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Subsequent Primary Neoplasms

Risks, Risk Factors, Surveillance, and Future Research



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KEYWORDS

- Subsequent primary neoplasms • Second malignant neoplasms
- Second primary cancers • Secondary cancers • Radiotherapy • Chemotherapy
- Genetic variation • Genetic susceptibility • Surveillance • Screening
- Follow-up guidelines

KEY POINTS

- Risks of subsequent primary neoplasms after childhood, teenage, and young adult cancer are provided from large-scale cohorts which yield the most reliable estimates.
- Radiotherapy and chemotherapy for childhood cancer are each evaluated as a risk factor for subsequent primary neoplasms.
- New investigations are evaluating whether genomic variants modify treatment-related subsequent neoplasm risk among childhood cancer survivors.

Continued

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Continued

- Surveillance, screening, and clinical follow-up guidelines for subsequent primary neoplasms after childhood cancer are each considered.
- Priorities for future research concerning subsequent primary neoplasms after childhood cancer are briefly summarized.

RISKS OF SUBSEQUENT PRIMARY NEOPLASMS AFTER CHILDHOOD, ADOLESCENT, AND YOUNG ADULT CANCER

Survivors of childhood cancer experience substantial premature mortality; in the British Childhood Cancer Survivor Study (BCCSS) cohort, by 50 years from diagnosis 30% of 5-year survivors had died when 6% were expected to have died from mortality rates in the general population.¹ Analysis of the same cohort revealed that among survivors at least 45 years from diagnosis 51% of excess number of deaths were caused by subsequent primary neoplasm (SPN).¹ However, efforts to reduce therapeutic exposures in more recent decades has contributed to a decline in late mortality in general and from SPN in particular, among 5-year survivors of childhood cancer.²

In this article the authors consider the risks of SPN after childhood cancer and compare these risks with those observed after adolescent and young adult (AYA) cancer; the carcinogenic impact of treatment of childhood cancer with radiotherapy and chemotherapy; the influence of inherited genetic susceptibility on the development of SPNs; and the role of surveillance, screening, and clinical follow-up guidelines.

Risks of Subsequent Primary Neoplasms After Childhood Cancer

The most reliable estimates of risk of SPN result from large-scale cohort studies with systematic long-term follow-up. Two large-scale population-based registry ascertained cohorts of childhood cancer survivors have been established: one in the United Kingdom, the BCCSS,³⁻⁵ and another including all of the Nordic countries.⁶ Three large-scale hospital-based cohorts of childhood cancer survivors have also been established: the North American Childhood Cancer Survivor Study (CCSS),⁷⁻⁹ the Dutch Childhood Cancer Oncology Group—Long-Term Effects After Childhood Cancer (DCOG-LATER) cohort,^{10,11} and the French Childhood Cancer Survivor Study (FCCSS).^{12,13} A recent review of risk estimates resulting from the initial 4 of these cohorts concluded that beyond age 40 years the standardized incidence ratio (SIR) was consistently at least 2-fold that expected and the absolute excess risk (AER) increased with attained age.¹⁴ Both SIRs and AERs were similar across cohorts younger than 40 years.¹⁴

Types of SPN observed in excess of that expected from the general population varies substantially by both attained age and interval from diagnosis. For example, within the BCCSS brain tumors and sarcomas, as an SPN accounted for 63% of the excess number of SPNs observed among survivors aged 5 to 19 years; in contrast 52% of the excess number of SPNs observed among survivors older than 40 years were carcinomas of digestive, genitourinary, respiratory, and breast sites.³

A pan-European collaboration has been initiated to exploit the advantages that Europe has relating to the establishment of population-based cancer registration in the Nordic countries and United Kingdom during the 1940s, 1950s, and 1960s, depending on the country. In the PanCare Childhood and Adolescent Cancer Survivor Care and Follow-up Studies (PanCareSurFup) SPN cohort comprises the largest ever

assembled SPN cohort comprising 69,460 5-year survivors of cancer diagnosed before age 20 years in 12 European countries within which there was systematic ascertainment of all SPNs diagnosed.^{15,16}

There was particular focus on subsequent primary bone, soft tissue sarcoma, digestive and genitourinary cancers because these 4 cancer types account for a substantial proportion of the excess number of SPNs observed in the short and long term. Approximately 300 subsequent primary cancers of each of these 4 types have been included in 4 nested case-control studies (1200 cases in total) to investigate the extent to which cumulative dose of radiation from radiotherapy, cumulative dose of specific cytotoxics, and particular genomic factors extracted from saliva are related to risk of developing specific types of SPN. So far the authors have published the cohort studies relating to bone¹⁷ and soft tissue sarcoma.¹⁸

Risks of Subsequent Primary Neoplasms After Adolescent and Young Adult Cancer

Large-scale studies of survivors of AYA cancer have tended to focus on risks of SPNs after specific common cancers such as lymphoma, testes, or breast cancer. Only 2 studies have investigated the risks of developing any SPN after each type of AYA cancer. One study was based on Surveillance, Epidemiology and End Results (SEER) registry data and the main finding from this study was that AYA cancer survivors had a higher absolute risk of developing an SPN compared with childhood or mature adult cancer survivors.¹⁹ This study did not investigate the risks of specific SPNs after each AYA cancer.¹⁹ Recently published is the largest ever study to investigate the risks of SPNs after each specific AYA cancer and the first to provide excess risks of specific types of SPN after each of 16 types of AYA cancer, the Teenage and Young Adult Cancer Survivor Study.²⁰ The Teenage and Young Adult Cancer Survivor Study is a population-based cohort of 200,945 5-year survivors of cancer diagnosed when aged 15 to 39 years in England and Wales from January 1971 to December 2006. During 2,631,326 person-years of follow-up 12,321 SPNs were diagnosed in 11,565 survivors.²⁰

The recent publication relating to the Teenage and Young Adult Cancer Survivor Study illustrates 2 key new findings.²⁰ Firstly, in individuals who survived at least 30 years from diagnosis of cervical cancer, testicular cancer, Hodgkin lymphoma in women, breast cancer, and Hodgkin lymphoma in men, the authors identified a small number of specific SPNs that account for 82%, 61%, 58%, 45%, and 41% of the total excess number of neoplasms, respectively, and provides an evidence base to inform priorities for clinical long-term follow-up.²⁰ Secondly, lung cancer accounted for a substantial proportion of the excess number of neoplasms across all AYA groups investigated and indicates need for further work aimed at preventing and reducing the risk of this cancer among current and future survivors. This latter finding is in marked contrast to survivors of childhood cancer who do not experience such substantial excess risks of lung cancer, and this may relate to the evidence that survivors of AYA cancer smoke notably in excess of that expected from the general population, and in contrast survivors of childhood cancer smoke much less than that expected from the general population.²⁰

SUBSEQUENT PRIMARY NEOPLASM RISK RELATED TO RADIOTHERAPY

Radiotherapy exposure has been recognized as a risk factor for SPNs among childhood cancer survivors for decades. One of the first comprehensive reports on SPNs among childhood cancer survivors demonstrated that most of the SPNs developed in previously irradiated sites.²¹

Breast Cancer

Radiotherapy exposure to the chest is an important risk factor for female breast cancer.^{22–25} Recent studies suggest that even at lower absorbed doses to the breast (<20 Gy), breast cancer risk can be substantially elevated,^{24,26} especially among survivors who were exposed to a large volume of the breast, such as whole lung irradiation for pulmonary metastases in Wilms tumor or Ewing sarcoma survivors.²⁴ A linear dose-response relation has been observed in several studies.^{22,24,26} Hormonal exposure can modify the radiation-related risk of breast cancer. Survivors who also received radiation to the ovaries were reported to have lower radiation-related breast cancer risks.^{22,26} Furthermore, the effect of radiation has been suggested to be stronger when administered near menarche.⁸ In addition, there is some recent evidence for a stronger effect of radiation among those who also received anthracyclines.²⁶

Sarcoma

Sarcoma risk is increased in childhood cancer survivors and both radiotherapy and chemotherapy have been implicated to contribute to this excess risk.^{4,25,27–29} A nested case-control study within the CCSS cohort found a linear dose-response for any sarcoma.²⁷ Several reports evaluated the radiation dose-response for *bone* sarcoma or *soft tissue* sarcoma specifically. For *bone* sarcoma, an increased risk with increasing dose has been observed.^{4,29–31} However, some of these reports suggested a decline in relative risk at doses above 40 Gy.^{4,30} Among the studies on *soft tissue* sarcoma, results were consistent with a linear dose-response relationship between radiation dose and risk.^{12,28,32} In general, the dose-related risk seemed somewhat higher for *bone* sarcoma than for *soft tissue* sarcoma.³³

Thyroid Cancer

A pooled analysis, consisting of data from 2 cohort studies and 2 case-control studies among childhood cancer survivors, showed that the relative risk of thyroid cancer increased linearly with radiation dose up to 10 Gy, after which the risk plateaued.³⁴ At doses higher than 30 Gy, the risk seems to decline, possibly because of cell-killing effects. The dose-response relationship was stronger among those exposed to radiotherapy at a younger age.

