

# Quick biochemical markers for assessment of quality control of intraoperative cell salvage : a prospective observational study

Dong, Peng; Che, Ji; Li, Xiuliang; Tian, Ming; Smith, Fang Gao

DOI:

[10.1186/1749-8090-9-86](https://doi.org/10.1186/1749-8090-9-86)

License:

Creative Commons: Attribution (CC BY)

*Document Version*

Publisher's PDF, also known as Version of record

*Citation for published version (Harvard):*

Dong, P, Che, J, Li, X, Tian, M & Smith, FG 2014, 'Quick biochemical markers for assessment of quality control of intraoperative cell salvage : a prospective observational study', *Journal of Cardiothoracic Surgery*, vol. 9, no. 1, 86. <https://doi.org/10.1186/1749-8090-9-86>

[Link to publication on Research at Birmingham portal](#)

**Publisher Rights Statement:**

Eligibility for repository : checked 24/06/2014

**General rights**

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

**Take down policy**

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact [UBIRA@lists.bham.ac.uk](mailto:UBIRA@lists.bham.ac.uk) providing details and we will remove access to the work immediately and investigate.

RESEARCH ARTICLE

Open Access

# Quick biochemical markers for assessment of quality control of intraoperative cell salvage: a prospective observational study

Peng Dong<sup>1</sup>, Ji Che<sup>1</sup>, Xiuliang Li<sup>1</sup>, Ming Tian<sup>1\*</sup> and Fang Gao Smith<sup>2\*</sup>

## Abstract

**Background:** Intraoperative Cell Salvage (ICS), hereby referred to 'mechanical red cell salvage', has been widely used in adult elective major surgeries to reduce requirement for homologous red blood cell transfusion and its associated complications. However, amount of free haemoglobin (fHb) from ICS has been shown related to incidence of renal failure. fHb is the most important indicator of quality control of cell salvaged blood, thus monitoring the fHb concentration is imperative to minimise renal injury. However, currently there has been lacking quick biochemical markers to monitor the levels of fHb during ICS. The aim of this study was to screen quick biochemical markers for evaluating the amount of fHb during use of intraoperative cell salvage.

**Methods:** Twenty patients undergoing elective cardiovascular surgery were enrolled. Blood was collected and processed using a Fresenius continuous auto-transfusion system device. The concentration of fHb, albumin (Alb), and calcium (Ca) in three washing modes were measured, and their clearance rates were calculated. The correlations among the clearances and concentrations of fHb, albumin, and calcium were analysed.

**Results:** In three washing modes, concentrations of albumin and calcium are significantly associated with amount of fHb:  $fHb(g/L) = 0.111Alb(g/L) - 0.108$ ,  $R = 0.638$ ,  $p = 0.000$ ;  $fHb(g/L) = 1.721Ca(mmol/L) + 0.091$ ,  $R = 0.514$ ,  $p = 0.000$ . Furthermore, the clearance rates of albumin and calcium significantly predict clearance of fHb,  $CR_{fHb} = 0.310CR_{ALB} + 0.686$ ,  $R = 0.753$ ,  $p = 0.000$ ,  $CR_{fHb} = 0.073 CR_{Ca} + 0.913$ ,  $R = 0.497$ ,  $p = 0.000$ .

**Conclusions:** In clinic practice, clearance rates of albumin, or calcium can be used to evaluate the quality of salvaged blood, fHb. Bed-side measurement of calcium could offer a more feasible means for clinicians to undertake a real-time assessment of fHb.

**Keywords:** Cell salvage, Clearance rate, Free haemoglobin, Albumin, Calcium

## Background

In the surgeries involving major blood loss, allogeneic red blood cell transfusion is often life-saving. However, allogeneic red blood cell transfusion has been shown to be associated with short-term and long-term complications of infectious diseases, non-infectious serious hazards of transfusion (NISHOTs) [1], or increased mortality [2-4]. This has led to the development of intraoperative cell salvage (ICS) and autologous red blood cell transfusion as important and effective blood conservation methods to

reduce or avoid the need for homologous red blood cell transfusion and its associated complications.

ICS has now been widely used, as a substitution for allogeneic red blood cell transfusion, in adult elective major surgeries [5-12]. Over the years, ICS has been shown to be safe and efficacious to reduce the amount of allogeneic red blood cell transfusion required [13]. Thus, the research focus on ICS has moved to the issue about how to ensure the best quality of salvaged blood to be transfused back to the patient. High concentration of free haemoglobin (fHb) during ICS can result in haemoglobinuria that is associated with increased incidence of renal failure [14]. fHb is the most important indicator for quality cell salvaged blood, thus monitoring

\* Correspondence: 13601167650@139.com; f.g.smith@bham.ac.uk

<sup>1</sup>Department of Anaesthesiology, Beijing Friendship Hospital, Capital Medical University, No. 95, Yong'an Road, Xicheng District, 100050 Beijing, China

