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Multi-scale engineering properties of tomato fruits related to harvesting, simulation and textural evaluation

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| 1 | Multi-scale engineering properties of tomato fruits related to |
| 2 | harvesting, simulation and textural evaluation |
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| 7 | Abstract |
| 8 | In this study, multi-scale engineering properties related to the harvesting, simulation and textural evaluation |
| 9 | of two tomato cultivars at six ripening stages were simultaneously investigated. A potential ripening scale based |
| 10 | on the ratio of R:G:B for a given ripening stage was suggested. The geometric mean diameter was most closely |
| 11 | correlated with the fruit mass. Tomato fruit feature an irregular shape and asymmetric internal structure at the |
| 12 | macro-scale, non-unique tissue thickness at the meso-scale and an irregular change of size, shape and arrangement |
| 13 | of single cells at the micro-scale. The hardness and shear strength of fruit at different scales and the single cell |
| 14 | mechanics varied with the fruit ripening stage but not the chosen cultivars. The contribution of exocarp to the |
| 15 | hardness of whole fruit gradually increased with fruit ripeness. The hardness and shear strength of fruit tissues and |
| 16 | the fruit's single cells varied between 0.37 and 2.25 MPa and 0.04 and 11.58 MPa, respectively. This puncture |
| 17 | experimental method is well-suited to measure the hardness and shear strength of tomato fruit at different scales |
| 18 | and single tomato cell mechanics. |

19 Keywords: Solanum lycopersicum; Tomato cell; Multi-scale biomechanics; Ripeness; Puncture test

20 **1 Introduction**

Numerous large-scale tomato-growing farms are in operation worldwide because tomatoes are a component of the diet of millions of people. Because the harvesting season is short and harvesting work is concentrated during a brief period of time, labor shortages tend to limit the farm acreage (Tanigaki, Fujiura, Akase, & Imagawa,

2008). Additionally, given the long distance between farms and sale markets, the design and development of

24 intelligent equipment for mechanical harvesting, packaging and transport have received increasing attention 25 26 (Kondo, Yata, Taniwaki, Tanihara, Monta, & Kurita, 2007; Li, Li, Yang, & Wang, 2013a). Furthermore, fruits with 27 a distinct multi-scale nature are very susceptible to mechanical damage during postharvest handling; thus, multi-scale modeling, internal damage simulation and postharvest textural evaluation are extremely important 28 (Genard et al., 2007; Mebatsion, Verboven, Ho, Verlinden, & Nicolai, 2008; Ghysels, Samaey, Van Liedekerke, 29 30 Tijskens, Ramon, & Roose, 2010; Ho et al., 2013). Determining the multi-scale engineering properties of tomato 31 fruits is essential to achieve these aims.

32 Some engineering properties of tomato fruits have been previously investigated. Arazuri et al. (2007), Li et al. (2011) and Sirisomboon et al. (2012) reported the geometric and mechanical macro-properties of tomato fruits at 33 three different stages of ripeness (Arazuri, Jaren, Arana & Perez De Ciriza, 2007; Li, Li, & Liu, 2011; 34 Sirisomboon, Tanaka, & Kojima, 2012). Hetzroni et al. (2011) and Li et al. (2012a) determined the physical and 35 biomechanical properties of the peels and internal tissues of five tomato cultivars at the meso-scale (Hetzroni, 36 Vana, & Mizrach, 2011; Li, Li, Yang, Liu, & Xu, 2012a). Rancic et al. (2010) presented the geometric 37 38 characteristics of the fruits and tissues of two tomato genotypes during fruit development (Rancic, Quarrie, & 39 Pecinar, 2010). Bargel and Neinhuis (2005) focused on the morphology and biomechanics of skin and enzymatically isolated the cuticular membranes of three tomato cultivars during fruit growth and ripening (Bargel 40 & Neinhuis, 2005). 41

42 Tomato fruits are hierarchically structured at the macro-scale, consisting of different tissue types at the 43 meso-scale, each of which is a highly structured arrangement of cells at the micro-scale (Li & Thomas, 2014a, b). 44 However, some important engineering parameters for multi-scale modeling and simulation, such as the geometry 45 of the whole fruit, the size and shape of the different tissues, the cell sizes in each tissue and the multi-scale 46 biomechanics, have never been fully determined for single fruit, even though these characteristics significantly

differ. Additionally, little is known about the relationship between different physical parameters and the micro-mechanics at the cell level. Therefore, a clear gap in knowledge exists between the published data on fruit engineering properties and the necessary data related to intelligent harvesting, multi-scale simulation and postharvest textural evaluation. The objective of this study was to investigate the multi-scale engineering properties of tomato fruits.