Colorectal Cancer

Risk of colorectal cancer has been shown to be elevated among childhood cancer survivors, and abdominal radiotherapy has been implicated as a risk factor.^{3,13,35,36} Researchers from the French Childhood Cancer Survivor Cohort and the St Jude Lifetime Cohort (SJL) found a radiation dose-dependent effect on colorectal cancer risk,^{13,36} and the results of the SJL study also suggested an effect of radiation volume, as the risk increased with an increasing number of colonic segments irradiated.³⁶ Cumulative incidence of colorectal cancer was shown to be similar to that among individuals with 2 or more first-degree relatives with colorectal cancer in the British Childhood Cancer Survivor Study.³

Central Nervous System Tumors

Central nervous system (CNS) tumors occur in excess among childhood cancer survivors.^{10,37,38} Nearly all meningiomas and most of the gliomas present in survivors treated with cranial or craniospinal irradiation for brain tumors or acute lymphoblastic leukemia.^{10,37,38} For both gliomas and meningiomas, a linear dose-response relation has been observed, which seems to be stronger for meningioma (range of excess

relative risks (ERRs): 0.30–5.1 per Gy)^{10,37,38} than for gliomas (range of ERRs: 0.079–0.33 per Gy).^{37,38}

Nonmelanoma Skin Cancer

Nonmelanoma skin cancer, particularly basal cell carcinoma, is the most frequently observed SPN among childhood cancer survivors. Most of the basal cell carcinomas occur among previously irradiated patients.^{11,39,40} A study in the Dutch LATER cohort observed that basal cell carcinoma risks increased with increasing skin surface area exposed.¹¹ A nested case-control study in the CCSS cohort demonstrated a linear radiation dose-response relation, with an ERR of 1.09 per Gy.⁴⁰

Salivary Gland Tumors

Salivary gland tumor risks are elevated among childhood cancer survivors and a linear radiation dose-response relation was observed in a study in the CCSS cohort (ERR = 0.36 per Gy).⁴¹

Leukemia

In addition to the strong effects of chemotherapy on leukemia risk among childhood cancer survivors,^{5,42–47} there is some evidence that radiotherapy exposure might add to the increased risk of subsequent leukemia.^{5,44}

The results presented earlier mainly represent data from patients with childhood cancer treated decades ago, because those patients have sufficient follow-up time to evaluate risk of SPNs. In recent decades, radiotherapy practices have changed. Where possible, radiotherapy has been avoided or fields and doses have been reduced. For example, technological advances have led to the introduction of new techniques such as intensity-modulated radiotherapy (IMRT) and proton radiotherapy. These techniques aim to reduce the radiotherapy dose to the surrounding tissue, which might reduce the risk of SPNs.^{48,49} However, with IMRT, the larger volume exposed to radiation (although at lower dose) can potentially increase SPN risk.^{50,51} Proton therapy leads to an improvement in dose distribution by reducing the entrance dose and having virtually no exit dose.⁵² However, there are some concerns regarding the secondary dose from neutron scatter with proton therapy, which might lead to an increased SPN risk compared with photon therapy.^{53,54} It is important to carefully monitor patients with childhood cancer treated with those modern radiotherapy techniques and evaluate SPN risks in this population.

SUBSEQUENT PRIMARY NEOPLASM RISK RELATED TO CHEMOTHERAPY

Recent work has continued to highlight the independent influence of chemotherapy on the risk of SPNs in childhood cancer survivors. In the CCSS, among survivors exposed to only chemotherapy there was a 2.8-fold increased SPN risk compared with the general population (95% confidence interval [CI]: 2.5–3.2).⁵⁵ Chemotherapy increases the risk of both hematologic and solid SPN, depending on type and cumulative dose.

Chemotherapy and Subsequent Hematologic Malignancy

The most well-established association between chemotherapy and SPN relates to therapy-related acute myeloid leukemia (t-AML) and myelodysplastic syndrome (t-MDS).⁵ Dose-dependent risks for t-AML/t-MDS are high (>10-fold increased) after almost all alkylating agents and topoisomerase II inhibitors.^{5,45,56} Notably, the

leukemogenicity of different agents in these chemotherapy families varies substantially, and the absolute excess risk is low due to the low background risk in the age-matched general population. Development of t-AML after alkylating agent exposure typically arises after a latency of 5 to 8 years, is frequently preceded by MDS, and often has a complex karyotype with chromosome 5/7 abnormalities.⁵ In contrast, t-AML after topoisomerase II inhibitor exposure typically arises less than 3 years following therapy, is rarely preceded by MDS, and is most frequently characterized by 11q23 rearrangements.⁵⁷

Chemotherapy and Subsequent Solid Tumors

Chemotherapy increases risk for solid SPN, which often occur at least 10 years after exposure.¹⁴ Several classes of chemotherapy directly or indirectly affect the risk of development of these SPNs.

Alkylating agents

Alkylating agent exposures increase risk for gastrointestinal, thyroid, lung, breast, and bladder cancers; melanomas; and sarcomas.^{4,29,35,36,55,58–62} Specifically, cyclophosphamide increases sarcoma risk in a dose-dependent manner.^{4,25,27,29} Likewise, cyclophosphamide equivalent doses of greater than 18,000 mg/m² increase breast cancer risk by 3-fold (SIR, 3.0; 95% CI, 1.2–7.7).⁶⁰ Procarbazine and platinum have been associated with 3.2 (95% CI, 1.1–9.4) and 7.6-fold (95% CI, 2.3–25.5) increased risks, respectively, of gastrointestinal SPNs.³⁵ Procarbazine-related risks for the gastrointestinal tract may be related to direct exposure of the mucosa,^{27,36,58} whereas the mechanisms of carcinogenesis for agents administered intravenously are unknown.

