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1 Organic coating reduces hygroscopic growth of phase
2 separated aerosol particles

3

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13

14 **Abstract**

15 A large fraction of secondary aerosol particles is liquid-liquid phase separation with
16 organic shell and inorganic core. This has the potential to regulate the hygroscopicity
17 of such particles, with significant implications for their optical properties, reactivity,
18 and lifetime. However, it is unclear how this phase separation affects the hygroscopic
19 growth of the particles. Here, we showed a large variation in hygroscopic growth (e.g.,
20 1.14 -1.32 under RH 90%) of particles from forest and urban atmosphere, which had
21 different average core-shell ratios. For this reason, a controlled laboratory experiment
22 further quantify the impact of organic shell on particle growth with different RHs.
23 Laboratory experiments demonstrated that $(\text{NH}_4)_2\text{SO}_4$ particles with thicker secondary
24 organic shells have a lower growth factor at RH below 94%. Organic shells started to
25 deliquesce first (RH>50%) and the phase changes of sulfate cores from solid to liquid
26 took place at a RH larger than 80% as deliquescence relative humidity (DRH) of pure
27 $(\text{NH}_4)_2\text{SO}_4$. Our study provides the first direct evidence on an individual particle basis
28 that hygroscopic growth behavior of phase-separated particles is dependent on the
29 thickness of organic shells, highlighting the importance of organic coating in water
30 uptake and possible heterogeneous reactions of the phase-separated particles.

31

32 **Keywords:** Hygroscopicity, liquid-liquid phase separation, aerosol, particle phase

33

34 **Synopsis:** The study significantly extends our understanding of liquid-liquid phase-
35 separated particles, how organic shell influences water uptake in the air, and constrains

36 hygroscopic growth of inorganic core.

37 **Introduction**

38 Atmospheric particles can directly or indirectly interact with sunlight and change
39 the energy balance of Earth's atmosphere, with the water uptake process greatly altering
40 the optical properties and the ability to act as cloud condensation nuclei (CCN) of these
41 aerosol particles, thus influencing their climate properties.¹⁻³ In addition, the water
42 uptake also influences the reactivity and lifetime of ambient particles, thereby affecting
43 air quality and human health.^{4,5} For instance, condensed water provides a medium for
44 the multiphase chemical reactions to happen within the particles^{6,7}, which leads to
45 changes in the gas uptake efficiency (e.g., N₂O₅ hydrolysis),⁸⁻¹⁰ metal solubility,^{11,12}
46 and the formation of secondary aerosol particles.^{1,13} Hygroscopicity is a key parameter
47 to characterize the water uptake ability of aerosol particles.^{2,14,15}

48 In the past decade, various advanced technologies have been used to investigate
49 the hygroscopicity of aerosol particles collected both from the laboratory and the field
50 campaign.¹⁶ The hygroscopic tandem differential mobility analyzer (HTDMA) is one
51 of the most commonly used techniques to investigate hygroscopic growth of aerosol
52 particles under various environments.¹⁷⁻¹⁹ Although the HTDMA significantly
53 enhanced our understanding in the hygroscopicity of ambient aerosol particles, it
54 provides an average growth factor (GF) of size-resolved aerosol populations.¹ Thus, it
55 is unable to reveal the heterogeneity of the hygroscopicity within an aerosol population.
56 Such heterogeneity often significantly impacts certain atmospheric processes, including
57 the CCN activation of ambient particles.² To address this issue, a single particle
58 technique has been used for hygroscopicity studies to understand the mixing state and

59 hygroscopicity of individual particles.^{14, 20-24} These results^{14, 20-24} show that individual
60 ambient aerosol particles often exhibit as a mixture of inorganic sulfate, nitrate, and
61 organic species of different ratios following the particle aging processes through
62 different environments in the atmosphere. The large differences in the hygroscopicity
63 of sulfate, nitrate, and organics,^{14, 25} together with the composition differences of each
64 individual particle, will likely lead to a wide range of growth factors of aerosol
65 particles^{26, 27} that needs to be considered in models for the formation, optical properties,
66 and CCN activity of aerosol particles.^{1, 2, 26}