<sup>2</sup>Perioperative, Critical Care and Trauma Trials Group, University of Birmingham, Edgbaston B15 2WB, Birmingham, UK

the fHb concentration is imperative to minimise renal injury. However, the measurement of fHb is not a routine laboratory test and also time consuming, which hampers improvement of quality control of cell salvaged blood. Albumin (ALB) and calcium (Ca) mainly exist in the plasma, the destroyed RBC during cell salvage has less impact on concentration of albumin and calcium. The molecular weight of albumin and calcium, the same as fHb, are lighter than erythrocyte, they should be eliminated together with the fHb during the procedure of centrifuging, so the clearance rates of these two should be expected to be similar with fHb. Therefore, the aim of this study was to assess whether quick biochemical markers, albumin and calcium, that can be used clinically to reflect the clearance of fHb for evaluating the quality control of salvaged blood.

## Methods

### Study participants

The institutional Ethics Committee of the Beijing Friendship Hospital approved the study proposal. Written informed consent was obtained from all participants. Adult patients undergoing elective cardiovascular surgeries with an estimated intraoperative blood loss of more than 800 ml were enrolled in this study. Exclusion criteria included the patients with haematopathy, malignancy, a contaminated operative field, or requirement for allogenic red blood cell transfusion during current surgery.

### Cell salvage procedure

As we previously reported [15], Fresenius Continuous Auto Transfusion System device (CATS, Germany) was used for this study. The standard operative procedures were performed according to the manufacturer's recommended protocols. Trained executive operators performed the blood salvage process. Intraoperative shed blood was collected through a special suction tube for blood collection. The vacuum pressure of the suction was maintained below 0.02 MPa to minimise haemolysis by suction. Blood was heparinized with physiological saline (30000 IU heparin in 1000 mL of 0.9% saline). The initial speed of the anticoagulant was 60 drops/min, which was continuously delivered to the tip of the suction catheter. The aspirated blood was filtered and stored in the blood reservoir. When enough blood was collected, the cell-washing device was operated according to the manufacturer's recommended parameters. As the blood enters the processing chamber, the chamber is spinning at a speed of 1400–2400 RPM to generate centrifugal force. Separation of salvaged blood components mainly depends on the balance between densities of the various constituents of blood. Because red cells are heavier than other blood components, they will stay against the walls of the chamber while the smaller, lighter

particles will be closer to the core of the chamber. In the following washing phase, RBCs are resuspended with 0.9% saline and blood plasma is further removed. At last, the saline is removed and the RBCs are concentrated and emptied into a reinfusion bag, and when appropriate, reinfused into the patient. CATS designed several washing programmes for different operations. Big flow is recommended for the operation with massive blood loss in a short time, standard mode is for the operation with normal blood loss speed, and high quality mode is for the shed blood with more debris.

### Blood sample collection

Three phases are involved in the cell salvage. During phase 1, blood is collected from the operating field to the reservoir using heparinised physical saline as the anticoagulant. During phase 2, three modes: big flux mode, standard mode and high quality mode, were used to separate, wash and concentrate the cells to reinfusion bags. In phase 3, concentrated washed red cells are then reinfused into the patient.

The blood samples were collected from the following time points and sites. The first time blood samples (3 ml) were obtained from venous blood. The second blood samples (10 ml) obtained from the reservoir following the blood in the reservoir was well mixed during phase 1. The third blood samples (5 ml ×3) were collected from the reinfusion bag after each mode was completed during phase 2. The volume of shed blood and the volume of reinfusion bag of each washing mode were recorded for calculating clearance rate. All blood samples were analysed within 20 min after collection.

### Analysis of blood samples

Free haemoglobin concentrations were measured using a HemoCue Plasma Low/Hb Photometer (HemoCue, Inc., Angelholm, Sweden). Haemoglobin (Hb) concentrations were measured using a HemoCueHb 201+ Analyser (Kuvettgatan 1, Angelholm, Sweden). Albumin concentrations were measured using HITACHI 7600 automatic biochemical analyser (Japan), and the concentration of calcium and haematocrit (Hct) were measured using GEM Premier 3000 blood gas machine (USA). The internal quality control of HITACHI 7600 automatic biochemical analyser was done with the standard samples

**Table 1 Concentration of fHb, clearance rate of fHb and percentage of haemolysis of washed blood**

n = 20	Washed blood
Concentration of fHb	0.40 ± 0.27 g/L
Clearance rate of fHb	97% ± 2%
Percentage of haemolysis	0.21% ± 0.11%

**Table 2 The results of the univariate regression analyses between fHb (g/L) and albumin (ALB, g/L)**

Washing mode	Regression analysis between fHb and albumin	r	p
Big flow	fHb = 0.118ALB - 0.166	0.780	0.000
Standard mode	fHb = 0.068ALB - 0.082	0.659	0.002
High quality mode	fHb = 0.175ALB - 0.366	0.663	0.001
Combined three modes	fHb = 0.111ALB - 0.108	0.638	0.000

from Wako Pure Chemical Industries every morning, and external quality control was done with the samples provided by National Centre for Clinical Laboratories every four months. The quality control of GEM Premier 3000 blood gas machine was done with the standard samples from Instrumentation Laboratory.