Anthracyclines

Risk for breast cancer and other solid malignancies, including sarcoma, are increased after anthracycline exposure.^{25,27,60,63} In the CCSS cohort, risk for breast cancer in survivors treated with greater than 250 mg/m² of anthracycline and without chest radiotherapy exposure was increased by nearly 4-fold compared with the general population (SIR, 3.8; 95% CI 1.7–8.3).⁶⁰ Both the DCOG-LATER cohort and the SJL cohorts reported similar findings. The DCOG-LATER cohort reported a dose-dependent relationship between breast cancer risk and doxorubicin ($P_{trend} < 0.001$).²⁵ The SJL cohort reported an increasing breast cancer risk in both those exposed to 1 to 249 mg/m² (hazard ratio [HR] = 2.6, 95% CI 1.1–6.2, $P = .034$) and those exposed to greater than 250 mg/m² (HR = 13.4, 95% CI 5.5–32.5, $P < .001$) of anthracyclines.^{25,63} In both the CCSS and DCOG-LATER reports, breast cancer risk was highest after Li Fraumeni syndrome–associated cancers, suggesting a possible interaction between chemotherapy and genetic predisposition.^{25,60} However, with whole-exome sequencing available in the SJL cohort, the risk of breast cancer remained elevated in survivors exposed to greater than 250 mg/m² excluding those survivors with an identified cancer predisposition gene.⁶³

Indirect associations of chemotherapy and subsequent primary neoplasm risk

Chemotherapy can indirectly affect SPN risk. In Hodgkin lymphoma survivors,^{59,64,65} higher cumulative procarbazine exposure was associated with a greater reduction of breast cancer risk, with 30% and 67% risk reductions for regimens with less than 8.4 g/m² and greater than 8.4 g/m² procarbazine, respectively.^{64,65} This risk reduction seems to reflect the higher frequency of premature menopause in more intensively chemotherapy-treated patients, and their resultant reduced exposure to ovarian hormones.^{65–67} Similarly, high cumulative alkylator exposure significantly reduced breast

cancer risk in the CCSS cohort,⁸ in contrast to earlier CCSS results that did not show a reduced breast cancer risk after alkylator therapy.²³ Breast cancer risk also increases in women with more than 10 years of ovarian function after chest radiotherapy compared with those with less.^{8,65,67}

RISK OF SUBSEQUENT PRIMARY NEOPLASM AND GENOMICS

Inherited genetic susceptibility has long been known to play a role in SPN risk based on familial syndromes that predispose individuals to developing multiple primary neoplasms. Indeed, the occurrence of multiple primary tumors in an individual, particularly at a young age, was one of the earliest clues to inherited cancer predisposition syndromes.⁶⁸ Key examples of these syndromes include Li Fraumeni syndrome and hereditary retinoblastoma, which are caused by rare, highly penetrant germline mutations in the tumor suppressor genes *TP53* and *RB1*, respectively. In Li Fraumeni syndrome, overall, half of the women develop a first cancer by age 31 years and more than half of the men by age 46 years; of these individuals, approximately half will develop an SPN after a median of 10 years.⁶⁹ In contrast, in hereditary retinoblastoma nearly all individuals who inherit a germline mutation develop retinoblastoma in early childhood, typically within the first year of life. More than one-third of individuals are estimated to develop an SPN by age 40 years, although this estimate has been shown to vary by treatment exposure, specific *RB1* mutation, and family history of retinoblastoma.^{70–73}

The field of cancer genomics has expanded rapidly in the last decade. Advances in technology and reductions in laboratory costs have now made it possible to broadly interrogate the entire genome using high-throughput microarray genotyping or next-generation sequencing in increasingly larger study populations. These advances are essential for enabling sufficient sample size to identify new disease-associated genes. In the general population, large-scale, international collaborative efforts to study breast cancer exemplify the discoveries that are possible with these new approaches. Genome-wide association studies (GWAS) using microarray genotyping for common single nucleotide polymorphisms have identified greater than 170 loci associated with breast cancer risk.⁷⁴ Although each of these individual loci has a very weak effect on risk (relative risks typically <1.2), combining the loci into a polygenic score provides dramatic risk stratification.⁷⁵ Large-scale sequencing studies also are demonstrating substantial heterogeneity in breast cancer risk associated with specific, rare mutations in *BRCA1* and *BRCA2*.⁷⁶

Although these advances are only now beginning to be applied to assess genetic susceptibility to SPNs, as reviewed recently,^{68,77,78} the future holds tremendous promise for advancing this research area to provide biological insights into SPN development and potentially changing clinical practice through front-line therapy decision-making and risk stratification for long-term patient follow-up. Paralleling research in the general population, most of the earliest studies focused on single nucleotide polymorphisms in candidate genes. However, unlike the general population, where specific exposure-disease relationships rarely have been taken into account in genetic association studies, initial studies in cancer survivors focused on genes in pathways such as DNA repair that mediate response to treatment exposures, which are the primary drivers of SPN risk. Although some of these reports have been promising, few have been replicated in independent study populations, thus further research is needed to clarify the role of common variation in DNA repair genes in SPN risk.

More recently, several GWAS or large-scale genotyping studies have been conducted to identify loci involved in SPN risk after childhood cancer, including SPN overall,

therapy-related acute myeloid leukemia, breast cancer, and basal cell carcinoma.^{79–83} Those studies each have identified novel putative loci associated with SPN risk, with one study also suggesting that the genetic risk factors for breast cancer as an SPN overlap at least somewhat with those in the general population.⁸¹ Although further replication of these findings will be essential before clinical translation because of the substantial risk of false-positive findings when broadly interrogating the genome, the common frequency of the risk allele for many of the identified variants (2% to >30% of the population) demonstrates the substantial potential for applying these results in clinical practice.

Broader understanding of the role of rare variants in SPN risk also is warranted because they may be associated with high risks, even if they account for a relatively small fraction of SPN. The first large-scale sequencing study of SPN after childhood cancer demonstrated that fewer than 10% of childhood cancer survivors harbor rare, damaging mutations in a known cancer predisposition gene.⁸⁴ Ongoing analyses of additional large-scale sequencing studies are expected in the coming years and promise to shed light on the role of rare variants in SPN risk.

ROLE OF SURVEILLANCE, SCREENING, AND CLINICAL FOLLOW-UP GUIDELINES

Rationale for Surveillance

As a consequence of past treatments, behavioral factors such as smoking and alcohol, and host factors such as genetics, specific groups of childhood cancer survivors have a 10-fold increased risk of developing an SPN.^{85,86} Given the significant morbidity and risk for premature mortality resulting from SPNs, risk-adapted surveillance protocols have been developed with the goal of detecting SPNs at an earlier and more treatable stage. In other patient groups at high risk of malignancy, such as individuals with cancer predisposition syndromes, adherence to risk-adapted surveillance protocols have been shown to reduce mortality from SPNs.^{85–87} The same is assumed to be true for childhood cancer survivors, but this has never been established in a clinical trial.⁸⁸ For survivors at elevated SPN risk, surveillance for a given neoplasm is warranted if surveillance modalities exist that do not cause significant morbidity, allow for earlier identification and intervention that might reduce the SPN's impact, and do not cause an excess of false-positive results that lead to unnecessary further testing or intervention.^{89,90}

Surveillance Guidelines

Numerous organizations have developed recommendations for SPN surveillance in childhood cancer survivors.⁹¹ Substantial variation exists between guidelines, but as a general principle, periodic follow-up by a physician that includes a history and physical examination focused on evaluation of irradiated structures is warranted for all survivors. There is also a general consensus that breast cancer surveillance is appropriate for female survivors who have received chest irradiation, but the specifics of the required surveillance vary. North American organizations (The Children's Oncology Group and The National Comprehensive Cancer Network) uniquely recommend colorectal cancer surveillance for survivors who have received abdominal and/or pelvic radiation.^{92,93} In an attempt to create a common strategy for SPN surveillance, the International Guideline Harmonization Group (IGHG)⁹⁴ was formed. The IGHG has published recommendations for breast⁹⁵ and thyroid cancer⁹⁶ surveillance (available at <http://www.ighg.org/>) and is currently developing guidelines for CNS and colorectal cancer surveillance as well.