67 Recently, the liquid-liquid phase separation of inorganic and organic components
68 within a single particle has been examined as such effect can affect the water and gas
69 uptake of the particles.^{15, 28-30} Previous studies^{20, 25, 28, 31, 32} confirmed that the core-shell
70 phase-separation morphology widely exhibited in ambient particles through
71 measurements performed by the transmission and scanning electron microscopy
72 (TEM/SEM). However, the current studies analyzed the hygroscopicity of bulk
73 collected samples under the typical air pollution events without examining the effects
74 of phase separations.^{19, 33, 34} Recent laboratory research on liquid-liquid phase
75 separation showed that organic materials occurring as shells on the inorganic core of
76 the particles are shown to have different impacts on early deliquescence of particle
77 surface^{15, 35, 36} as well as water and gas uptake (e.g., N₂O₅).^{8, 9, 35} However, there is a
78 lack of systematic study on the effects of organic shell (also called as organic coating)
79 and inorganic core on hygroscopicity under different RH conditions. For instance,
80 online instruments such as an aerosol time-of-flight mass spectrometer (ATOFMS) in

81 tandem to a HTDMA could observe mixing state and hygroscopicity of individual
82 particles without providing the evidence of liquid-liquid phase separation.^{1, 18, 23} Other
83 studies on single particles often used a mixture of organic surrogates and inorganic
84 species without further examining the effects of phase separations.^{27, 28, 37-40} In addition,
85 some studies reported the hygroscopic properties of the laboratory generated surrogate
86 particles which are not consistent with the core-shell structure of the ambient particles.^{27,}
87 ³⁸ These results highlight the necessity to systematically examine how phase separation
88 alters the hygroscopic properties of individual particles under different RHs using a
89 single particle technique.

90 The aim of this study is to understand hygroscopic properties of liquid-liquid
91 phase-separated particles. To achieve this aim, we first investigated the core-shell ratios
92 and hygroscopic growth in airborne particles collected in different environmental air.
93 We then found that the coating thickness might be a key factor affecting hygroscopic
94 growth. To obtain direct evidence, we built a photochemical flow tube to produce
95 organic aerosols to generate the internally mixed particles with ammonium sulfate core
96 and organic shell. The thickness of the organic shells was controlled by changing the
97 reaction time. Then an individual particle hygroscopic (IPH) system is constructed to
98 continuously observe the hygroscopic growth processes of the phase-separated particles
99 during 3-94% RH.

100 **2. Experiments**

101 **2.1 Field campaigns**

102 Aerosol particle samples were collected at three locations representing different

103 atmospheric environments: a forest site in the Lesser Khingan Mountains of
104 northeastern China, urban Beijing of northern China, and urban Hangzhou of eastern
105 China. Here we selected six typical samples which commonly include the core-shell
106 particles for hygroscopic experiments. Table S1 shows meteorological data and
107 sampling period of each sample. Aerosol samples are collected both onto copper TEM
108 grids coated with carbon film and silicon wafers using a DKL-2 single particle sampler
109 with a single-stage cascade impactor equipped with a 0.3 mm diameter jet nozzle at a
110 flow of 1.0 liters per minute (lpm). The cutoff size of the impactor (50% collection
111 efficiency) is 0.1 μm (aerodynamic diameter, assuming a material density of 2 g cm^{-3}).
112 The collection time varied between 2-20 min, depending on the number concentrations
113 of aerosol particles. After the aerosol collection, the samples were placed in sealed, dry,
114 plastic capsules to prevent contamination. Then the samples were stored in a desiccator
115 at 20 °C and $20 \pm 3\%$ RH for subsequent analysis.

116 **2.2 TEM analysis**

117 Imaging of the particles with cryo-TEM was conducted on a Talos F200C equipped
118 with a Ceta 4k \times 4k camera at the Center of Cryo-Electron Microscopy, Zhejiang
119 University (Hangzhou, China), which is operated at 200 kV accelerating voltage.
120 Samples were cooled in the cryo-TEM to approximately 104 K using a liquid nitrogen
121 cooling cryo-TEM holder before exposure to electron radiation to reduce damage to the
122 particles. The cryo-TEM is one efficient way to observe the original state of individual
123 secondary particles on the substrate because the secondary nitrates or sulfates receive
124 minimum damage from the electron radiation when the images are taken. To further

125 confirm composition and mixing state of individual ambient particles collected on TEM
126 grids, a JEOL JEM-2100 transmission electron microscope operated at 200 kV with
127 energy-dispersive X-ray spectrometry (TEM/EDX) was conducted. The scanning TEM
128 (STEM) function was adopted to obtain line scanning of elemental composition on
129 individual particles which can directly reflect different aerosol components to form the
130 core-shell structure. Finally, these microscopic observations are used to explain
131 hygroscopic behaviors of individual aerosol particles.

