/509) encoded hypothetical proteins. As expected, the most significantly upregulated gene in response to meropenem treatment was *ampC* (log₂ fold change=7.75, Padj=1.9510⁻⁴²), which encodes a beta-lactamase, important for carbapenem resistance. GO term enrichment analysis of the differentially regulated genes confirmed that meropenem resulted in the significant upregulation of genes involved in the maintenance of the bacterial cell membrane, cell wall, biofilm formation and extracellular matrix production, siderophore production, type IV pilus formation and type VI secretion system, while genes involved in putrescine transport were significantly downregulated (Fig. 3a). KEGG pathway analysis identified the over-representation of genes involved in siderophore biosynthesis, vancomycin resistance, amino acid metabolism, biofilm formation and bacterial secretion in genes that were significantly upregulated and pathways associated with thiamine metabolism, mismatch repair, pyrimidine metabolism and purine metabolism were significantly downregulated (Fig. 3b).

The presence of *C. albicans* in dual-species biofilms results in mild changes in the *P. aeruginosa* transcriptome

To determine how the presence of *C. albicans* affected *P. aeruginosa*, we compared the transcriptional profile of *P. aeruginosa* single-species biofilms to dual-species biofilms. The presence of *C. albicans* resulted in the differential regulation of 392 genes, with 265 genes being significantly upregulated and 127 genes being significantly downregulated (Figs 2 and S2b, Table 3). Although significant, we note that the log₂ values for the majority of these genes were <1, suggesting that *C. albicans* has a mild impact on the transcription profile of *P. aeruginosa*. The most significantly differentially regulated genes were PA4097

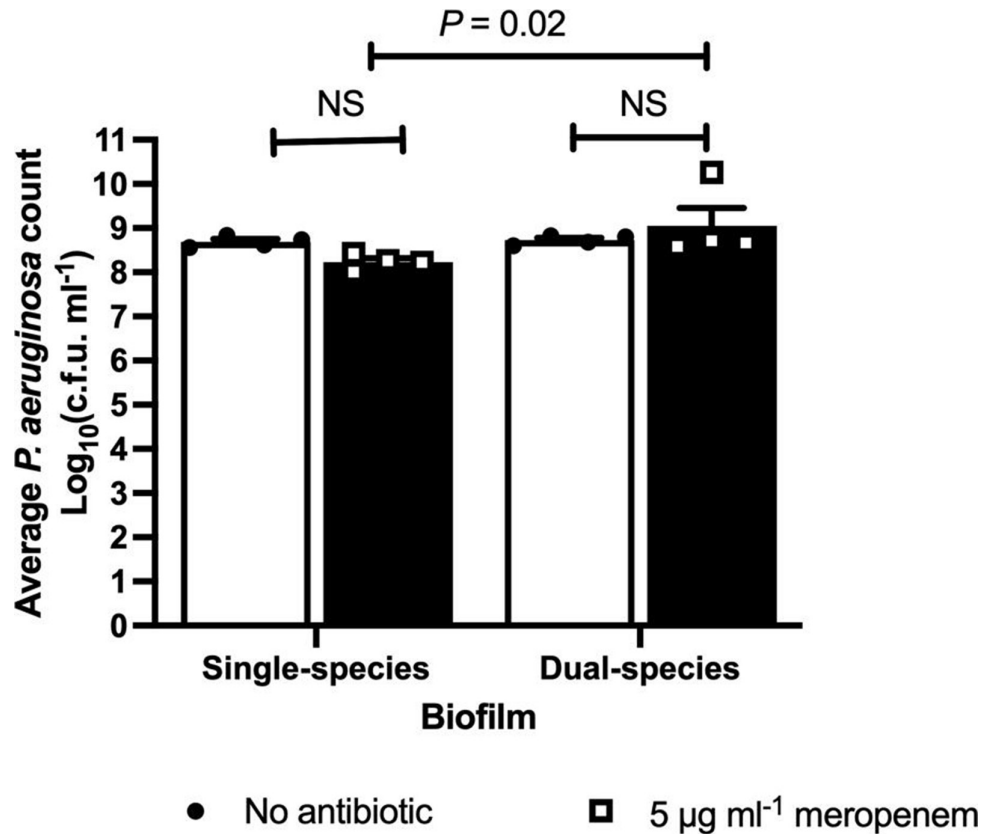


Fig. 1. *C. albicans* enhances the tolerance of *P. aeruginosa* to meropenem after short exposures. Preformed 24 hr biofilms were incubated in Mueller Hinton broth containing 0 or 5 µg/mL meropenem, for 4 hr and viable *P. aeruginosa* counts quantified. Data are the mean ± the SEM from 4 biological replicates. Data were analysed using 2-way ANOVA and Holm-Sidak's multiple comparisons test (ns not significant; * $P < 0.05$).

(a lipolytic protein) and PA5384 (an alcohol dehydrogenase), which were both twofold upregulated in the presence of *C. albicans*. GO term enrichment analysis (Fig. 4a) and KEGG pathway analysis (Fig. 4b) of the 392 differentially regulated genes identified that phenazine biosynthesis, *Pseudomonas* quinolone signal (PQS) production, pyridocal phosphate biosynthesis, and siderophore transport and pyoverdine biosynthesis were significantly upregulated, while genes involved in type IV pilus biogenesis, the general secretory (Sec) pathway and amino acid biosynthesis were significantly downregulated. Therefore, *P. aeruginosa* appears to upregulate processes that might provide the bacterium with a competitive advantage when growing in the presence of *C. albicans*.

To identify whether the presence of the fungus was inducing a similar transcriptional response to the drug treatment we compared DEGs identified in *P. aeruginosa* single-species biofilms treated with meropenem to the list of DEGs identified in untreated dual-species biofilms. Seven genes (PA10907, PA4830, PA4518, PA1209, *rimK*, PA0881 and PA3758) were upregulated, while five genes (*pyrD*, *aroC*, PA4633, PA3526 and PA4637) were downregulated under both conditions. However, as the majority of the genes encode hypothetical proteins, the impact of these transcriptional responses is unknown.

Impact of *C. albicans* on the *P. aeruginosa* transcriptional response to meropenem

To determine whether the presence of *C. albicans* affected the response of *P. aeruginosa* to meropenem, we compared the transcriptional profile of untreated dual-species biofilms to that of meropenem-treated dual-species biofilms. In response to meropenem, a total of 620 genes were differentially regulated, with 304 genes being significantly upregulated and 316 being significantly downregulated (Figs 2 and S2c, Table 4). As was the case for the single-species biofilms, *ampC* was the most significantly upregulated gene ($\log_2=5.52$, $\text{Padj}=1.9510^{-47}$). GO term enrichment analysis (Fig. 5a) and KEGG pathway analysis (Figs 4 and 5b) confirmed that as with single-species biofilms, treatment of *P. aeruginosa* dual-species biofilms with meropenem resulted in the upregulation of genes involved in cell division, peptidoglycan biosynthesis, biofilm formation, extracellular matrix production, siderophore biosynthesis and type VI secretion. Likewise, the processes that were significantly downregulated in single-species biofilms in response to meropenem (i.e. putrescine transport and mismatch repair) were shared with dual-species biofilms (Fig. 5).

