

# Twin-twin transfusion syndrome is associated with alterations in the metabolic profile of maternal plasma in early gestation

Yang, Yang ; Wen, Li; Han, Ting-Li; Zhang, Lan; Fu, Huijia; Gan, Jie; Saffery, Richard; Tong, Chao; Li, Junnan; Qi, Hongbo; Baker, Philip; Kilby, Mark

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1 **Twin-Twin Transfusion Syndrome Is Associated with Alterations in the**  
2 **Metabolic Profile of Maternal Plasma in Early Gestation: A Pilot Study**

3 Yang Yang<sup>1,2,3#</sup>, Li Wen<sup>1,2,3#</sup>, Ting-li Han<sup>1,2,3</sup>, Lan Zhang<sup>1,2,3</sup>, Huijia Fu<sup>4</sup>, Jie Gan<sup>1,2,</sup>  
4 <sup>3</sup>, Richard Saffery<sup>5,6</sup>, Chao Tong<sup>1,2,3\*</sup>, Junnan Li<sup>1,2,3\*</sup>, Hongbo Qi<sup>1,2,3\*</sup>, Philip N.  
5 Baker<sup>1,7</sup>, Mark D. Kilby<sup>8,9</sup>

6 <sup>1</sup>Department of Obstetrics, The First Affiliated Hospital of Chongqing Medical  
7 University, Chongqing 400016, China;

8 <sup>2</sup>International Collaborative Laboratory of Reproduction and Development, Ministry  
9 of Education, Chongqing Medical University, Chongqing 400016, China;

10 <sup>3</sup>State Key Laboratory of Maternal and Fetal Medicine of Chongqing Municipality,  
11 Chongqing, The First Affiliated Hospital of Chongqing Medical University,  
12 Chongqing 400016, China;

13 <sup>4</sup>Department of Reproduction Health and Infertility, The First Affiliated Hospital of  
14 Chongqing Medical University, Chongqing 400016, China;

15 <sup>5</sup>Cancer, Disease and Developmental Epigenetics, Murdoch Children's Research  
16 Institute, Parkville, VIC 3052, Australia;

17 <sup>6</sup>Department of Pediatrics, University of Melbourne, Parkville, VIC 3052, Australia;

18 <sup>7</sup> College of Life Sciences, University of Leicester, Leicester LE1 7RH, UK.

19 <sup>8</sup> Institute of Metabolism and System Research, University of Birmingham, Edgbaston,  
20 Birmingham, B15 2TT, UK;

21 <sup>9</sup>Fetal Medicine Centre, Birmingham Women's & Children's foundation Trust,

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22 Birmingham B152TG, UK;

23 #These authors contributed equally to this work.

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25 \*Correspondence to Dr. Chao Tong, Dr. Junnan Li and Dr. Hongbo Qi.

26 Address: Department of Obstetrics, The First Affiliated Hospital of Chongqing

27 Medical University, 1 Youyi Road, Yuzhong District, Chongqing 400016, China (H.Q

28 & J.L & C.T);

29 Tel: +86 23 89011101 (H.Q & J.L); +86 23 89011800 (C.T).

30 Fax: +86 23 89011102 (H.Q & J.L); +86 23 89011791(C.T);

31 Email: chaotongcqmu@163.com (C.T); summerbolo@163.com (J.L);

32 qihongbo728@163.com (H.Q).

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47

48 **Bulleted statements:**

49 **What's already known about this topic?**

- 50 ● Twin-twin transfusion syndrome (TTTS) is associated with fetal and perinatal  
51 mortality and early intervention by fetoscopic laser coagulation of the placental  
52 arteriovenous anastomoses may increase the chance of a favorable outcome and  
53 reduce morbidity.
- 54 ● Maternal circulating “biomarkers” for aiding the prediction of TTTS in the first  
55 trimester are still lacking.

56 **What does this study add?**

- 57 ● Based on a large prospective longitudinal twin birth cohort, we used a  
58 prospectively collected case-controlled cohort of monochorionic twins that  
59 developed TTTS, an altered metabolome of maternal plasma before 16 weeks was  
60 noted. This principally involved fatty acids, TCA cycle intermediates and amino  
61 acids those involved in angiogenesis and energy metabolism.
- 62 ● Metabolic perturbations can be detected in maternal plasma before ultrasound  
63 features that are diagnostic of TTTS, and could therefore be a promising approach

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64 for early screening of monochorionic twin pregnancies.

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66 **Availability of data and materials**

67 The LOTiS datasets used and/or analyzed during the current study are available from

68 the corresponding author on reasonable request.

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87 **Abstract**

88 **Objective:** Twin-twin transfusion syndrome (TTTS) causes perinatal mortality and  
89 morbidity in monochorionic (MC) twins. Early recognition and interventional therapy  
90 for TTTS is associated with a more favorable overall prognosis. However, the  
91 prediction by ultrasound in the first trimester has relatively poor sensitivity and  
92 specificity. This study aims to profile maternal metabolic changes before the clinical  
93 onset of TTTS and identify potential metabolic biomarkers to aid ultrasound  
94 screening.

95 **Method:** Maternal plasma was prospectively collected between 11-15 weeks of  
96 gestation in apparently uncomplicated MCDA pregnancies. This cohort was divided  
97 into: i) patients subsequently diagnosed using ultrasound with TTTS and ii)  
98 uncomplicated matched controls. Gas chromatography-mass spectrometry was used  
99 for metabolomic profiling.