52 2 Materials and Methods

53 2.1 Materials

The experiments were conducted in May 2014 at Henan Polytechnic University. Fruits of two tomato 54 varieties, Fendu 79 and Omeiya 333, were used for this study. The tomato fruits were hand-harvested from the 55 Jiaozuo Manfeng Vegetable Planting Base at six ripening stages (green, breaker, turning, pink, light red and red) 56 according to the USDA standards (USDA, 1991), as shown in Fig. 1a. These fruits were inspected to ensure that 57 58 they were not damaged or infested with insects prior to transport to the laboratory. Subsequently, the fruits' 59 surfaces were manually cleaned and dried. In total, 60 tomato fruits (5 samples \times 2 varieties \times 6 ripening stages) were used to measure the multi-scale geometric characteristics, and 12 tomato fruits (1 sample \times 2 varieties \times 6 60 ripening stages) were used for the puncture test. 61

62 **2.2 Quantification of ripeness**

One tomato fruit was randomly selected from each ripening stage (green, breaker, turning, pink, light red and red). These tomatoes were grouped and placed on a blank paper with the support of wedges, as shown in Fig. 1a. A JPEG photo of tomato fruits from front view was then obtained using a digital camera (Canon 95IS, Photo size: 3648×2736 pixels). Subsequently, ten pixel points were randomly grabbed from each tomato fruit using a color picker software (ColorPix version 1.1, http://www.colorschemer.com/colorpix_info.php). The three primary color values of grabbed points, namely Red-Green-Blue (RGB), were then based on the automatic transformation provided by the software.

70 2.3 Multi-scale geometric characteristics measurement

71 The sampled fruits were labeled and then cut into halves with a sharp knife along the stem-blossom axis (Fig. 72 2a). One half of each fruit was cut again along the equatorial axis (Fig. 2b). Further descriptions of the tomato 73 fruit anatomy are given in Thomas (1996). The following were measured using an electronic digital caliper (to an accuracy of 0.01 mm): height above the fruit's equatorial axis (section) (H_1) ; height below the fruit's equatorial 74 axis (H_2); diameter of the equatorial section (D_f); maximum thickness (W_{mmax}), minimum thickness (W_{mmin}), 75 76 middle thickness of the mesocarp tissue (W_{mmid}); thickness of the septa tissue (W_s); and columella diameter (D_{ct}). Subsequently, rectangular tissue blocks, including the exocarp and some of the adhering mesocarp, were excised 77 and soaked in boiling water for 5 minutes. The exocarp samples remained after the mesocarp was carefully 78 scraped off using a razor blade. The thicknesses of the exocarp samples (W_e) were then measured with an 79 electronic digital caliper. 80

As shown in Fig. 2a, some tissue blocks, including the exocarp and mesocarp and some columella tissue 81 82 blocks, were excised from the sample zones shown in the other half of the fruits. The mesocarp and columella tissue samples were cut into thin rectangular slices (length \times width \times thickness: 15 mm \times 10 mm \times 0.5 mm) using 83 84 a razor blade. The exocarp samples remained after the adhering mesocarp sample was carefully scraped off. These 85 tissue samples were made into temporary mounts and then vertically observed using a Belona BL-SM1280 biological microscope that featured a 130w electronic eyepiece (Captured image size: 1280x1024 pixels, 86 Resolution: 96 PPI). The diameters (D_c) (Fig. 2c) of cells in different tomato fruit tissue types were measured with 87 88 a virtual cross ruler using the image processing software Future WinJoe of the eyepiece (Assumption: spherical cell). The reported values of the cell geometric characteristics are the means of 5 cells in corresponding tissues. 89 90 The multi-scale geometric characteristics of tomato fruits were measured within 24 h of sampling at room 91 temperature ($23 \pm 1^{\circ}$ C, 56-58 % RH).

92 2.4 Puncture test

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93 The puncture test, which involves compression and shear components, is one of the most widely used methods for the objective measurement of the biomechanics of fruits. The compression component results 94 95 from the compression effect of a probe tip plate that contacts an area of planar tissue at a puncture point. The peak puncture force of the biomaterial (e.g. fruit, tissue and cell) at a unit compression area in a puncture test 96 has been considered to represent the biomaterial hardness in studies of apple fruit by Harker et al. (2002) and 97 studies of tomato fruit by Biswas et al. (2014) (Harker, Maindonald, Murray, Gunson, Hallett & Walker, 98 99 2002; Biswas, East, Hewett, & Heyes, 2014). The shear component results from the shear effect of the probe tip edge, which contacts a toroidal tissue area at a puncture depth. The peak puncture force of the biomaterial 100 (e.g. fruit, tissue and cell) at a unit shear area in a puncture test is considered the shear strength of the 101

102 biomaterial.

The puncture test of fruits and their exocarp, mesocarp and columella tissues were used to determine the multi-scale hardness and shear strength of tomato fruits as related to texture evaluation. The tests utilized a GY-4 manual fruit sclerometer with a 3.5-mm diameter flat-head stainless steel cylindrical probe (Fig. 3a). The compression speed was approximately 1 mm/s, which was measured using a DM6236P Digital Velometer (Resolution: 0.2 mm/s). A 5-mm diameter counter bore was created in the base plate of the manual test stand. The test was conducted as follows:

First, a tomato fruit was placed on a base plate with the stem-blossom axis parallel to the flat plate. Six points (Fig. 3-b1) on the equatorial section of the fruit were punctured to a depth of 10 mm. The fruit hardness, $P_{\rm fh}$, was calculated with 6 points (replications) using the equation in Fig. 3-c1, and its mean value is reported.