**Calculation of clearance rate**

For the calculation of clearance rate, the concentrations of fHb, albumin and calcium in the reservoir and reinfusion bag, the volumes of shed blood in the reservoir for different washing modes, the corresponding volumes of washed blood in the reinfusion bag, the concentrations of Hb in the reinfusion bags of different washing modes were measured and recorded.

The efficacy of collection, centrifugation, and washing was assessed by the clearance rate (CR) of fHb, albumin

and calcium, which were calculated using the following equations

$$\text{Clearance Rate (\%)} = \left( 1 - \frac{\text{Solute}_{\text{after}} \times \text{Volume}_{\text{after}}}{\text{Solute}_{\text{before}} \times \text{Volume}_{\text{before}}} \right) \times 100\%$$

Where  $\text{solute}_{\text{after}}$  was the concentration of fHb, albumin or calcium of the washed blood in the reinfusion bag,  $\text{volume}_{\text{after}}$  was the volume of washed blood in the reinfusion bag,  $\text{solute}_{\text{before}}$  was the concentration of fHb, albumin or calcium of blood collected in the reservoir, and  $\text{volume}_{\text{before}}$  was the volume of blood collected in the reservoir.

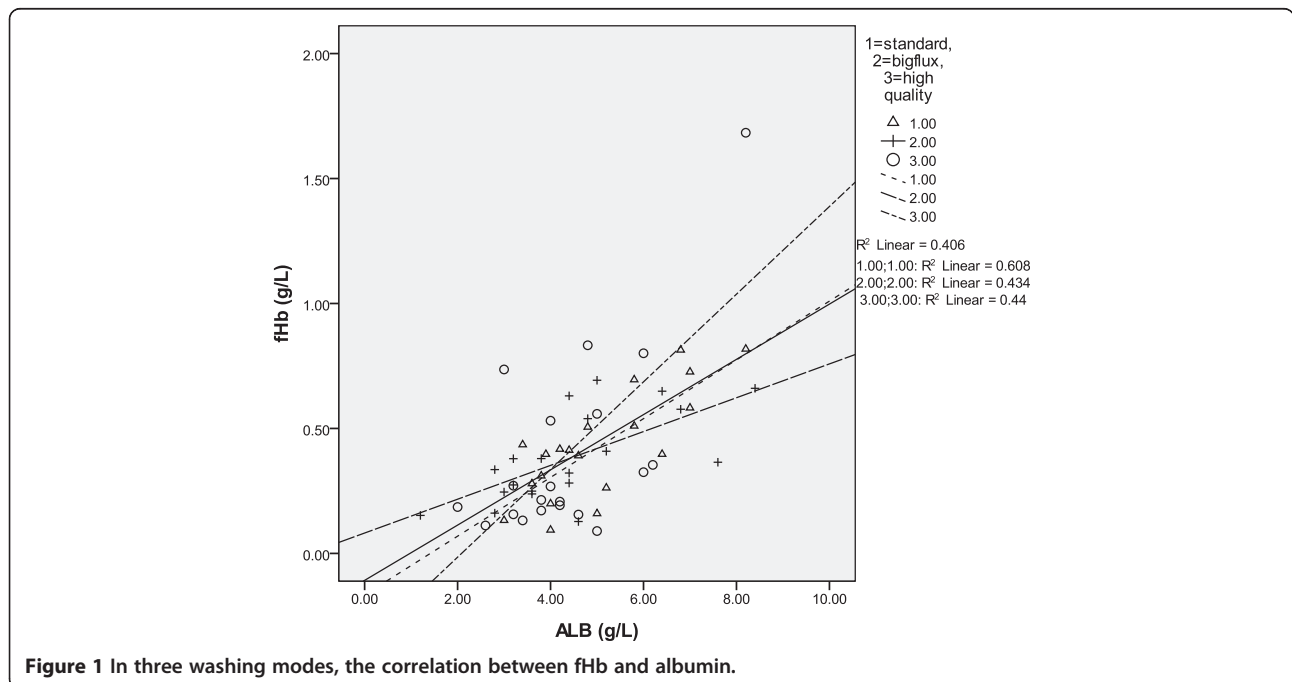
The percentage of haemolysis was calculated using the following formula [14]:

$$\begin{aligned} \text{Percentage of haemolysis(\%)} \\ = [(\text{fHb}/\text{Hb}) \times (1-\text{Hct})] \times 100\% \end{aligned}$$

Where fHb, Hb and Hct were the concentrations of fHb, Hb and haematocrit (Hct) in the reinfusion bag.