132 **2.3 Hygroscopic experiment**

133 In this study, an individual particle hygroscopicity (IPH) system was built to
134 investigate the hygroscopic properties of individual particles at different RHs. The
135 experimental process is composed of three steps: (1) Introducing N₂ gas with controlled
136 flow by a mass flow controller into a chamber; (2) Mounting the TEM grid or silicon
137 wafer with particles on the bottom of an environmental microscopic cell (Gen-RH
138 Mcell, UK), which can change RH while maintaining a constant temperature of 20 ± 1
139 °C; (3) Taking images at different RH through an optical microscope (Olympus BX51M,
140 Japan) with a camera (Canon 650D). A similar experimental set-up is described
141 elsewhere.^{26, 41} (4) The equivalent circle diameter of each individual particle in images
142 were analyzed using image analysis software (Radius, EMSIS GmbH, Germany). We
143 manually obtained sizes of the same particle at different RHs and further calculated its
144 GF.

145 The pure (NH₄)₂SO₄ particles with the size range of 0.5-20 μm was made in
146 laboratory and collected on silicon wafer substrates and TEM grids. Hygroscopic

147 behaviors of these $(\text{NH}_4)_2\text{SO}_4$ particles were used to test the IPH system at different
148 RHs from 3% to 94%. The deliquescence relative humidity (DRH) of all particles were
149 observed to be 80 ± 2 %, which is consistent with the theoretical DRH of $(\text{NH}_4)_2\text{SO}_4$
150 (Figure S2). Following the control study, nine laboratory-generated samples and six
151 samples collected in the ambient air were chosen to observe particle hygroscopic
152 growth in the IPH system. The images taken by the IPH system were used to compare
153 their morphology and size at different RH values (e.g., Figure S2). Particle GF from
154 this study is calculated using Eqn. 1 below.

$$155 \quad GF(RH) = \frac{D(RH)}{D_0} \quad (1)$$

156 Where $D(RH)$ is the diameter of particles at a given RH, D_0 is the diameter of dry
157 particles at 5% RH. The GF curve of individual particles can be used to understand
158 changes of particle phase and size growth under different RHs.²⁶ Finally, we tried to
159 select 5-8 core-shell particles of each sample in the optical images and calculated their
160 average GF.

161 **2.4 Laboratory experiment**

162 Individual $(\text{NH}_4)_2\text{SO}_4$ particles on silicon wafers and TEM grids are prepared
163 following the procedure reported by Sun et al. (2018).²⁶ After leaving the $(\text{NH}_4)_2\text{SO}_4$
164 sample in an enclosed container for two hours to allow equilibrium with the
165 environment, the prepared samples are placed at the outflow tip of the flow tube. Figure
166 S1 shows the flow tube including three main parts: the gas inlet, the reaction tube, and
167 the outlet. The similar flow tube was tested and widely used to study secondary aerosol
168 formation in the laboratory experiment.⁴² In this study, 0.1 lpm of air was used to

169 vaporize liquid α -pinene while 1.5 lpm of pure nitrogen and 1.5 lpm O_3 were mixed
170 within a 2-liter glass tube with UV-B lights on to react and produce the secondary
171 organic materials that can be coated onto the existing $(NH_4)_2SO_4$ particles. The organic
172 materials from oxidizing α -pinene are observed to form a layer of film on the substrate
173 as the control experiment. All the samples contain individual particles with organic shell
174 and sulfate core on the substrate after the 11h, 15h, and 20h of secondary organic
175 materials exposure in the flow tube, which is used to produce organic shell with
176 different thickness in different periods. Upon the flow tube experiments, the samples
177 were conditioned at three hygroscopic cycles ramping from 3-94% RH in the IPH.
178 During the hygroscopic growth of individual particles, the organic matter surrounding
179 the inorganic sulfate core always maintained its stable organic shell on individual
180 particles. In total, three parallel experiments of each with six silicon wafers samples are
181 collected for hygroscopic experiments and nine TEM grids are collected for chemical
182 analysis.

