

# Effect of surface roughness and temperature on stainless steel - whey protein interfacial interactions under pasteurisation conditions

Avila-Sierra, Alejandro; Zhang, Jason; Fryer, Peter

DOI:

[10.1016/j.jfoodeng.2021.110542](https://doi.org/10.1016/j.jfoodeng.2021.110542)

[10.1016/j.jfoodeng.2021.110542](https://doi.org/10.1016/j.jfoodeng.2021.110542)

License:

Creative Commons: Attribution-NonCommercial-NoDerivs (CC BY-NC-ND)

*Document Version*

Peer reviewed version

*Citation for published version (Harvard):*

Avila-Sierra, A, Zhang, J & Fryer, P 2021, 'Effect of surface roughness and temperature on stainless steel - whey protein interfacial interactions under pasteurisation conditions', *Journal of Food Engineering*, vol. 301, 110542. <https://doi.org/10.1016/j.jfoodeng.2021.110542>, <https://doi.org/10.1016/j.jfoodeng.2021.110542>

[Link to publication on Research at Birmingham portal](#)

## General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

## Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact [UBIRA@lists.bham.ac.uk](mailto:UBIRA@lists.bham.ac.uk) providing details and we will remove access to the work immediately and investigate.

2 **Title**

3 **Effect of surface roughness and temperature on stainless steel - whey protein interfacial**  
4 **interactions under pasteurisation conditions**

5 **Authors**

6 Alejandro Ávila-Sierra, Zhenyu Jason Zhang, and Peter J. Fryer\*

7 **Address**

8 School of Chemical Engineering, University of Birmingham, Edgbaston, Birmingham B15  
9 2TT, United Kingdom

10 **Corresponding authors**

11 Peter J. Fryer *E-mail address:* p.j.fryer@bham.ac.uk

12 **Keywords**

13 Milk fouling, surface free energy, temperature, roughness, adhesion, polarity.

14 **Acknowledgements**

15 The research team acknowledges the School of Chemical Engineering, University of  
16 Birmingham, for financial support and provision of a studentship. We are also grateful for  
17 funding from the CSEF project that is funded by EPSRC (EP/K011820/1). ZJZ thanks EPSRC  
18 for financial support (EP/P007864/1).

## 20 **Abstract**

21 The effects of the surface characteristics of 316L stainless steel (SS316L), including chemical  
22 composition before and after foulant deposition, surface roughness, and wall temperature, on  
23 both the liquid-solid and the solid-solid interfacial interactions have been investigated using  
24 contact angle measurements and atomic force microscopy respectively. Wettability of the metal  
25 surface was favoured by increased surface roughness (in the range-limited for food contact  
26 applications) and wall temperature (within the temperature range used for pasteurisation). A  
27 fine surface finish (i.e. mirror) could be an effective intervention to reduce liquid adhesion and  
28 the subsequent foulant deposition, especially under thermal treatment. The surface free energy  
29 (SFE) of SS316L and its polar and disperse components remained constant from ambient to  
30 pasteurisation temperatures ( $< 80^{\circ}\text{C}$ ). However, as fouling develops, the surface free energy  
31 evolved: upon foulant deposition, SFE decreased. An increased polarity (3.4% from 25 to  
32  $80^{\circ}\text{C}$ ) of the fouled surface could be related to the exposure the hydrophobic core of reversibly  
33 adsorbed  $\beta$ -Lactoglobulin toward the foulant-air interface. Both surface adhesion and Young's  
34 modulus at sub-micron spatial resolution confirmed that the packing within the foulant and  
35 molecular orientation on the foulant surface were affected by the temperature of the underlying  
36 substrate. Temperature also affected the wetting behaviour of cleaning solutions on surface  
37 foulant; as the surface temperature increased from  $25^{\circ}\text{C}$  to  $75^{\circ}\text{C}$ , the contact angle on WPC  
38 increased, suggesting an enhanced surface hydrophobicity. Overall, this work highlights the  
39 importance of surface parameters on governing the interfacial interactions that are competing  
40 for the control of the complex fouling phenomena.

41

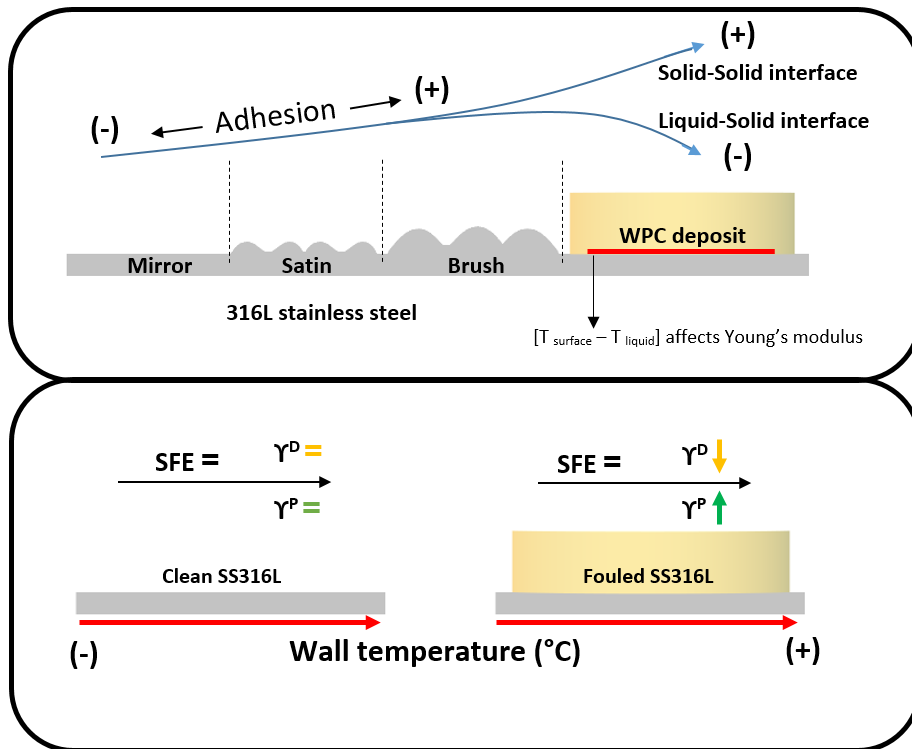
42

43 **Highlights**

- 44 • Surface free energy and polarity of stainless steel remains constant over the range of
- 45 operational temperatures (25 ~ 80°C).
- 46 • Surface free energy decreases upon foulant deposition.
- 47 • Polarity of the model proteinaceous foulant increases as temperature increases.
- 48 • Foulant hydrophobicity increases as the wall temperature increases.
- 49 • Liquid-surface temperature difference affects the mechanical characteristics of the deposit
- 50 formed.

51

52 **Graphical Abstract**



53

54

## 55 **1. Introduction**

56 In the dairy industry, surface fouling within pasteurisation equipment results in serious  
57 challenges for maintaining the performance of processing lines. Extensive cleaning operations  
58 are often required, which has a significant impact on the total production cost (Van Asselt et  
59 al., 2005). The cost to address issues related to heat-exchanger fouling for industrialised  
60 countries was estimated as 0.25% of the country gross national product (GNP) (Garrett-Price  
61 et al., 1985). This financial cost, alongside other issues such as product contamination,  
62 environmental impact, and industrial sustainability, emphasises the urgent need to understand  
63 fouling.

64 Food manufacture commonly involves equipment made of stainless steel, amongst which  
65 austenitic 304 and 316L are mostly used due to their chemical neutrality and physical durability  
66 (Schmidt et al., 2012). A range of studies have been carried out to understand and fabricate  
67 hierarchically structured surfaces, with much less attention paid to realistic engineering  
68 surfaces (Kubiak et al., 2011). The surfaces involved in a manufacturing process, including the  
69 welded joints, would be polished to meet the hygienic criteria of the installation, but even  
70 though surface roughness is well defined for food applications ( $R_a \leq 0.80 \mu\text{m}$ ) (Frantsen and  
71 Mathiesen, 2009), subtle variations could considerably affect interfacial interactions, especially  
72 under processing conditions.

73 Fouling results from interactions between the products being processed and the surfaces in  
74 contact with them, whose characteristics govern deposition and the magnitude of interfacial  
75 adhesion. At the macroscopic scale, interfacial adhesion is commonly related to surface  
76 wettability, the ability of a liquid to wet a solid surface, and contact angle measurements are  
77 used to predict the amount of foulant deposited (Handojo et al., 2009; Williams et al., 2005).  
78 The wettability of a solid substrate is determined by the balance between cohesive forces (Work

79 of cohesion:  $W_c$ ) and adhesive forces of the liquid on a solid surface (Work of adhesion:  $W_a$ )  
80 (Choi et al., 2002): if  $W_a > W_c$ , the liquid spreads over the surface, and vice versa. However,  
81 surface wettability can be altered by surface characteristics such as topography (Avila-Sierra  
82 et al., 2019; Zhang et al., 2015) and temperature gradients (Karapetsas et al., 2017).-Kubiak et  
83 al. (2011) investigated a broad spectrum of surfaces, including metallic, ceramic and polymeric  
84 ones, reporting that there was a minimum contact angle below  $Ra < 1 \mu m$  associated with the  
85 droplet spreading along the polishing grooves.

86 Contact angle measurements can be also used to quantify surface free energy (SFE), viewed as  
87 a critical fouling precursor. The SFE of a given substrate offers a direct measure of the  
88 intermolecular interactions at the interface (Zhao et al., 2004), and strongly influences the  
89 adsorption/adhesion behaviour of compounds (such as proteins (Boxler et al., 2013;  
90 Rosmaninho and Melo, 2008), cells and bacteria (Baier, 1980; Tsibouklis et al., 2000), starches  
91 (Białopiotrowicz, 2003) and minerals (Boxler et al., 2013; Rosmaninho and Melo, 2008,  
92 2006a)). A correlation was established between the electron-donor component of the substrate  
93 and the final amount of deposit formed (Rosmaninho and Melo, 2006b), where surfaces with  
94 low energy less favourable for binding (Rosmaninho and Melo, 2006b; Tsibouklis et al., 2000;  
95 Zhao et al., 2007). The weaker binding at the interface, the easier the cleaning process (Akeso  
96 et al., 2009). Baier et al. (1985) demonstrated the importance of both temperature and SFE on  
97 bacterial adhesion at 37°C. However, very few SFE studies have studied common engineering  
98 surfaces, particularly at working temperatures. Zhao et al. (2004) measured the surface free  
99 energy of 304 stainless steel, alongside some other amorphous carbon surfaces, from 20 to  
100 95°C, and reported that there were significant SFE variations when the testing temperature was  
101 above 80°C. In addition to determining the effects that surface free energy and temperature  
102 might have on surface fouling, it is critical to understand the influence that other physical (e.g.