Second, the centerlines of the puncture probe and the counter bore were adjusted on the same straight line. The conjoint exocarp and mesocarp, mesocarp and columella tissues were cut into several standard cuboid samples (Fig. 3-b2, b3) and then fully punctured to counter bore. The tissue sample thickness, d_2 , was measured using an electronic digital caliper. The hardness, P_{th} , and shear strength, σ_{tf} , of the mesocarp and columella tissues

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| 116 | were calculated with 3 samples (replications) using the equation in Fig. 3-c2, and their mean values are reported. |
|-----|---|
| 117 | Because the puncture resistance of the exocarp tissue always exceeded that of the mesocarp tissue (Li et al., |
| 118 | 2012a), the peak puncture force of the exocarp tissue was retained (obtained by the fruit sclerometer) when the |
| 119 | conjoint exocarp and mesocarp sample was punctured (Fig. 3-b2). Each tissue (e.g., exocarp, mesocarp or |
| 120 | columella) consists of cells. To simplify the later analysis and the multi-scale modeling in future, we assumed that |
| 121 | i) the hardness or shear strength values of each tissue and its single cells are the same during the puncture test, |
| 122 | namely $P_{\text{th}}=P_{\text{ch}}$, $\sigma_{\text{tf}}=\sigma_{\text{cf}}$; and that ii) the hardness and the shear strength for single cells in each tissue are |
| 123 | approximations. |

Third, the tissue was assumed to consist of multilayers of regularly arranged cells (Fig. 3-b3). The mean or maximum compression force applied to a single cell, F_{ch} , and the mean shear force applied to a single cell, F_{cf} , were calculated using 3 samples (replications) and the equation in Fig. 3-c3, and their mean values are reported. The labels n_1 and n_2 are the numbers of cells compressed and sheared by the probe during the tissue puncture test, respectively. n_1 represents the number of first-layer cells compressed directly by the probe.

129 2.5 Statistical analysis

- 130 The results were analyzed for statistical significance using a variance analysis in the SAS9.1 software, with a
- 131 significance level of α =0.05.

132 **3 Results and Discussion**

133 **3.1 Ripening stage**

According to the United States Standards for Grades of Fresh Tomatoes, the six ripening stages of tomato fruits include green, breaker, turning, pink, light red, and red. "Green" indicates a completely green color on the fruit surface; "breaker" indicates fruit that is not fully green and features a red area on the surface (< 10 %); more than 10% but less than 30% of the surface of "turning" fruit is red; more than 30% but less than 60% of the surfaces of "pink" fruit are red; more than 60% but less than 90% of the surfaces of "light red" are red; and "red"

denotes fruit whose surfaces are more than 90% red. The above ripeness classification is a simple qualitative recognition method. The RGB values and their standard deviations corresponding to six ripening stages are presented in Fig. 1b. A ripening scale based on the ratio of R:G:B obtained from quantifying the RGB value has been proposed. The standard deviations showed that the three primary colors, RGB, fluctuated for a given ripening stage. The RGB threshold values of different ripening stages contribute to quantitative recognition and are vital for the color-sorting device of a tomato harvester and other quality detection systems (Li, Kan, Tan, Zhang, Sui, & Chen, 2012b).

As the fruit ripened, the color gradually changed from green to red, mainly because of the increased lycopene 146 and decreased chlorophyll content in fruit tissues (Salunkhe et al., 1974), while the hardness gradually decreased 147 148 due to multiple coordinated processes, including the disassembly of polysaccharide in the primary cell wall and middle lamella and transpirational water/turgor loss (Saladie, Jadav, & Yu, 2007). Therefore, tomato fruits at the 149 pink and light-red stages are considered optimal for harvesting due to the long distance transportation from farms 150 151 to markets and thus are the most important fruit for biomechanical measurements and simulation analysis. After quantitative transformation, the R, G, and B values \pm standard deviations were 150 \pm 31, 110 \pm 27, and 70 \pm 18 for 152 pink tomatoes and 152±15, 78±13, and 54±9 for light red tomatoes, respectively. 153

154 **3.2 Multi-scale geometry**

155 **3.2.1 Fruit**

The height, diameter and mass of the two tomato cultivars varied from 51.75-61.86 mm, 58.56-74.19 mm and 89.49-224.56 g, respectively, which further illustrated that the shapes of tomato fruits are irregular. Based on these data, the finger grasping stroke and the surface width should exceed 61.86 mm and 37.09, respectively, for the two-finger harvesting robot designed by Monta, Kondo, & Ting. (1998) and Li et al. (2013a). The centrifugal force of the separation device of the tomato harvester designed by Li et al. (2012b) should be sufficient to separate the largest (224.56 g) fruits from tomato stems; and the grid gap of its conveyor chain should not exceed 51.75

mm. Fig. 4a shows the height and diameter of the two tomato cultivars at the six ripening stages. The height and diameter of the two tomato cultivars at the six ripening stages did not significantly differ because the fruits had already been selected during manual harvesting. Therefore, the macro size of the fruits will not affect the

- 165 following physical-mechanical characteristics in section 3.3 and 3.4 .
- Fig. 4b shows the fruit heights above and below the equatorial section. The height above the equatorial 166 section of tomato fruits was 21.93±3.73 mm, and the height below the equatorial section was 33.47±4.03 mm. The 167 168 fruit height below the equatorial section was larger than the height above the equatorial section. This finding further illustrates that the macro-structure of tomato fruits is asymmetrical and that consistent, significant 169 170 differences in the fruit mechanics can be expected during quasi-static compression, dynamic impact or free drop experiments from different force action points, as reported by Li et al. (2011) and Van Linden, Scheelinck, Desmet, 171 & De Baerdemaeker. (2006). Therefore, a real multi-scale fruit model is more valuable for a mechanical handling 172 (such as harvesting, transporting and packaging) simulation than an ideal model. 173

Fig. 4c shows the regression relationships between the fruit mass and height and the equatorial diameter and geometric mean diameter. The fruit mass and its geometric mean diameter most closely correlated according to the determination coefficient, R^2 . of the regression functions (n=12). When a robot harvests fruit, the stable grasp force of fingers needs to be calculated from the friction coefficient and mass (Chen, Hasegawaa, & amashita, 2006; Li et al., 2013a), but the fruit mass cannot be directly measured with a balance. According to the Fig. 4c, the geometric mean diameter can be considered as the most accurate parameter for predicting the fruit mass for robot harvesting.