**Statistical analysis**

All statistical analyses were performed using SPSS 13.0 software (SPSS, Inc., Chicago, IL, USA). Descriptive statistics were calculated on all measures to determine the characteristics of the sample, to check normality assumptions and to ensure adequate variability. All results are presented as mean ± standard deviation (SD) or



**Table 3 The results of the univariate regression analyses between fHb(g/L) and calcium(Ca, mmol/L)**

Washing mode	Regression analysis between fHb and Ca	r	p
Big flow	fHb = 1.608Ca + 0.122	0.564	0.010
standard mode	fHb = 0.967Ca + 0.211	0.469	0.037
High quality mode	fHb = 2.676Ca - 0.075	0.571	0.011
Combined three modes	fHb = 1.721Ca + 0.091	0.514	0.000

incidence (n,%). Univariate linear regression analysis was applied to quantify the association of the clearance rates of fHb, albumin and calcium. A P-value of 0.05 was considered to be statistically significant.

**Results**

From March to November 2012, 20 patients (13 males) undergoing off-pump Coronary Artery Bypass Grafting surgeries (n = 13) and vascular surgeries with an American Society of Anaesthesiology physical status II-III were enrolled to the study. The mean age of the patients was 60.0 ± 14.0 years old (49–81 years old). The mean pre-operative concentration of albumin was 38.3 ± 3.1 g/L (31.0-42.4 g/L), and the mean of calcium was 2.20 ± 0.09 mmol/L (2.09-2.44 mmol/L), which were in the normal range. The concentration of albumin and calcium were 18.4 ± 5.1 g/L (9.8-31.4 g/L) and 0.61 ± 0.17 mmol/L (0.33-0.99 mmol/L) in the reservoirs, 4.6 ± 1.6 g/L (1.2-8.4 g/L) and 0.19 ± 0.08 mmol/L (0.10-0.37 mmol/L) in the reinfusion bags. After the procedure of cell salvage, the concentration of fHb, clearance rate of fHb and the percentage of haemolysis see Table 1.

**Correlation between concentrations fHb and albumin**

In each of these three modes, (big flux, standard and high quality modes) there existed positive correlation between

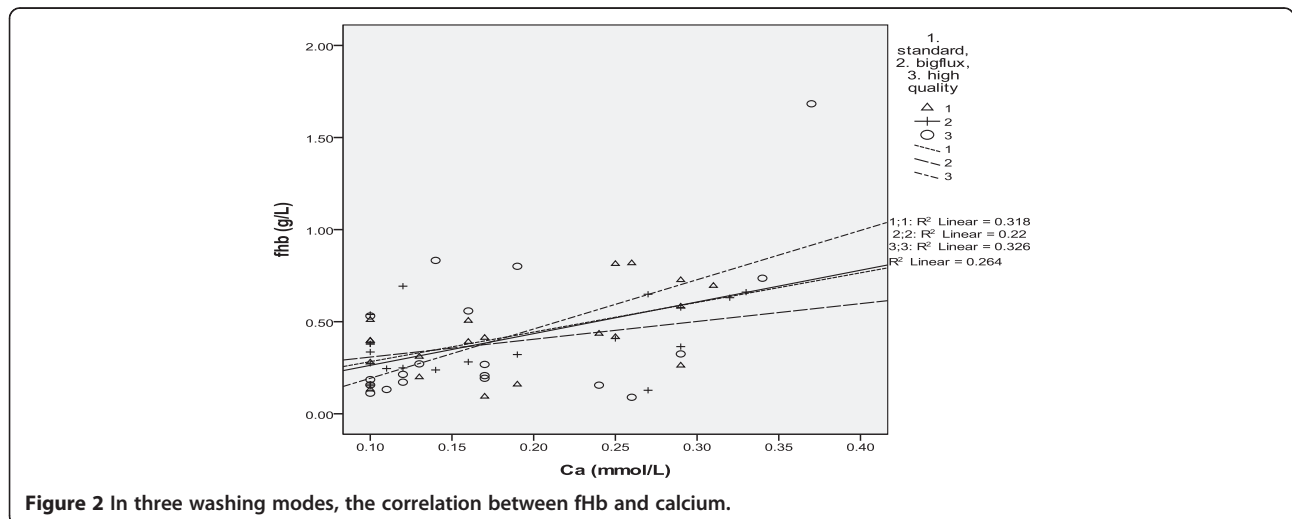
the concentration of albumin and that of fHb in the reinfusion bag. In the mode of big flow, fHb(g/L) = 0.118ALB(g/L) - 0.166, r = 0.780, p = 0.000; in the standard mode, fHb(g/L) = 0.068ALB(g/L) - 0.082, r = 0.659, p = 0.002; in the high quality mode, fHb(g/L) = 0.175ALB(g/L) - 0.366, r = 0.663, p = 0.001. In combined three washing modes, the correlation between fHb and albumin in the reinfusing bag was fHb (g/L) = 0.111ALB(g/L) - 0.108, r = 0.638, p = 0.000 (see Table 2, Figure 1).