103 polishing process) and chemical transformations (e.g. foulant deposition) on the metal surface  
104 might have on the interfacial interactions involved amid food processing.

105 During milk thermal treatment,  $\beta$ -Lactoglobulin ( $\beta$ -Lg) unfolds and exposes its hydrophobic  
106 core containing reactive disulphide and sulfhydryl bonds (Claeys et al., 2001) that can react  
107 rapidly with the processing equipment and other bulk fluid compounds (Bansal and Chen,  
108 2006). At the nanoscale, atomic force microscopy (AFM) can determine the force/work of  
109 adhesion between a wide variety of surfaces (e.g. stainless steel, ceramic, mineral, glass and  
110 poly(tetrafluoroethylene)) (Akhtar, 2010; Navabpour et al., 2010; Sauerer et al., 2016; Verran  
111 et al., 2000) and has been used to study food confectionary foulants such as Turkish delight,  
112 caramel and sweetened condensed milk (Akhtar et al., 2010). These works highlighted  
113 differences between adhesion forces as a function of surface type, while others showed that  
114 adhesion of substrates can be measured by AFM at real process temperatures (Capella and  
115 Stark, 2006; Goode et al., 2013). When temperature increases, the work of adhesion tends to  
116 increase, especially for proteinaceous deposits that are denatured upon heating (Goode et al.,  
117 2013).

118 The influence of surface parameters (i.e. roughness, SFE, and temperature) on surface fouling  
119 is clear, but identifying their synergetic effects, especially under realistic conditions, will  
120 provide further insights to reduce industrial fouling. This work studies the effects of surface  
121 characteristics of 316L stainless steel on surface fouling, from nano- to macro-scale, by  
122 characterisation of surface properties before and after foulant deposition under simulated  
123 pasteurisation conditions. The objectives are (i) to determine the influence of surface  
124 characteristics on adhesion between liquid and solid (SS or foulant), (ii) to determine the extent  
125 of surface hydrophobicity as a function of surface fouling, and (iii) to establish connections  
126 between the mechanical properties of both substrates, stainless steel and foulant, and the  
127 surface parameters examined.

## 128 2. Materials & methods

### 129 2.1 Surface characterisation

130 Stainless steel 316L surfaces (2.54 x 2.54 cm) were prepared by using different sandpapers grit  
131 (600, 240, and 180 for mirror, satin and brush grades respectively). The process produces  
132 unidirectionally oriented substrates within the standard roughness limit defined by the 3-A  
133 Sanitary Standards (3-A SSI) and the European Hygienic Engineering & Design Group  
134 (EHEDG) for dairy industries ( $R_a < 0.8 \mu\text{m}$ ) (Frantsen and Mathiesen, 2009). Surface  
135 roughness ( $R_a$ ) was determined by White Light Interferometry (WLI) (MicroXAM2,  
136 Omniscan, U.K.) from at least four locations on each sample.

137 Stainless steel coupons were cleaned by the method detailed in Phinney et al., (2017): 2.0%  
138 (wt./wt.) NaOH aqueous solution at 80°C under stirring for 1 h to achieve complete removal  
139 of potential contaminants, and cooled to room temperature using a water bath. The substrates  
140 were subsequently rinsed by 1.0% (vol./vol.) HCl solution, soaked in hexane for 5 min and  
141 then acetone for another 5 min before dried by an air stream. All solvents used are HPLC grade.

### 142 2.2 Fouling material and procedure

143 A commercial whey protein concentrate (WPC) (CARBELAC 35, Carbery, Cork, Ireland) was  
144 used as received to prepare a model foulant solution (10% wt./wt.) to which the polished  
145 stainless steel coupons were exposed. Specifications of the WPC powder used are listed in  
146 **Table 1**. The model solution was prepared by mixing the WPC powder with de-ionised water  
147 at room temperature for an hour. Attention was paid to minimise aeration, foam formation, and  
148 proteins denaturation of the solution following the procedure developed in Phinney et al.,  
149 (2017). To mimic relevant industrial conditions (pasteurisation temperatures, protein  
150 denaturation and surface contact time allowing ageing of the deposit), 1 ml of the prepared



151 solution was placed on the cleaned coupons (temperature kept at 25°C before deposition, unless  
 152 otherwise stated) and maintained at 75°C for 1 h in an oven, and then cooled. Time and  
 153 temperature profiles were used to minimise bubble formation, allowing gelation of the solution  
 154 (Phinney et al., 2017). Average fouling thickness and roughness were measured by WLI from  
 155 at least four different areas, and surface topography characterised using an AFM (Dimension  
 156 3100, Veeco, Cambridge, UK) in Tapping mode using silicon cantilevers (HQ:NSC15/AIBS  
 157 AFM tip; ApexProbes, UK).

158 **Table 1.** Chemical composition and protein profile of the commercial WPC powder. For chemical specification,  
 159 percentage is expressed by grams of component per 100 g of WPC powder. For protein profile, percentage is  
 160 expressed by grams of proteins per 100 g of True Protein. Data supplied by Carbery (Ballineen, Co Cork, Ireland).

161

Chemical Specification	Total concentration (%)
Protein	35.0
Total Nitrogen	5.5
Moisture	5.0
Fat	4.0
Ash	6.0
Lactose	50.0
<b>Protein profile</b>	
Glycomacropeptide / Caseinomacropeptide	27.1
$\alpha$ -lactalbumin	7.5
Blood Serum Albumin	4.5
$\beta$ -lactoglobulin	56.7
Lactoferrin	2.0
Immunoglobulin G	2.1

162

### 163 2.3 Contact angle measurements and surface free energy characterisation

164 The sessile drop method was deployed to measure the equilibrium contact angle (ECA) for  
 165 wall temperatures between 25 to 80°C. ECA was quantitatively measured on a stage where the  
 166 influence of convective motion is negligible. A small liquid droplet was placed on the solid  
 167 substrate while contact angle evolution was recorded in real-time (1000 fps) by a high-speed  
 168 camera (FastCam SA2, Photron Europe, Bucks, United Kingdom). Stainless steel coupons

169 (with or without foulant) were placed on a heating stage monitored by a digital thermometer  
170 and controlled by a thermal bath. A pipette was used to place 10  $\mu\text{L}$  droplets of the testing  
171 liquids on the substrate when the surface temperature is constant. ImageJ software was used  
172 for image processing.

173 The set of test liquids in **Table 2** has been selected to emphasize specific molecular interactions  
174 of the surfaces of interest: two non-polar liquids (diiodomethane and 1-bromonaphthalene)  
175 were selected to characterise non-polar interactions, while a polar liquid (Ethylene glycol) is  
176 used to model the solid surface as having two components to its surface energy, polar and non-  
177 polar.

178 The **Wu method** (Wu, 1973, 1971) was selected for calculating the surface free energy (SFE)  
179 of a solid substrate by dividing it into polar and disperse components. This Harmonic mean  
180 model provides reliable values of both disperse and polar parts. The liquids used were 1-  
181 Bromonaphthalene and ethylene glycol. The equations used for calculations are:

$$182 \quad \gamma_{sl} = \gamma_s + \gamma_l - \frac{4\gamma_s^d \gamma_l^d}{\gamma_s^d + \gamma_l^d} - \frac{4\gamma_s^p \gamma_l^p}{\gamma_s^p + \gamma_l^p} \quad [1]$$

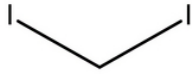
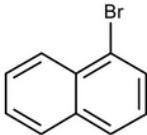
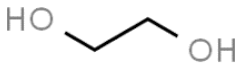
183 Combining [1] with Young's equation, the following equation can be obtained:

$$184 \quad \gamma_l(1 + \cos\text{ECA}) = \frac{4\gamma_s^d \gamma_l^d}{\gamma_s^d + \gamma_l^d} - \frac{4\gamma_s^p \gamma_l^p}{\gamma_s^p + \gamma_l^p} \quad [2]$$

185 Where ECA is the equilibrium contact angle,  $\gamma_{sl}$  is the interfacial tension between the solid  
186 and the liquid,  $\gamma_s$  is the overall surface energy of the solid, and  $\gamma_l$  is the overall surface tension  
187 of the wetting liquid, along with their corresponding disperse ( $\gamma^D$ ) and polar components ( $\gamma^P$ ).

188

189 **Table 2.** Properties of liquids used to characterise the equilibrium contact angle. Properties listed as a function of  
 190 temperature: total surface tension of the liquid ( $\gamma_L^T$ ), and corresponding disperse ( $\gamma_L^D$ ) and polar components  
 191 ( $\gamma_L^P$ ).  
 192

Liquid	T (°C)	Formula	$\gamma_L^T$ (mN/m)	$\gamma_L^D$ (mN/m)	$\gamma_L^P$ (mN/m)
Diiodomethane (Landolt and Börnstein, 1961)	25		50.0	50.0	0.0
	80		42.5	42.5	0.0
1-Bromonaphthalene (Rulison, 2005)	25		44.6	44.6	0.0
	80		42.2	42.2	0.0
Ethylene glycol (MEGlobal, 2008)	25		47.5	28.7	18.8
	80		43.6	26.3	17.3

193

#### 194 2.4 Nano-mechanical characterisation of substrates

195 Nano-mechanical properties of all surfaces were quantified by atomic force microscope (AFM)  
 196 (Dimension 3100, Veeco, Cambridge, UK) based force spectroscopy. A borosilicate  
 197 microsphere, with a nominal diameter of 5.9  $\mu\text{m}$  (Thermo Fisher Scientific, Loughborough,  
 198 UK), was fixed to an AFM cantilever (ApexProbes, UK) using an epoxy adhesive (Araldite,  
 199 UK) that is chemically inert. Spring constant of each cantilever was quantified using the  
 200 thermal method (Hutter and Bechhoefer, 1993). Force measurements were carried out over four  
 201 different locations per sample, with at least 50 force curves at each location. Adhesion force  
 202 was quantified by the hysteresis upon retraction of the particle from the surface in contact.  
 203 Indentation Analysis (NanoScope Analysis), using the Hertz model (spherical indenter) and  
 204 fitting by the Contact Point Based method, was used to calculate Young's modulus of the  
 205 substrates of interest. This method emphasises the minimum force at the contact point while  
 206 minimising the influence of noise and interferences. Poisson's ratio was assumed to be 0.477  
 207 for Whey protein gels (Langley and Green, 1989) and 0.270 for SS316L surface (AZoNetwork  
 208 UK Ltd, Manchester, United Kingdom).