100 **Results:** The levels of fatty acids, organic acids, oxaloacetic acid, and beta-alanine  
101 were significantly lower in the maternal plasma of TTTS at 11-15 weeks of gestation,  
102 while methionine and glycine were higher ( $p < 0.05$ ,  $FDR < 0.12$ ). Generally, in TTTS  
103 pregnancies, metabolism of amino acid, carbohydrate, cofactors, vitamins, and purine  
104 were 'down-regulated'; whilst bile secretion and pyrimidine metabolism were  
105 'upregulated'.

106 **Conclusions:** Metabolomics scanning of early gestation maternal plasma may  
107 identify those pregnancies that subsequently develop TTTS, especially,

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108 downregulated fatty acid level may be biologically plausible to be  
109 implicated in the pathogenesis of TTTS.

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111 **Keywords:** Twin-twin transfusion syndrome, GC-MS, metabolite, biomarker

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115 thanks for all participants who volunteered to provide blood samples.

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## 120 **1. Introduction**

121 Twin-twin transfusion syndrome (TTTS) complicates 15% of monochorionic  
122 diamniotic (MCDA) twin pregnancies and the underlying etiology and predisposition  
123 is the presence of unidirectional placental arteriovenous anastomoses causing  
124 imbalanced intertwin hemodynamics<sup>1,2</sup>. This leads to the “recipient fetus” having a  
125 “*hyperdynamic*” circulation with increased cardiac afterload with increased  
126 ventricular and systemic blood pressure<sup>3</sup>, and secondary endocrine dysfunction,  
127 involving the renin-angiotensin-aldosterone, atrial natriuretic factor and endothelin-1  
128 systems<sup>4</sup>. In contrast, the “donor fetus” rarely demonstrates significant change in  
129 cardiac function and has relatively poor perfusion, hypovolemia with associated  
130 oliguria and oligohydramnios<sup>5</sup>. If untreated, TTTS leads to perinatal death before 26  
131 weeks in over 90% of diagnosed cases<sup>4</sup>. It is also associated with severe fetal  
132 morbidity and neurodevelopmental sequelae, preterm birth, growth restriction,  
133 cardiomyopathy, and hydrops fetalis. Fetoscopic laser coagulation (FLC) of the  
134 pathologic placental vascular anastomoses is the optimal fetal therapy, leading to  
135 survival of at least one fetus in 90% of pregnancies and minimizing both preterm birth  
136 and neurologic morbidity<sup>5,6,7</sup>.

137

138 Early prediction of ‘at risk’ monochorionic twin pregnancies to aid the timely  
139 diagnosis of TTTS is clinically important, as early intervention, before there is  
140 significant cardiac dysfunction, may result in a more favorable overall prognosis<sup>8</sup>

141 There has been much interest in the role of first trimester (11<sup>+0</sup> - 13<sup>+6</sup> weeks)



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142 ultrasound biometric measurement of fetal crown rump length (CRL) and nuchal  
143 translucency (NT) in the prediction of adverse outcome in monochorionic twinning.  
144 However, the individual measurements of CRL and NT and the intertwin differences  
145 have only a moderate predictive value for the development of TTTS, selective growth  
146 restriction (sGR) or indeed fetal demise; only 52% of cases deemed high risk  
147 ultimately develop TTTS<sup>9</sup>. This may be further refined by measurement of Ductus  
148 Venosus velocimetry<sup>10</sup> (as noted in small cohort studies) but overall the predictive  
149 value of first trimester ultrasound alone is disappointing. In the second trimester, the  
150 ultrasound detection of mild discordance in deepest vertical pool measurement in the  
151 amniotic sacs has a moderate ability to predict later onset of TTTS but again, as in the  
152 first trimester, sensitivity and specificity of prediction is not high<sup>11,12</sup>. Therefore, to  
153 improve early detection, novel approaches for screening are required using other  
154 modalities in addition to ultrasound.

155

156 Metabolomics is a powerful tool for investigating the final downstream products of  
157 genotype and environmental interactions and may be employed to investigate the  
158 complex interactions between specific metabolites and others as well as disease<sup>13</sup>. In a  
159 comparison of pre- and post-laser coagulation, Dunn and colleagues observed  
160 perturbations in carbohydrate and fatty acid levels in the fetal “recipient” amniotic  
161 fluid of monochorionic pregnancies complicated by TTTS<sup>14</sup>, indicating that this  
162 pathological process may be associated with aberrant metabolism. Moreover, a  
163 profound change in the first-trimester metabolite profile of maternal plasma has been

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164 noted in women who subsequently developed early-onset preeclampsia<sup>15</sup>, indicating  
165 that metabolic changes in the first trimester may be an indicator of the occurrence or  
166 development of diseases.

167

168 In this ‘pilot’ study, Gas Chromatography-Mass Spectrometry (GC-MS) was used in a  
169 metabolomics analysis of plasma collected at 11-15 weeks of gestation from pregnant  
170 women with monochorionic, diamniotic twin pregnancies (identified using ultrasound)  
171 that subsequently developed TTTS later in gestation. The finding from biochemical  
172 compounds and metabolic networks may enhance understanding of the maternal  
173 metabolic changes in early pregnancy associated with the occurrence of TTTS.