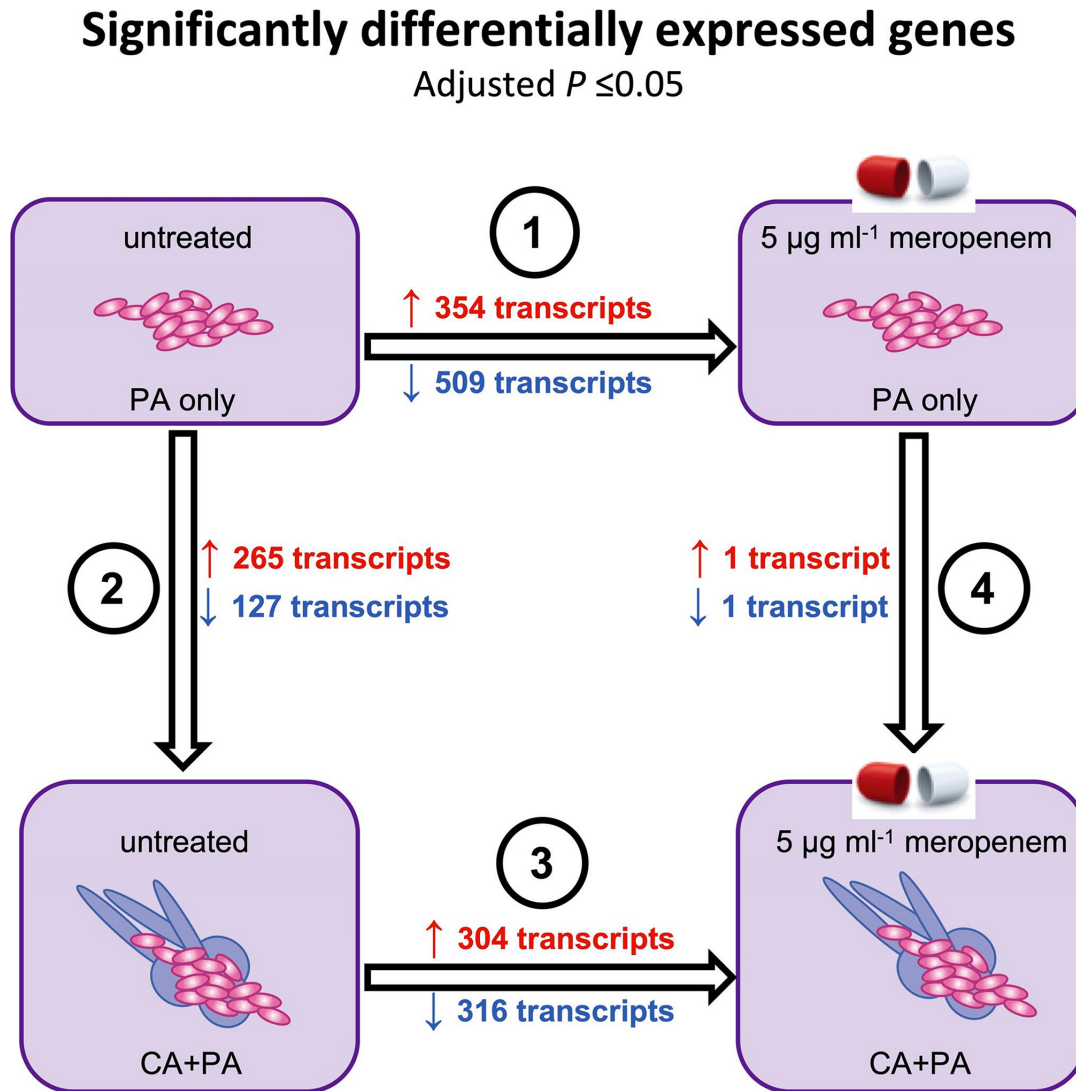


Fig. 2. Summary of transcriptional responses of *P. aeruginosa* biofilm cells to *C. albicans* and to meropenem treatment. Numbers of significantly differentially expressed *P. aeruginosa* transcripts ($P_{adj} \leq 0.05$) in 1) *P. aeruginosa* single-species biofilms treated with 5 µg ml⁻¹ meropenem; 2) dual species untreated biofilms; 3) dual species biofilms treated with 5 µg ml⁻¹ meropenem; 4) single vs dual species biofilms treated with 5 µg ml⁻¹ meropenem. PA: *P. aeruginosa*; CA: *C. albicans*. Red: numbers of significantly upregulated transcripts; Blue: numbers of significantly downregulated transcripts.

Although *C. albicans* does not affect the general processes that are differentially regulated in response to meropenem, it is possible that the underlying DEGs are different. Therefore, we compared the DEGs between single- and dual-species biofilms in response to meropenem. Of the genes that were significantly upregulated in response to meropenem, 181 (37.9%) were upregulated in both single- and dual-species biofilms, while 173 (36.3%) and 123 (25.8%) were unique to single- and dual-species biofilms, respectively (Fig. 6a). Among the upregulated genes that were unique to the dual-species meropenem response, three were linked to outer-membrane vesicles (OMVs; *pagL*, *galU* and PA5441) [20], one was involved in cell wall synthesis (*ddlA*), three were involved in biofilm formation (*pslH*, *pslI* and *algR*) and three were involved in efflux (PA1809, *mexC* and PA3314). This suggests that the presence of *C. albicans* may increase the ability of *P. aeruginosa* to form robust biofilms, to secrete molecules via OMVs and upregulate the MexCD-OprJ efflux pump, which may play a role in increasing the tolerance of the bacterium to meropenem.

Of the genes that were significantly downregulated in response to meropenem, 282 (51.9%) genes were shared in common between single- and dual-species *P. aeruginosa* biofilm cells, while 227 (41.8%) and 34 (6.3%) were unique to single- and dual-species biofilms, respectively (Fig. 6b). Among the few downregulated genes that were unique to the dual-species meropenem response, there were six involved in efflux or membrane transport (PA0603, PA0604, *mexE*, *mexF*, PA0860 and

Table 2. Top 20 differentially regulated genes in *P. aeruginosa* untreated single biofilms vs single-species biofilms treated with 5 µg ml⁻¹ meropenem