209 **2.5 Liquid cohesion and adhesion work**

210 The work of cohesion ( $W_c$ ) is defined as the work per unit area produced in dividing a pure  
211 liquid (**Eq. 3**), while the work of adhesion ( $W_a$ ) is defined (**Eq. 4**) as the work required to  
212 separate two adjacent phases, in this case, a liquid-solid system (Ebnesajjad, 2006). If the ratio  
213  $W_c/W_a$  is below one, the liquid spreads along the surface because adhesion work is larger than  
214 the cohesive one.

215  $W_c = 2\gamma_L$  [3]

216  $W_a = \gamma_L(1 + \cos ECA)$  [4]

217 **2.6 Statistical analysis: ANOVA**

218 One-way analysis of variance (ANOVA) (Gelman, 2005) of both liquid contact angle and SFE  
219 of stainless steel surfaces was carried out as a function of both surface roughness and  
220 temperature to identify statistical differences between the means of two or more groups.

221

## 222 3. Results & Discussion

### 223 3.1 Effect of surface temperature and roughness on stainless steel wettability

224 Stainless steel (316L) coupons were processed to achieve three different surface finishes based  
225 on their roughness level ( $R_a$ ): mirror ( $0.03 \pm 0.01 \mu\text{m}$ ); satin ( $0.31 \pm 0.01 \mu\text{m}$ ); and brush ( $0.83$   
226  $\pm 0.13 \mu\text{m}$ ), for which the wettability was measured as a function of both wall temperature and  
227 liquid type. Droplets of three different liquids, ethylene glycol (EG), bromonaphthalene (BN),  
228 and diiodomethane (DM), were placed on the stainless steel coupons for contact angle  
229 measurements. ECA values are summarised in **Figure 1**. It was assumed that liquid droplets  
230 completely wet the metal surface according to Wenzel (1936) (no air entrapped).

231 Temperature directly influences liquid properties such as surface tension, density and viscosity  
232 (Escobedo and Mansoori, 1996; Wandschneider et al., 2008). At room temperature, contact  
233 angles decreased according to liquid surface tension; DM showed the highest contact angle  
234 ( $43.0 \pm 1.8^\circ$ ; **Figure 1b**). At higher temperatures (25-80°C), ECAs decreased. The contact  
235 angles of EG and BN were most reduced as surface temperature increased. One-way ANOVA  
236 analysis was performed (**Table 3**), and shows significant ECA differences for EG and BN as a  
237 function of temperature. However, the wetting properties of DM did not seem to change with  
238 temperature despite its surface tension being more sensitive to the temperature than the other  
239 liquids (**Table 2**).

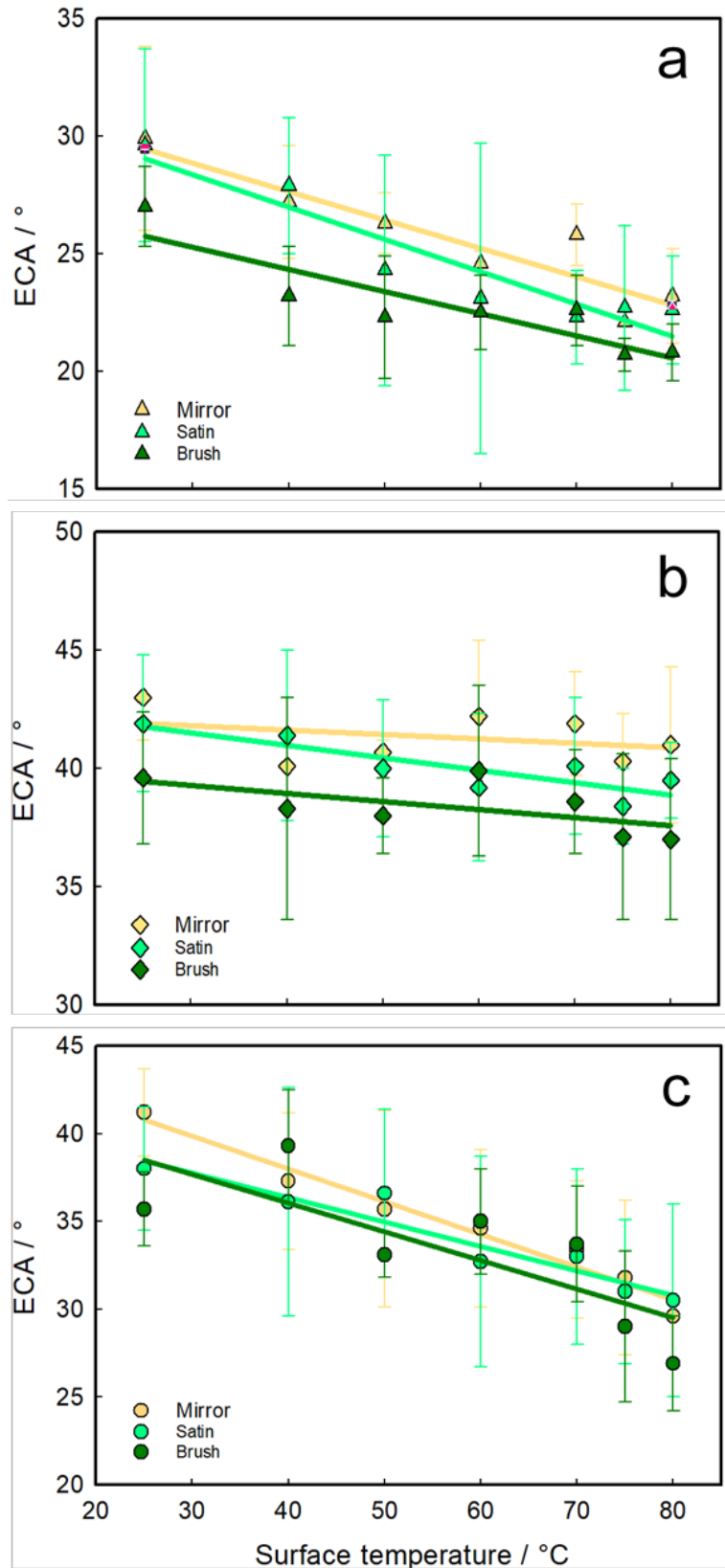
240 Surface roughness ( $R_a < 0.83 \mu\text{m}$ ) appeared to affect the ECA measurements under the testing  
241 conditions: the rougher the surface, the greater the wetting observed. ANOVA analysis shows  
242 insignificant differences for ECA values of both EG and BN as a function of substrate  
243 roughness. However, the ECA results of DM was very responsive to roughness variations.  
244 These differences may be related to the spreading factor of those liquids (Kubiak et al., 2011).

245 Surface parameters play an important role in interfacial adhesion. Wetting of SS316L increases  
246 as a function of both the surface roughness and temperature. Industrially, these results imply  
247 that polishing surfaces to a high finish is effective in reducing liquid adhesion, and subsequent  
248 fouling: this agrees with practice, as well as previous experimental works where significant  
249 fouling reduction was observed using a mirror-finish surface instead of an unpolished one  
250 (Zouaghi et al., 2018).

251

252 **Figure 1.** Equilibrium contact angle (ECA) as a function of both SS316L roughness and temperature. Three  
 253 classes of surface finish have been used: mirror, satin, and brush. The mean values of liquid contact angles of at  
 254 least three different drops per liquid are showed along standard deviation. The liquid used are 1-bromonaphthalene  
 255 (a), diiodomethane (b) and ethylene glycol (c). Lines show linear regression fit to facilitate data visualisation.

256



257

258 **Table 3.** One-way ANOVA analysis of both Equilibrium Contact Angle (ECA) and Surface Free Energy (SFE)  
 259 measurements of clean and fouled 316L stainless steel as a function of surface temperature and roughness. F-  
 260 value and p-value refer to the ratio of the variance of the group means to pooled within group variance and the  
 261 probability of obtaining an F-value, respectively. P-value must be <0.05 to show a statistical significant difference  
 262 between groups for the studied conditions.

	Temperature dependence		Roughness dependence	
	<i>F-Value</i>	<i>p-Value</i>	<i>F-Value</i>	<i>p- Value</i>
<b>ECA upon SS316L</b>				
Diodomethane	1.0996	0.4098	6.9074	0.0059
1-Bromonaphthalene	5.7781	0.0033	2.4257	0.1167
Ethylene glycol	8.1499	0.0006	0.6341	0.5418
<b>SFE of SS 316L</b>				
Total	0.4156	0.8566	10.1399	0.0011
Disperse	0.5533	0.7599	12.5021	0.0004
Polar	1.4503	0.2645	0.3753	0.6924
<b>ECA upon foulant</b>				
Diodomethane	0.4958	0.6952		
1-Bromonaphthalene	0.0349	0.9906		
Ethylene glycol	4.8457	0.0330		
<b>SFE of WPC foulant</b>				
Total	0.0915	0.9627		
Disperse	0.4989	0.6933		
Polar	6.3722	0.0163		