181 **3.2.2 Tissue**

182 The results of the two-factor analysis of variance indicated significant differences in the thickness of the 183 exocarp tissue, W_e , but no significant differences in the other geometric parameters of the tissues of the two 184 tomato cultivars. Furthermore, the measured geometric parameters of the tissues did not correlate with ripeness.

185 This lack of correlation may have been due to the fact that the geometric sizes of tomato fruits no longer rapidly 186 grow as a result of insignificant cell enlargement 5~8 weeks after anthesis, when the tomato fruit has accumulated 187 the majority of its final mass from the mature green stage (Thomas, 1996).

- The thicknesses of Fendu 79 and Omeiya 333 exocarp tissue were 0.17±0.02 mm and 0.14±0.03 mm, 188 respectively. The thicknesses (W_{mmax} , W_{mmin} and W_{mmid}) of the mesocarp tissue, the thickness (W_s) of the septa 189 tissue and the columella diameter (D_{cl}) were 7.56±1.02 mm, 5.17±0.63 mm, 4.17±0.84 mm, 6.38±0.62 mm and 190 191 9.76±0.75 mm, respectively, which further illustrated non-unique tissue thickness at the meso-scale. All of the exocarp tissues were thicker than those examined by Hetzroni et al. (2011) and Bargel & Neinhuis (2005). Lahaye, 192 193 Devaux, Poole, Seymour, & Causse (2013) proposed that tomato pericarp tissue thickness ranged from 5.2 to 9.3 mm. These measured geometric data are essential for tissue geometric modeling in multi-scale simulations but 194 have not been fully determined for individual fruit cultivars. The measured geometric sizes of tissues demonstrate 195 the complex internal structural characteristics of tomato fruits. Structural failure has been suggested as another 196 197 type of mechanical damage to tomato fruits in addition to the failure of tissues (Li, Li, Yang, & Liu, 2013b).
- 198 **3.2.3 Cell**

The cell diameters in different types of tissues of the two tomato cultivars at six ripening stages are presented in Fig. 5a. The sizes of cells in the mesocarp and columella tissues significantly varied, as illustrated by several large standard deviation values. The results of the analysis of variance show that the two tomato cultivars and the three tissue types (e.g. exocarp, mesocarp, and columella) markedly affected the cell diameter, D_c . The mean cell diameters of the exocarp, mesocarp and columella tissues were 32 µm, 327 µm and 340 µm, respectively, for *Fendu* 79 tomato fruits and 35 µm, 389 µm and 429 µm, respectively, for *Omeiya* 333 tomato fruits.

Fig. 5b shows a picture of the microstructure of the exocarp, mesocarp and columella tissues of *Fendu* 79 tomato fruits at the breaker stage. The mesocarp and columella tissues contained significantly fewer cells per unit volume than the exocarp tissue. The cells from the exocarp tissue were small irregular polygons that were

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| 208 | compactly and densely arranged due to the cutin membrane, while the cells from the mesocarp and columella |
|-----|--|
| 209 | tissues were large and sparsely and loosely arranged. Therefore, the failure stress and elastic modulus of exocarp |
| 210 | tissues are always exceed those of the internal mesocarp and columella tissues based on the biomaterial structure |
| 211 | and components, as reported by Li et al. (2012a) and Bargel & Neinhuis (2005). |
| 212 | Furthermore, the cell diameters in the tissues did not correlate with fruit ripeness in the experimental results. |
| 213 | During fruit development, slow cell division and enlargement occurs for 2~3 weeks, followed by rapid cell |
| 214 | enlargement for another 3~5 weeks. Subsequently, cell enlargement continues but will not significantly change |
| 215 | from the mature green stage (Thomas, 1996). Therefore, the cell size did not markedly differ between the six |
| 216 | ripening stages. |
| 217 | 3.3 Multi-scale biomechanics |
| 218 | The results of the analysis of variance showed that the hardness, single cell mechanics and shear strength |

of fruit at different scales (e.g. fruit, exocarp, mesocarp, columella and cell) were not significantly different between the two cultivars. Therefore, one of the fruit cultivars, *Fendu* 79, was used as an example to illustrate the multi-scale mechanical properties in this section.