183 **3 Results and Discussion**

184 **3.1 Phase separation of organic and sulfate in secondary aerosols**

185 Cryo-TEM and TEM observations both revealed a common core-shell mixing
186 structure (i.e., liquid-liquid phase separation) in the aerosol particles collected in
187 different environmental conditions, as shown in Figures 1 and S3. EDS shows that the
188 core part mainly contains S, O, N elements and the shell part is dominant by the C and
189 minor O elements (Figure S3). Figure 1a-b also clearly shows various intact secondary
190 organic-inorganic phase separated particles that were collected from ambient

191 environments. The line scanning in STEM shows that the core exhibits high
192 concentration of O and S and the shell exhibits more C. Based on the distribution of
193 elemental composition and mixing state of individual particles, we can conclude that
194 the cores of these ambient particles are dominated by inorganic sulfates/nitrates and the
195 shells mainly contain secondary organic matter, agreeing with the liquid-liquid phase
196 separation reported in previous studies.^{15, 20, 25}

197 In this study, we found that over 59% of secondary inorganic aerosols have the
198 organic shells in background forest particles and 36-45% in the urban particles in
199 summer (Table S1). Recently, Yu et al., (2019)³¹ revealed that 74% of secondary
200 inorganic aerosols have thick organic shell in remote Arctic air. These results suggest
201 that a large portion of the secondary aerosol particles is phase-separated under ambient
202 conditions. This phase separation has the potential to impact water uptake of inorganic
203 cores at selected RH levels and thus influencing the heterogeneous reactions and CCN
204 activities of individual particles.^{2, 8, 19}

205 **3.2 Hygroscopic behavior of ambient phase-separated particles**

206 Figure 2a lists the optical image of ambient particles collected from ambient
207 environment, showing that organic shells in individual secondary particles all start to
208 grow up at RH=50% in all the samples, whether from the forest or urban air. As the RH
209 increases, the particles collected in forest air grow continuously and then are converted
210 into liquid phase when the RH reaches 84% (Figure S4). For particles collected under
211 the urban environment, the continuous growth of the inorganic cores and organic shells
212 become liquid phase at 83% RH (Figure 2a and Figures S5-6). We noticed that the urban

213 aerosol particles have more abrupt hygroscopic growth above RH=60% compared with
214 the forest aerosol particles. Given that Sun et al., (2018)²⁶ found that phase of secondary
215 aerosol particles start changes at $60 \pm 1\%$ RH due to the presence of ammonium nitrate
216 and the wide presence of inorganic nitrate in urban aerosols,²⁰ it is likely that these
217 analyzed particles also contain inorganic nitrate. The presence of ammonium nitrate in
218 individual particles might slightly enhance GF of individual urban particles at 60-80%
219 RH, as shown in Figure 2a.

220 Figure 2a shows the average GFs of aerosol particles at RH=90%: ~1.14 in forest
221 environment, 1.21-1.32 in urban environments, indicating the GF of secondary aerosol
222 particles collected in forest environment is much lower than those in the urban
223 environment. Moreover, the GFs of urban aerosol particles are much lower than that of
224 the pure ammonium nitrate and ammonium sulfate particles (1.61-1.75) measured by
225 the IPH techniques.²⁶ We noticed that the average thicknesses of organic shells in the
226 core-shell particles collected in Beijing, Hangzhou, and lesser Khingan mountain air
227 were 60 nm, 86 nm, and 110 nm, respectively (Table S1). Therefore, we hypothesize
228 that the organic shells contribute to the lowering of GFs among different liquid-liquid
229 phase separation particles.

230 This hypothesis cannot be tested in airborne particles because it is not feasible to
231 characterize the same individual particles with both IPH and TEM, which provide
232 growth and shell thickness separately. For this reason, a controlled laboratory
233 experiment was carried out to systematically quantify the impact of organic shell on
234 particle growth with different RHs (see below).