**Correlation between concentrations fHb and calcium**

In three modes, there existed correlation between the concentration of calcium and that of fHb in the reinfusion bag. In the mode of big flow, fHb(g/L) = 1.608Ca (mmol/L) + 0.122, r = 0.564, p = 0.010; in the standard mode, fHb(g/L) = 0.967Ca(mmol/L) + 0.211, r = 0.469, p = 0.037; in the high quality mode, fHb(g/L) = 2.676Ca (mmol/L) - 0.075, r = 0.571, p = 0.011. In combined three washing modes, the correlation between fHb and calcium in the reinfusing bag was fHb(g/L) = 1.721Ca(mmol/L) + 0.091, r = 0.514, p = 0.000 (see Table 3, Figure 2).

**Regression relationship between clearance of fHb and clearance of albumin**

In the three modes of washing, there existed univariate regression relationship between the clearance rate of fHb(CR<sub>fHb</sub>) and that of albumin(CR<sub>ALB</sub>). In the mode of big flow, CR<sub>fHb</sub> = 0.295CR<sub>ALB</sub> + 0.699, r = 0.638, p = 0.008; In the standard mode, CR<sub>fHb</sub> = 0.324CR<sub>ALB</sub> + 0.675, r = 0.896, p = 0.000; In the high quality mode, CR<sub>fHb</sub> = 0.275CR<sub>ALB</sub> + 0.719, r = 0.540, p = 0.017. In combined three washing modes, the regression relationship of clearance of fHb and clearance of albumin in the reinfusing bag was CR<sub>fHb</sub> = 0.310CR<sub>ALB</sub> + 0.686, r = 0.753, p = 0.000 (See Table 4, Figure 3).



**Figure 2 In three washing modes, the correlation between fHb and calcium.**

**Table 4 The results of the univariate regression analyses between CR<sub>fHb</sub> and CR<sub>ALB</sub>**

Washing mode	Regression analysis between CR <sub>fHb</sub> and CR <sub>ALB</sub>	r	p
Big flow	CR <sub>fHb</sub> = 0.295CR <sub>ALB</sub> + 0.699	0.638	0.008
Standard mode	CR <sub>fHb</sub> = 0.324CR <sub>ALB</sub> + 0.675	0.896	0.000
High quality mode	CR <sub>fHb</sub> = 0.275CR <sub>ALB</sub> + 0.719	0.540	0.017
Combined three modes	CR <sub>fHb</sub> = 0.310CR <sub>ALB</sub> + 0.686	0.753	0.000

**Regression relationship between clearance of fHb and clearance of calcium**

Under three modes of washing, there was linear regression relationship between the clearance rate of fHb and that of calcium (CR<sub>Ca</sub>). In the mode of big flow, CR<sub>fHb</sub> = 0.261 CR<sub>Ca</sub> + 0.734, r = 0.746, p = 0.000; In the standard mode, CR<sub>fHb</sub> = 0.111CR<sub>Ca</sub> + 0.894, r = 0.760, p = 0.000; In the high quality mode, CR<sub>fHb</sub> = -0.100CR<sub>Ca</sub> + 0.997, r = 0.592, p = 0.008. In combined three washing modes, the regression relationship between clearance of fHb and clearance of calcium was CR<sub>fHb</sub> = 0.073 CR<sub>Ca</sub> + 0.913, r = 0.497, p = 0.000 (see Table 5, Figure 4).

**Discussion**

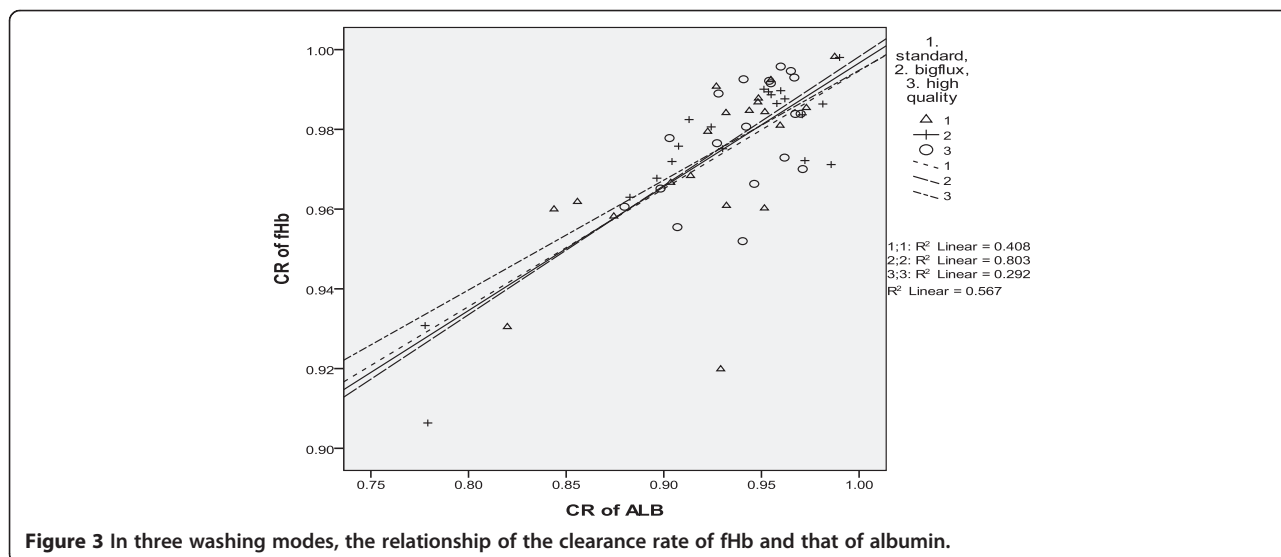
Thus far, there has been no information on the quality standards of fHb for both mechanical red cell salvaged blood from autologous red blood cell transfusion and for stored blood from allogeneic red blood cell transfusion. The quality standard level of haemolysis rate has been referred to stored blood < 1% in the USA or < 0.8% in the Europe [16] as there has been no such information available for salvaged blood. Most studies regard salvaged blood with more than 90% clearance rate of fHb as good quality control [12,15]. Our study results, produced under good quality control of intraoperative cell