263

## 264 3.2 Alteration of surface wettability upon deposition of WPC foulant

### 265 3.2.1 Roughness of the surface foulant

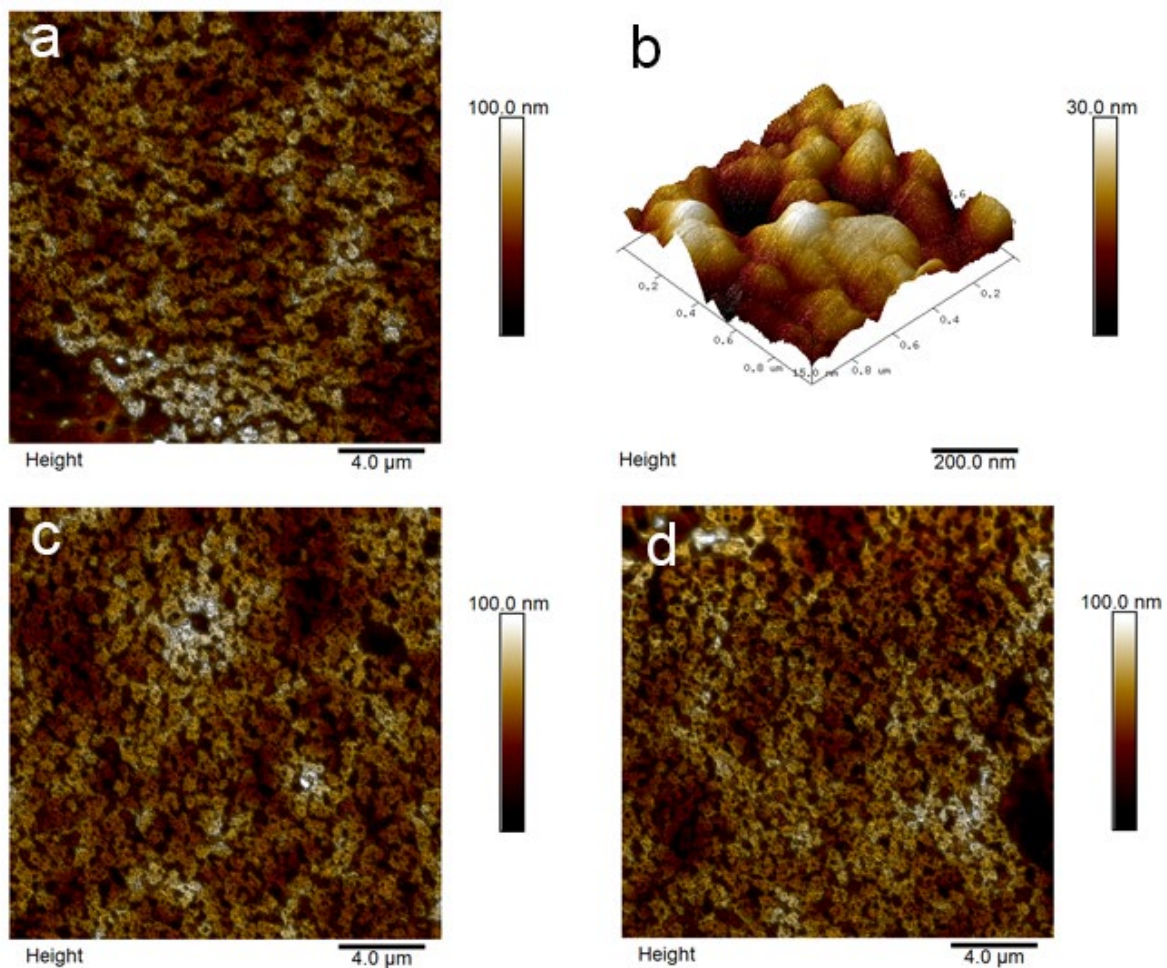
266 Whey protein foulant was prepared on the polished stainless steel coupons, simulating a well-  
 267 formed proteinaceous layer of similar characteristics to pasteuriser deposits. The areal density  
 268 and averaged thickness of this model foulant were 17.67 mg/cm<sup>2</sup> (Phinney et al., 2017) and  
 269 105.8 ± 8.6 µm respectively, which is consistent with the values for averaged fouling of raw  
 270 milk after eight hours of pasteurisation (12.73 ± 0.65 mg/cm<sup>2</sup> ; SS316L plate of Ra 0.46 ± 0.2  
 271 µm) found by Barish and Goddard (2013). Surface morphology of the whey protein foulant,  
 272 acquired by AFM in ambient conditions, are presented in **Figure 2**. The mean foulant  
 273 roughness measured by WLI is 23 ± 6 nm, 23 ± 10 nm and 22 ± 8 nm on SS substrates with  
 274 mirror, satin, and brush finishes respectively, close to that measured by AFM (12.4 ± 0.8 nm



275 over a  $20 \times 20 \mu\text{m}$  area). A high resolution 3D scan (**Figure 2b**) shows that the clusters are of  
276 sizes less than  $0.3 \mu\text{m}$ , agreeing with the previous work (Jimenez et al., 2013). The consistent  
277 surface roughness values of foulants suggests that the influence of the surface finish of the  
278 underlying substrate is negligible for the model foulants formed. This is likely because the  
279 thickness of the foulant far exceeds the magnitude of the roughness of the coupons used. As  
280 such, the effect of WPC foulant roughness was neglected for contact angle measurements.

281 **Figure 2.** Representative surface morphology images of WPC foulant prepared at  $75^\circ\text{C}$  for 1 hour on stainless  
282 steel coupons of (a) mirror ( $20 \times 20 \mu\text{m}$ ); (b) mirror (3D image  $1 \times 1 \mu\text{m}$ ); (c) satin and (c) brush finishes.

283



284

285

### 286 3.2.2 Contact angle measurements of pure liquid as a function of foulant temperature

287 Denaturation and aggregation reactions of  $\beta$ -Lactoglobulin ( $\beta$ -Lg) that occur at pasteurisation  
288 temperatures lead to reactions of  $\beta$ -Lg with processing equipment and with other bulk  
289 compounds.  $\beta$ -Lg adsorption and its adhesion force are favoured by increased surface  
290 temperature (Santos et al., 2003). This section aims to examine the wetting characteristics of  
291 the model foulant layer as a function of wall temperature and liquid type.

292 **Figure 3** presents the wettability of both stainless steel and foulant as a function of temperature  
293 for the three testing liquids, which shows temperature dependence for both substrates. The  
294 ECA of non-polar liquids remained constant as the temperature increases: DM showed greater  
295 contact angle than BN ( $41.90 \pm 3.24^\circ$  and  $37.72 \pm 2.69^\circ$  respectively) throughout the  
296 temperature range examined. While the ECA of DM was constant for both substrates, that of  
297 BN on the foulant was almost 10 degrees greater than on the bare metal surface. Although the  
298 contact angle of BN on the stainless steel coupons decreased by ca.  $6^\circ$  when the temperature  
299 was increased from 25 to  $80^\circ\text{C}$ , it remained nearly constant ( $\pm 0.38^\circ$ ) on the WPC deposit over  
300 the same temperature range.

301 The polar liquid, EG, showed the highest value at room temperature ( $60.0 \pm 2.0^\circ$ ), suggesting  
302 a significant reduction of surface energy at the foulant-liquid interface. When the temperature  
303 of the substrate increased, the magnitude of reduction in the ECA of EG was similar for both  
304 metal and foulant. However, the ECA of EG on stainless steel was ca. 20 degrees greater than  
305 on the formed foulant. Of the three liquids tested, EG is the only one that shows such significant  
306 statistical differences with increased temperature (**Table 3**), which suggests that changes of  
307 polar and disperse interactions could be important once foulant is formed. A proteinaceous  
308 foulant layer can alter the wettability of a substrate as a function of both liquid composition  
309 and wall temperature.

310 Surface wettability is determined by the balance between adhesive and cohesive interactions at  
311 the solid-liquid interface. The ratio between the works of cohesion and adhesion of the liquids  
312 tested over the mirror-finish stainless steel substrates is presented in **Table 4**.

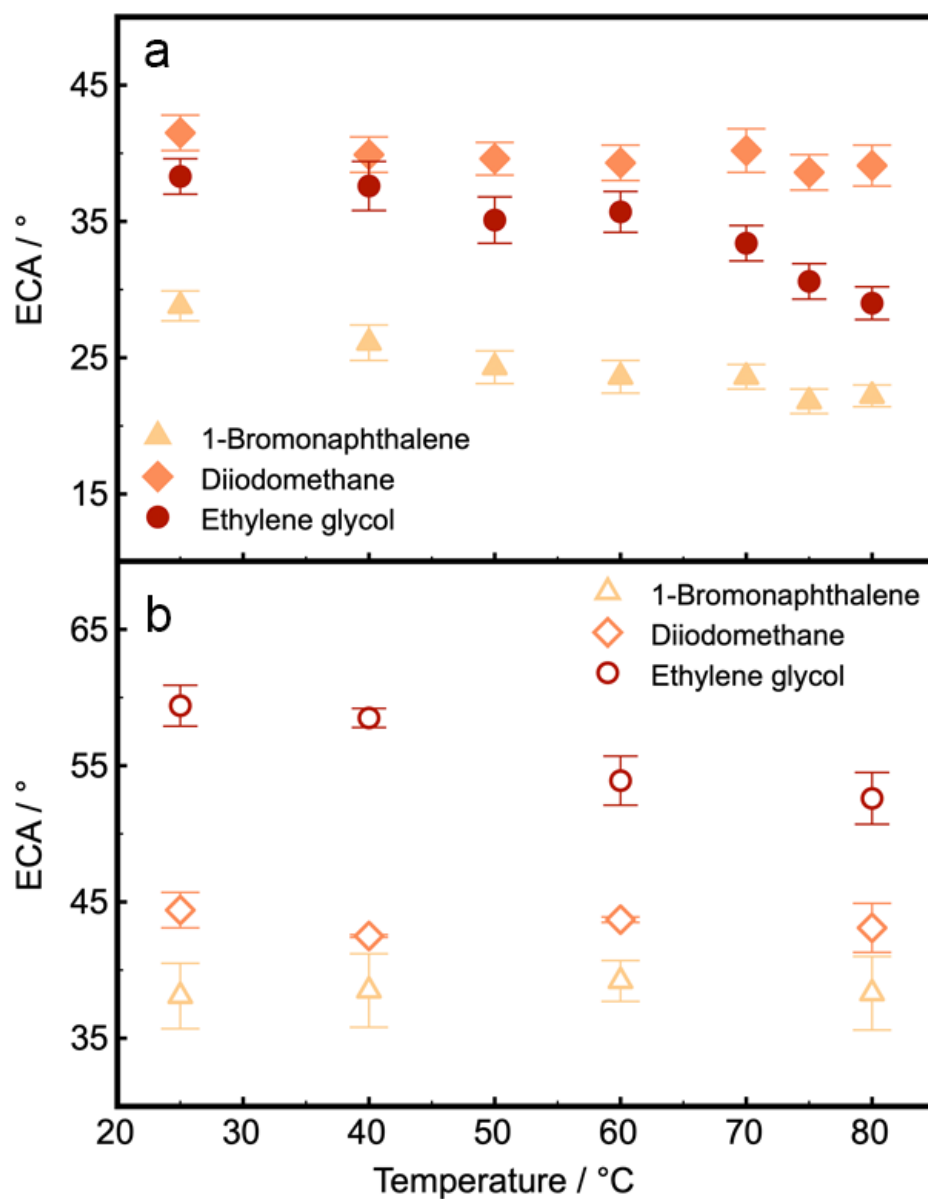
313 For non-polar liquids, there is a reduction of the disperse forces inside the liquid drop as  
314 temperature increases, promoting liquid spreading across the metal substrate. The constant  
315 contact angles of the non-polar liquids on the foulant is likely due to reduction of the disperse  
316 interactions, that compensate the temperature influence. For the polar liquid, increasing  
317 temperature reduces the polar and disperse bonds inside the liquid, reducing cohesive  
318 interactions and favouring surface wetting. It is clear that temperature has a much greater  
319 influence on the ECA on the foulant than on the SS substrate, related to the increased polarity  
320 at the interface.