235 3.3 Hygroscopic effects of laboratory generated phase-separated organic- 236 inorganic particles

237 To achieve the above aim, we prepared laboratory generated liquid-liquid phase
238 separation particles with different thickness of organic shells through a flow tube
239 (Figure S1). TEM observation confirmed that the laboratory generated particles clearly
240 displayed a typical organic shell and sulfate core structure (Figure 3). Figure 4 shows
241 similar shell thickness among different particles in the optical images of laboratory
242 generated particles (11 hours aging) exposed to RH ranging from 3-94%. Organic shell
243 started to uptake water and increased its thickness at 50% RH but never fully
244 deliquesced while the sulfate cores did not grow until 81% RH. The core then gradually
245 changed from solid to liquid phase (Figure 2b) at 83%, which is different and higher
246 than the abrupt phase change at 81% for the pure $(\text{NH}_4)_2\text{SO}_4$. This indicates that the
247 hygroscopic growth of sulfate cores was inhibited by the secondary organic shells.

248 Two additional experiments were carried out to investigate whether the thickness
249 of the organic shell may affect the hygroscopic growth behavior of ammonium sulfate.
250 Figures 2b (green line) and S7 show that phase separated particles (15 h of exposure in
251 the flow tube) started to change at 50% and sulfate cores started to grow at 81% but did
252 not completely deliquesce until 84.5% RH. Figures 2b and S8 show that phase separated
253 particles (21 h of exposure in the flow tube) started to change at 50% RH and sulfate
254 cores grew at 86% RH but did not completely deliquesce until at 90% RH. In addition,
255 the small error bars show that the GFs among different particles have similar values
256 (Figure 2b), suggesting that the laboratory-generated particles display similar

257 hygroscopic properties in the same sample. Therefore, we can conclude that the
258 increasing thickness of the organic shells (Figures 2b and S7-8) raised the DRH of the
259 sulfate cores.

260 The optical images show that individual particles clearly demonstrated different
261 deliquescence changes regarding the core and shell, contrary to the results reported in
262 previous bulk-measured studies that observed changes of the whole particles as a
263 function of RHs.^{1, 27, 43} This difference could be explained by the inherent inaccuracy
264 in the aerosol population, which does not account for the thickness of organic shell and
265 mixing state, e.g., using the HTDMA.

266 We repeated the three RH cycles for each sample at the range of 3-94% RH and
267 observed the same GFs each time, while the GFs of individual particles decrease as the
268 time of exposure increases in the flow tube. For instance, Figure 2b shows that
269 individual particles exposed for 11, 15, and 21 hrs in the flow tube displayed GFs at 1.5,
270 1.4, and 1.2 under the RH=90%, respectively, suggesting that thicker organic shells
271 suppress the GF of the individual particles. The GF values of the phase separated
272 particles varied between the 1.07-1.67,⁴⁴ with the upper value equal to the GF of the
273 pure (NH₄)₂SO₄ (e.g., 1.67 at RH=85% shown in Figure 2b). Moreover, the inorganic
274 sulfate cores show a slower growth curve as the shell thickness of the organic shells
275 increase (Figure 2b), when comparing with an abrupt phase change of the pure
276 (NH₄)₂SO₄ at DRH=81%. Nevertheless, Figures 2 and 4 show that hygroscopic growth
277 is mainly associated with the deliquescence of the sulfate cores while the organic shell
278 has limited influence. The above result indicates that secondary organic shells

279 significantly affect the deliquescence and growth of the inorganic particles. The organic
280 shell suppresses the water uptake of sulfate cores, that is, limiting the water diffusion
281 to the particle core.

282 Once the particles deliquesced following water uptake, they might keep water in
283 the particles until 43% RH during the dehydration. The reason is that sulfate cores in
284 the phase-separated particles effloresced from liquid to solid phase until the RH down
285 to 41-43%, which is lower than the efflorescence relative humidity at 48% of the pure
286 ammonium sulfate (Figures S9-S10). In other words, the organic shells can limit water
287 evaporation from the sulfate cores.

288 In summary, our study confirmed the hypothesis that the thickness of the organic
289 shells contributes to the lowering of GFs among different liquid-liquid phase separation
290 particles. We recognize that chemical composition of the coating also has the potential
291 to affect the hygroscopic growth. This should be investigated further in future field and
292 laboratory experiments.