salvage (0.21% haemolysis rates and 97% clearance rate of fHb), should be generalizable.

During the process of cell salvage, red blood cells (RBC) may be destroyed by the suction with negative pressure and centrifugation. These destroyed RBC will increase fHb. When the fHb level in plasma is > 1 g/L, the metabolic load may exceed the tolerance in adults, and methaemoglobin may form to cause organ injury. In particular, circulating fHb may precipitate within renal tubules, which leads to haemoglobinuria nephropathy and tubular dysfunction [14]. However, fHb > 4 g/L frequently occurs in washed RBC following severe haemolysis. Thus, it is crucial to monitor fHb concentrations to assess salvaged blood quality in order to prevent patients from renal and other organ injury.

However, in clinical settings, measurements of concentrations of fHb have to be undertaken in a specialised haematological laboratory, which usually takes more than two hours before the results become available. This is useless for clinicians to be able to manipulate the process of cell salvage to improve the quality of cell salvaged blood before reinfusion. Bed-side fHb analyser is currently available to be able to assay the concentration of fHb accurately in 60 seconds. But the instrument is too expensive to be applied widely.

The basic mechanism of autotransfusion system device is to eliminate the components that the molecular weight lighter than that of erythrocyte through the centrifuging. The molecular weight of albumin and fHb are 66000 Dalton and 67000 Dalton respectively, lighter than erythrocyte. Molecules of Alb and fHb should stay at the same layer and be eliminated together during the procedure of centrifuging, so the clearance rates of these two should be expected to be similar. Our study suggests there is a linear relationship between the concentrations



**Figure 3** In three washing modes, the relationship of the clearance rate of fHb and that of albumin.

**Table 5 The results of the univariate regression analyses between CR<sub>fHb</sub> and CR<sub>Ca</sub>**

Washing mode	Regression analysis between CR <sub>fHb</sub> and CR <sub>Ca</sub>	r	p
Big flow	CR <sub>fHb</sub> = 0.261 CR <sub>Ca</sub> + 0.734	0.746	0.000
Standard mode	CR <sub>fHb</sub> = 0.111 CR <sub>Ca</sub> + 0.894	0.760	0.000
High quality mode	CR <sub>fHb</sub> = -0.100 CR <sub>Ca</sub> + 0.997	0.592	0.008
Combined three modes	CR <sub>fHb</sub> = 0.073 CR <sub>Ca</sub> + 0.913	0.497	0.000