	Gene ID	log ₂ fold change	P-value	Padj
Upregulated	<i>ampC</i>	5.745634677	3.54E-46	1.95E-42
	<i>PA4111</i>	3.141752798	2.08E-25	1.27E-22
	<i>hcpI</i>	2.299594301	9.57E-20	2.64E-17
	<i>PA0126</i>	2.010278388	4.77E-15	6.75E-13
	<i>clpV1</i>	1.832477469	5.89E-09	3.65E-07
	<i>PA4280.2</i>	1.825681731	0.0007200374797	0.008213485384
	<i>PA0084</i>	1.81927681	9.59E-07	3.60E-05
	<i>PA0083</i>	1.812545239	2.64E-06	8.69E-05
	<i>PA4690.2</i>	1.811643426	0.0006939310453	0.00798165271
	<i>PA5369.2</i>	1.811071763	0.0007048977579	0.008057433792
	<i>PA0668.4</i>	1.770903731	0.0005529877155	0.006709989401
	<i>PA0070</i>	1.759966355	2.86E-06	9.28E-05
	<i>PA0277</i>	1.706435238	2.22E-11	2.04E-09
	<i>vgrG1</i>	1.692796239	4.23E-08	2.12E-06
	<i>PA0466</i>	1.68245735	3.45E-06	0.0001075909918
	<i>PA0089</i>	1.645610422	8.37E-08	3.89E-06
	<i>PA0050</i>	1.596317219	3.73E-05	0.0007623159776
	<i>rplX</i>	1.577791178	0.003955658099	0.03020634629
	<i>rpsQ</i>	1.514302023	0.003135531829	0.0252719288
	<i>rplN</i>	1.482472974	0.006174262969	0.04190492586
Downregulated	<i>PA0627</i>	-2.563807744	7.94E-27	6.26E-24
	<i>PA0629</i>	-2.554150934	7.27E-33	1.34E-29
	<i>PA0641</i>	-2.55319332	1.46E-27	1.34E-24
	<i>PA0628</i>	-2.534582315	1.07E-31	1.47E-28
	<i>PA0638</i>	-2.46361566	3.36E-33	9.27E-30
	<i>PA0636</i>	-2.444427878	2.44E-22	1.04E-19
	<i>PA0635</i>	-2.415846681	1.41E-22	6.50E-20
	<i>gfnR</i>	-2.353411699	6.68E-14	8.58E-12
	<i>PA0630</i>	-2.310960008	2.67E-21	8.68E-19
	<i>PA0639</i>	-2.28619203	5.70E-25	2.86E-22
	<i>PA3638</i>	-2.285364587	4.83E-11	4.17E-09
	<i>PA0634</i>	-2.272274644	9.09E-19	2.09E-16
	<i>PA1168</i>	-2.27006418	2.26E-06	7.65E-05
	<i>PA0637</i>	-2.231450235	9.63E-22	3.80E-19
	<i>PA0616</i>	-2.222013132	1.13E-25	7.79E-23
	<i>PA0625</i>	-2.213888097	1.84E-21	6.78E-19
	<i>ptrB</i>	-2.203953211	6.10E-17	1.05E-14
	<i>PA0631</i>	-2.195277667	2.16E-20	6.63E-18
	<i>PA0613</i>	-2.187796631	4.07E-18	8.32E-16
	<i>PA0618</i>	-2.143020354	3.73E-18	7.91E-16

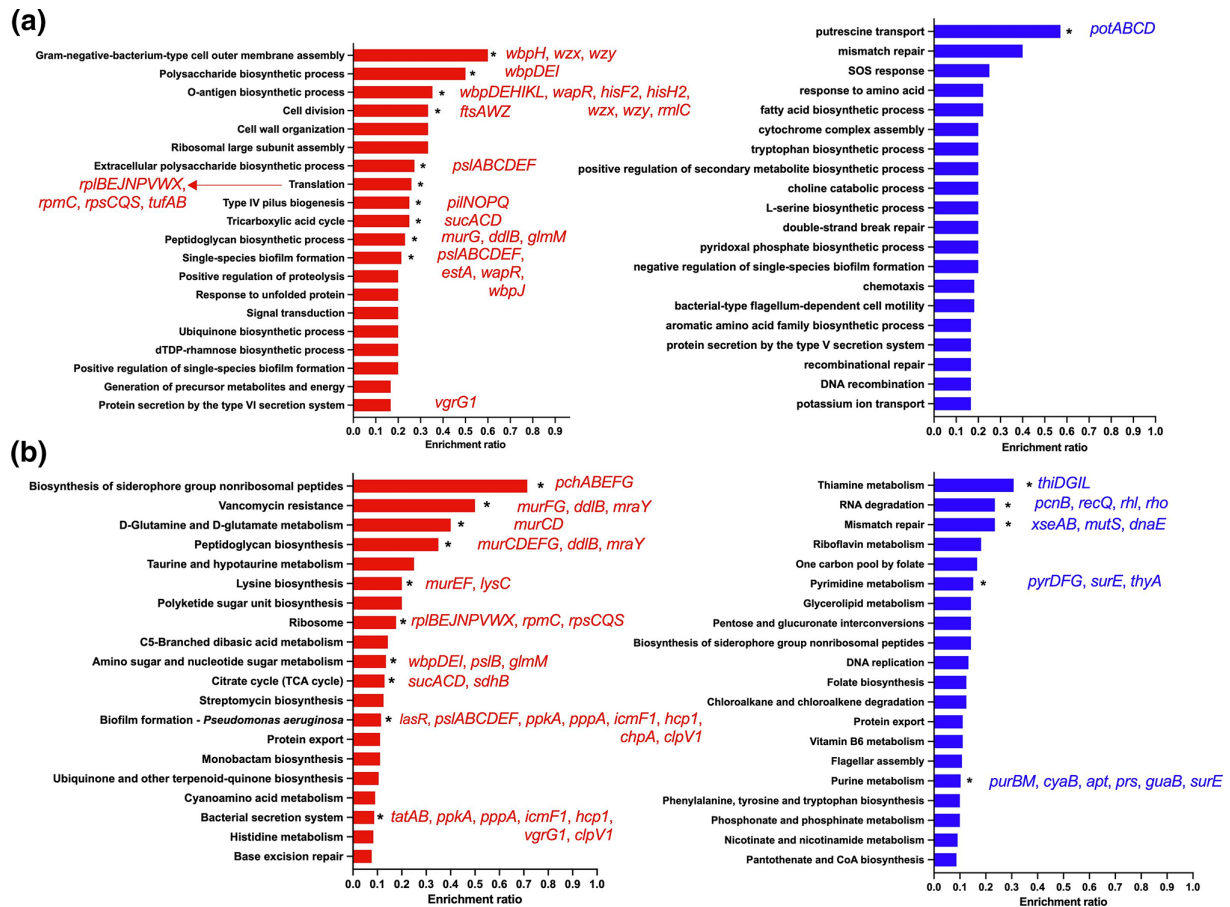


Fig. 3. Meropenem treatment increases the expression of genes required for biofilm formation and iron homeostasis. (a) GO term enrichment analysis and (b) KEGG analysis of genes significantly differentially regulated in *P. aeruginosa* single-species biofilms by meropenem. Bars in red represent pathways/processes that were significantly upregulated, while bars in blue represent pathways/processes that were significantly downregulated. Data were analysed using Fisher's exact test and the Benjamini–Hochberg method for FDR correction (*, FDR \leq 0.05).

PA1051). This suggests that the presence of *C. albicans* may result in reduced production of the MexEF-OprN efflux pump in *P. aeruginosa* biofilm cells.

DISCUSSION

Biofilms are medically important, as they result in antimicrobial resistance and protect microbes from the actions of the immune system, making infection harder to treat. Furthermore, biofilms are normally mixed-species communities, with our understanding of how microbe–microbe interactions within these communities affect disease progression being limited. We have previously shown that *C. albicans* enhances the tolerance of *P. aeruginosa* to meropenem, a commonly used carbapenem for the treatment of chronic *P. aeruginosa* infections. Therefore, transcriptional analysis was carried out to understand how *C. albicans* promotes meropenem tolerance.