222 3.3.1 Multi-scale mechanical properties obtained from compression component

223 The multi-scale mechanical properties of tomato fruits at six ripening stages obtained from compression testing are given in Table 1. The ripening stage significantly affected the hardness of fruit at different scales. The 224 225 hardness of fruit at different scales negatively correlated with fruit ripeness. The hardness of whole fruits did not 226 significantly differ from that of exocarp tissue, and the hardness of exocarp tissue and its cells exceeded those of the mesocarp and columella tissues and their cells. This finding illustrates that the exocarp tissue is the most 227 228 important factor for fruit hardness and would have a largest impact on the protection of internal tissues during 229 mechanical handling. Similarly, some puncture studies reported tomato hardness values on the order of 1~2 MPa 230 for six varieties of ripe fruit (Stommel, Abbott, Campbell, & Francis, 2005), decreases from 1.2 to 0.1 MPa for

mesocarp and columella tissues as the fruit ripened (Wu & Abbott, 2002), and values of 0.4~0.7 MPa for the mesocarp and columella of ripe fruit punctured by probes of three sizes (Lana, Tijskens, Theije, Dekker, & Barrett, 2007). These published results are similar to those reported here. Furthermore, the difference in the hardness between whole fruit and exocarp gradually decreased as the fruit ripened, which further illustrated that the contribution of the exocarp to the hardness of whole fruit gradually increased as the fruit ripened. This viewpoint is also supported by Jackman & Stanley (1994).

Hardness indicates the strength with which fruit can resist to external compression force. The transporter, seller and consumer can use these forces to accurately evaluate the quality of postharvest fruit. In material science, the hardness of metal and plastic materials is always measured based on a national standard method. However, such a measurement method for biomaterials is currently lacking. The peak puncture force (Goyal, Kingsly, Kumar, & Walia, 2007) and peak compression stress (Wu & Abbott, 2002) have been used to characterize the fruit hardness in previous studies, and this approach differs from that utilized in this study. Therefore, comparing the data presented herein with previously published results for further analysis is difficult.

In micro-scale research, the mean numbers of compressed cells in the exocarp, mesocarp and columella 244 tissue samples were 63553, 1614 and 1503, respectively. During the puncture test, the applied mean compression 245 246 force to single cells in the mesocarp and columella tissues was significantly larger than that experienced by single cells in the exocarp tissue. The numbers of cells in the exocarp, mesocarp and columella tissue samples, which 247 directly contacted the probe end, were 11963, 115 and 106, respectively. The applied maximum compression force 248 to single cells in the mesocarp and columella tissues was at least 110 times larger than that applied to single cells 249 250 in the exocarp tissue. Some previous studies have reported bursting force ranges of single cells in ripe tomato 251 mesocarp tissue of 0~24 mN and a mean bursting force of 3.6 mN (Blewett, 2000). In contrast, the data in Table 2 252 show that the cells that directly contacted the probe end at the first several layers will burst, and their protoplasts 253 flow out during the puncture test. Conspicuously, a piece of tissue that was dropped into the counter bore

- 254 contained some damaged cells in its upper layer and some undamaged cells in its lower layer, and some liquids
 255 remained on the end of the probe and the base plate after puncture.
- **3.3.2 Multi-scale mechanical properties obtained from the shear component**

257 The multi-scale mechanical properties of tomato fruit at six ripening stages obtained from the shear component are listed in Table 2. The shear strength of fruit at different scales significantly differed by the ripening 258 stage. The shear strength of fruit at different scales (e.g., fruit, exocarp, mesocarp, columella and cells) negatively 259 260 correlated with fruit ripeness. At the meso- and micro-scale, the shear strengths of the mesocarp and columella tissues did not significantly differ, and the shear strength of the exocarp tissue and its single cells was an order of 261 magnitude larger than those of the mesocarp, columella tissues and their single cells. This finding illustrates that 262 263 the mesocarp, columella tissues and their single cells were prone to shear failure at a smaller external force than the exocarp tissue and its single cells. Some previous studies reported shear strength values of light-red tomato 264 exocarp and mesocarp tissues of 2.98±1.03 MPa and 0.07±0.02 MPa, respectively, as determined by a shear 265 experiment (Li et al., 2012a). In contrast, Table 2 shows that shear strength values in mesocarp that are close to 266 these values, but shear strength values in the exocarp that are approximately 0.5 times. In micro-scale research, 267 268 the mean numbers of sheared cells in the exocarp, columella and mesocarp tissues were 2324, 603 and 584, 269 respectively. The applied mean shear force to single cells in the mesocarp and columella tissues was larger than that applied to single cells in the exocarp tissue during a puncture test. 270

- The shear strength, especially the maximum shear stress of biomaterials at failure (damage), is another important characteristic of materials. This parameter can be helpful for the development of a fruit processing machines, such as an auto-slicer; the design of packaging methods to prevent puncture damage during transporting; and the textural assessment of crunchiness during chewing.
- 275 4 Conclusions

276

In this study, the multi-scale engineering properties of two tomato cultivars at six ripening stages were

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277 simultaneously investigated. Our results show that the geometric mean diameter is the most suitable index to predict fruit mass. A potential ripening scale based on the ratio of R:G:B obtained by quantifying the RGB value 278 279 for a given ripening stage has been proposed. The mechanical properties of tomato fruit, their tissues and their single cells are heterogeneous and anisotropic due to the irregular shape and asymmetric internal structure at the 280 macro-scale, the non-unique tissue thickness at the meso-scale and the irregular change of size, shape and 281 arrangement of single cells at the micro-scale. The hardness and shear strength of fruit at different scales and the 282 283 single cell mechanics varied with the fruit ripening stage but not the chosen cultivars. The contribution of the exocarp to the hardness of whole fruit gradually increased with fruit ripeness. This puncture experimental method 284 is well suited to measure the hardness and shear strength of multi-scale tomato fruit and the mechanics of single 285 tomato cells. The measured multi-scale engineering parameters are extremely important for intelligent harvesting, 286 multi-scale damage simulation and the postharvest textural evaluation of tomato fruits. 287