293 **3.4 Summary and atmospheric implications**

294 The synthetic experiment demonstrated that organic shells in the ambient liquid-
295 liquid phase separation particles, commonly present in forest and urban environments,
296 act as barriers for the water vapor exchange to the inorganic cores. This also indicates
297 that organic coating reduces the mass accommodation coefficient for gases (e.g., N_2O_5)
298 at surfaces¹⁰, leading to a reduction of the formation of secondary organic aerosols.⁴⁵
299 The presence of organic species at the particle surface also influences the CCN
300 activation by altering the surface tension and the water activity, leading to changes in

301 the equilibrium vapor pressure of water over these solution droplets.^{2, 29, 46, 47}

302 Previous studies also showed that organic shells can be semi-solid with high
303 viscosity,^{45, 48, 49} leading to slower heterogeneous reaction rates. The very small growth
304 factor of the laboratory particles at 50-80% (Figure 2) implies that the organic shells
305 are semi-solid (T=295 K), consistent with reported viscosity of secondary organic
306 aerosols in field and laboratory experiments.⁵⁰⁻⁵³ Once the organic shell becomes
307 aqueous phase at above 50% RH, the wet organic shell may further influence the
308 formation of sulfate and nitrate in the aqueous phase (aqSOA) from photo-oxidation.^{54,}
309 ⁵⁵ Information regarding phase-separated particles should be incorporated into the
310 atmospheric modeling for heterogeneous chemistry as well as particle hygroscopicity
311 and growth to improve predictions on air quality and indirect effects of aerosol-climate
312 interactions.

313 **ASSOCIATED CONTENT**

314 **Supporting information**

315 Detailed description of hygroscopic growth and data analysis (Table S1 and Figure S1-
316 S8)

317

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339 **Notes**

340 The authors declare no competing financial interest.

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346 All the data are presented in the paper.

347

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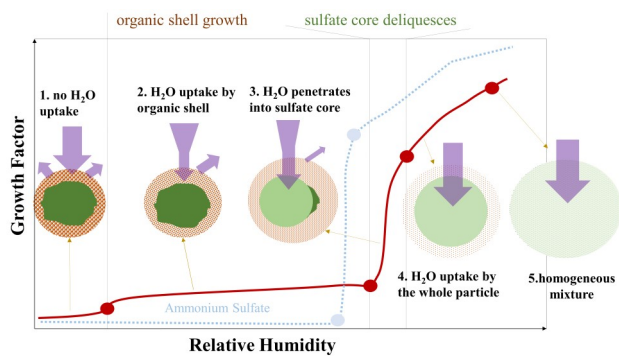
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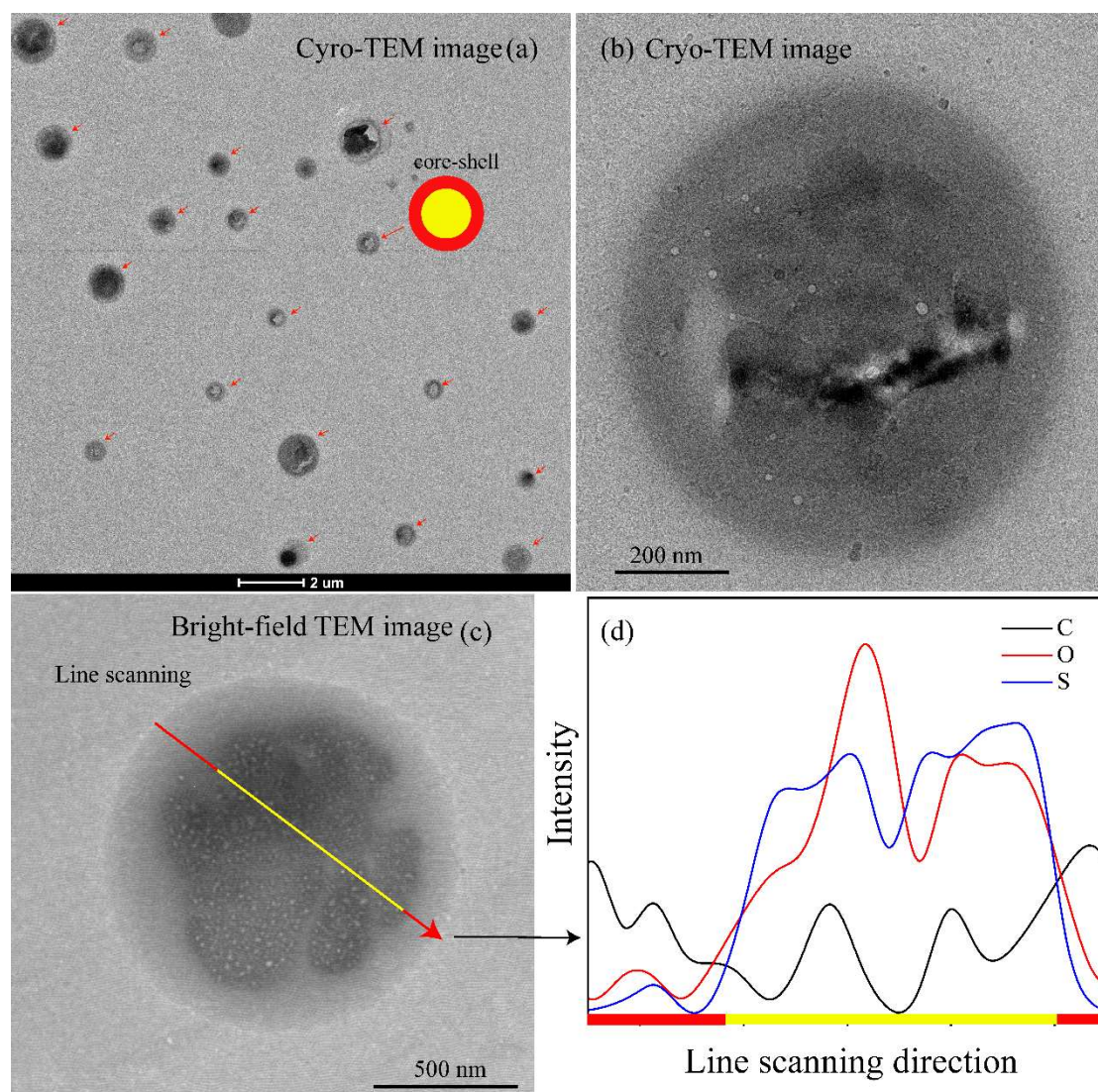
Abstract Graph