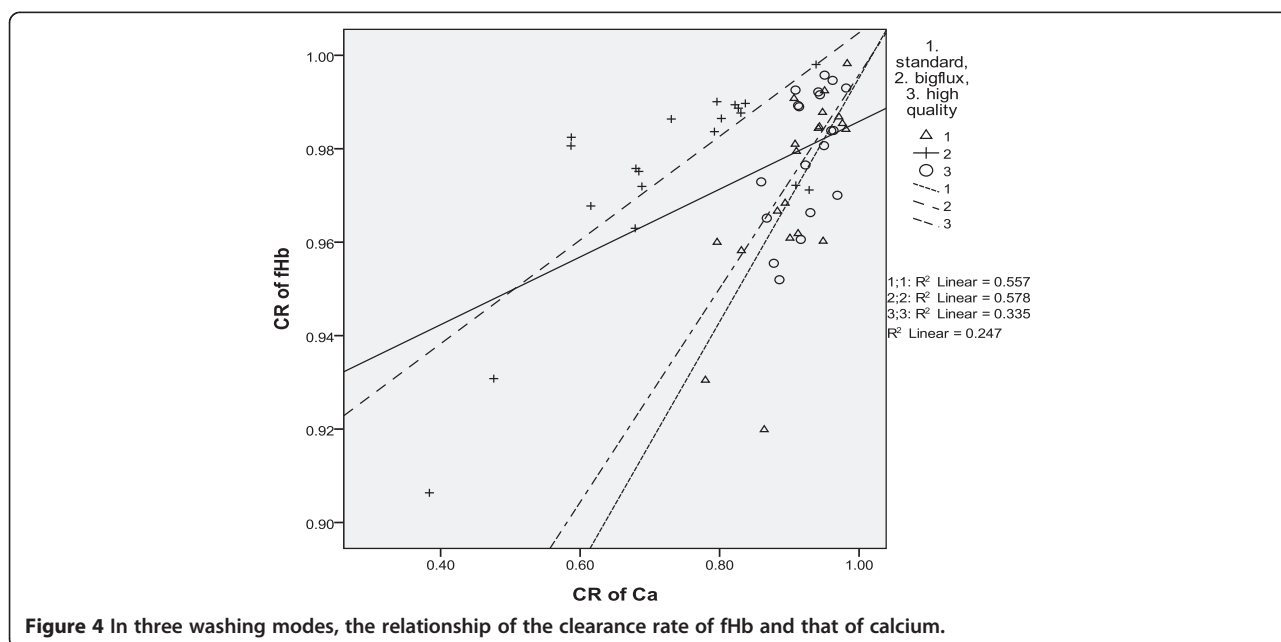
of fHb and albumin. Therefore, the regression equation will allow us to calculate the concentration of fHb from the concentrations of albumin. Also, clearance rate of fHb is positively correlated with clearance of albumin and their correlation coefficient *r* 0.638, 0.896 and 0.540 for three washing modes (big flux, standard or high quality) respectively. This association again has offered clinicians a feasible mean to learn the clearance rate of fHb from the clearance rate of albumin. However, like lactate dehydrogenase that was strongly correlated to fHb [17], assays of albumin concentrations can only be undertaken in a biochemical laboratory although this is easier and less expensive than assays of fHb, the measurements have to be one-off or intermittent without a real time monitoring using a blood gas machine.

Potassium assay was suggested as a biochemical marker for evaluating the clearance of fHb, but the elimination of fHb during the washing process was not correlated with the residual concentration of potassium [17,18]. This finding may be explained by the fact that intracellular compartment of the red blood cells contains very high

concentration of potassium and so degree of increase in fHb concentrations was not proportional to degree of increase in potassium concentrations when the RBC were destroyed. In a similar way, the elimination of fHb was not correlated to elimination of potassium. In contrast, calcium mainly exists in the plasma, the destroyed RBC during cell salvage has less impact on concentration of calcium. The molecular weight of calcium is lighter than the erythrocyte, so calcium, like fHb, is eliminated by centrifugation. Our study has proven that the clearance rate of fHb can be predicted by the known clearance of calcium, the latter can be calculated if the concentrations of calcium in the reservoir and reinfusion bag are measured. In the clinic practice, the concentrations of calcium can be measured in several minutes using a bed-side blood gas machine.

Either heparin or citrate can be used for anticoagulation during red blood cell salvage. There existed controversy about which anticoagulant is best. Also heparin anticoagulation increased the fHb level compared to citrate, but the haemolysis rate was still in normal range [19]. Because of its low price and ready availability, heparin is most commonly used [20].

Our study has limitations, firstly, the fHb concentrations of patients' blood were not tested before and after the transfusion of cell salvaged red blood cells; secondly, the study has a small sample size in cardiovascular surgery only. These findings are promising and further studies with larger sample size in a number of types of surgeries are needed to confirm these outcomes and to establish whether these clinical biochemical markers will offer utility in this regard.



## Conclusion

In summary, during the use of Continuous Auto Transfusion System device for cardiac and vascular surgeries, the concentration and clearance rate of albumin, and calcium can be employed as useful biochemical markers to evaluate the quality control of cell salvaged blood, fHb, in the washing modes of big flux, standard and high quality. Because the concentration of calcium can be assayed quickly using a blood gas machine in the operation room, bed-side measurement of calcium could offer a more feasible means for clinicians to undertake a real-time assessment of fHb.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

MT had the concept, PD designed and set up the study with supervision from MT and FGS. PD, JC and XL obtained the consents, collected all the samples and undertook the measurements. PD completed analysing the data with statistical support from a statistician and drafted the manuscript. FGS was chiefly responsible for the acquisition and interpretation of data and for finalising the manuscript. MT, FGS and PD also were involved in critical review of the manuscript. PD completed revision after submission. All authors read and approved the final manuscript.

## Acknowledgement

Thanks to Yan Zhang, Associate Professor in Medical Statistics, Shanghai Institute of Planned Parenthood Research, for the statistical advice. Thanks to all the surgeons, anaesthetists, nursing staff and technicians for their help with data collection and sample analysis. Thanks to all the patients who willingly donated blood samples and contributed the data.