321 Once total spreading of liquid was observed on the prepared foulant, drops of the two non-  
322 polar liquids were stable at short contact times (minutes), and there was no significant  
323 dissolution on the protein layer. However, the solubility of the polar liquid was favoured  
324 considerably over contact time. This supports the hypothesis that polarity might be critical for  
325 understanding the foulant-liquid interface.

326

327 **Figure 3.** Equilibrium contact angle (ECA) of the three selected liquids, 1-Bromonaphthalene, Diiodomethane,  
328 and Ethylene glycol as a function of temperature. Comparison of ECA evolution upon both substrates, (a) stainless  
329 steel and (b) WPC foulant. Error bars represent the standard error from at least three measurements.

330



331

332

333 **Table 4.** Ratio of cohesion and adhesion work (**section 2.5**) for liquids as a function of temperature on the  
 334 substrates of interest (stainless steel and WPC fouling). Large ratio ( $W_{\text{cohesion}}/W_{\text{adhesion}} > 1$ ) suggests that the liquid  
 335 has less tendency to spread on the substrate under the given condition, and vice versa.

336

	T [°C ]	[ $W_c / W_a$ ]		
		<i>DM</i>	<i>BN</i>	<i>EG</i>
SS316L (mirror)	25	1.16	1.07	1.14
	40	1.13	1.06	1.13
	50	1.12	1.06	1.12
	60	1.10	1.05	1.11
	70	1.08	1.05	1.10
	75	1.07	1.04	1.10
	80	1.07	1.04	1.09
	<i>Reduction</i>	<i>0.09</i>	<i>0.03</i>	<i>0.05</i>
WPC foulant	25	1.15	1.12	1.33
	40	1.15	1.12	1.31
	60	1.15	1.12	1.27
	80	1.14	1.12	1.23
	<i>Reduction</i>	<i>0.01</i>	<i>0.00</i>	<i>0.10</i>

337

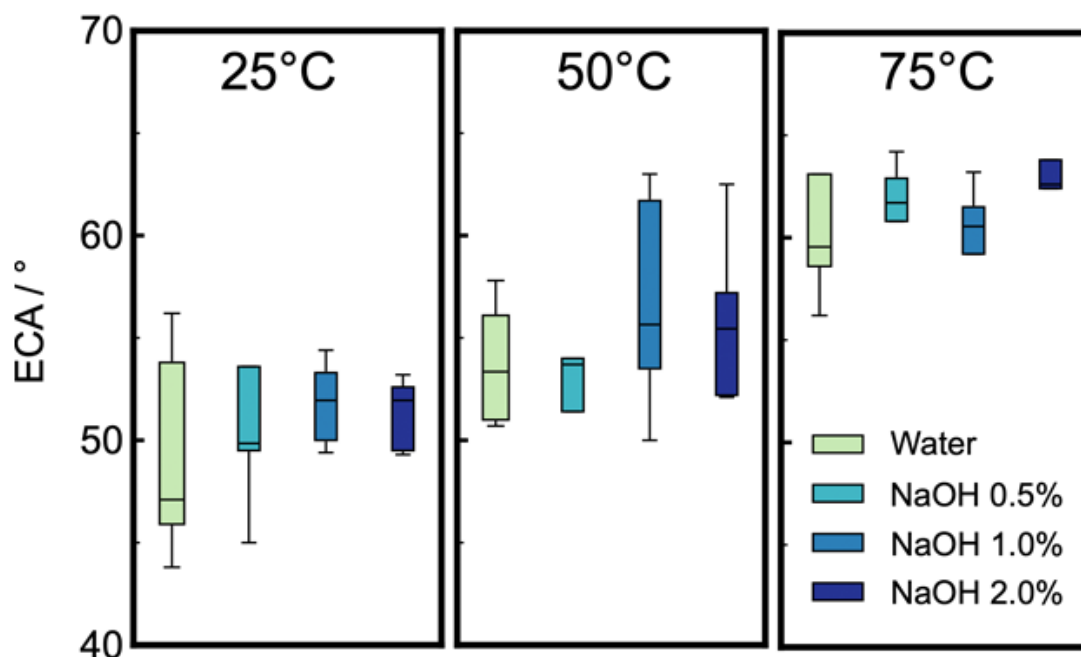
### 338 3.2.3 Contact angles of cleaning solution on WPC foulant

339 As shown in **Figures 1** and **3**, it is likely that temperature will have a significant influence on  
 340 the wetting behaviour of the cleaning solutions on the surface foulant, which determines the  
 341 removal mechanisms. **Figure 4** shows the contact angles of different cleaning formulations,  
 342 including water and aqueous solutions of different NaOH concentration (0.5%, 1% and 2%  
 343 wt./wt.), on the WPC foulant as a function of the surface temperature (25, 50 and 75°C).

344 At room temperature (25°C), water contact angle on the WPC foulant was found to be  $49.0 \pm$   
 345  $5.2^\circ$ , slightly less than on a clean stainless steel surface ( $66.8 \pm 9.0^\circ$  for mirror and  $52.4 \pm 5.4^\circ$   
 346 for brush finishes). This is very likely due to solvation of the proteinaceous film upon contact  
 347 with water. As the temperature increased from 25°C to 75°C, the contact angle of water on  
 348 WPC increased, suggesting an enhanced surface hydrophobicity, a different characteristic to  
 349 that observed on bare stainless steel (**Figure 3**) where high temperature facilitated surface  
 350 wetting of water. Because the prepared WPC foulant consists of densely packed proteins, we

351 speculate that the adsorbed  $\beta$ -Lactoglobulin either exposed its hydrophobic core to the foulant-  
352 air interface or denaturated at increased temperature, giving increased surface hydrophobicity  
353 of the WPC. Contact angles of cleaning solutions followed a similar behaviour, and according  
354 to the ANOVA test in **Table 5**, there was no significant difference between ECAs of water and  
355 the cleaning solutions tested. This observation highlights the critical role of molecular  
356 configuration on the foulant surface in determining its wettability. Previous work suggested  
357 that fast foulant removal was observed at a high temperatures (Phinney et al., 2017), which  
358 confirms that cleaning is a complex process determined by not only the surface wettability of  
359 the foulant, but its cohesiveness and its adhesion to the supporting substrate (stainless steel  
360 here).

361 **Figure 4.** Contact angle measurements of cleaning liquids upon WPC foulant as a function of wall temperature  
362 (25, 50, and 75°C). The cleaning solutions are water, NaOH 0.5%, NaOH 1%, and NaOH 2%. ANOVA analysis  
363 shows non-significant differences between CAs of the cleaning solutions tested.



364  
365

366 **Table 5.** One-way ANOVA analysis of the effect of both cleaning formulations and substrate temperature on the  
 367 wettability of WPC fouled stainless steel 316L. F-value and p-value refer to the ratio of the variance of the group  
 368 means to pooled within group variance and the probability of obtaining an F-value, respectively. P-value must be  
 369 <0.05 to show a statistical significant difference between groups for the studied conditions.

<b>Temperature dependence</b>		
	<i>F-Value</i>	<i>p-Value</i>
Water	3.6823	0.0006
NaOH 0.5%	3.6823	0.0005
NaOH 1%	3.6823	0.0010
NaOH 2%	3.6823	0.0000

<b>Effect of cleaning formulation</b>		
	<i>F-Value</i>	<i>p-Value</i>
25°C	3.0984	0.5085
50°C	3.0984	0.4525
75°C	3.0984	0.5069

370

371 **3.3 Surface free energy of stainless steel and WPC foulant**

372 **Figure 5** shows that the total surface free energy of the SS316L substrate, as well as its disperse  
 373 and polar components, are independent of surface temperature ( $45.4 \pm 0.6$ ,  $39.4 \pm 0.5$ , and  $6.0$   
 374  $\pm 0.4$  mN/m respectively). The measured values of SFE are in agreement with those reported  
 375 at room temperature (Barish and Goddard, 2013; Zhao et al., 2004). ANOVA analysis (**Table**  
 376 **3**) suggests that the polar component has no notable dependence on surface roughness and  
 377 temperature, whilst the disperse component is affected by surface roughness. A previous study  
 378 concerning the effects of both roughness and temperature on SFE (Avila-Sierra et al., 2019)  
 379 showed that the greater surface roughness, the higher the surface energy.

380 Our results confirm that the SFE of stainless steel surfaces is constant in the operational  
 381 window of industrial pasteurisation processes, which implies that the attractive interactions  
 382 between stainless steel and the liquid being processed remain constant. SFE at room  
 383 temperature could be used to estimate the free energy of the substrate under 80°C. However,  
 384 other parameters such as surface roughness or alterations of liquid properties do affect the  
 385 interfacial interactions.