Current Guideline Adherence

Unfortunately, most adult survivors of childhood cancer are not adherent to the recommended SPN surveillance, potentially resulting in preventable morbidity and mortality. In one study of North American survivors enrolled in the CCSS, adherence to SPN surveillance was 12.6%, 37.0%, and 22.3% for breast, colorectal, and skin cancer surveillance, respectively (Yan A and Nathan P, unpublished data, 2019). Survivor reported barriers to surveillance include lack of time, forgetting, a perception that surveillance is not important, concerns about insurance coverage and cost, and lack of physician recommendation for surveillance.^{97,98} Psychosocial barriers include poor mental health, lower socioeconomic status, and lower educational level.^{99–101} In 2012, only 12% of US general internists¹⁰² and 9% of US and Canadian family doctors¹⁰³ felt at least “somewhat familiar” with care guidelines for childhood cancer survivors. A lack of primary care provider comfort with care guidelines likely contributes to poor adherence as well. Regular engagement with the health care system, receipt of a treatment summary, and patient-provider communication discussing the need for surveillance have been associated with better adherence to surveillance guidelines.^{104–108}

Mechanisms to Improve Adherence

To address barriers to receiving risk-adapted surveillance, the United States Institute of Medicine and the European collaboration PanCare have recommended that all childhood cancer survivors receive a treatment summary and survivorship care plan (SCP) that documents their cancer treatment–related health risks and the recommended surveillance.^{109–111} The impact of SCPs on surveillance outcomes in childhood cancer survivors is unclear.¹¹² In fact, little is known about how best to increase the completion of recommended surveillance testing. A recent systematic review that evaluated interventions to improve surveillance adherence only identified one randomized trial where the intervention significantly increased SPN surveillance.^{98,113} In this trial, mailed information coupled with motivational telephone interviewing increased adherence to mammography in women at risk for subsequent breast cancer.⁹⁸ Other interventions that have been tried with less success include motivational telephone counseling, SCP provision, web-based virtual information, and mailing of health risk information.^{98,114–116}

Cancer Predisposition Syndromes

In addition to the risk of SPNs as a consequence of cancer therapy, a subset of survivors is at high risk of SPNs secondary to an underlying cancer predisposition syndrome. Nearly 10% of childhood cancer survivors have an actionable germline genetic mutation, making yearly review of family cancer history and subsequent referral to genetics when necessary imperative.¹¹⁷ Specific guidelines have been created for the more common pediatric cancer predisposition syndromes, such as Li-Fraumeni syndrome¹¹⁸ and Beckwith Wiedemann syndrome.¹¹⁹ When no specific recommendations exist for a given syndrome, the American Association of Cancer Research recommends screening for malignancy if effective screening modalities exist and the overall risk exceeds 5% in the first 20 years of life. In addition, they recommend that when the overall risk is between 1% and 5%, screening can be considered on an individual basis.¹²⁰

Summary

Despite the availability of numerous guidelines that guide health care providers in providing surveillance for SPNs in childhood cancer survivors, very few survivors

are currently adherent to recommendations, and few interventions have been successful in increasing surveillance. Further studies that develop and test interventions to improve adherence are needed.

PRIORITIES FOR FUTURE RESEARCH

Observational Studies to Address Specific Gaps in Knowledge

Subsequent primary neoplasms in survivors of adolescent and young adult cancer

A large population-based study described the risk of SPN in survivors of AYA cancer, reporting that a small number of specific SPNs account for a large proportion of the overall excess, with a prominence of lung cancer.²⁰ However, the association between therapeutic exposures and the risk of SPNs after AYA cancer remains unstudied, as does the role of lifestyle factors, which could have greater impact among AYA survivors than among childhood cancer survivors.²⁰

Subsequent primary neoplasm risk in patients treated with immunotherapy

Targeted immunotherapy has emerged as an effective treatment option especially in pediatric malignancies.^{121,122} Although the early toxicities are clearly described, there remains a significant gap in knowledge regarding the development of delayed complications, especially SPNs. Systematic, long-term follow-up of patients treated with targeted immunotherapy is needed to address this gap.

Solid subsequent primary neoplasm risk in patients treated with chemotherapy

Although the association between radiation and solid SPNs (thyroid, breast, brain, colorectal) is well established,^{22,37,123} as is the risk between specific chemotherapeutic agents and therapy-related leukemia, there is emerging evidence regarding the role of adjuvant chemotherapy.⁵⁵ For example, treatment with anthracyclines may be a risk factor for thyroid cancer^{55,61} and breast cancer.⁵⁵ These findings are based on small numbers of SPNs developing after exposure to a specific chemotherapy class. This gap could be addressed by pooling large well-characterized cohorts and case-control studies of survivors.

Subsequent primary neoplasm risk: interaction of behavioral factors or infections with genotoxic exposures

The risk of lung cancer is significantly increased in patients treated for Hodgkin lymphoma. Both chemotherapy and radiation contribute to the risk. Cigarette smoking multiplies the risk associated with both chemotherapy and radiation.^{124,125} However, interaction between smoking and therapeutic exposures has not been examined for other types of SPN, such as esophageal, oropharyngeal, and gastric carcinoma. Furthermore, behavioral factors such as excessive alcohol consumption or a diet rich in processed meats has not been examined in this population. Finally, the interaction between chronic viral infections (hepatitis B virus, hepatitis C virus, human papillomavirus, Epstein-Barr virus) and prolonged immune suppression due to genotoxic exposures in increasing the risk of SPNs remains unstudied.

Temporal Changes in Subsequent Primary Neoplasm Risk with Changes in Treatment Strategies

With the decrease in the proportion of patients receiving radiation as well as a progressive reduction in the dose and field of radiation, the relative rates of meningioma and nonmelanoma skin cancers have declined over the past several decades.⁹ Additional follow-up using pooled data from other well-characterized cohorts is needed to understand whether the decline in SPNs is limited to just these 2 specific types of SPNs, or

whether smaller samples precluded the ability to detect trends for other SPNs, such as breast cancer. Further, these trends need to be placed within the context of increasing use of chemotherapy and changes in surveillance practice. Most importantly, as the cancer survivor population ages, it is important to understand the lifelong risk of SPN and particularly the types of SPNs that account for most of the excess observed later in life.

Identification of Survivors at Highest Risk of Subsequent Primary Neoplasm and Potential for Targeted Interventions

Although the magnitude of association between radiation exposure and SPN risk is moderate-to-large (3.1-fold to 15.9-fold)¹²⁶ with clear evidence for a dose-response relationship,^{22,37,123} there is wide variation in individual susceptibility, suggesting the role of genetic susceptibility in modifying this association.^{32,127–136} Genetic variants may modify the association between radiation and SPN risk or increase the risk of SPNs even in the absence of radiation.⁶⁰ Indeed, cancer survivors who carry a deleterious, high-penetrance mutation are at increased risk for SPNs.^{132–134,137,138} However, the low frequency of these mutations in the general population¹³⁹ suggests that the attributable risk is likely small. The interindividual variability in risk of SPNs is more likely related to common polymorphisms in low-penetrance genes that regulate drug metabolism or those responsible for DNA repair.^{140,141} Although there is significant effort currently expended on identifying genetic variants and their association with SPNs, an equally important aspect of this discovery currently lagging involves understanding the functional relevance of the identified genetic variants. Although we can speculate about the relevance of a specific genetic variant, it is critical to delve into the functional aspects of the identified variant in order to understand the mechanistic basis of SPNs; this is critical in order to develop risk-reducing interventions. An equally important, yet underutilized opportunity is the use of demographic, clinical (therapeutic exposures), behavioral, and genetic information to determine the individual risk of SPN. An example is the risk prediction model developed for survivors at risk for radiation-related brain tumors,¹⁴² where the sensitivity and specificity of predicting survivors of childhood cancer at highest or lowest risk of subsequent CNS tumors was 87.5% and 83.5%, respectively.

Radiation continues to serve as a critical backbone of treatment of childhood cancer, and although there may be options to use alternative treatments on a case-by-case basis (for patients at highest risk of SPN), the pediatric oncology community is reluctant to replace radiation with alternative treatments for *all patients*. In addition, among childhood cancer survivors already exposed to radiation, offering screening or behavioral/pharmacologic interventions based on personal risk could be cost-effective and better accepted by the survivor population. Finally, a deeper understanding of the mechanistic basis of radiation-related SPNs would lead us closer to developing targeted interventions.