Mechanisms of resistance to the carbapenem class of antibiotics include decreased outer membrane permeability, beta-lactamase expression, increased efflux and target modification. In agreement with this, the addition of meropenem to mature *P. aeruginosa* single-species biofilms resulted in the significant induction of *ampC*, a beta-lactamase precursor. Differential regulation of *ampC* is linked to the natural resistance of *P. aeruginosa* to this class of antibiotic [21]. Imipenem, another carbapenem used to treat *P. aeruginosa* infections, also induces the expression of *ampC* [22], highlighting *ampC* induction as a conserved response to carbapenems. In addition to *ampC*, the outer-membrane porin, OprD, has also been associated with meropenem resistance. OprD facilitates entry of carbapenems into bacterial cells, and is therefore frequently downregulated in meropenem-resistant strains [21]. In our study, *oprD* was not differentially regulated in response to meropenem treatment. However, there is often a poor correlation between *oprD* mRNA levels and protein levels [21]. Therefore, it is possible that OprD protein levels are downregulated in our biofilms to reduce meropenem uptake.

Table 3. Top 20 differentially regulated *P. aeruginosa* genes in dual-species verses single-species biofilms

	Gene ID	log ₂ fold change	P-value	Padj
Upregulated	PA5384	1.136608573	4.28E-06	0.0002779376198
	PA4097	1.000660434	3.17E-05	0.001214321657
	<i>cdhB</i>	0.9811955186	5.21E-06	0.0003198899691
	PA2161	0.9810583138	0.0003525619038	0.008808371695
	<i>cdhC</i>	0.936864827	9.18E-07	7.92E-05
	<i>cdhA</i>	0.9176339588	1.17E-09	4.57E-07
	<i>ssrS</i>	0.9064141369	8.59E-13	6.76E-10
	PA0698	0.9014266847	1.42E-05	0.0006654679167
	PA2349	0.8950640392	2.59E-05	0.001067775173
	PA2181	0.8814108328	9.08E-08	1.36E-05
	PA2090	0.8800287264	1.03E-07	1.50E-05
	PA2324	0.867636224	1.23E-05	0.0005976988392
	PA2213	0.8532908266	7.35E-16	1.35E-12
	PA2180	0.8294884121	1.24E-09	4.57E-07
	PA4088	0.8176661995	7.06E-09	1.77E-06
	<i>czcC</i>	0.8158365062	9.15E-08	1.36E-05
	<i>phzC2</i>	0.7984412293	3.58E-07	4.21E-05
	<i>phzC1</i>	0.788317377	3.04E-05	0.00117677973
	PA4098	0.7866314328	0.0005949449343	0.01341483781
	<i>phzA1</i>	0.7731714023	4.59E-06	0.0002943805348
Downregulated	PA3572	-0.9273548369	0.003059228691	0.0450911657
	<i>argB</i>	-0.8972467441	3.86E-09	1.02E-06
	<i>rnpB</i>	-0.8480655754	3.66E-08	5.77E-06
	PA1746	-0.7808094482	2.35E-05	0.0009898437811
	PA4351	-0.7261070956	2.78E-05	0.00110592551
	PA5303	-0.6697670409	3.36E-13	3.09E-10
	<i>fisY</i>	-0.5672143027	1.04E-08	2.29E-06
	PA4276.1	-0.5616231924	1.53E-05	0.0006970521393
	PA4690.3	-0.5391253382	0.002180107776	0.03467439429
	PA1137	-0.5377222343	1.98E-11	9.95E-09
	PA3133.3	-0.5289323628	0.0003527179099	0.008808371695
	<i>ssrA</i>	-0.5226447106	3.43E-06	0.0002369715977
	PA3573	-0.5188361793	6.23E-05	0.002219824419
	PA4421	-0.5151637325	1.87E-11	9.95E-09
	PA1796.1	-0.4988013205	0.001465215919	0.02591835467
	PA4637	-0.4958503362	0.001596304616	0.02744549898
	PA1510	-0.4875966167	3.42E-08	5.55E-06
	PA3951	-0.4870639148	9.51E-09	2.25E-06
	PA2943	-0.4748718073	1.76E-06	0.0001407992501
	PA3277	-0.4616098795	0.0007070580094	0.01530295354