386 The pasteurisation process, however, is more dynamic than the contact angle measurements  
387 carried out in the present study – SFE of the solid substrate would evolve as the foulant  
388 develops. It is therefore critical to evaluate the SFE of a model proteinaceous layer as the  
389 function of temperature, upon which the underpinning formation mechanism of the foulant can  
390 be established. The effect of temperature on the liquid-foulant interface is of particular interest.  
391 Harmonic mean approach (**section 2.3**) was implemented to evaluate the SFE variations up to  
392 80°C, with data of **Figure 5** confirming that the total SFE of the foulant remained constant  
393 ( $38.0 \pm 0.1$  mN/m), consistent with the observation made on stainless steel. However, the  
394 dispersive and polar components of the SFE changed: there is a slight decrease of the dispersive  
395 part while the polar part increases significantly, showing an increase of the foulant polarity  
396 around 3.4% from 25 to 80°C (calculated as % of  $\gamma_{\text{SPolar}}/\gamma_{\text{STotal}}$ ). ANOVA analysis (**Table 3**)  
397 shows significant differences for the polar part once temperature increased. These findings  
398 support our hypothesis that the adsorbed  $\beta$ -Lactoglobulin could adjust its molecular  
399 configuration so as to expose the hydrophobic core, leading to an increased surface polarity.

400 Some previous studies confirmed the relationship between an increased amount of foulant and  
401 the polar component of a wide variety of surfaces (e.g. diamond-like carbon (DLC) coatings  
402 (Boxler et al., 2013), imbedded  $\text{MoS}_2^{2+}$  ions,  $\text{SiO}_x$  and DLC–Si–O films, Ni–P matrix with  
403 PTFE particles (Rosmaninho and Melo, 2008), TiN layers (Rosmaninho et al., 2005), and  
404 implantation of  $\text{SiF}_3^+$  ions (Rosmaninho et al., 2007; Rosmaninho and Melo, 2006b)), where a  
405 secondary protein layer could develop on the initially bound protein film through polar  
406 interactions (Addesso and Lund, 1997). However, temperature not only affects fouling rate and  
407 polarity of the deposit, it also affects deposit itself (Burton, 1968). As a result of increased wall  
408 temperature, the increased polarity of the surface foulant might accelerate the interactions  
409 between compounds at the bulk fluid and the pre-deposited material, which explains why  
410 minerals tend to present in the first layer of protein deposit (Belmar-Beiny and Fryer, 1993),



411 forming a compacted structure over the processing time (Pappas and Rothwell, 1991).

412 During milk processing, the rate of heat transfer decreases with time due to the build-up of

413 surface foulant (Kukulka and Leising, 2009). Alharthi (Alharthi, 2014) identified how the

414 concentration of proteins and minerals can affect such reduction. Therefore, after the

415 development of the surface deposit, heat transfer will decrease due to the deposit

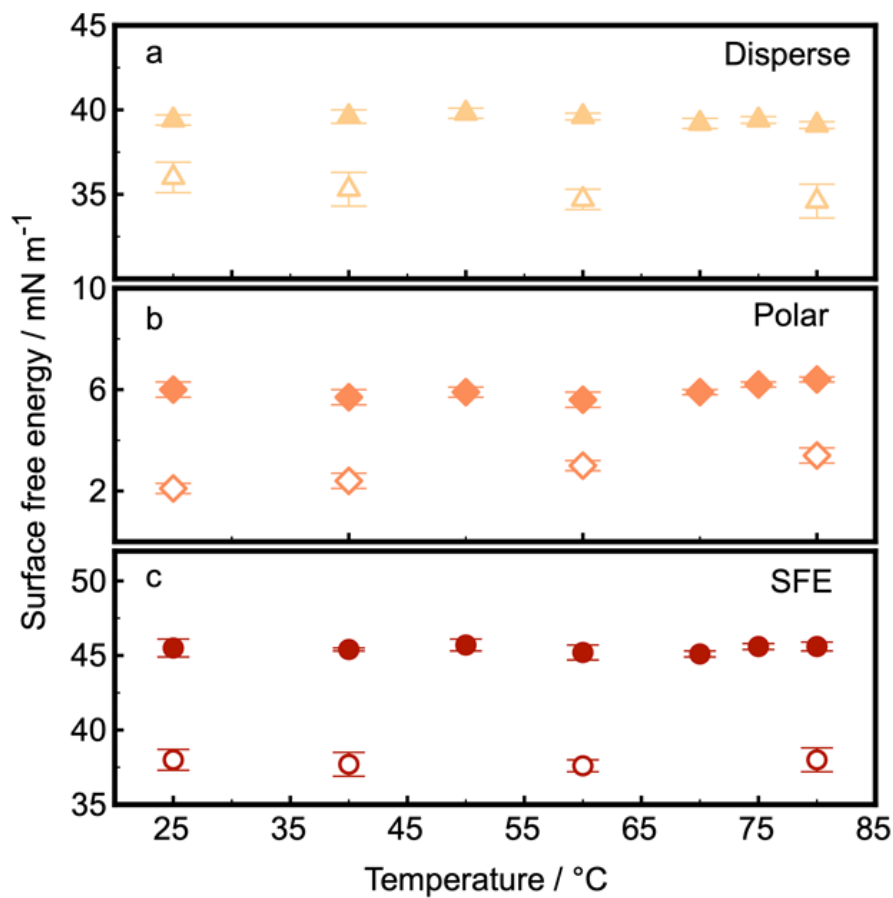
416 thickness/composition, generating a gradient of temperature inside the deposit. This implies

417 alterations of temperature that would limit the molecular interactions at the interface, and hence

418 minimise the fouling rate over time.

419 **Figure 5.** (a) Disperse, (b) Polar, and (c) Total Surface Free Energy of both SS316L (filled) and WPC foulant  
 420 (empty) as a function of wall temperature. Liquids tested: Ethylene glycol and 1-Bromonaphthalene. Error bars  
 421 represent the standard error of at least three measurements.

422



423

424

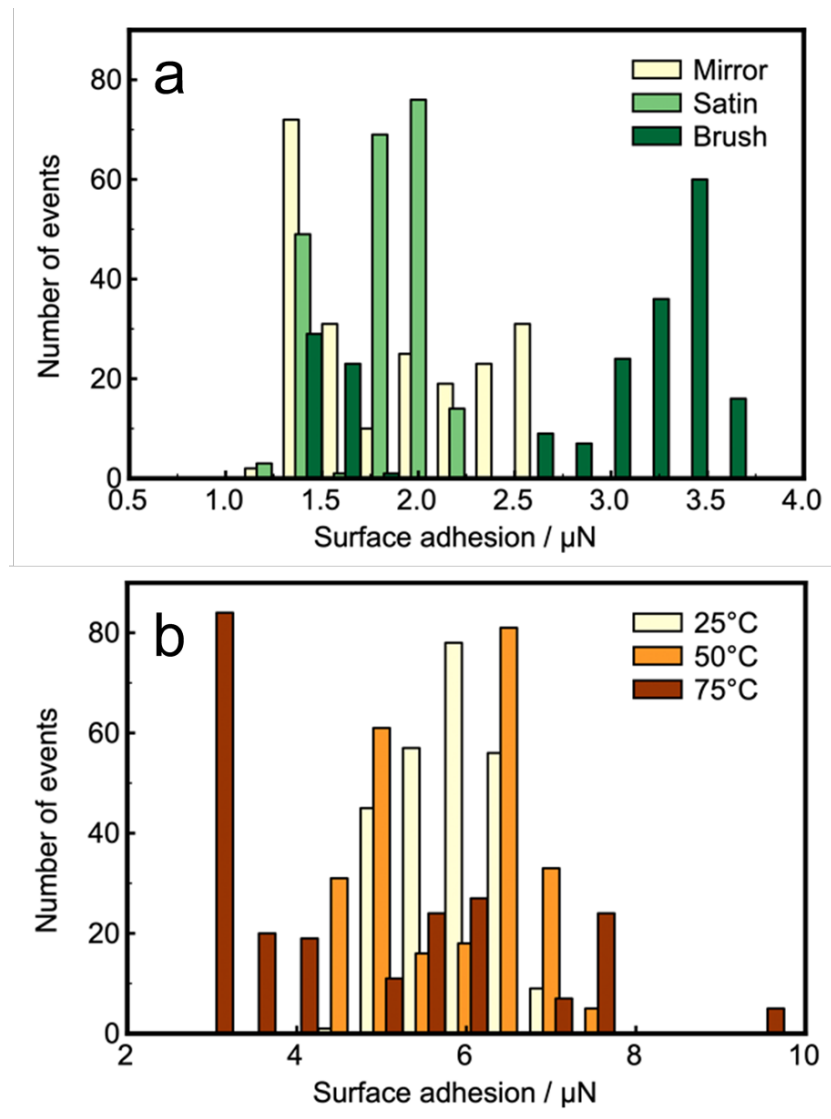
### 425 **3.4 Effect of the roughness and deposition temperature on the nanomechanical** 426 **properties of the substrate**

427 Results suggest that the liquid-solid interface is controlled by surface parameters such as  
428 roughness and wall temperature, whilst the surface free energy data confirms that the  
429 temperature of the solid substrate influences the characteristics of the formed foulant in terms  
430 of polarity and hydrophobicity. To further decouple the effects of chemistry and roughness on  
431 the surface free energy, force spectroscopy experiments based on AFM were carried out in  
432 ambient on SS316L substrates of different finishing grades, data presented in **Figure 6a**. As  
433 demonstrated (Sauerer et al., 2016), such a technique can be effectively used as an alternative  
434 to conventional contact angle experiments with significantly improved spatial resolution. The  
435 adhesion measured between a colloidal probe (diameter ca. 6  $\mu\text{m}$ ) and substrate in an ambient  
436 environment is primarily determined by the capillary force that is controlled by the humidity  
437 of the environment, chemical composition, roughness, and modulus of the substrate.

438 For stainless steel samples, both environmental conditions and chemical composition were kept  
439 constant, and the contact area is approximately 0.056  $\mu\text{m}^2$ , assuming Hertzian contact  
440 mechanics. Adhesion force on the SS substrates with mirror finish was in the range 1.5-2.5  $\mu\text{N}$ ,  
441 consistent with that on SS of satin finishing, but with a slightly broader distribution, as shown  
442 in **Figure 6a**. The similar range of adhesion measured on the mirror and satin samples suggests  
443 that the effect of roughness on surface energy at sub-micron scale was insignificant between  
444 those two finishes. The averaged surface adhesion increased to 3.5  $\mu\text{N}$ , with a broad  
445 distribution, on the SS substrate with brush finish. Enhanced surface adhesion was likely due  
446 to the elevated contact area between the colloidal probe and the solid surface, as the result of  
447 increased surface roughness, evidenced by both the surface morphology and the scattered  
448 distribution of the adhesion force.