Screening recommendations for early detection of subsequent primary neoplasms in childhood cancer survivors

The primary goal of risk-based surveillance is to facilitate early detection of treatment-related complications (including SPNs) in childhood cancer survivors.^{89,94} However, there is an opportunity to examine the cost-effectiveness of screening recommendations^{95,96} that tailor the intensity of screening based on personal SPN risk. Simulation, using Markov health states is currently being used to address cost-effectiveness of breast cancer screening recommendations.¹⁴³

Interventions to reduce subsequent primary neoplasm risk in childhood cancer survivors

Breast cancer, brain tumors, sarcoma, thyroid cancer, and gastrointestinal malignancies constitute most of the non-skin cancer SPNs. All these SPNs are radiation related, with a clear dose-response relationship. Understanding the pathogenesis of each of these tumors could inform specific interventions, which when applied in those at highest risk would significantly improve the efficacy of such an intervention. As an example, Bhatia and colleagues¹⁴⁴ recently completed a pharmacologic intervention for reducing the risk of radiation-related breast SPN in childhood cancer survivors, using a randomized, double-blinded, placebo-controlled trial design. The biological premise is based on the fact that endogenous estrogens play a role in radiation-related breast carcinogenesis.

REFERENCES

1. Reulen RC, Winter DL, Frobisher C, et al. Long-term cause-specific mortality among survivors of childhood cancer. *JAMA* 2010;304(2):172–9.
2. Armstrong GT, Chen Y, Yasui Y, et al. Reduction in late mortality among 5-year survivors of childhood Cancer. *N Engl J Med* 2016;374:833–42.
3. Reulen RC, Frobisher C, Winter DL, et al. Long-Term Risks of Subsequent Primary Neoplasms Among Survivors of Childhood Cancer. *JAMA* 2011;305(22):2311–9.
4. Hawkins MM, Wilson LM, Burton HS, et al. Radiotherapy, alkylating agents, and risk of bone cancer after childhood cancer. *J Natl Cancer Inst* 1996;88:270–8.
5. Hawkins MM, Wilson LM, Stovall MA, et al. Epipodophyllotoxins, alkylating agents, and radiation and risk of secondary leukaemia after childhood cancer. *BMJ* 1992;304:951–8.
6. Olsen JH, Moller T, Anderson H, et al. Lifelong cancer incidence in 47,697 patients treated for childhood cancer in the Nordic countries. *J Natl Cancer Inst* 2009;101:806–13.
7. Turcotte LM, Whitton J, Friedman D, et al. Risk of subsequent neoplasms during the fifth and sixth decades of life in the Childhood Cancer Survivor Cohort. *J Clin Oncol* 2015;33:3568–75.
8. Moskowitz CS, Chou JF, Sklar CA, et al. Radiation-associated breast cancer and gonadal hormone exposure: a report from the Childhood Cancer Survivor Study. *Br J Cancer* 2017;117:290–9.
9. Turcotte LM, Liu Q, Yasui Y, et al. Temporal trends in treatment and subsequent neoplasm risk among 5-year survivors of childhood cancer, 1970–2015. *JAMA* 2017;317:814–24.
10. Kok JL, Teepen JC, van Leeuwen FE, et al. Risk of benign meningioma after childhood cancer in the DCOG-LATER cohort: contributions of radiation dose, exposed cranial volume, and age. *Neuro Oncol* 2019;21:392–403.
11. Teepen JC, Kok JL, Kremer LC, et al. Long-term risk of skin cancer among childhood cancer survivors: a DCOG-LATER cohort study. *J Natl Cancer Inst* 2019;111(8):845–53.
12. Menu-Branthomme A, Rubino C, Shamsaldin A, et al. Radiation dose, chemotherapy and risk of soft tissue sarcoma after solid tumours during childhood. *Int J Cancer* 2004;110:87–93.
13. Allodji RS, Haddy N, Vu-Bezin G, et al. Risk of subsequent colorectal cancers after a solid tumor in childhood: Effects of radiation therapy and chemotherapy. *Pediatr Blood Cancer* 2019;66:e27495.

14. Turcotte LM, Neglia JP, Reulen RC, et al. Risk, risk factors, and surveillance of subsequent malignant neoplasms in survivors of childhood cancer: a review. *J Clin Oncol* 2018;36(21):2145–52.
15. Grabow D, Kaiser M, Hjorth L, et al. The PanCareSurFup cohort of 83,333 5-year survivors of childhood cancer: a cohort from 12 European countries. *Eur J Epidemiol* 2018;33(3):335–49.
16. Byrne J, Alessi D, Allodji RS, et al. The PanCareSurFup consortium: research and guidelines to improve lives for survivors of childhood cancer. *Eur J Cancer* 2018;103:238–48.
17. Fidler MM, Reulen RC, Winter DL, et al. Risk of subsequent primary bone cancers among 69,460 5-year survivors of childhood and adolescent cancer in Europe. *J Natl Cancer Inst* 2017;110(2):183–94.
18. Bright CJ, Hawkins MM, Winter DL, et al. Risk of soft-tissue sarcoma among 69,460 5-year survivors of childhood cancer in Europe. *J Natl Cancer Inst* 2017;110(6):649–60.
19. Lee JS, DuBois SG, Coccia PF, et al. Increased risk of second malignant neoplasms in adolescents and young adults with cancer. *Cancer* 2016;122(1):116–23.
20. Bright CJ, Reulen RC, Winter DL, et al. Risk of subsequent primary neoplasms in survivors of adolescent and young adult cancer (Teenage and Young Adult Cancer Survivor Study): a population-based cohort study. *Lancet Oncol* 2019;20(4):531–45.
21. Meadows AT, Baum E, Fossati-Bellani F, et al. Second malignant neoplasms in children: an update from the late effects study group. *J Clin Oncol* 1985;3:532–8.
22. Inskip PD, Robison LL, Stovall M, et al. Radiation dose and breast cancer risk in the childhood cancer survivor study. *J Clin Oncol* 2009;27:3901–7.
23. Kenney LB, Yasui Y, Inskip PD, et al. Breast cancer after childhood cancer: a report from the Childhood Cancer Survivor Study. *Ann Intern Med* 2004;141:590–7.
24. Moskowitz CS, Chou JF, Wolden SL, et al. Breast cancer after chest radiation therapy for childhood cancer. *J Clin Oncol* 2014;32:2217–23.
25. Teepen JC, van Leeuwen FE, Tissing WJ, et al. Long-term risk of subsequent malignant neoplasms after treatment of childhood cancer in the DCOG LATER study cohort: role of chemotherapy. *J Clin Oncol* 2017;35:2288–98.
26. Veiga LH, Curtis RE, Morton LM, et al. Association of breast cancer risk after childhood cancer with radiation dose to the breast and anthracycline use: a report from the childhood cancer survivor study. *JAMA Pediatr* 2019;173(12):1171–9.
27. Henderson TO, Rajaraman P, Stovall M, et al. Risk factors associated with secondary sarcomas in childhood cancer survivors: a report from the childhood cancer survivor study. *Int J Radiat Oncol Biol Phys* 2012;84:224–30.
28. Jenkinson HC, Winter DL, Marsden HB, et al. A study of soft tissue sarcomas after childhood cancer in Britain. *Br J Cancer* 2007;97:695–9.
29. Tucker MA, D'Angio GJ, Boice JD Jr, et al. Bone sarcomas linked to radiotherapy and chemotherapy in children. *N Engl J Med* 1987;317:588–93.
30. Le Vu B, de Vathaire F, Shamsaldin A, et al. Radiation dose, chemotherapy and risk of osteosarcoma after solid tumours during childhood. *Int J Cancer* 1998;77:370–7.