174

## 175 **2. Methods**

### 176 **2.1. Patient recruitment**

177 430 women with twin pregnancies were recruited in the Chongqing Longitudinal  
178 Twin Study (LoTiS)<sup>16</sup>, which has been registered with the Chinese Clinical Trial  
179 Registry (ChiCTR-OOC-16008203). Of the 430 twin pregnancies, 117 were  
180 monochorionic diamniotic (MCDA), and 7 of the MCDA twin pregnancies developed  
181 TTTS (6%) (Figure 1). High-resolution fetal ultrasound was used to diagnose TTTS.  
182 The diagnosis was made using international criteria; if a monochorionic pregnancy  
183 had markedly discordant amniotic fluid volumes by a deepest vertical pool  
184 (DVP) >10 cm (100mm) at >20<sup>+0</sup> weeks’ and >8 cm (80mm) at <20<sup>+0</sup> weeks’  
185 gestation in the recipient twin together with a DVP <2 cm (20mm) in the co-twin<sup>17</sup>.

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186 However, sGR was diagnosed when one twin had an estimated fetal weight of  $<10\%$ <sup>18</sup>.  
187 As there are different diagnostic criteria for TTTS and sGR, sGR was excluded.  
188 Uncomplicated MCDA twin pregnancies were assigned to the non-TTTS group as  
189 controls; the gestation age was matched between TTTS and control groups when the  
190 plasma sample was collected in the first trimester (always in the morning with similar  
191 timing in the TTTS and control groups). The control group excluded MCDA twin  
192 pregnancies with the serious complications, of sGR, malformation and of course TTTS  
193 (Figure 1). Participants were recruited at a median gestation of  $13^{+1}$  (range  $12^{+4}$  -  $13^{+5}$ )  
194 weeks of gestation; five of the seven patients were diagnosed with Quintero stage I  
195 TTTS in the second trimester at a median gestation of  $19^{+5}$  ( $19, 22^{+6}$ ) week<sup>19</sup>. These  
196 MCDA twins were treated by fetoscopic laser coagulation and the outcome  
197 monitored until delivery (with overall 80% perinatal survival). A further two MCDA  
198 twin pregnancies were complicated by Stage III TTTS, diagnosed in the third  
199 trimester at  $34^{+1}$  or  $36^{+1}$  weeks and were delivered by cesarean section; all fetuses  
200 survived (100% survival).

201

## 202 **2.2 Collection of samples**

203 Maternal blood samples were collected into tubes with ethylene diamine tetra-acetic  
204 acid (EDTA) between 11-15 weeks (in the mornings) of gestation and then  
205 centrifuged at  $4^{\circ}\text{C}$  and 3000 rpm for 10 min. The plasma was stored at  $-80^{\circ}\text{C}$  for  
206 future use.

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### 208 **2.3 Sample preparation**

209 To extract the metabolites from plasma, 100  $\mu\text{l}$  of plasma was added into the tubes  
210 with 400  $\mu\text{l}$  of pre-chilled methanol; an internal standard (20  $\mu\text{l}$  of 2,3,3,3- $\text{d}_4$ -alanine  
211 (10 mM, Sigma-Aldrich, Missouri, USA)) was then added. The mixture was vortexed  
212 for 1 min and incubated at  $-20^\circ\text{C}$  for 30 min to precipitate protein. The supernatant  
213 was collected after centrifugation (17,000 g, 15 min), dried by using a SpeedVac  
214 (Labconco, Kansas, USA), and stored at  $-20^\circ\text{C}$  for derivatization.

215

### 216 **2.4 Methyl chloroformate derivatization and GC-MS analysis**

217 Samples were chemically derivatized using a methyl chloroformate (MCF) method to  
218 extract metabolites according to the protocol published by Smart *et al*<sup>20</sup>. Briefly, 200  
219  $\mu\text{L}$  of sodium hydroxide (1 M) was added to the dried samples, and 167  $\mu\text{L}$  of  
220 methanol and 34  $\mu\text{L}$  of pyridine were added. Then, 20  $\mu\text{L}$  of MCF was added with 30s  
221 of vortexing, and the addition of another 20  $\mu\text{L}$  of MCF was followed by 30s of  
222 vortexing. Then, 400  $\mu\text{L}$  of chloroform and 400  $\mu\text{L}$  of sodium bicarbonate (50 mM)  
223 were added and vortexed for 10s. The lower chloroform phase was used for GC-MS  
224 analysis.

225

226 The samples were analyzed in a GC7890 system with an MSD5975 mass selective  
227 detector (Agilent, California, USA). The MSD5975 mass selective detector (Agilent)  
228 was a ZB-1701 GC capillary column (30 m x 250  $\mu\text{m}$  id x 0.15  $\mu\text{m}$  with 5-m guard  
229 column, Phenomenex, California, USA). One microliter of sample was injected into

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230 the GC inlet. Helium gas was the carrier gas flow at 1 mL/min. The auxiliary  
231 temperature, MS quadrupole, and MS source were 250°C, 230°C, and 150°C,  
232 respectively, and the scan speed was 1.562 u/s.