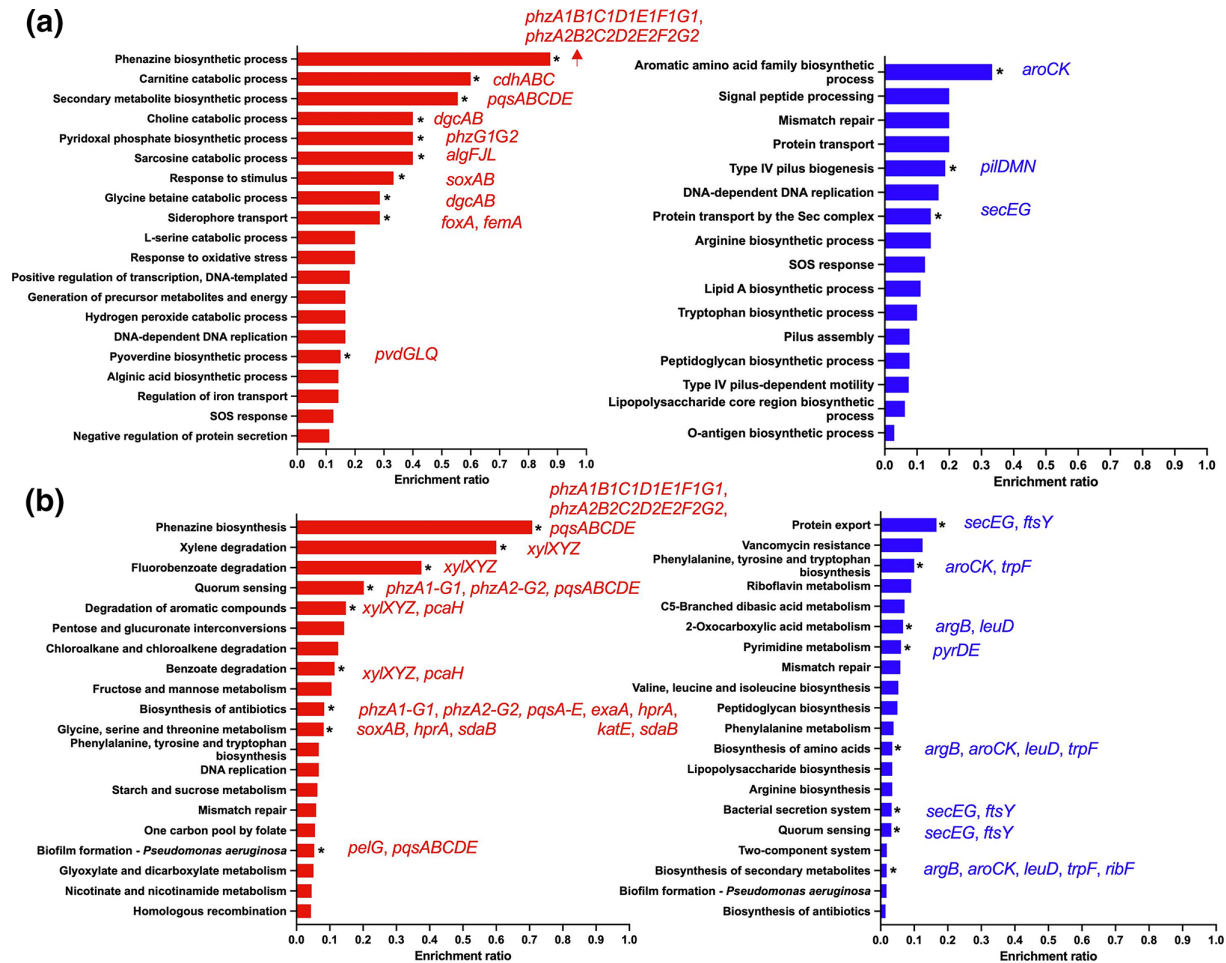


Fig. 4. *P. aeruginosa* upregulates genes that provide a competitive fitness advantage during growth in dual-species biofilms. (a) GO term enrichment analysis and (b) KEGG analysis of *P. aeruginosa* genes significantly differentially regulated in dual-species biofilms. Bars in red represent pathways/processes that were significantly upregulated, while bars in blue represent pathways/processes that were significantly downregulated. Data were analysed using Fisher's exact test and the Benjamini-Hochberg method for FDR correction (*, FDR \leq 0.05).

Interestingly, treatment of *P. aeruginosa* biofilms with either meropenem or imipenem, results in the significant induction of biofilm associated genes [22]. For example, in response to meropenem extracellular polysaccharide biosynthetic genes such as *pslB*, *pslD*, *pslE* and *pslF* were significantly upregulated. The induction of these polysaccharide genes results in enhanced intracellular and cell-substrate interactions, and the expression of subsets of these genes are commonly enhanced in biofilm-forming clinical isolates. Psl is thought to enhance the tolerance of *P. aeruginosa* biofilms to a range of antimicrobial agents including colistin, tobramycin and ciprofloxacin [23].

In response to meropenem, genes involved in the type 6 secretion system (T6SS) were also significantly upregulated, including *ppkA*, *pppA*, *clpV1*, *icmF1*, *vgrG1* and *hcp1*. The structure of the T6SS bears similarities to the cell-puncturing needle of bacteriophage viruses; the roles of the T6SS in *P. aeruginosa* include virulence within hosts, delivery of toxins to neighbouring microbes that are competing for resources, and biofilm formation [24, 25]. Although there has been no previous research linking antibiotic treatment to upregulation of T6SS activity in *P. aeruginosa*, the T6SS has been implicated in drug resistance in *A. baumannii* and *K. pneumoniae* [26, 27]. However, in *P. aeruginosa*, the T6SS is upregulated during competition with other bacteria as a result of kin cell lysis [28]. Therefore, it is possible that the observed increase in transcription of components of the T6SS is a result of meropenem-induced cell lysis.

During growth in dual-species biofilms, *P. aeruginosa* upregulates genes required for quorum sensing, with all genes from the *pqsABCDE* operon being significantly upregulated. The *pqs* operon encodes several enzymes required for production of the *Pseudomonas* quinolone signal (PQS) [29]. *C. albicans*-dependent regulation of PQS production is complex. The fungal quorum-sensing molecule, farnesol, inhibits PQS production in planktonic interactions through modulation of PqsR-dependent transcription of the *pqs* operon [30]. However, in *lasR*-deficient *P. aeruginosa* strains, *C. albicans* restores

Table 4. Top 20 differentially regulated *P. aeruginosa* genes in dual-species biofilms treated with 5 µg ml⁻¹ meropenem vs untreated dual-species biofilms

	Gene ID	log ₂ fold change	P-value	Padj
Upregulated	<i>ampC</i>	5.515387278	7.06E-51	1.95E-47
	<i>PA4280.5</i>	3.700797106	7.82E-06	0.0002259666295
	<i>PA0668.1</i>	3.67825942	8.05E-06	0.0002315796869
	<i>PA4690.5</i>	3.49778215	2.92E-05	0.0007129969543
	<i>PA5369.5</i>	3.485074044	3.01E-05	0.0007281076683
	<i>PA4111</i>	2.963866877	8.30E-18	1.99E-15
	<i>PA4690.2</i>	2.605990986	0.003138749477	0.03261549984
	<i>PA5369.2</i>	2.604799667	0.003142239391	0.03261549984
	<i>PA4280.2</i>	2.596702641	0.003658501903	0.03633497754
	<i>PA0668.4</i>	2.58797494	0.003177734857	0.03286039678
	<i>hcp1</i>	2.353244054	1.14E-22	9.02E-20
	<i>PA0466</i>	1.908730924	1.39E-11	1.30E-09
	<i>pchG</i>	1.899939854	1.07E-08	6.09E-07
	<i>PA0126</i>	1.883859984	5.23E-13	6.57E-11
	<i>PA0050</i>	1.820434031	2.26E-06	7.78E-05
	<i>clpV1</i>	1.75537326	1.53E-11	1.37E-09
	<i>vgrG1</i>	1.730154359	3.27E-12	3.37E-10
	<i>PA4222</i>	1.708263864	2.26E-12	2.45E-10
	<i>PA0563</i>	1.691305244	1.68E-11	1.47E-09
	<i>PA0086</i>	1.677560955	4.89E-10	3.46E-08
Downregulated	<i>PA0627</i>	-2.524464134	1.52E-15	2.79E-13
	<i>gfnR</i>	-2.418533396	8.72E-23	8.02E-20
	<i>PA0641</i>	-2.404904976	2.04E-20	7.49E-18
	<i>PA0638</i>	-2.364308134	4.30E-21	1.98E-18
	<i>PA0628</i>	-2.298437047	1.97E-20	7.49E-18
	<i>PA0635</i>	-2.263949045	1.95E-16	3.84E-14
	<i>PA0629</i>	-2.260720569	5.93E-19	1.92E-16
	<i>PA0639</i>	-2.220599378	2.18E-21	1.21E-18
	<i>PA0636</i>	-2.211742782	4.17E-18	1.15E-15
	<i>PA0625</i>	-2.099597686	3.91E-17	8.30E-15
	<i>PA0616</i>	-2.045775386	2.09E-15	3.72E-13
	<i>PA3638</i>	-2.041401724	2.33E-18	7.14E-16
	<i>PA0634</i>	-1.982462548	1.30E-17	3.00E-15
	<i>PA1168</i>	-1.971155931	1.76E-14	2.56E-12
	<i>PA0631</i>	-1.92975403	4.75E-15	7.95E-13
	<i>PA0619</i>	-1.919212071	1.02E-13	1.31E-11
	<i>PA3631</i>	-1.891142906	4.13E-19	1.43E-16
	<i>PA0626</i>	-1.882097335	5.95E-17	1.22E-14
	<i>PA0630</i>	-1.820934828	4.77E-11	3.71E-09
	<i>PA0637</i>	-1.809932956	1.14E-14	1.75E-12