31. Schwartz B, Benadjaoud MA, Clero E, et al. Risk of second bone sarcoma following childhood cancer: role of radiation therapy treatment. *Radiat Environ Biophys* 2014;53:381–90.
32. Wong FL, Boice JD Jr, Abramson DH, et al. Cancer incidence after retinoblastoma. Radiation dose and sarcoma risk. *JAMA* 1997;278:1262–7.
33. Berrington de Gonzalez A, Kutsenko A, Rajaraman P, et al. Sarcoma risk after radiation exposure. *Clin Sarcoma Res* 2012;2:18.
34. Veiga LH, Lubin JH, Anderson H, et al. A pooled analysis of thyroid cancer incidence following radiotherapy for childhood cancer. *Radiat Res* 2012;178:365–76.
35. Henderson TO, Oeffinger KC, Whitton J, et al. Secondary gastrointestinal cancer in childhood cancer survivors: a cohort study. *Ann Intern Med* 2012;156:757–66. W-260.
36. Nottage K, McFarlane J, Krasin MJ, et al. Secondary colorectal carcinoma after childhood cancer. *J Clin Oncol* 2012;30:2552–8.
37. Neglia JP, Robison LL, Stovall M, et al. New primary neoplasms of the central nervous system in survivors of childhood cancer: a report from the Childhood Cancer Survivor Study. *J Natl Cancer Inst* 2006;98:1528–37.
38. Taylor AJ, Little MP, Winter DL, et al. Population-based risks of CNS tumors in survivors of childhood cancer: the British Childhood Cancer Survivor Study. *J Clin Oncol* 2010;28:5287–93.
39. Perkins JL, Liu Y, Mitby PA, et al. Nonmelanoma skin cancer in survivors of childhood and adolescent cancer: a report from the childhood cancer survivor study. *J Clin Oncol* 2005;23:3733–41.
40. Watt TC, Inskip PD, Stratton K, et al. Radiation-related risk of basal cell carcinoma: a report from the Childhood Cancer Survivor Study. *J Natl Cancer Inst* 2012;104:1240–50.
41. Boukheris H, Stovall M, Gilbert ES, et al. Risk of salivary gland cancer after childhood cancer: a report from the Childhood Cancer Survivor Study. *Int J Radiat Oncol Biol Phys* 2013;85:776–83.
42. Allard A, Haddy N, Le Deley MC, et al. Role of radiation dose in the risk of secondary leukemia after a solid tumor in childhood treated between 1980 and 1999. *Int J Radiat Oncol Biol Phys* 2010;78:1474–82.
43. Allodji RS, Schwartz B, Veres C, et al. Risk of subsequent leukemia after a solid tumor in childhood: impact of bone marrow radiation therapy and chemotherapy. *Int J Radiat Oncol Biol Phys* 2015;93:658–67.
44. Haddy N, Le Deley MC, Samand A, et al. Role of radiotherapy and chemotherapy in the risk of secondary leukaemia after a solid tumour in childhood. *Eur J Cancer* 2006;42:2757–64.
45. Le Deley MC, Leblanc T, Shamsaldin A, et al. Risk of secondary leukemia after a solid tumor in childhood according to the dose of epipodophyllotoxins and anthracyclines: a case-control study by the Societe Francaise d'Oncologie Pediatric. *J Clin Oncol* 2003;21:1074–81.
46. Tucker MA, Meadows AT, Boice JD Jr, et al. Leukemia after therapy with alkylating agents for childhood cancer. *J Natl Cancer Inst* 1987;78:459–64.
47. Advani PG, Schonfeld SJ, Curtis RE, et al. Risk of therapy-related myelodysplastic syndrome/acute myeloid leukemia after childhood cancer: a population-based study. *Leukemia* 2019;33:2947–78.
48. Cotter SE, McBride SM, Yock TI. Proton radiotherapy for solid tumors of childhood. *Technol Cancer Res Treat* 2012;11:267–78.

49. Veldeman L, Madani I, Hulstaert F, et al. Evidence behind use of intensity-modulated radiotherapy: a systematic review of comparative clinical studies. *Lancet Oncol* 2008;9:367–75.
50. Casey DL, Friedman DN, Moskowitz CS, et al. Second cancer risk in childhood cancer survivors treated with intensity-modulated radiation therapy (IMRT). *Pediatr Blood Cancer* 2015;62:311–6.
51. Schneider U, Lomax A, Pemler P, et al. The impact of IMRT and proton radiotherapy on secondary cancer incidence. *Strahlenther Onkol* 2006;182:647–52.
52. Miralbell R, Lomax A, Cella L, et al. Potential reduction of the incidence of radiation-induced second cancers by using proton beams in the treatment of pediatric tumors. *Int J Radiat Oncol Biol Phys* 2002;54:824–9.
53. Eaton BR, MacDonald SM, Yock TI, et al. Secondary malignancy risk following proton radiation therapy. *Front Oncol* 2015;5:261.
54. Hall EJ. Intensity-modulated radiation therapy, protons, and the risk of second cancers. *Int J Radiat Oncol Biol Phys* 2006;65:1–7.
55. Turcotte LM, Liu Q, Yasui Y, et al. Chemotherapy and risk of subsequent malignant neoplasms in the childhood cancer survivor study cohort. *J Clin Oncol* 2019;37(34):3310–9.
56. Pui CH, Ribeiro RC, Hancock ML, et al. Acute myeloid leukemia in children treated with epipodophyllotoxins for acute lymphoblastic leukemia. *N Engl J Med* 1991;325(24):1682–7.
57. Pendleton M, Lindsey RH, Felix CA, et al. Topoisomerase II and leukemia. *Ann N Y Acad Sci* 2014;1310:98–110.
58. Morton LM, Dores GM, Curtis RE, et al. Stomach cancer risk after treatment for Hodgkin Lymphoma. *J Clin Oncol* 2013;31(27):3369–77.
59. Schaapveld M, Aleman BM, van Eggermond AM, et al. Second cancer risk up to 40 years after treatment for Hodgkin's lymphoma. *N Engl J Med* 2015;373(26):2499–511.
60. Henderson TO, Moskowitz CS, Chou JF, et al. Breast cancer risk in childhood cancer survivors without a history of chest radiotherapy: a report from the childhood cancer survivor study. *J Clin Oncol* 2016;34(9):910–8.
61. Veiga LHS, Bhatti P, Ronckers CM, et al. Chemotherapy and thyroid cancer risk: a report from the childhood cancer survivor study. *Cancer Epidemiol Biomarkers Prev* 2012;21(1):92–101.
62. Travis LB, Gospodarowicz M, Curtis RE, et al. Lung cancer following chemotherapy and radiotherapy for Hodgkin's disease. *J Natl Cancer Inst* 2002;94(3):182–92.
63. Ehrhardt MJ, Howell CR, Hale K, et al. Subsequent Breast Cancer in Female Childhood Cancer Survivors in the St Jude Lifetime Cohort Study (SJLIFE). *J Clin Oncol* 2019;37(19):1647–56.
64. Swerdlow AJ, Cooke R, Bates A, et al. Breast cancer risk after supradiaphragmatic radiotherapy for Hodgkin's Lymphoma in England and Wales: A National Cohort Study. *J Clin Oncol* 2012;30(22):2745–52.
65. De Bruin ML, Sparidans J, van't Veer MB, et al. Breast cancer risk in female survivors of Hodgkin's lymphoma: lower risk after smaller radiation volumes. *J Clin Oncol* 2009;27(26):4239–46.
66. Cooke R, Jones ME, Cunningham D, et al. Breast cancer risk following Hodgkin lymphoma radiotherapy in relation to menstrual and reproductive factors. *Br J Cance* 2013;108(11):2399–406.