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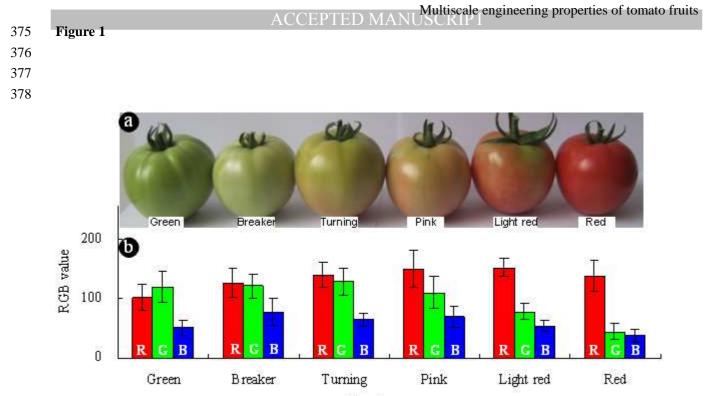
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365 Figure and Table Captions

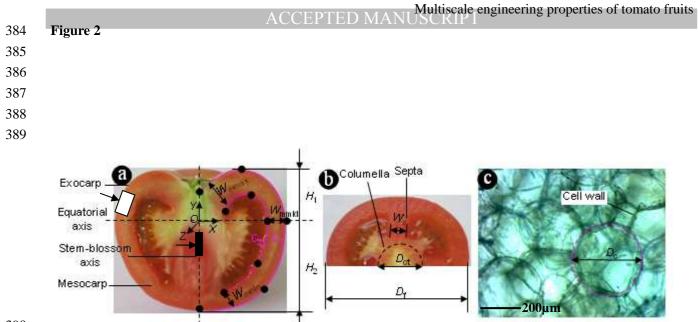
- **Figure 1** Tomato fruits at six ripening stages and their corresponding RGB values.
- **Figure 2** Multi-scale geometrical characteristics of tomato fruits.
- **Figure 3 -** Puncture test of tomato fruits and the calculation of the mechanical parameters.
- **Figure 4** Geometric characteristics of tomato fruits at the macro level.
- **Figure 5** Cell diameters in different types of tissues of two tomato cultivars with microstructure pictures.
- **Table 1** Multi-scale mechanical properties of *Fendu* 79 tomato fruit obtained from compression testing.
- 373 Table 2 Multi-scale mechanical properties of *Fendu* 79 tomato fruit obtained from shear testing



Rip ening stage

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Fig. 1 Tomato fruits at six ripening stages and their corresponding RGB values. (a) Six ripening stages of tomato fruits. (b) RGB values of tomato fruits at the six ripening stages, R-Red, G-Green, B-Blue. Data are expressed as the mean \pm SD (*n*=10).



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Fig. 2 Multi-scale geometrical characteristics of tomato fruits. (a) Longitudinal section of a tomato fruit along the stem-apex axis. The white rectangle shows the sampling zone in exocarp and mesocarp tissue, and the arrow is pointing to the angle of observation. The black rectangle shows the sampling zone in columella tissue, and the arrow shows the angle of observation. (b) Transverse equatorial half-section of a tomato fruit. (c) Cells in the columella tissue of breaker tomato fruit. H_1 -Height above the fruit's equatorial axis, H_2 -Height below the fruit's equatorial axis, D_f -Diameter of the equatorial section, W_{mmax} , W_{mmin} and W_{mmid} . Maximum, minimum and middle thickness of the mesocarp tissue, W_s -Thickness of the septa tissue, D_{ct} -Columella diameter, D_c -Cell diameter.

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400 Figure 3

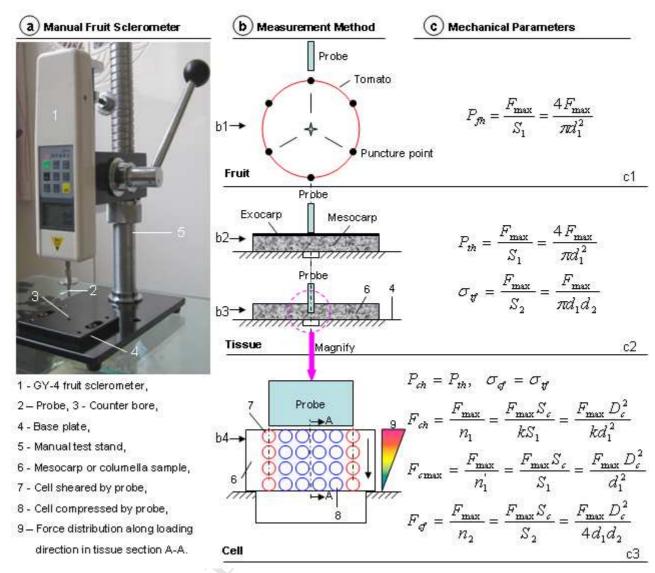
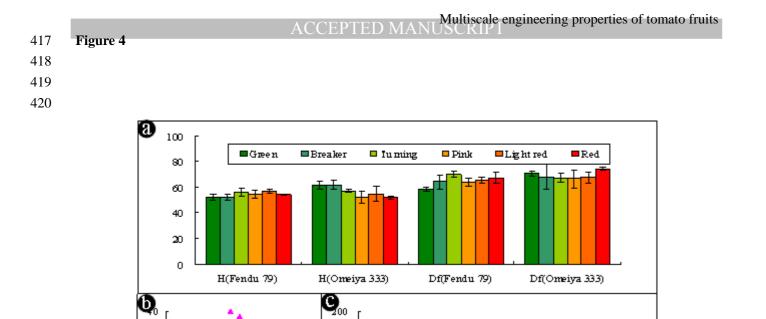