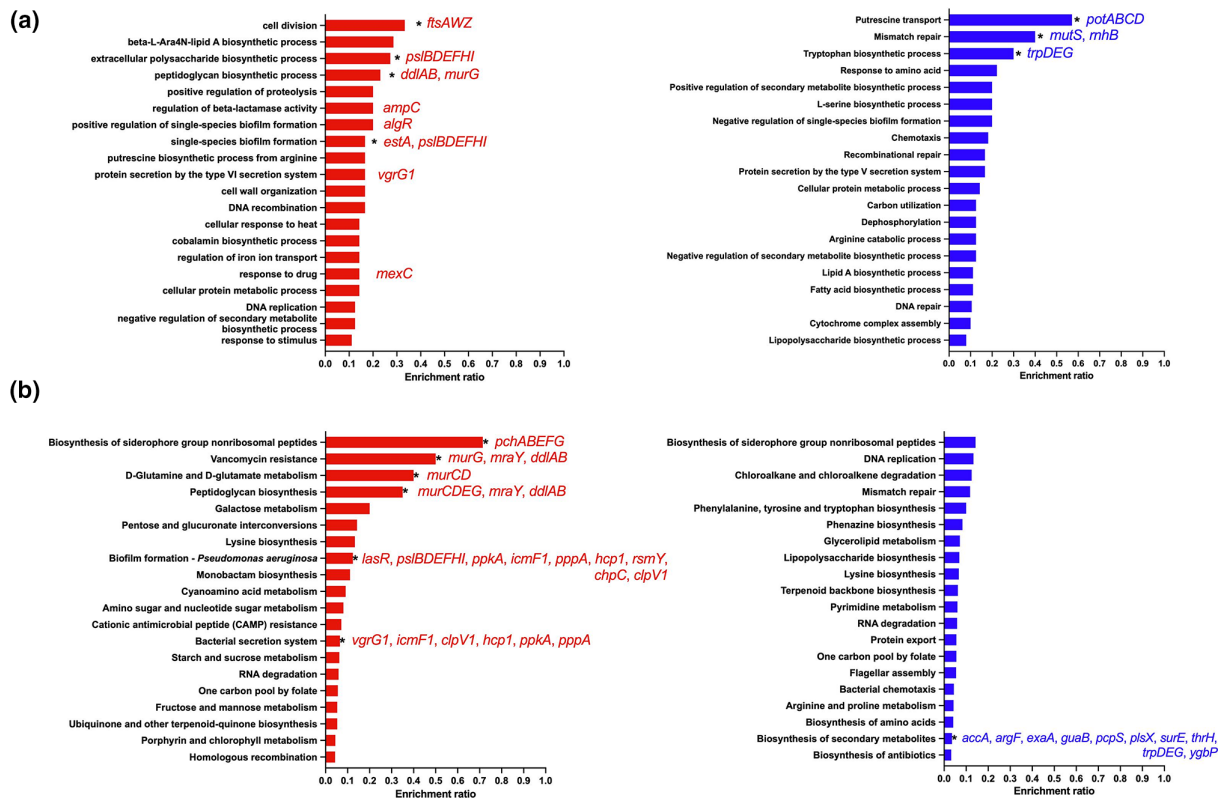


Fig. 5. Dual-species biofilms do not perturb the *P. aeruginosa* transcriptional response to meropenem. (a) GO term enrichment analysis and (b) KEGG analysis of *P. aeruginosa* genes significantly differentially regulated in dual-species biofilms in response to meropenem treatment. Bars in red represent pathways/processes that were significantly upregulated, while bars in blue represent pathways/processes that were significantly downregulated. Data were analysed using Fisher's exact test and the Benjamini-Hochberg method for FDR correction (*, FDR \leq 0.05).

PQS production through farnesol-induced ROS-dependent production of C4-HSL [31]. Increased C4-HSL results in the induction of *pqsH* and therefore restores PQS and phenazine production [31]. However, PQS is also induced under iron-limiting conditions [32], resulting in increased biosynthesis of phenazines and siderophores, which both scavenge iron from the environment.

Phenazines are redox-active molecules that affect the redox balance of cells, the uptake of metabolites and gene expression, and have also been shown to enhance *P. aeruginosa* tolerance to the antibiotic ciprofloxacin [33]. Phenazines are also able to facilitate electron transfer within biofilms [34]. Pathways associated with the biosynthesis of these molecules were the most significantly upregulated biological process in our dual-species biofilms, and in other studies highlighting their importance in interkingdom interactions.

Our transcriptional analysis identified several RND and ABC transporters to be differentially regulated in dual-species biofilms in the presence of meropenem. Little is known about the role of ABC transporters in *P. aeruginosa* drug resistance [35], but it is clear that the presence of *C. albicans* alters the expression of several efflux pumps, which may contribute to carbapenem tolerance in dual-species biofilms. The *P. aeruginosa* genome encodes for at least 12 RND efflux pumps [36], 3 of which (*MexAB-OprM*, *MexXY-OprM* and *MexCD-OprJ*) have been linked to meropenem resistance [37, 38]. *MexC* was induced in dual-species biofilms in response to meropenem treatment, suggesting that *P. aeruginosa* may increase efflux through *MexCD-OprJ*. Phenazines and their derivatives have been shown to directly regulate the expression of several efflux pumps, including *mexG* [39, 40]. Therefore, increased synthesis of phenazines in these biofilms could alter drug efflux and increase the tolerance of *P. aeruginosa* to meropenem. In agreement with this, enhanced production of the phenazine, pyocyanin, promotes resistance to the beta-lactam class of antibiotics through reduced drug influx [41].

Other differences between the transcriptional profiles for single- and dual-species biofilms in response to meropenem include genes involved in biofilm formation (*pslH*, *pslI* and *algR*) and outer-membrane vesicles (OMVs). *AlgR* is a transcriptional regulator of genes involved in alginate production [42]. Therefore, in addition to enhanced extracellular polysaccharide production through *Psl* biosynthesis, dual-species biofilms will also contain increased levels of alginate. Increased alginate