## Funding

The project were supported by Beijing Medicine Research and Development Foundation(No.2007-1039), Beijing Natural Science Foundation(No.7122050) and Beijing Friendship Hospital Science Foundation (No.yyqdk2011-25).

Received: 19 March 2014 Accepted: 6 May 2014

Published: 14 May 2014

## References

1. Hendrickson JE, Hillyer CD: Noninfectious serious hazards of transfusion. *Anesth Analg* 2009, **108**:759–769.
2. Rao SV, Jollis JG, Harrington RA, Granger CB, Newby LK, Armstrong PW, Moliterno DJ, Lindblad L, Pieper K, Topol EJ, Stamler JS, Califf RM: Relationship of bloodtransfusion and clinical outcomes in patients with acute coronary syndromes. *JAMA* 2004, **292**:1555–1562.
3. Tinmouth A, Fergusson D, Yee IC, Hébert PC, ABLE Investigators; Canadian Critical Care Trials Group: Clinical consequences of red cell storage in the critically ill. *Transfusion* 2006, **46**:2014–2027.
4. Engoren M, Mitchell E, Perring P, Sferra J: The effect of erythrocyte blood transfusions on survival after surgery for hip fracture. *Trauma* 2008, **65**:1411–1415.
5. Wang G, Bainbridge D, Martin J, Cheng D: The efficacy of an intraoperative cell saver during cardiac surgery: a meta-analysis of randomized trials. *Anesth Analg* 2009, **109**:320–330.
6. Takagi H, Sekino S, Kato T, Matsuno Y, Umemoto T: Intraoperative autotransfusion in abdominal aortic aneurysm surgery: meta-analysis of randomized controlled trials. *Arch Surg* 2007, **142**:1098–1101.
7. Bridgens JP, Evans CR, Dobson PM, Hamer AJ: Intraoperative red blood cellsalvage in revision hip surgery. A case-matched study. *J Bone Joint Surg Am* 2007, **89**:270–275.
8. Waters JH, Dyga RM, Waters JF, Yazer MH: The volume of returned red blood cells in a large blood salvage program: where does it all go? *Transfusion* 2011, **51**:2126–2132.

9. Sinclair KC, Clarke HD, Noble BN: Blood management in total knee arthroplasty: a comparison of techniques. *Orthopedics* 2009, **32**:19.
10. Thomas D, Wareham K, Cohen D, Hutchings H: Autologous blood transfusion in total knee replacement surgery. *Br J Anaesth* 2001, **86**:669–673.
11. Phillips SD, Maguire D, Deshpande R, Muiresan P, Bowles MJ, Rela M, Heaton ND: A prospective study investigating the cost effectiveness of intraoperative blood salvage during liver transplantation. *Transplantation* 2006, **81**:536–540.
12. Liang H, Zhao Y, Wang D, Wang B: Evaluation of the quality of processed blood salvaged during craniotomy. *Surg Neurol* 2009, **71**:74–80.
13. Carless PA, Henry DA, Moxey AJ, O'Connell D, Brown T, Fergusson DA: Cell salvage for minimising perioperative allogeneic blood transfusion. *Cochrane Database Syst Rev* 2010, **17**:CD001888.
14. Toshiharu Y: Haptoglobin therapy for possible prevention of renal failure following thermal injury: a clinical study. *J Trauma* 1985, **25**:281–287.
15. Che J, Tian M, Ding G, Huai Q, Dong P, Li Y, Li S: Effects of cell salvage on erythrocyte 2,3-disphosphoglycerate and G-6-PD levels and phosphatidylserine expression. *Int J Lab Hematol* 2013, **35**:385–392.
16. Sowemimo-Coker SO: Red blood cell hemolysis during processing. *Trans Med Rev* 2002, **16**:46–60.
17. Sullivan IJ, Faulds JN: Lactate dehydrogenase and haemolysis index as quality control markers of haemolysis in intra-operative cell salvage. *Transfus Med* 2013, doi:10.1111/tme.12065. [Epub ahead of print].
18. Szpisjak D, Edgell D, Bissonnette B: Potassium as a surrogate marker of debris in cell-salvaged blood. *Anesth Analg* 2000, **91**:40–43.
19. Oller DW, Rice CL, Herman CM, Cochran RC, Homer LD, John DA, Kingsly JR, Fouty WJ: Heparin versus citrate anticoagulation in autotransfusion. *J Surg Res* 1976, **20**:333–340.
20. Waters JH: Intraoperative Blood Recovery. *ASAIO J* 2013, **59**:11–17.

doi:10.1186/1749-8090-9-86

Cite this article as: Dong et al.: Quick biochemical markers for assessment of quality control of intraoperative cell salvage: a prospective observational study. *Journal of Cardiothoracic Surgery* 2014 **9**:86.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