Fig. 3 Puncture test of tomato fruits and the calculation of the mechanical parameters. $P_{\rm fh}$, $P_{\rm th}$ and $P_{\rm ch}$ are the hardness of fruits, tissues and single cells, respectively, subjected to probing; σ_{tf} and σ_{cf} are the shear strength of tissues and single cells subjected to probing; F_{max} is the applied peak puncture force of the probe; F_{ch} is the applied mean compression force to single cells during the puncture test; F_{cmax} is the applied maximum compression force to single cells during the puncture test; F_{cf} is the applied mean shear force to single cell during the puncture test; S_1 and S_2 are the valid compression and shear areas, respectively; D_c and S_c are the diameter and equatorial section areas of a cell; d_1 and d_2 are the probe diameter and tissue sample thickness, respectively; n_1 and n_2 are the numbers of cells compressed by probe and sheared by probe, respectively, and k is the number of cell layers ($k = d_2/D_c$); n'_1 is the number of first-layer cells compressed by the probe.



M=5.49H-166.24 R²=0.64 ▼

M=4.76D+181.34

 $R^2 = 0.58$

80

70

M=736D₈324

 $R^2 = 0.88$

50

60

421

20

0

0

• H1

▲ H2

10

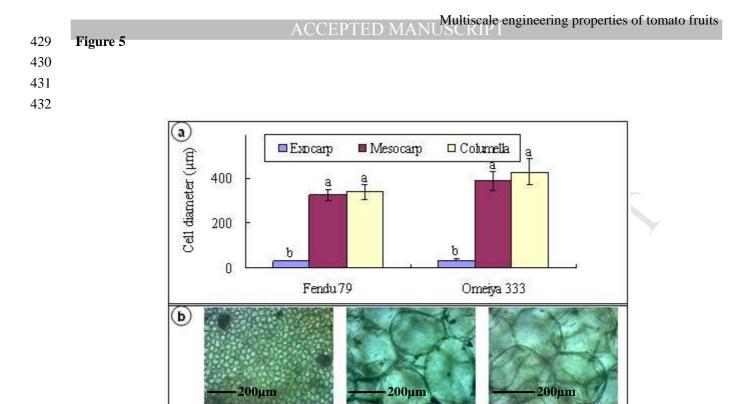
Fig. 4 Geometric characteristics of tomato fruits at the macro level. (a) Heights and Equatorial diameters of tomato fruits at the six ripening stages, ordinate unit: mm. *H*-Fruit height, D_f -Equatorial diameter. Data are expressed as the mean \pm SD (*n*=5). (b) Fruit height above or below the equatorial section, ordinate unit: mm, abscissa unit: number. *H*1-Fruit height above the equatorial section, *H*2-Fruit height below the equatorial section. (c) Regression relationships between *M* and *H*, D_f , D_g , respectively. *M*-fruit mass, D_g -geometric mean diameter, ordinate unit: g, abscissa unit: mm.

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Fig. 5 Cell diameters in different types of tissues of two tomato cultivars and microstructural pictures. (a) Cell diameters of *Fendu* 79 and *Omeiya* 333 tomato fruits. Data are expressed as the mean \pm SD (*n*=5). For each variety, the letters above the error bar indicate significant differences (*P* < 0.05) according to Student's *t*-test. (b)