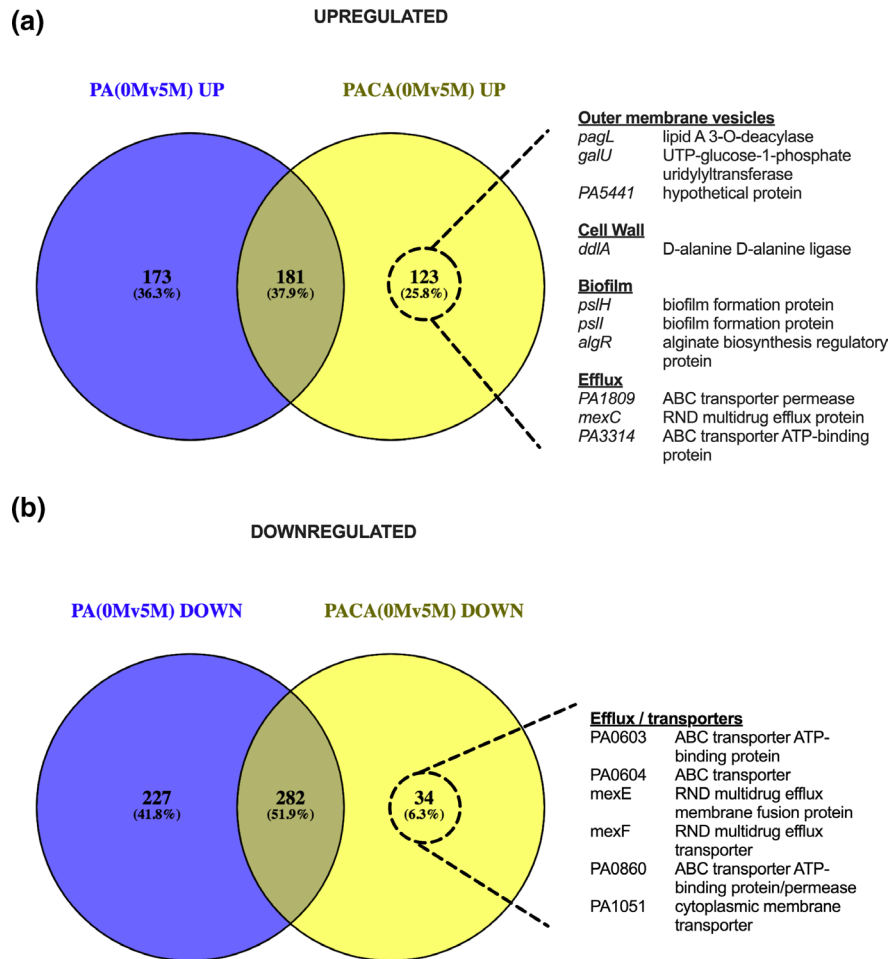


Fig. 6. Identification of overlapping differentially expressed genes in *P. aeruginosa* mono and dual species biofilms in response to meropenem. Numbers of significantly upregulated (a) and downregulated (b) genes ($P_{adj} \leq 0.05$), in response to treatment with 5 $\mu\text{g}/\text{mL}$ meropenem, are compared between single- and dual-species biofilms (PA *P. aeruginosa*; CA *C. albicans*). Genes of interest, unique to the dual-species condition, are listed on the right-hand side.

production is known to enhance biofilm production [22], but has not been directly linked to antibiotic resistance. OMVs play important roles in biofilms, including the secretion of PQS, extracellular DNA and beta-lactamase [43, 44]. Therefore, enhance OMV production in the presence of *C. albicans* may function to increase the composition and complexity of the extracellular matrix, which together with enhanced beta-lactamase secretion would increase the tolerance of *P. aeruginosa* to meropenem.

In summary, in response to meropenem, *P. aeruginosa* biofilms upregulate *ampC* and biofilm formation to provide protection from meropenem. Although dual-species biofilms exhibit enhanced tolerance to meropenem, the *P. aeruginosa* transcriptional responses from dual-species biofilms exposed to meropenem were similar to those of single-species biofilms treated with meropenem, suggesting that the presence of *C. albicans* in dual-species biofilms may limit the diffusion of or sequester the antibiotic, leading to increased bacterial tolerance (Figs 1–6).

Funding information

Work in the Hall Lab is supported by an MRC Career Development Award (MR/L00903X/1) and the BBSRC (BB/R00966X/1, BB/R00966X/2). Work in the Blair Lab is support via a David Phillips Fellowship to J.M.A.B. (BB/M02623X/1). F.A. is supported by the Wellcome Trust Antimicrobials and Antimicrobial Resistance (AAMR) doctoral training programme (108876/Z/15/Z).

Author contributions

R.A.H and J.B conceived and designed the experiments, F.A performed the experiments and analysed the data. R.A.H wrote the manuscript.

Conflicts of interest

The authors declare that there are no conflicts of interest.