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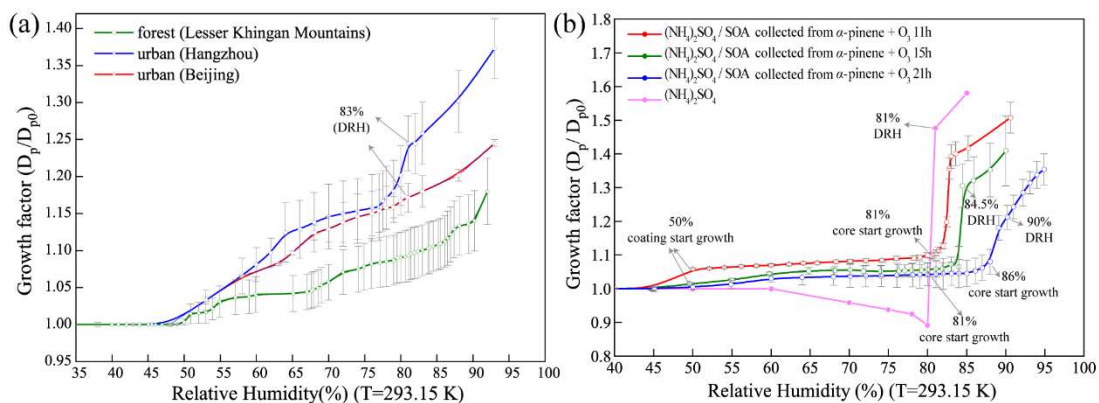
525 **Figure 1** Morphology and composition of secondary particles in the atmosphere. (a)

526 Low magnification cryo-TEM image showing the core-shell structure (i.e., liquid-liquid

527 phase separation) of secondary particles collected at Lesser Khingan Mountain site; (b)