67. Krul IM, Opstal-van Winden AWJ, Aleman BMP, et al. Breast cancer risk after radiation therapy for hodgkin lymphoma: influence of gonadal hormone exposure. *Int J Radiat Oncol Biol Phys* 2017;99(4):843–53.
68. Morton LM, Savage SA, Bhatia S, et al. *Schottenfeld & Fraumeni: cancer epidemiology & prevention*. 4th edition. New York: Oxford University Press; 2017. p. 1155–92.
69. Mai PL, Best AF, Peters JA, et al. Risks of first and subsequent cancers among TP53 mutation carriers in the National Cancer Institute Li-Fraumeni syndrome cohort. *Cancer* 2016;122(23):3673–81.
70. Dommering CJ, Marees T, van der Hout AH, et al. RB1 mutations and second primary malignancies after hereditary retinoblastoma. *Fam Cancer* 2012;11: 225–33.
71. Kleinerman RA, Yu CL, Little MP, et al. Variation of second cancer risk by family history of retinoblastoma among long-term survivors. *J Clin Oncol* 2012;30: 950–7.
72. Marees T, Moll AC, Imhof SM, et al. Risk of second malignancies in survivors of retinoblastoma: more than 40 years of follow-up. *J Natl Cancer Inst* 2008;100: 1771–9.
73. Marees T, van Leeuwen FE, Schaapveld M, et al. Risk of third malignancies and death after a second malignancy in retinoblastoma survivors. *Eur J Cancer* 2010;46:2052–8.
74. Michailidou K, Lindstrom S, Dennis J, et al. Association analysis identifies 65 new breast cancer risk loci. *Nature* 2017;551:92–4.
75. Maas P, Barrdahl M, Joshi AD, et al. Breast cancer risk from modifiable and non-modifiable risk factors among white women in the United States. *JAMA Oncol* 2016;2:1295–302.
76. Cline MS, Liao RG, Parsons MT, et al. BRCA challenge: BRCA Exchange as a global resource for variants in BRCA1 and BRCA2. *PLoS Genet* 2018;14: e1007752.
77. Bhatia S. Genetic variation as a modifier of association between therapeutic exposure and subsequent malignant neoplasms in cancer survivors. *Cancer* 2015;121:648–63.
78. Gramatges MM, Bhatia S. Evidence for genetic risk contributing to long-term adverse treatment effects in childhood cancer survivors. *Annu Rev Med* 2018; 69:247–62.
79. Sapkota Y, Turcotte LM, Ehrhardt MJ, et al. Genome-wide association study in irradiated childhood cancer survivors identifies HTR2A for subsequent basal cell carcinoma. *J Invest Dermatol* 2019;139:2042–5.e8.
80. Morton LM, Sampson JN, Armstrong GT, et al. Genome-wide association study to identify susceptibility loci that modify radiation-related risk for breast cancer after childhood cancer. *J Natl Cancer Inst* 2017;109(11):dix058.
81. Opstal-van Winden AWJ, de Haan HG, Hauptmann M, et al. Genetic susceptibility to radiation-induced breast cancer after Hodgkin lymphoma. *Blood* 2019; 133:1130–9.
82. Knight JA, Skol AD, Shinde A, et al. Genome-wide association study to identify novel loci associated with therapy-related myeloid leukemia susceptibility. *Blood* 2009;113:5575–82.
83. Best T, Li D, Skol AD, et al. Variants at 6q21 implicate PRDM1 in the etiology of therapy-induced second malignancies after Hodgkin's lymphoma. *Nat Med* 2011;17:941–3.

84. Wang Z, Wilson CL, Easton J, et al. Genetic risk for subsequent neoplasms among long-term survivors of childhood cancer. *J Clin Oncol* 2018;36:2078–87.
85. Villani A, Shore A, Wasserman JD, et al. Biochemical and imaging surveillance in germline TP53 mutation carriers with Li-Fraumeni syndrome: 11 year follow-up of a prospective observational study. *Lancet Oncol* 2016;17(9):1295–305.
86. Koskenvuo L, Pitkaniemi J, Rantanen M, et al. Impact of Screening on Survival in Familial Adenomatous Polyposis. *J Clin Gastroenterol* 2016;50(1):40–4.
87. Gareth ED, Nisha K, Yit L, et al. MRI breast screening in high-risk women: Cancer detection and survival analysis. *Breast Cancer Res Treat* 2014;145:663–72.
88. Hodgson DC, Cotton C, Crystal P, et al. Impact of Early Breast Cancer Screening on Mortality Among Young Survivors of Childhood Hodgkin's Lymphoma. *J Natl Cancer Inst* 2016;108(7):djw010.
89. Landier W, Bhatia S, Eshelman DA, et al. Development of risk based guidelines for pediatric cancer survivors: the Children's Oncology Group Long Term Follow Up Guidelines from the Children's Oncology Group Late Effects Committee and Nursing Discipline. *J Clin Oncol* 2016;22:4979–90.
90. Barratt A, Irwin L, Glasziou P, et al. How to use guidelines and recommendations about screening. *JAMA* 1999;281(2):2029–34.
91. Fidler MM, Frobisher C, Hawkins MM, et al. Challenges and opportunities in the care of survivors of adolescent and young adult cancers. *Pediatr Blood Cancer* 2019;66:e27668.
92. Long-Term Follow-Up Guidelines for Survivors of Childhood, Adolescent and Young Adult Cancers, version 4.0. Available at: <http://www.survivorshipguidelines.org>. Accessed November 9, 2019.
93. National Comprehensive Cancer Network Clinical Practice Guidelines in Oncology; Adolescent and Young Adult Oncology Version 1.2020. Available at: https://www.nccn.org/professionals/physician_gls/pdf/aya.pdf. Accessed November 9, 2019.
94. Kremer LC, Mulder RL, Oeffinger KC, et al. A worldwide collaboration to harmonize guidelines for the long-term follow-up of childhood and young adult cancer survivors: A report from the International Late Effects of Childhood Cancer Guideline Harmonization Group. *Pediatr Blood Cancer* 2013;60:543–9.
95. Mulder RL, Kremer LCM, Hudsom MM, et al. Recommendations for breast cancer surveillance for female survivors of childhood, adolescent, and young adult cancer given chest radiation: a report from the International Late Effects of Childhood Cancer Guideline Harmonization Group. *Lancet Oncol* 2013;14:e621–9.
96. Clement SC, Kremer LCM, Verburg FA, et al. Balancing the benefits and harms of thyroid cancer surveillance in survivors of Childhood, adolescent and young adult cancer: Recommendations from the international Late Effects of Childhood Cancer Guideline Harmonization Group in collaboration with the PanCare-SurFup Consortium. *Cancer Treat Rev* 2018;63:28–39.
97. Hudson MM, Leisenring W, Stratton KK, et al. Increasing cardiomyopathy screening in at-risk adult survivors of pediatric malignancies: a randomized controlled trial. *J Clin Oncol* 2014;32(35):3974–81.
98. Oeffinger KC, Ford J, Moskowitz CS, et al. The EMPOWER study: promoting breast cancer screening—a randomized controlled trial (RCT) in the childhood cancer survivor study (CCSS). *J Clin Oncol* 2016;34(15_suppl):10506.
99. Berg CJ, Stratton E, Esiashvili N, et al. Young adult cancer survivors' experience with cancer treatment and follow-up care and perceptions of barriers to engaging in recommended care. *J Cancer Educ* 2016;31(3):430–42.