References

- Lyczak JB, Cannon CL, Pier GB. Lung infections associated with cystic fibrosis. *Clin Microbiol Rev* 2002;15:194–222.
- Ramakrishnan M, Putli Bai S, Babu M. Study on biofilm formation in burn wound infection in a pediatric hospital in Chennai, India. *Ann Burns Fire Disasters* 2016;29:276–280.
- Chatterjee M, Anju CP, Biswas L, Anil Kumar V, Gopi Mohan C, et al. Antibiotic resistance in *Pseudomonas aeruginosa* and alternative therapeutic options. *Int J Med Microbiol* 2016;306:48–58.
- Bjarnsholt T, Jensen PØ, Fiandaca MJ, Pedersen J, Hansen CR, et al. *Pseudomonas aeruginosa* biofilms in the respiratory tract of cystic fibrosis patients. *Pediatr Pulmonol* 2009;44:547–558.
- Fazli M, Bjarnsholt T, Kirketerp-Møller K, Jørgensen B, Andersen AS, et al. Nonrandom distribution of *Pseudomonas aeruginosa* and *Staphylococcus aureus* in chronic wounds. *J Clin Microbiol* 2009;47:4084–4089.
- Alhede M, Bjarnsholt T, Jensen PØ, Phipps RK, Moser C, et al. *Pseudomonas aeruginosa* recognizes and responds aggressively to the presence of polymorphonuclear leukocytes. *Microbiology* 2009;155:3500–3508.
- Mah TF, O'Toole GA. Mechanisms of biofilm resistance to antimicrobial agents. *Trends Microbiol* 2001;9:34–39.
- Guilhen C, Charbonnel N, Parisot N, Gueguen N, Iltis A, et al. Transcriptional profiling of *Klebsiella pneumoniae* defines signatures for planktonic, sessile and biofilm-dispersed cells. *BMC Genomics* 2016;17:237.
- Doern GV, Brogden-Torres B. Optimum use of selective plated media in primary processing of respiratory tract specimens from patients with cystic fibrosis. *J Clin Microbiol* 1992;30:2740–2742.
- Fourie R, Pohl CH. Beyond antagonism: the interaction between *Candida* species and *Pseudomonas aeruginosa*. *JoF* 2019;5:34.
- Brand A, Barnes JD, Mackenzie KS, Odds FC, Gow NAR. Cell wall glycans and soluble factors determine the interactions between the hyphae of *Candida albicans* and *Pseudomonas aeruginosa*. *FEMS Microbiol Lett* 2008;287:48–55.
- Hogan DA, Kolter R. *Pseudomonas-Candida* interactions: an ecological role for virulence factors. *Science* 2002;296:2229–2232.
- Alam F, Catlow D, Di Maio A, Blair JMA, Hall RA. *Candida albicans* enhances meropenem tolerance of *Pseudomonas aeruginosa* in a dual-species biofilm. *J Antimicrob Chemother* 2020;75:925–935.
- Harriott MM, Noverr MC. *Candida albicans* and *Staphylococcus aureus* form polymicrobial biofilms: effects on antimicrobial resistance. *Antimicrob Agents Chemother* 2009;53:3914–3922.
- Kong EF, Tsui C, Kucharíková S, Andes D, Van Dijk P, et al. Commensal protection of *Staphylococcus aureus* against antimicrobials by *Candida albicans* biofilm matrix. *mBio* 2016;7:e01365-16.
- De Brucker K, Tan Y, Vints K, De Cremer K, Braem A, et al. Fungal β -1,3-glucan increases ofloxacin tolerance of *Escherichia coli* in a polymicrobial *E. coli/Candida albicans* biofilm. *Antimicrob Agents Chemother* 2015;59:3052–3058.
- Mitchell KF, Zarnowski R, Sanchez H, Edward JA, Reinicke EL, et al. Community participation in biofilm matrix assembly and function. *Proc Natl Acad Sci U S A* 2015;112:4092–4097.
- Nett J, Lincoln L, Marchillo K, Massey R, Holoyda K, et al. Putative role of beta-1,3 glucans in *Candida albicans* biofilm resistance. *Antimicrob Agents Chemother* 2007;51:510–520.
- Xie C, Mao X, Huang J, Ding Y, Wu J, et al. KOBAS 2.0: a web server for annotation and identification of enriched pathways and diseases. *Nucleic Acids Res* 2011;39:W316–22.
- Choi D-S, Kim D-K, Choi S-J, Lee J, Choi J-P, et al. Proteomic analysis of outer membrane vesicles derived from *Pseudomonas aeruginosa*. *Proteomics* 2011;11:3424–3429.
- Khaledi A, Schniederjans M, Pohl S, Rainer R, Bodenhofer U, et al. Transcriptome profiling of antimicrobial resistance in *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 2016;60:4722–4733.
- Bagge N, Schuster M, Hentzer M, Ciofu O, Givskov M, et al. *Pseudomonas aeruginosa* biofilms exposed to imipenem exhibit changes in global gene expression and beta-lactamase and alginate production. *Antimicrob Agents Chemother* 2004;48:1175–1187.
- Billings N, Millan M, Caldara M, Rusconi R, Tarasova Y, et al. The extracellular matrix component Psl provides fast-acting antibiotic defense in *Pseudomonas aeruginosa* biofilms. *PLoS Pathog* 2013;9:e1003526.
- Chen L, Zou Y, Kronfl AA, Wu Y. Type VI secretion system of *Pseudomonas aeruginosa* is associated with biofilm formation but not environmental adaptation. *MicrobiologyOpen* 2020;9:e991.
- Chen L, Zou Y, She P, Wu Y. Composition, function, and regulation of T6SS in *Pseudomonas aeruginosa*. *Microbiol Res* 2015;172:19–25.
- Liu L, Ye M, Li X, Li J, Deng Z, et al. Identification and characterization of an antibacterial type VI secretion system in the carbapenem-resistant strain *Klebsiella pneumoniae* HS11286. *Front Cell Infect Microbiol* 2017;7:442.
- Wang J, Zhou Z, He F, Ruan Z, Jiang Y, et al. The role of the type VI secretion system vgrG gene in the virulence and antimicrobial resistance of *Acinetobacter baumannii* ATCC 19606. *PLoS One* 2018;13:e0192288.
- LeRoux M, Kirkpatrick RL, Montauti EI, Tran BQ, Peterson SB, et al. Kin cell lysis is a danger signal that activates antibacterial pathways of *Pseudomonas aeruginosa*. *Elife* 2015;4.
- Higgins S, Heeb S, Rampioni G, Fletcher MP, Williams P, et al. Differential regulation of the phenazine biosynthetic operons by quorum sensing in *Pseudomonas aeruginosa* PA01-N. *Front Cell Infect Microbiol* 2018;8:252.
- Cugini C, Calfee MW, Farrow JM 3rd, Morales DK, Pesci EC, et al. Farnesol, a common sesquiterpene, inhibits PQS production in *Pseudomonas aeruginosa*. *Mol Microbiol* 2007;65:896–906.
- Cugini C, Morales DK, Hogan DA. *Candida albicans*-produced farnesol stimulates *Pseudomonas quinolone* signal production in LasR-defective *Pseudomonas aeruginosa* strains. *Microbiology* 2010;156:3096–3107.
- Oglesby AG, Farrow JM 3rd, Lee J-H, Tomaras AP, Greenberg EP, et al. The influence of iron on *Pseudomonas aeruginosa* physiology: a regulatory link between iron and quorum sensing. *J Biol Chem* 2008;283:15558–15567.
- Schiessl KT, Hu F, Jo J, Nazia SZ, Wang B, et al. Phenazine production promotes antibiotic tolerance and metabolic heterogeneity in *Pseudomonas aeruginosa* biofilms. *Nat Commun* 2019;10:762.
- Saunders SH, Tse ECM, Yates MD, Otero FJ, Trammell SA, et al. Extracellular DNA promotes efficient extracellular electron transfer by pyocyanin in *Pseudomonas aeruginosa* biofilms. *Cell* 2020;182:919–932.
- Hulen C, Racine P-J, Chevalier S, Feuilloley M, Lomri N-E. Identification of the PA1113 gene product as an ABC transporter involved in the uptake of Carbenicillin in *Pseudomonas aeruginosa* PA01. *Antibiotics* 2020;9:596.
- Scoffone VC, Trespidi G, Barbieri G, Irudal S, Perrin E, et al. Role of RND efflux pumps in drug resistance of cystic fibrosis pathogens. *Antibiotics* 2021;10:863.
- Fusté E, López-Jiménez L, Segura C, Gainza E, Vinuesa T, et al. Carbapenem-resistance mechanisms of multidrug-resistant *Pseudomonas aeruginosa*. *J Med Microbiol* 2013;62:1317–1325.
- Hassuna NA, Darwish MK, Sayed M, Ibrahim RA. Molecular epidemiology and mechanisms of high-level resistance to meropenem and imipenem in *Pseudomonas aeruginosa*. *Infect Drug Resist* 2020;13:285–293.
- Dietrich LEP, Price-Whelan A, Petersen A, Whiteley M, Newman DK. The phenazine pyocyanin is a terminal signalling factor in the quorum sensing network of *Pseudomonas aeruginosa*. *Mol Microbiol* 2006;61:1308–1321.
- Sakhtah H, Koyama L, Zhang Y, Morales DK, Fields BL, et al. The *Pseudomonas aeruginosa* efflux pump MexGHI-OpmD transports a natural phenazine that controls gene expression and biofilm development. *Proc Natl Acad Sci U S A* 2016;113:E3538–47.

41. Zhao X, Jin Y, Bai F, Cheng Z, Wu W, et al. Mutation of PA4292 in *Pseudomonas aeruginosa* increases β -lactam resistance through upregulating pyocyanin production. *Antimicrob Agents Chemother* 2022;66:e00421–00422.
42. Deretic V, Dikshit R, Konyecsni WM, Chakrabarty AM, Misra TK. The algR gene, which regulates mucoidy in *Pseudomonas aeruginosa*, belongs to a class of environmentally responsive genes. *J Bacteriol* 1989;171:1278–1283.
43. Bomberger JM, Maceachran DP, Coutermarsh BA, Ye S, O'Toole GA, et al. Long-distance delivery of bacterial virulence factors by *Pseudomonas aeruginosa* outer membrane vesicles. *PLoS Pathog* 2009;5:e1000382.
44. Florez C, Raab JE, Cooke AC, Schertzer JW. Membrane distribution of the *Pseudomonas* quinolone signal modulates outer membrane vesicle production in *Pseudomonas aeruginosa* *mBio* 2017;8:e01034–01017.

Edited by: R. Manganelli and D. R Neill

Five reasons to publish your next article with a Microbiology Society journal

1. When you submit to our journals, you are supporting Society activities for your community.
2. Experience a fair, transparent process and critical, constructive review.
3. If you are at a Publish and Read institution, you'll enjoy the benefits of Open Access across our journal portfolio.
4. Author feedback says our Editors are 'thorough and fair' and 'patient and caring'.
5. Increase your reach and impact and share your research more widely.

Find out more and submit your article at microbiologyresearch.org.