233

## 234 **2.5 Data analysis and statistics**

235 Automated Mass Spectral Deconvolution & Identification System (AMDIS) software  
236 was used to de-convolute GC-MS chromatograms and identify metabolites with our  
237 in-house MCF. The metabolite values were normalized to the abundance of the  
238 internal standard and total ion count. Significant metabolites were analyzed with the  
239 limma R package. Principal component analysis (PCA) was based on data from all  
240 metabolites to visualize the differences between TTTS and control group. Log  
241 transformation was used to make the distributions of the data more similar to a  
242 Gaussian distribution. Mann–Whitney U-test, Chi square test, Student’s t-test and box  
243 plot were used in R and SPSS 24.0. The discrepancy in the CRL of the twins was  
244 calculated by subtracting the CRL of the smaller fetus from that of the larger one and  
245 expressing this as a quotient of the larger fetus CRL<sup>21</sup>. Power analysis was applied in  
246 website (<https://www.metaboanalyst.ca/>). PAPI enrichment approach was applied to  
247 compare metabolic pathways activities based on R-software package, then t-test was  
248 used to investigate difference between TTTS and control group <sup>22</sup>. The metabolic  
249 pathways with p-values less than 0.05 and q-values less than 0.30 were considered  
250 statistically significant. Based on KEGG database, Cytoscape <sup>23</sup> was used to visualize  
251 and interpret metabolomic data in the context of metabolic networks.

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### 252 **3. Results**

#### 253 *Population characteristics*

254 Seven TTTS pregnancies and seven control MCDA pregnancies from LoTiS were  
255 included in this pilot study. A retrospective calculation of a sample size of 7 cases and  
256 7 controls available in this study, indicated we had a power of 0.67 to detect a  
257 difference of 1.2 folds in metabolite abundance at a false discovery rate of 5%  
258 (supplementary Figure 1). The gestational age at diagnosis of the five second  
259 trimester TTTS patients was 19<sup>+5</sup> weeks (19, 22<sup>+6</sup>). The other two patients were  
260 diagnosed with late-onset acute TTTS stage III at 34<sup>+1</sup> and 36<sup>+6</sup> weeks', respectively.  
261 Clinical maternal and fetal characteristics are outlined in Table 1. There were no  
262 significant differences in maternal age, nuchal translucency (NT), crown–rump length  
263 (CRL) discrepancy, gestational age and fetal gender, between the two groups at  
264 sample collection. Maternal pre-pregnancy BMI, CRL, gestational age at delivery  
265 (32<sup>+2</sup>-35<sup>+3</sup>, 37<sup>+2</sup> -37<sup>+3</sup> weeks'), larger and smaller amniotic fluid volume, amniotic  
266 fluid volume difference, and birth weight of the large fetus were significantly different  
267 between TTTS and control groups.

268

#### 269 *Metabolome of maternal plasma in the control versus TTTS group*

270 In the maternal plasma samples that were collected between the 11<sup>th</sup> and 15<sup>th</sup> weeks of  
271 gestation, there were over 200 individual spectral peaks separated by gas  
272 chromatography. PCA analysis showed that the metabolites were notably separated  
273 (Figure 2). Significance is assessed via moderated t-tests implemented in the limma

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274 package of Bioconductor, 17 metabolites were significantly differed in plasma from  
275 the control and TTTS groups (Figure 3). Specifically, the levels of saturated fatty  
276 acids (palmitic acid, pentadecanoic acid and myristic acid), unsaturated fatty acids  
277 (9-heptadecenoic acid and linolelaidic acid), organic acids (4-aminobutyric acid,  
278 D-fumaric acid, 2-methyloctadecanoic acid and cabamic acid), TCA cycle and  
279 intermediate acids (oxaloacetic acid), alkanes (pentadecane), nicotinamide, an amino  
280 acid derivative (beta-alanine) and cysteine were decreased in the TTTS group  
281 compared to the control group, while the levels of amino acid (methionine and glycine)  
282 were increased in the TTTS group.

283

284 **Disrupted metabolic pathways in the maternal plasma of TTTS pregnancies**  
285 **from 11-16 weeks of gestation**

286 To elucidate the metabolic pathways that were different in pregnancies complicated  
287 by TTTS, all of the identified metabolites were subjected to pathway analysis, and the  
288 data suggested that 13 metabolic pathways were significantly altered in  
289 TTTS-complicated pregnancies from 11-15 weeks of gestation (Figure 4a): amino  
290 acid metabolism, carbohydrate metabolism, cofactors, vitamins, and purine were  
291 downregulated, and bile secretion and pyrimidine metabolism were upregulated. A  
292 metabolic network was constructed based on these results (Figure 4b). Intriguingly,  
293 glycine and cysteine were the nexus of the network. The correlation between  
294 identified metabolites and metabolic pathways is illustrated in Figure 4c, in which the  
295 involved pathway is predicted. Taken together, these findings suggest that

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296 perturbations in metabolic pathways of maternal blood during early gestation may be  
297 associated with subsequent TTTS development.