449 **Figure 6.** Adhesion force between an AFM colloidal probe and both (a) 316L stainless steel with mirror, satin,  
 450 and brush surface finishes and (b) the WPC foulant generated on 316L SS with mirror finish under controlled  
 451 surface temperature: 25°C, 50°C and 75°C.

452



453

454

455 **Figure 6b** shows the histograms of adhesion force acquired from the proteinaceous foulant  
 456 developed on SS substrates of mirror finish at three different substrate temperatures. In the  
 457 presence of the WPC foulant, it is clear that the surface adhesion was increased to a range of  
 458 3-10 μN: of the several parameters that determine the surface adhesion, foulant roughness  
 459 probably plays only a small role, as evidenced by the morphology in **Figure 2**. The polar groups  
 460 on the surface of the foulant, are likely the major contributing factor for increased adhesion,

461 consistent with the contact angle results presented in **Figure 3**. Although there was only minor  
462 difference between average adhesion measured on foulants formed at 25 and 50°C, there was  
463 an increased range of adhesion force on the latter. This increased further on foulant prepared  
464 at 75°C: adhesion force spanned a broad range, implying a heterogeneous surface, likely the  
465 result of increasingly random molecular orientation.

466 The cohesiveness of the formed foulant and its correlation with the surface parameters and the  
467 processing conditions, can be quantified by using AFM based nanoindentation. The Young's  
468 modulus (YM) of the foulant was quantified as a function of temperature. The synergistic effect  
469 of surface roughness and deformability (Young's modulus) determines the contact area  
470 between two surfaces (Halvey et al., 2018; Rabinovich et al., 2000). At room temperature, there  
471 is a reduction in YM from  $3.9 \pm 0.7$  GPa to  $3.3 \pm 1.3$  GPa for clean and fouled mirror SS  
472 substrates respectively, where both materials can be viewed as hard substrates (Halvey et al.,  
473 2018). For proteinaceous foulants deposited on the SS with the three different metal finishes,  
474 the averaged YM remains practically constant ( $3.3 \pm 1.3$  GPa and  $3.2 \pm 0.4$  GPa for both fouled  
475 mirror and brush metal surfaces respectively), independent of the roughness of the supporting  
476 substrate. However, Young's modulus of the WPC foulant increased with temperature:  $3.3 \pm$   
477  $1.3$  GPa,  $3.7 \pm 0.3$  GPa and  $3.9 \pm 0.7$  GPa for 25, 50 and 75°C respectively, likely due to the  
478 configuration of protein molecules during deposition. This again highlights the impact the first  
479 foulant layer could have on the overall deposit characteristics. It is probable that the WPC  
480 proteins would form a densely packed foulant layer, with less uniform molecular orientation,  
481 when exposed to a mirror polish SS substrate at high temperature (75°C), whilst they would  
482 construct a less densely packed and more homogeneous surface film at 25°C.

483

484

#### 485 4. Conclusions

486 This work demonstrates that surface roughness, temperature, changes in surface composition,  
487 as well as the temperature difference between liquid and substrate govern the interfacial  
488 interactions in fouling, and therefore will control initial and subsequent formation of surface  
489 layers. Wettability of 316L stainless steel is favoured by increased surface roughness and wall  
490 temperature, showing how fine surface finishes are effective in reducing fouling. The surface  
491 free energy (SFE) of SS316L and its components remain constant between ambient and  
492 pasteurisation temperatures. However, as fouling develops, the SFE evolves. Upon foulant  
493 deposition, SFE decreases, and there was a polarity increase (3.4% from 25 to 80°C) of the  
494 fouled surface that might relate to the opening of the hydrophobic core of  $\beta$ -Lactoglobulin  
495 toward the foulant-air interface. Both surface adhesion and Young's modulus at sub-micron  
496 spatial resolution confirm that the molecular packing within the foulant and the molecular  
497 orientation on the foulant surface are affected by the temperature of the underlying substrate,  
498 showing how temperature variations in an industrial heat exchanger can result in different  
499 surface deposits.

500

501

502 **References**

- 503 Addesso, A., Lund, D.B., 1997. Influence of soild surface energy on protein adsorption. *J. Food Process. Preserv.*  
504 21, 319–333. <https://doi.org/10.1111/j.1745-4549.1997.tb00786.x>
- 505 Akesso, L., Pettitt, M.E., Callow, J.A., Callow, M.E., Stallard, J., Teer, D., Liu, C., Wang, S., Zhao, Q., D’Souza,  
506 F., Willemsen, P.R., Donnelly, G.T., Kocijan, A., Jenko, M., Jones, L.A., Guinaldo, P.C., 2009. The potential  
507 of nano-structured silicon oxide type coatings deposited by PACVD for control of aquatic biofouling.  
508 *Biofouling* 25, 55–67. <https://doi.org/10.1080/08927010802444275>
- 509 Akhtar, N., Bowen, J., Asteriadou, K., Robbins, P.T., Zhang, Z., Fryer, P.J., 2010. Matching the nano- to the  
510 meso-scale: Measuring deposit–surface interactions with atomic force microscopy and micromanipulation.  
511 *Food Bioprod. Process.* 88, 341–348. <https://doi.org/10.1016/j.fbp.2010.08.006>
- 512 Akhtar, N.R., 2010. The fundamental interactions between deposits and surfaces at nanoscale using atomic force  
513 microscopy. PhD thesis, University of Birmingham.
- 514 Alharthi, M., 2014. Fouling and cleaning studies of protein fouling at pasteurisation temperatures. PhD thesis,  
515 University of Birmingham.
- 516 Avila-Sierra, A., Zhang, Z.J., Fryer, P.J., 2019. Effect of surface characteristics on cleaning performance for CIP  
517 system in food processing. *Energy Procedia* 161, 115–122. <https://doi.org/10.1016/j.egypro.2019.02.067>
- 518 Baier, R.E., 1980. Substrata influences on the adhesion of microorganisms and their resultant new surface  
519 properties. G. Bitton, K.S. Marshall (Eds.), *Adsorption of Microorganisms to Surfaces*, Wiley-Interscience  
520 Publishers, New York, 59–104.
- 521 Baier, R.E., Depalma, V.A., Goupil, D.W., Cohen, E., 1985. Human platelet spreading on substrata of known  
522 surface chemistry. *J. Biomed. Mater. Res.* 19, 1157–1167. <https://doi.org/10.1002/jbm.820190922>
- 523 Bansal, B., Chen, X.D., 2006. A Critical Review of Milk Fouling in Heat Exchangers. *Comprehensive reviews in*  
524 *food science and food safety.* 5, 27–33. <https://doi.org/10.1111/j.1541-4337.2006.tb00080.x>
- 525 Barish, J.A., Goddard, J.M., 2013. Anti-fouling surface modified stainless steel for food processing. *Food*  
526 *Bioprod. Process.* 91, 352–361. <https://doi.org/10.1016/j.fbp.2013.01.003>
- 527 Belmar-Beiny, M.T., Fryer, P.J., 1993. Preliminary stages of fouling from whey protein solutions. *J. Dairy Res.*

528 6, 467–483. <https://doi.org/10.1017/S0022029900027837>

529 Białopiotrowicz, T., 2003. Wettability of starch gel films. *Food Hydrocoll.* 17, 141–147.  
530 [https://doi.org/10.1016/S0268-005X\(02\)00046-2](https://doi.org/10.1016/S0268-005X(02)00046-2)

531 Boxler, C., W., A., Scholl, S., 2013. Fouling of milk components on DLC coated surfaces at pasteurization and  
532 UHT temperatures. *Food Bioprod. Process.* 91, 336–347. <https://doi.org/10.1016/j.fbp.2012.11.012>

533 Burton, H., 1968. Reviews of the progress of dairy science. *J. Dairy Res.* 35, 317–330.  
534 <https://doi.org/10.1017/S0022029900019038>

535 Capella, B., Stark, W., 2006. Adhesion of amorphous polymers as a function of temperature probed with AFM  
536 force–distance curves. *J. Colloid Interface Sci.* 296, 507–514. <https://doi.org/10.1016/j.jcis.2005.09.043>

537 Choi, W.Y., Park, H.J., Ahn, D.J., Lee, J., Lee, C.Y., 2002. Wettability of chitosan coating solution on “Fuji”  
538 apple skin. *J. Food Sci.* 67, 2668–2672. <https://doi.org/10.1111/j.1365-2621.2002.tb08796.x>

539 Claeys, W.L., Ludikhuyze, L.R., Van Loey, A.M., Hendrickx, M.E., 2001. Inactivation kinetics of alkaline  
540 phosphatase and lactoperoxidase, and denaturation kinetics of  $\beta$ -lactoglobulin in raw milk under isothermal  
541 and dynamic temperature conditions. *J. Dairy Res.* 68, 95–107.  
542 <https://doi.org/10.1017/S002202990000460X>

543 Ebnesajjad, S., 2006. 2 - Surface Tension and Its Measurement. *Surf. Treat. Mater. Adhes. Bond.* 9–28.  
544 <https://doi.org/10.1016/B978-081551523-4.50004-3>

545 Escobedo, J., Mansoori, G.A., 1996. Surface Tension Prediction for Pure Fluids. *AIChE J.* 42, 1425–1433.  
546 <https://doi.org/10.1002/aic.690420523>

547 Frantsen, E., Mathiesen, J.T., 2009. Specifying Stainless Steel Surfaces for the Brewery, Dairy and  
548 Pharmaceutical Sectors, in: *NACE Corrosion*. p. 9573.

549 Garrett-Price, B., Smith, B., Watts, R., Knudsen, J., Marner, W., Suitor, J., 1985. Fouling of Heat Exchangers,  
550 Characteristics, Costs, Prevention, Control and Removal. Noyes Publ. Park Ridge, NJ.

551 Gelman, A., 2005. Analysis of variance—why it is more important than ever. *Ann. Stat.* 33, 1–53.  
552 <https://doi.org/10.1214/009053604000001048>

553 Goode, K.R., Bowen, J., Akhtar, N., Robbins, P.T., Fryer, P.J., 2013. The effect of temperature on adhesion forces

554 between surfaces and model foods containing whey protein and sugar. *J. Food Eng.* 118, 371–379.  
555 <https://doi.org/10.1016/j.jfoodeng.2013.03.016>

556 Halvey, A.K., Macdonald, B., Dhyani, A., Tuteja, A., 2018. Design of surfaces for controlling hard and soft  
557 fouling. *Philos. Trans. A* 377, 2138. <https://doi.org/10.1098/rsta.2018.0266>.