Mesocarp

Columella

437 Microstructure of exocarp, mesocarp and columella tissues of *Fendu* 79 tomato fruits at the breaker stage

Exocarp

| Λ | 35 | 2 | |
|---|------|---|--|
| 4 | ·.)(| • | |

Table 1

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| M14:1- | Mechanical | | Ripening stage | | | | | |
|--------------------------------|-------------------------|------------|-----------------|-----------------|------------|-----------------|---------------|--|
| Multi-scale | Parameters | Green | Breaker | Turning | Pink | Light red | Red | |
| ANO | VA: | а | ab | abc | abc | bc | с | |
| Multiple-co | omparison | | | | abe | UC | C | |
| Macro-scale (MPa) $P_{\rm fh}$ | Fruit [*] | 2.35±0.20 | 2.02±0.31 | 1.92±0.21 | 1.45±0.10 | 1.20±0.17 | 1.15±0.06 | |
| Meso-/Micro- | Exocarp* | 2.25±0.23 | 1.97 ± 0.07 | 1.87 ± 0.09 | 1.45±0.09 | 1.14±0.11 | 1.12±0.17 | |
| Scale (MPa) | Mesocarp ** | 1.36±0.06 | 1.15±0.35 | 0.80 ± 0.27 | 0.72±0.27 | 0.53 ± 0.06 | 0.40 ± 0.02 | |
| $P_{\rm th}$ and $P_{\rm ch}$ | Columella ^{**} | 1.82±0.14 | 1.04 ± 0.09 | 0.60 ± 0.06 | 0.51±0.22 | 0.41±0.07 | 0.37±0.07 | |
| | Cell in exocarp |)++ | | | | | | |
| | F | $1.808\pm$ | 1.580± | 1.505± | 1.162± | 0.920± | 0.903± | |
| | $F_{\rm cmax}$ | 0.188 | 0.059 | 0.075 | 0.068 | 0.085 | 0.133 | |
| | $F_{\rm ch}$ | $0.340\pm$ | $0.297 \pm$ | 0.283± | 0.219± | $0.173\pm$ | $0.170\pm$ | |
| | | 0.035 | 0.011 | 0.014 | 0.013 | 0.016 | 0.025 | |
| | Cell in mesoca | rp^+ | | | | | | |
| Micro-scale | F | 113.62± | 109.86± | 89.57± | $60.00\pm$ | 33.04± | 32.17± | |
| (mN) | $F_{\rm cmax}$ | 5.24 | 29.02 | 22.61 | 22.88 | 5.21 | 1.81 | |
| | $F_{\rm ch}$ | 8.16±0.97 | 7.76±0.69 | 7.53±0.80 | 4.99±1.92 | 2.50±0.42 | 2.00±0.89 | |
| | Cell in colume | lla^+ | | | | | | |
| | $F_{\rm cmax}$ | 164.78± | 94.65± | $54.40\pm$ | 44.34± | 37.11± | 33.96± | |
| | | 13.08 | 8.51 | 5.69 | 19.66 | 6.42 | 6.80 | |
| | $F_{\rm ch}$ | 11.66±1.28 | 5.42±0.50 | 5.06±0.25 | 3.98±1.11 | 3.50±0.34 | 1.58±0.32 | |

443 Date are expressed as the mean \pm SD (n=6 for macro-scale, n=3 for meso- and micro-scale). Different letters (namely a, ab, abc, bc, c) in the same row indicate significant differences (P < 0.05) according to Student's *t*-test. 444 445 The superscript marks (*) in the macro- and meso-scale column indicate significant differences (P < 0.05), and the 446 superscript marks (+) in the micro-scale column indicate significant differences (P < 0.05).

 $P_{\rm fh}$, $P_{\rm th}$ and $P_{\rm ch}$ are the hardness of fruits, tissues and single cells, respectively, subjected to probing; $F_{\rm ch}$ is the 447 448 applied mean compression force to a single cell during a puncture test; F_{cmax} is the applied maximum compression 449 force to single cell during the puncture test.

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Table 2

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Table 2 Multi-scale mechanical properties of Fendu 79 tomato fruit resulted from shear component

| Multi-scale | scale Biomaterials | Ripening stage | | | | | |
|---------------------------------------|-----------------------------------|------------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Multi-scale | | Green | Breaker | Turning | Pink | Light red | Red |
| ANOVA: Multiple-comparison | | а | ab | abc | abc | bc | с |
| Meso-/Micro- | Exocarp [*] | 11.58 ± 1.20 | 10.12±0.38 | 9.63±0.48 | 7.44 ± 0.44 | 5.89 ± 0.54 | 5.78 ± 0.85 |
| Scale (MPa) | Mesocarp ^{**} | 0.26 ± 0.03 | 0.25 ± 0.02 | 0.24 ± 0.03 | 0.16 ± 0.06 | 0.09 ± 0.01 | 0.06 ± 0.03 |
| $\sigma_{ m tf}$ and $\sigma_{ m cf}$ | Columella ^{**} | 0.33±0.04 | 0.15 ± 0.01 | 0.14 ± 0.01 | 0.11±0.03 | 0.10 ± 0.01 | 0.04 ± 0.01 |
| | Cell in $exocarp^+$ | 9.31±0.97 | 7.82±0.30 | 7.74±0.39 | 5.98 ± 0.35 | 4.73±0.44 | 4.65±0.68 |
| Micro-scale (mN) | Cell in mesocarp ⁺⁺ | 21.82±2.60 | 20.77±1.84 | 20.14±2.15 | 13.34±5.14 | 6.70±1.12 | 5.37±2.39 |
| $F_{ m cf}$ | Cell in columella ⁺⁺ | 29.99±3.30 | 13.94±1.28 | 13.03±0.63 | 10.26±2.86 | 9.01±0.88 | 4.06±0.83 |

456 Date are expressed as the mean \pm SD (*n*=3). Different letters (namely a, ab, abc, bc, c) in the same row indicate

457 significant difference (P < 0.05) according to Student's *t*-test. The superscript marks (*) in the meso-scale column

458 indicate significant differences (P < 0.05), and the superscript marks (+) in the micro-scale column indicate 459 significant differences (P < 0.05).

460 $\sigma_{\rm tf}$ and $\sigma_{\rm cf}$ are the shear strength of tissues and single cells subjected to probing; $F_{\rm cf}$ is the applied mean shear force

to a single cell during the puncture test.

Highlights

- Geometric mean diameter was most closely correlated with tomato fruit mass
- Multi-scale geometry of tomatoes showed heterogeneous and anisotropic properties
- The contribution of exocarp to the hardness of whole fruit increased with ripeness
- Puncture experiment is suited to measure the multi-scale mechanics of fruit

Chillip Marine