298

### 299 **Discussion**

300 Pregnancy is characterized by its complex metabolic processes that can impact fetal  
301 development<sup>24</sup>. Given the potential high fetal mortality of TTTS without appropriate  
302 fetal therapy, clinical evidence suggests that the optimal time frame for fetoscopy in  
303 TTTS is 16-26 weeks of gestation.<sup>6</sup> Predictive metabolic biomarkers (before 16  
304 weeks) may aid timely identification of “at risk” monochorionic twin pregnancies and a  
305 potentially diagnosis of TTTS at a lower Quintero stage with improved double survival  
306 rates<sup>8</sup>. This ‘proof of principle study’ noted metabolic perturbations in maternal plasma  
307 at 11-15 weeks of gestation in apparently uncomplicated monochorionic, diamniotic  
308 twin pregnancies prior to the subsequent ultrasound diagnosis of TTTS.

309

310 Dunn and colleagues have previously reported higher levels of fatty acids and lower  
311 levels of carbohydrates in amniotic fluid of post-FLC; they suggested that metabolic  
312 disorders existed in TTTS, and the changes were associated with fetal or placental  
313 energy metabolism and the echocardiographic measures of recipient cardiac  
314 function<sup>14</sup>.

315

316 A role for fatty acids in the pathogenesis of TTTS is biologically plausible. Fatty acids  
317 can be imported, oxidized, synthesized, and exported by endothelial cells, and fatty



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318 acid oxidation contributes to approximately 5% of total cellular ATP production in  
319 endothelial cells<sup>25</sup>. Oxidation of fatty acids is irreplaceable for endothelial cell  
320 proliferation because carbons are provided for de novo nucleotide synthesis<sup>26</sup>. Hence,  
321 fatty acids are a metabolic determinant of the angiogenic process. Many studies have  
322 shown that fatty acids are key to pregnancy outcomes. Lipid metabolic genes are  
323 upregulated in the first trimester, and fatty acid uptake by placental trophoblast cells is  
324 associated with angiogenic processes<sup>27</sup>. Fatty acids are not only a substantial energy  
325 source but also have effects on membrane fluidity, intracellular cell-signaling cascades  
326 and susceptibility to oxidative injury<sup>28</sup>. Linolelaidic acid has been shown to exhibit a  
327 proliferative effect on human umbilical vein smooth muscle cells<sup>29</sup>. Trophoblasts  
328 oxidize palmitate and myristate for energy<sup>30</sup>. During normal pregnancy, vascular  
329 remodeling, fatty acid synthesis and transport, and lipid droplet formation are  
330 increased<sup>31</sup>. Fatty acids are key to angiogenesis and thus play an important role in  
331 human placentation<sup>32,33</sup>. Maternal angiogenic activity is decreased in severe TTTS<sup>34</sup>.  
332 However, in our study, we found that the decrease of saturated fatty acids and  
333 unsaturated fatty acids in TTTS group, which indicated that the utilization of fatty  
334 acids in TTTS pregnant women was increased. Increased oxidative utilization of fatty  
335 acids can provide energy for vascular remodeling of placenta. Fatty acids are  
336 important components of cell membrane. Therefore, increased utilization of fatty  
337 acids in the first trimester can promote the process of pathological placental vascular  
338 anastomosis in TTTS patients, and it may develop into an early metabolic disorder  
339 that disrupts placental angiogenesis and the fetal energy supplies.

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341  
342 Recently, amino acid metabolism was found to play important roles in regulating and  
343 maintaining various aspects of vascular function, such as vascular tone, coagulation  
344 and fibrinolysis, cell growth and differentiation, redox homeostasis, and immune and  
345 inflammatory responses<sup>35</sup>. In this study, we found that methionine and glycine were  
346 upregulated in the maternal plasma of pregnancies subsequently complicated by  
347 TTTS and that these metabolites positively correlated with abnormal amniotic fluid  
348 depth. Alterations in amino acid levels between donor and recipient twins in TTTS  
349 and between non-TTTS twin pairs have previously been reported, due to intertwin  
350 transfusion that impairs placental transport of amino acids<sup>36</sup>. In placental insufficiency,  
351 maternal cysteine and methionine levels are decreased, and maternal glycine is  
352 increased<sup>37</sup>. Amino acids enhance protein synthesis and cell proliferation in the  
353 placenta<sup>38</sup>. Glycine can promote angiogenesis<sup>39</sup>. Methionine, a sulfur-containing  
354 amino acid, is a major component in protein synthesis<sup>35</sup>, and blockade of methionine  
355 metabolism in endothelial cells leads to endothelial damage or inflammation<sup>40</sup>. In the  
356 metabolome of amniotic fluid, higher levels of carbohydrates in TTTS pregnancies  
357 before fetoscopic laser coagulation treatment and higher levels of fatty acids  
358 post-treatment<sup>14</sup>, indicate that this treatment can partially correct metabolic changes.  
359 These studies suggest that amino acids are associated with placentation and  
360 angiogenesis. In addition, upregulated methionine and glycine levels may be a sign  
361 that indicates early changes in the metabolic microenvironment during the first

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362 trimester in women whose pregnancies are subsequently complicated by TTTS.

363

364 Because of the low incidence of TTTS in monochorionic twin pregnancies (~10%), it  
365 is difficult to investigate the manifestations of TTTS risk in early pregnancy maternal  
366 blood. A prospective study in a large cohort of monochorionic twins pregnancies  
367 would be required and the improved identification of ‘at risk’ pregnancies when  
368 metabolic profiles are used alone or in combination with fetal ultrasound variable  
369 would be desirable<sup>41</sup>. The aim of this work was to evaluate the prognostic ability of  
370 individual first-trimester metabolomic biomarkers for TTTS in MC twins. For those  
371 metabolomic biomarkers where there was a statistically significant independent  
372 association with adverse outcome, calculating the absolute risk for each outcome  
373 using common values demonstrated that the markers may be useful clinically,  
374 although this would depend on a change in management consequent on the risk  
375 assessment<sup>42</sup>.