558 Handojo, A., Zhai, Y., Frankel, G., Pascall, M.A., 2009. Measurement of adhesion strengths between various milk  
559 products on glass surfaces using contact angle measurement and atomic force microscopy. *J. Food Eng.* 92,  
560 305–311. <https://doi.org/10.1016/j.jfoodeng.2008.11.018>

561 Hutter, J.L., Bechhoefer, J., 1993. Calibration of Atomic Force Microscope Tips. *Rev. Sci. Instrum.* 64, 1868.  
562 <https://doi.org/10.1063/1.1143970>

563 Jimenez, M., Delaplace, G., Nuns, N., Bellayer, S., Deresmes, D., Ronse, G., Alogaili, G., Collinet-Fressancourt,  
564 M., Traisnel, M., 2013. Toward the understanding of the interfacial dairy fouling deposition and growth  
565 mechanisms at a stainless steel surface: A multiscale approach. *J. Colloid Interface Sci.* 404, 192–200.  
566 <https://doi.org/10.1016/j.jcis.2013.04.021>

567 Karapetsas, G., Chamakos, N.T., Athanasios, O., Papathanasiou, G., 2017. Thermocapillary Droplet Actuation:  
568 Effect of Solid Structure and Wettability. *Langmuir* 33, 10838–10850.  
569 <https://doi.org/10.1021/acs.langmuir.7b02762>

570 Kubiak, K.J., Wilson, M.C.T., Mathia, T.G., Carra, S., 2011. Dynamics of contact line motion during the wetting  
571 of rough surfaces and correlation with topographical surface parameters. *Scanning* 33, 370–377.  
572 <https://doi.org/10.1002/sca.20289>

573 Kukulka, D.J., Leising, P., 2009. Evaluation of Surface Coatings on Heat Exchangers. *Chem. Eng. Trans.* 18,  
574 339–344.

575 Landolt, H., Börnstein, R., 1961. *Numerical Data and Functional Relationships in Science and Technology*. New  
576 Ser. 16, 144.

577 Langley, K.R., Green, M.L., 1989. Compression strength and fracture properties of model particulate food  
578 composites in relation to their microstructure and particle-matrix interaction. *J. Texture Stud.* 20, 191–207.  
579 <https://doi.org/10.1017/S0022029900026480>

580 MEGlobal, 2008. Ethylene Glycol Product Guide.



581 Navabpour, P., Teer, D., Su, X., Liu, C., Wang, S., Zhao, Q., Donik, C., Kocijan, A., Jenko, M., 2010.  
582 Optimisation of the properties of siloxane coatings as anti-biofouling coatings: Comparison of PACVD and  
583 hybrid PACVD–PVD coatings. *Surf. Coatings Technol.* 204, 3188–3195.  
584 <https://doi.org/10.1016/j.surfcoat.2010.03.011>

585 Pappas, C.P., Rothwell, J., 1991. The effects of heating, alone or in the presence of calcium or lactose, on calcium  
586 binding to milk proteins. *Food Chem.* 42, 183–201. [https://doi.org/10.1016/0308-8146\(91\)90033-K](https://doi.org/10.1016/0308-8146(91)90033-K)

587 Phinney, D.M., Goode, K.R., Fryer, P.J., Heldman, D., Bakalis, S., 2017. Identification of residual nano-scale  
588 foulant material on stainless steel using atomic force microscopy after clean in place. *J. Food Eng.* 214,  
589 236–244. <https://doi.org/10.1016/j.jfoodeng.2017.06.019>

590 Rabinovich, Y., Adler, J.J., Ata, A., Singh, R.K., Moudgil, B.M., 2000. Adhesion between Nanoscale Rough  
591 Surfaces. *J. Colloid Interface Sci.* 232, 10–16. <https://doi.org/10.1006/jcis.2000.7167>

592 Rosmaninho, R., Melo, L.F., 2008. Protein–calcium phosphate interactions in fouling of modified stainless-steel  
593 surfaces by simulated milk. *Int. Dairy J.* 18, 72–80. <https://doi.org/10.1016/j.idairyj.2007.06.008>

594 Rosmaninho, R., Melo, L.F., 2006a. The effect of citrate on calcium phosphate deposition from simulated milk  
595 ultrafiltrate (SMUF) solution. *J. Food Eng.* 73, 379–387. <https://doi.org/10.1016/j.jfoodeng.2005.02.017>

596 Rosmaninho, R., Melo, L.F., 2006b. Calcium phosphate deposition from simulated milk ultrafiltrate on different  
597 stainless steel-based surfaces. *Int. Dairy J.* 16, 81–87. <https://doi.org/10.1016/j.idairyj.2005.01.006>

598 Rosmaninho, R., Rizzo, G., Muller-Steinhagen, H., Melo, L.F., 2005. Anti-fouling stainless steel based surfaces  
599 for milk heating processes, in: *Proceedings of 6th International Conference on Heat Exchanger Fouling and*  
600 *Cleaning - Challenges and Opportunities.*

601 Rosmaninho, R., Santos, O., Nylander, T., Paulsson, M., Beuf, M., Benezech, T., Yiantsios, S., Andritsos, N.,  
602 Karabelas, A., Rizzo, G., Müller-Steinhagen, H., Melo, L.F., 2007. Modified stainless steel surfaces targeted  
603 to reduce fouling – Evaluation of fouling by milk components. *J. Food Eng.* 80, 1176–1187.  
604 <https://doi.org/10.1016/j.jfoodeng.2006.09.008>

605 Rulison, C., 2005. Effect of Temperature on the Surface Energy of Solids. KRUSS Application Note AN250e.

606 Santos, O., Nylander, T., Rizzo, G., Müller-Steinhagen, H., Trägårdh, C., Paulsson, M., 2003. Study of whey  
607 protein adsorption under turbulent flow rate, in: *Proceedings of Heat Exchanger Fouling and Cleaning—*

608 fundamentals and Applications. p. 24.

609 Sauerer, B., Stukan, M., Abdallah, W., Derkani, M.H., Fedorov, M., Buiting, J., Zhang, Z.J., 2016. Quantifying  
610 mineral surface energy by scanning force microscopy. *J. Colloid Interface Sci.* 472, 237–246.  
611 <https://doi.org/10.1016/j.jcis.2016.03.049>

612 Schmidt, R.H., Erickson, D.J., Sims, S., Wolff, P., 2012. Characteristics of Food Contact Surface Materials:  
613 Stainless Steel. *Food Prot. Trends* 32, 574–584.

614 Tsibouklis, J., Stone, M., Thorpe, A.A., Graham, P., Nevell, T.G., Ewen, R.J., 2000. Inhibiting bacterial adhesion  
615 onto surfaces: the non-stick coating approach. *Int. J. Adhes. Adhes.* 20, 91–96.  
616 [https://doi.org/10.1016/S0143-7496\(99\)00034-2](https://doi.org/10.1016/S0143-7496(99)00034-2)

617 Van Asselt, A.J., Vissers, M.M.M., Smit, F., De Jong, P., 2005. In-line control of fouling., *Heat Exchanger Fouling  
618 and Cleaning—Challenges and Opportunities*, Engineering Conferences International Kloster Irsee,  
619 Germany, 5-10 June (Engineering Conferences International, New York, USA).

620 Verran, J., Rowe, D.L., Cole, D., Boyd, R.D., 2000. The use of the atomic force microscope to visualise and  
621 measure wear of food contact surfaces. *Int. Biodeterior. Biodegradation* 42, 99–105.  
622 [https://doi.org/10.1016/S0964-8305\(00\)00070-6](https://doi.org/10.1016/S0964-8305(00)00070-6)

623 Wandschneider, A., Lehmann, J.K., Heintz, A., 2008. Surface Tension and Density of Pure Ionic Liquids and  
624 Some Binary Mixtures with 1-Propanol and 1-Butanol. *J. Chem. Eng. Data* 53, 596–599.  
625 <https://doi.org/10.1021/jc700621d>

626 Wenzel, R.N., 1936. Resistance of solid surfaces to wetting by water. *Ind. Eng. Chem. Res.* 28, 988.

627 Williams, A.M., Jones, J.R., Paterson, A.H.J., Pearce, D.L., 2005. Milks and milk concentrates: surface tension  
628 measurement. *Int. J. Food Eng.* 1, 1556–3758. <https://doi.org/10.2202/1556-3758.1002>

629 Wu, S., 1973. Polar and Nonpolar Interactions in Adhesion. *J. Adhes.* 5, 39–55.  
630 <https://doi.org/10.1080/00218467308078437>

631 Wu, S., 1971. Calculation of interfacial tension in polymer systems: Polymer Symposia. *J. Polym. Sci. Polym.  
632 Symp.* 34, 19–30. <https://doi.org/10.1002/polc.5070340105>

633 Zhang, P., Wang, S., Wang, S., Jiang, L., 2015. Superwetting Surfaces under Different Media: Effects of Surface  
634 Topography on Wettability. *Small nano micro* 11, 1939–1946. <https://doi.org/10.1002/sml.201401869>

635 Zhao, G., Raines, A.L., Wieland, M., Schwartz, Z., Boyan, B.D., 2007. Requirement for both micron- and  
636 submicron scale structure for synergistic responses of osteoblasts to substrate surface energy and  
637 topography. *Biomaterials* 28, 2821–2829. <https://doi.org/10.1016/j.biomaterials.2007.02.024>

638 Zhao, Q., Liu, Y., Abel, E.W., 2004. Effect of temperature on the surface free energy of amorphous carbon films.  
639 *J. Colloid Interface Sci.* 280, 174–183. <https://doi.org/10.1016/j.jcis.2004.07.004>

640 Zouaghi, S., Six, T., Nuns, N., Simon, P., Bellayer, S., Moradi, S., Hatzikiriakos, S.G., André, C., Delaplace, G.,  
641 Jimenez, M., 2018. Influence of stainless steel surface properties on whey protein fouling under industrial  
642 processing conditions. *J. Food Eng.* 228, 38–49. <https://doi.org/10.1016/j.jfoodeng.2018.02.009>

643