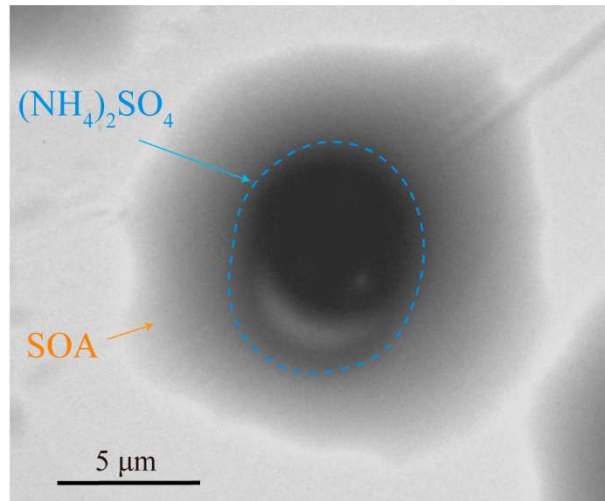
528 High magnification cryo-TEM image showing shell and core; (c-d) A core shell particle

529 and its line scanning in STEM showing C, O, S variations.



530

531 **Figure 2** Deliquesces and growth factors of ambient particles collected in different
 532 environment air (a) and coated $(NH_4)_2SO_4$ particles (after exposing in the
 533 photochemical chamber for 11h, 15h, and 21h). The data in Figs. 2a and 2b were
 534 obtained from the optical images of the size growth of individual particles under
 535 different RHs in Figures 4, S2, and S3.

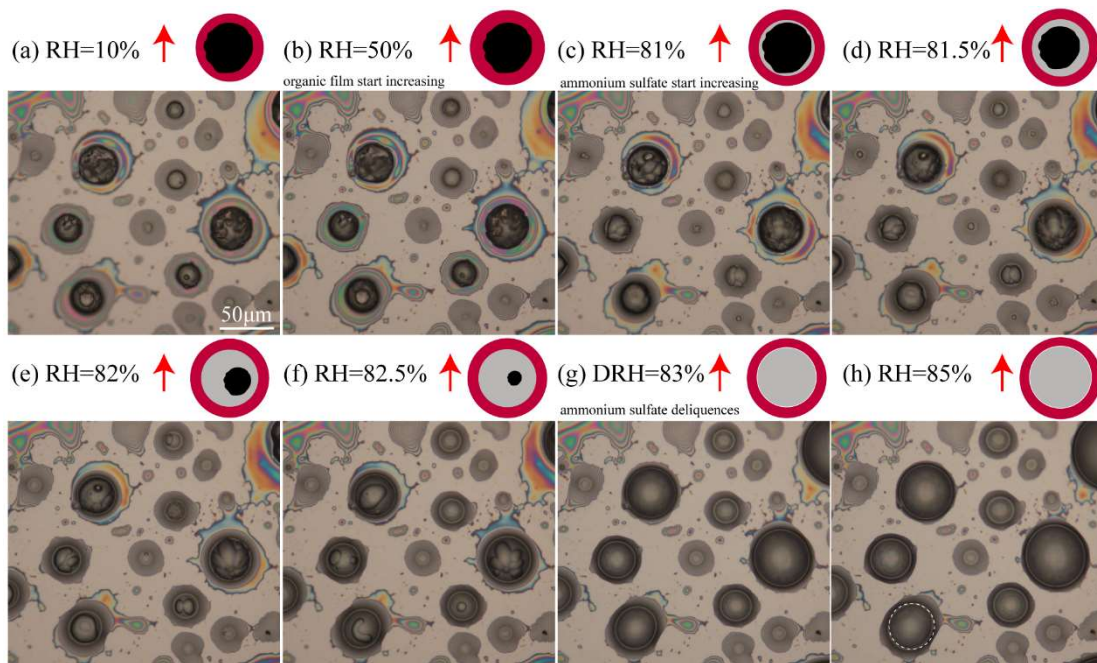


536

537 **Figure 3** TEM image showing the core-shell shape (i.e., liquid-liquid phase separation)

538 of an individual particle generated from the laboratory flow tube after 21 hrs. The core

539 is the ammonium sulfate and the shell is secondary organic aerosol (SOA).



540

541 **Figure 4** Optical images showing hygroscopic growth during hydration from 5% to 94%

542 RH of individual particles (a-h). The ammonium sulfate particles were coated with

543 organic shell in the flow tube after 11 hours of exposure under different RH at T=293.15

544 k. Up arrow (↑) represents the RH increase process.