### 376 **Limitations**

377 As the lower statistic power, we should expand a significantly larger prospective  
378 study of monochorionic twins to enable the development of a prognostic model.  
379 Secondly, another limit is that we only tested metabolites of maternal blood, rather the  
380 blood metabolome not likely reflects disrupted metabolism in unspecified uterus and  
381 fetal development. Thirdly, our study was not a randomized controlled study, but the  
382 results also could provide scientific support for future large randomized controlled  
383 cohort studies. Fourthly, early diagnosis is of great significance for the detection of

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384 clinical diseases, but we cannot exclude that monochorionic twins will not develop to  
385 TTTS just because there is no difference in metabolomic of blood samples in the first  
386 trimester of pregnancy. The next step of our future work is to replicate the study in  
387 independent samples. Finally, the study should be performed and analyzed according  
388 to robust prognostic methodology, which has been reported by McShane and  
389 colleagues for predictive markers studies<sup>43</sup>.

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### 391 **Conclusions**

392 Although the idea that TTTS can potentially be detected using this methodology is  
393 exciting, to our knowledge, this is the first report of a metabolomic profile with  
394 human plasma for predicting the occurrence of TTTS before 16 weeks. This study  
395 provides insights into the relationship between early metabolic changes and the  
396 occurrence of TTTS. Based on our results, we found that amino acid metabolism,  
397 carbohydrate metabolism and lipid metabolism were altered and that the abnormal  
398 changes may be related to subsequent placental development and angiogenesis.

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### 400 **Authors' contributions**

401 C.T, J.L and H.Q conceived the study, which was discussed with RS, MDK and PB at  
402 regular Chongqing 111 study meetings; Y.Y, L.W, T.H performed experiments and  
403 analyzed the data; L.Z, H.F and J.G collected samples and clinical data; Y.Y and C.T  
404 drafted the manuscript; R.S, M.D.K and P.B commented on the data and revised the  
405 manuscript.

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407 **Ethics approval and consent to participate**

408 This study was approved by The Ethics Committee of The First Affiliate Hospital of  
409 Chongqing Medical University (approval number: 201530; date of approval:  
410 2/29/2016). And each participant will sign the informed consent before  
411 re-investigation.

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413 **Consent for publication:** Not applicable.

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**416 References**

- 417 1. Society for Maternal-Fetal M, Simpson LL. Twin-twin transfusion syndrome. *Am J Obstet*  
418 *Gynecol.* 2013;208(1):3-18.
- 419 2. Simpson LL. What you need to know when managing twins: 10 key facts. *Obstet Gynecol*  
420 *Clin North Am.* 2015;42(2):225-239.
- 421 3. Van Mieghem T, Klaritsch P, Done E, et al. Assessment of fetal cardiac function before  
422 and after therapy for twin-to-twin transfusion syndrome. *Am J Obstet Gynecol.*  
423 2009;200(4):400 e401-407.
- 424 4. Lewi L, Jani J, Blickstein I, et al. The outcome of monochorionic diamniotic twin gestations  
425 in the era of invasive fetal therapy: a prospective cohort study. *Am J Obstet Gynecol.*  
426 2008;199(5):514 e511-518.
- 427 5. Behrendt N, Galan HL. Twin-twin transfusion and laser therapy. *Curr Opin Obstet*  
428 *Gynecol.* 2016;28(2):79-85.
- 429 6. Sago H, Ishii K, Sugibayashi R, Ozawa K, Sumie M, Wada S. Fetoscopic laser  
430 photocoagulation for twin-twin transfusion syndrome. *J Obstet Gynaecol Res.*  
431 2018;44(5):831-839.
- 432 7. Bautista TN, Krebs TL, Jnah A, Newberry D. Twin-to-Twin Transfusion Syndrome: A Case  
433 Report. *Neonatal Netw.* 2018;37(5):292-302.
- 434 8. Chmait RH, Kontopoulos EV, Korst LM, Llanes A, Petisco I, Quintero RA. Stage-based  
435 outcomes of 682 consecutive cases of twin-twin transfusion syndrome treated with laser  
436 surgery: the USFetus experience. *Am J Obstet Gynecol.* 2011;204(5):393 e391-396.
- 437 9. Mackie FL, Morris RK, Kilby MD. The prediction, diagnosis and management of  
438 complications in monochorionic twin pregnancies: the OMMIT (Optimal Management of  
439 Monochorionic Twins) study. *BMC Pregnancy Childbirth.* 2017;17(1):153.
- 440 10. Matias A, Montenegro N, Loureiro T, et al. Screening for twin-twin transfusion syndrome  
441 at 11-14 weeks of pregnancy: the key role of ductus venosus blood flow assessment.  
442 *Ultrasound Obstet Gynecol.* 2010;35(2):142-148.
- 443 11. Sebire NJ, Souka A, Skentou H, Geerts L, Nicolaides KH. Early prediction of severe  
444 twin-to-twin transfusion syndrome. *Hum Reprod.* 2000;15(9):2008-2010.
- 445 12. Mosquera C, Miller RS, Simpson LL. Twin-twin transfusion syndrome. *Semin Perinatol.*  
446 2012;36(3):182-189.
- 447 13. Dunn WB, Broadhurst DI, Atherton HJ, Goodacre R, Griffin JL. Systems level studies of  
448 mammalian metabolomes: the roles of mass spectrometry and nuclear magnetic  
449 resonance spectroscopy. *Chem Soc Rev.* 2011;40(1):387-426.
- 450 14. Dunn WB, Allwood JW, Van Mieghem T, et al. Carbohydrate and fatty acid perturbations  
451 in the amniotic fluid of the recipient twin of pregnancies complicated by twin-twin  
452 transfusion syndrome in relation to treatment and fetal cardiovascular risk. *Placenta.*  
453 2016;44:6-12.
- 454 15. Bahado-Singh RO, Akolekar R, Mandal R, et al. Metabolomics and first-trimester  
455 prediction of early-onset preeclampsia. *J Matern Fetal Neonatal Med.*  
456 2012;25(10):1840-1847.
- 457 16. Tong C, Wen L, Xia Y, et al. Protocol for a longitudinal twin birth cohort study to unravel  
458 the complex interplay between early-life environmental and genetic risk factors in health