100. Casillas J, Oeffinger KC, Hudson MM, et al. Identifying predictors of longitudinal decline in the level of medical care received by adult survivors of childhood cancer: a report from the childhood cancer survivor study. *Health Serv Res* 2015; 50(4):1021–42.
101. Nathan PC, Agha M, Pole JD, et al. Predictors of attendance at specialized survivor clinics in a population-based cohort of adult survivors of childhood cancer. *J Cancer Surviv* 2016;10(4):611–8.
102. Suh E, Daugherty CK, Wroblewski K, et al. General internists' preferences and knowledge about the care of adult survivors of childhood cancer: a cross-sectional survey. *Ann Intern Med* 2014;160(1):11–7.
103. Nathan PC, Daugherty CK, Wroblewski KE, et al. Family physician preferences and knowledge gaps regarding the care of adolescent and young adult survivors of childhood cancer. *J Cancer Surviv* 2013;7(3):275–82.
104. Cox CL, Oeffinger KC, Montgomery M, et al. Determinants of mammography screening participation in adult childhood cancer survivors: results from the Childhood Cancer Survivor Study. *Oncol Nurs Forum* 2009;36(3):335–44.
105. Daniel CL, Kohler CL, Stratton KL, et al. Predictors of colorectal cancer surveillance among survivors of childhood cancer treated with radiation: a report from the Childhood Cancer Survivor Study. *Cancer* 2015;121(11):1856–63.
106. Baxstrom K, Peterson BA, Lee C, et al. A pilot investigation on impact of participation in a long-term follow-up clinic (LTFU) on breast cancer and cardiovascular screening among women who received chest radiation for Hodgkin lymphoma. *Support Care Cancer* 2018;26(7):2361–8.
107. Oeffinger KC, Ford JS, Moskowitz CS, et al. Breast cancer surveillance practices among women previously treated with chest radiation for a childhood cancer. *JAMA* 2009;301(4):404–14.
108. Nathan PC, Ness KK, Mahoney MC, et al. Screening and surveillance for second malignant neoplasms in adult survivors of childhood cancer: a report from the childhood cancer survivor study. *J Cancer Surviv* 2019;13:713–29.
109. Institute of Medicine and National Research Council. From cancer patient to cancer survivor: lost in transition. Washington, DC: The National Academies Press; 2006. <https://doi.org/10.17226/11488>.
110. Haupt R, Essiaf S, Dellacasa C, et al. PanCareSurFup, ENCCA Working Group; ExPo-r-Net Working Group. The 'Survivorship Passport' for childhood cancer survivors. *Eur J Cancer* 2018;102:69–81.
111. Michel G, Mulder RL, van der Pal HJH, et al. Evidence based recommendations for the organization of long-term follow-up care for childhood and adolescent cancer survivors: a report from the PanCareSurFup Guidelines Working Group. *J Cancer Surviv* 2019;13(5):759–72.
112. Jacobsen PB, DeRosa AP, Henderson TO, et al. Systematic review of the impact of cancer survivorship care plans on health outcomes and health care delivery. *J Clin Oncol* 2018;36(20):2088–100.
113. Zabih V, Kahane A, O'Neill NE, et al. Interventions to improve adherence to surveillance guidelines in survivors of childhood cancer: a systematic review. *J Cancer Surviv* 2019;13:713–29.
114. Steele JR, Wall M, Salkowski N, et al. Predictors of risk-based medical follow-up: a report from the childhood cancer survivor study. *J Cancer Surviv* 2013;7(3): 379–91.
115. Kadan-Lottick NS, Ross WL, Mitchell H-R, et al. Randomized trial of the impact of empowering childhood cancer survivors with survivorship care plans. *J Natl Cancer Inst* 2018;110:1352–9.

116. Oeffinger KC, Hudson MM, Mertens AC, et al. Increasing rates of breast cancer and cardiac surveillance among high-risk survivors of childhood Hodgkin lymphoma following a mailed, one-page survivorship care plan. *Pediatr Blood Cancer* 2011;56(5):818–24.
117. Zhang J, Walsh MF, Wu G, et al. Germline mutations in predisposition genes in pediatric cancer. *N Engl J Med* 2015;373:2336–46.
118. Kratz CP, Achatz MI, Brugieres L, et al. Cancer screening recommendations for individuals with Li-Fraumeni syndrome. *Clin Cancer Res* 2017;23(11):e38–45.
119. Kalish JM, Doros L, Lee J, et al. Surveillance recommendations for children with overgrowth syndromes and predisposition to Wilms tumors and hepatoblastoma. *Clin Cancer Res* 2017;23(11):e115–22.
120. Brodeur GM, Nichols KE, Plon SE, et al. Pediatric cancer predisposition and surveillance: an overview, and a tribute to Alfred G. Knudson JR. *Clin Cancer Res* 2017;23(11):e1–5.
121. Majzner RG, Heitzeneder S, Mackall CL. Harnessing the Immunotherapy Revolution for the Treatment of Childhood Cancers. *Cancer Cell* 2017;31:476–85.
122. Mackall CL, Merchant MS, Fry TJ. Immune-based therapies for childhood cancer. *Nat Rev Clin Oncol* 2014;11:693–703.
123. Bhatti P, Veiga LH, Ronckers CM, et al. Risk of second primary thyroid cancer after radiotherapy for a childhood cancer in a large cohort study: an update from the childhood cancer survivor study. *Radiat Res* 2010;174:741–52.
124. Lorigan P, Radford J, Howell A, et al. Lung cancer after treatment for Hodgkin's lymphoma: a systematic review. *Lancet Oncol* 2005;6:773–9.
125. van Leeuwen FE, Klokman WJ, Stovall M, et al. Roles of radiotherapy and smoking in lung cancer following Hodgkin's disease. *J Natl Cancer Inst* 1995;87:1530–7.
126. Friedman DL, Whitton J, Leisenring W, et al. Subsequent neoplasms in 5-year survivors of childhood cancer: the Childhood Cancer Survivor Study. *J Natl Cancer Inst* 2010;102:1083–95.
127. Limacher JM, Frebourg T, Natarajan-Ame S, et al. Two metachronous tumors in the radiotherapy fields of a patient with Li-Fraumeni syndrome. *Int J Cancer* 2001;96:238–42.
128. Birch JM, Alston RD, McNally RJ, et al. Relative frequency and morphology of cancers in carriers of germline TP53 mutations. *Oncogene* 2001;20:4621–8.
129. Talwalkar SS, Yin CC, Naeem RC, et al. Myelodysplastic syndromes arising in patients with germline TP53 mutation and Li-Fraumeni syndrome. *Arch Pathol Lab Med* 2010;134:1010–5.
130. Kleinerman RA, Tucker MA, Tarone RE, et al. Risk of new cancers after radiotherapy in long-term survivors of retinoblastoma: an extended follow-up. *J Clin Oncol* 2005;23:2272–9.
131. Draper GJ, Sanders BM, Kingston JE. Second primary neoplasms in patients with retinoblastoma. *Br J Cancer* 1986;53:661–71.
132. Sharif S, Ferner R, Birch JM, et al. Second primary tumors in neurofibromatosis 1 patients treated for optic glioma: substantial risks after radiotherapy. *J Clin Oncol* 2006;24:2570–5.
133. Bhatia S, Chen Y, Wong FL, et al. Subsequent Neoplasms After a Primary Tumor in Individuals With Neurofibromatosis Type 1. *J Clin Oncol* 2019;37:3050–8.
134. Swift M, Morrell D, Massey RB, et al. Incidence of cancer in 161 families affected by ataxia-telangiectasia. *N Engl J Med* 1991;325:1831–6.

135. Bernstein JL, Haile RW, Stovall M, et al. Radiation exposure, the ATM Gene, and contralateral breast cancer in the women's environmental cancer and radiation epidemiology study. *J Natl Cancer Inst* 2010;102:475–83.
136. Brooks JD, Teraoka SN, Reiner AS, et al. Variants in activators and downstream targets of ATM, radiation exposure, and contralateral breast cancer risk in the WECARE study. *Hum Mutat* 2012;33:158–64.
137. Travis LB, Demark Wahnefried W, Allan JM, et al. Aetiology, genetics and prevention of secondary neoplasms in adult cancer survivors. *Nat Rev Clin Oncol* 2013;10:289–301.
138. Olsen JH, Hahneemann JM, Borresen-Dale AL, et al. Breast and other cancers in 1445 blood relatives of 75 Nordic patients with ataxia telangiectasia. *Br J Cancer* 2005;93:260–5.
139. Allan JM. Genetic susceptibility to radiogenic cancer in humans. *Health Phys* 2008;95:677–86.
140. Kalow W, Ozdemir V, Tang BK, et al. The science of pharmacological variability: an essay. *Clin Pharmacol Ther* 1999;66:445–7.
141. Evans WE, McLeod HL. Pharmacogenomics—drug disposition, drug targets, and side effects. *N Engl J Med* 2003;348:538–49.
142. Wang X, Sun CL, Hageman L, et al. Clinical and genetic risk prediction of subsequent CNS tumors in survivors of childhood cancer: a report from the COG ALTE03N1 Study. *J Clin Oncol* 2017;35:3688–96.
143. Furzer J, Tessier L, Hodgson D, et al. Cost-utility of early breast cancer surveillance in survivors of thoracic radiation-treated adolescent Hodgkin lymphoma. *J Natl Cancer Inst* 2020;112(1):63–70.
144. Bhatia S, Palomares M, Hageman L, et al. Low-Dose Tamoxifen (LDTam) As a Breast Cancer (BC) Risk-Reduction Strategy in Lymphoma Survivors Exposed to Chest Radiation Therapy (RT) during adolescence/young adulthood – a randomized, placebo- controlled double blinded phase IIb trial. *Blood* 2019; 134(Issue supplement_1):2843.