- 459 and disease: the Chongqing Longitudinal Twin Study (LoTiS). *BMJ Open*.  
460 2018;8(2):e017889.
- 461 17. Stirnemann J, Djaafri F, Kim A, et al. Preterm premature rupture of membranes is a  
462 collateral effect of improvement in perinatal outcomes following fetoscopic coagulation  
463 of chorionic vessels for twin-twin transfusion syndrome: a retrospective observational  
464 study of 1092 cases. *BJOG*. 2018;125(9):1154-1162.
- 465 18. Chon AH, Ma SY, Korst LM, Chmait HR, Purnell ME, Chmait RH. Antenatal course of  
466 referred monochorionic diamniotic twins complicated by selective intrauterine growth  
467 restriction (SIUGR) type III. *J Matern Fetal Neonatal Med*. 2019:1-7.
- 468 19. Quintero RA, Dickinson JE, Morales WJ, et al. Stage-based treatment of twin-twin  
469 transfusion syndrome. *Am J Obstet Gynecol*. 2003;188(5):1333-1340.
- 470 20. Smart KF, Aggio RB, Van Houtte JR, Villas-Boas SG. Analytical platform for metabolome  
471 analysis of microbial cells using methyl chloroformate derivatization followed by gas  
472 chromatography-mass spectrometry. *Nat Protoc*. 2010;5(10):1709-1729.
- 473 21. Bhide A, Sankaran S, Sairam S, Papageorghiou AT, Thilaganathan B. Relationship of  
474 intertwin crown-rump length discrepancy to chorionicity, fetal demise and birth-weight  
475 discordance. *Ultrasound Obstet Gynecol*. 2009;34(2):131-135.
- 476 22. Aggio RB, Ruggiero K, Villas-Boas SG. Pathway Activity Profiling (PAPi): from the  
477 metabolite profile to the metabolic pathway activity. *Bioinformatics*.  
478 2010;26(23):2969-2976.
- 479 23. Gao J, Tarcea VG, Karnovsky A, et al. Metscape: a Cytoscape plug-in for visualizing and  
480 interpreting metabolomic data in the context of human metabolic networks.  
481 *Bioinformatics*. 2010;26(7):971-973.
- 482 24. Lindsay KL, Hellmuth C, Uhl O, et al. Longitudinal Metabolomic Profiling of Amino Acids  
483 and Lipids across Healthy Pregnancy. *PLoS One*. 2015;10(12):e0145794.
- 484 25. Patella F, Schug ZT, Persi E, et al. Proteomics-based metabolic modeling reveals that  
485 fatty acid oxidation (FAO) controls endothelial cell (EC) permeability. *Mol Cell Proteomics*.  
486 2015;14(3):621-634.
- 487 26. Harjes U, Kalucka J, Carmeliet P. Targeting fatty acid metabolism in cancer and  
488 endothelial cells. *Crit Rev Oncol Hematol*. 2016;97:15-21.
- 489 27. Basak S, Duttaroy AK. Effects of fatty acids on angiogenic activity in the placental  
490 extravillous trophoblast cells. *Prostaglandins Leukot Essent Fatty Acids*.  
491 2013;88(2):155-162.
- 492 28. Wathes DC, Abayasekara DR, Aitken RJ. Polyunsaturated fatty acids in male and female  
493 reproduction. *Biol Reprod*. 2007;77(2):190-201.
- 494 29. Li XP, Luo T, Li J, et al. Linolelaidic acid induces a stronger proliferative effect on human  
495 umbilical vein smooth muscle cells compared to elaidic acid. *Lipids*. 2013;48(4):395-403.
- 496 30. Shekhawat P, Bennett MJ, Sadovsky Y, Nelson DM, Rakheja D, Strauss AW. Human  
497 placenta metabolizes fatty acids: implications for fetal fatty acid oxidation disorders and  
498 maternal liver diseases. *Am J Physiol Endocrinol Metab*. 2003;284(6):E1098-1105.
- 499 31. Johnsen GM, Basak S, Weedon-Fekjaer MS, Staff AC, Duttaroy AK. Docosahexaenoic acid  
500 stimulates tube formation in first trimester trophoblast cells, HTR8/SVneo. *Placenta*.  
501 2011;32(9):626-632.
- 502 32. Kaufmann P, Mayhew TM, Charnock-Jones DS. Aspects of human fetoplacental

- 
- 503            vasculogenesis and angiogenesis. II. Changes during normal pregnancy. *Placenta*.  
504            2004;25(2-3):114-126.
- 505    33.        Cahill LS, Rennie MY, Hoggarth J, et al. Feto- and utero-placental vascular adaptations to  
506            chronic maternal hypoxia in the mouse. *J Physiol*. 2018;596(15):3285-3297.
- 507    34.        Fox CE, Lash GE, Pretlove SJ, Chan BC, Holder R, Kilby MD. Maternal plasma and amniotic  
508            fluid angiogenic factors and their receptors in monochorionic twin pregnancies  
509            complicated by twin-to-twin transfusion syndrome. *Ultrasound Obstet Gynecol*.  
510            2010;35(6):695-701.
- 511    35.        Oberkersch RE, Santoro MM. Role of amino acid metabolism in angiogenesis. *Vascul*  
512            *Pharmacol*. 2019;112:17-23.
- 513    36.        Bajoria R, Hancock M, Ward S, D'Souza SW, Sooranna SR. Discordant amino acid profiles  
514            in monochorionic twins with twin-twin transfusion syndrome. *Pediatr Res*.  
515            2000;48(6):821-828.
- 516    37.        Pogorelova TN, Gunko VO, Nikashina AA, et al. [Influence of amino acid imbalance in  
517            maternal and fetal organisms on the development of placental insufficiency and the  
518            course of the neonatal period.]. *Klin Lab Diagn*. 2018;63(10):610-614.
- 519    38.        Wu G, Bazer FW, Burghardt RC, et al. Impacts of amino acid nutrition on pregnancy  
520            outcome in pigs: mechanisms and implications for swine production. *J Anim Sci*.  
521            2010;88(13 Suppl):E195-204.
- 522    39.        Guo D, Murdoch CE, Xu H, et al. Vascular endothelial growth factor signaling requires  
523            glycine to promote angiogenesis. *Sci Rep*. 2017;7(1):14749.
- 524    40.        Vijaya Lakshmi SV, Naushad SM, Rupasree Y, Seshagiri Rao D, Kutala VK. Interactions of  
525            5'-UTR thymidylate synthase polymorphism with 677C --> T methylene tetrahydrofolate  
526            reductase and 66A --> G methyltetrahydrofolate homocysteine methyl-transferase  
527            reductase polymorphisms determine susceptibility to coronary artery disease. *J*  
528            *Atheroscler Thromb*. 2011;18(1):56-64.
- 529    41.        Riley RD, Hayden JA, Steyerberg EW, et al. Prognosis Research Strategy (PROGRESS) 2:  
530            prognostic factor research. *PLoS Med*. 2013;10(2):e1001380.
- 531    42.        Mackie FL, Whittle R, Morris RK, Hyett J, Riley RD, Kilby MD. First-trimester ultrasound  
532            measurements and maternal serum biomarkers as prognostic factors in monochorionic  
533            twins: a cohort study. *Diagn Progn Res*. 2019;3:9.
- 534    43.        McShane LM, Altman DG, Sauerbrei W, et al. Reporting recommendations for tumor  
535            marker prognostic studies (REMARK). *J Natl Cancer Inst*. 2005;97(16):1180-1184.

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539 Table 1. Clinical characteristics of subjects.

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583 **Figure legend**

584 **Figure 1. Flowchart showing selection of TTTS and control participants**

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605 **Figure 2. Principal component analysis (PCA) of TTTS and controls.**

606 PC1 vs. PC2. Score plot shows separation between TTTS cases (green) and controls  
607 (red) based on significant differences in metabolite abundance ( $p < 0.01$ ) between the  
608 groups (n=7 in each group).

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627 **Figure 3. Altered metabolites in the maternal plasma of TTTS versus control**  
628 **pregnancies between 12 and 15 weeks of gestation.**

629 The boxplots show the significant metabolites with their relative concentrations. Blue  
630 boxes indicate TTTS group, red boxes indicate Control group. Only the metabolites  
631 with p-values less than 0.05 and q-values less than 0.12 are displayed in the figure.

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649 **Figure 4. Predicted alterations in metabolic pathways in TTTS between 11 and**  
650 **16 weeks of gestation.**

651 A. Predicted metabolic changes in TTTS compared to normal controls; red represents  
652 increased metabolism, whereas green represents downregulated metabolism. Only the  
653 metabolic pathways with p-values and q-values less than 0.05 are displayed in the  
654 heatmap. B. The metabolic network plotted according to the metabolites identified in  
655 maternal plasma collected at 12-15 weeks of gestational age is shown. The red dots  
656 represent metabolites that were significantly upregulated in TTTS, while the blue dots  
657 represent metabolites that were significantly downregulated in TTTS. The yellow dots  
658 represent the metabolites in plasma that showed no differences between TTTS and  
659 normal controls. The smaller yellow circles are unidentified metabolites that were  
660 directly connected to the identified metabolites. C. Circles indicate the connections  
661 between metabolic pathways and metabolites.